

Review Article

Type 2 immunity in intestinal homeostasis and inflammatory bowel disease

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Type 2 immune responses commonly emerge during allergic reactions or infections with helminth parasites. Most of the cytokines associated with type 2 immune responses are IL-4, IL-5, and IL13, which are mainly produced by T helper 2 cells (T_H2), eosinophils, basophils, mast cells, and group 2 innate lymphoid cells (ILC2s). Over the course of evolution, humans have developed type 2 immune responses to fight infections and to protect tissues from the potential collateral damage caused by inflammation. For example, worm parasites induce potent type 2 immune responses, which are needed to simultaneously clear the pathogen and to promote tissue repair following injury. Due to the strong type 2 immune responses induced by helminths, which can promote tissue repair in the damaged epithelium, their use has been suggested as a possible treatment for inflammatory bowel disease (IBD); however, the role of type 2 immune responses in the initiation and progression of IBD is not fully understood. In this review, we discuss the molecular and cellular mechanisms that regulate type 2 immune responses during intestinal homeostasis, and we briefly discuss the scarce evidence linking type 2 immune responses with the aetiology of IBD.

Introduction

The breakdown of intestinal homeostasis may lead to aberrant immune responses against luminal antigens and eventually lead to inflammatory bowel diseases (IBD), which includes Crohn's disease (CD) and ulcerative colitis (UC). Although the aetiology of IBD is not well understood, it is broadly accepted that genetic and environmental factors are key [1,2]. The incidence of IBD in developed countries and regions with superior hygiene standards is higher than in regions with notable parasite exposure [3]. The removal of these co-evolved parasites may alter the establishment of tolerogenic and immunoregulatory responses [3]. Thus, parasite exposure may benefit the immune system by conferring protection against IBD. Parasite infection also known as helminths infection such as intestinal residing hookworm [4] promotes strong type 2 immune responses, wherein T helper 2 cells (T_H2) and innate lymphoid cells class 2 (ILC2s) are the major drivers of such responses [5,6]. Type 2 immune responses play an important role in epithelial barrier tissues. The epithelium, specifically tuft cells [7], control type 2 immune responses, highlighting a bi-directional epithelium-immune circuit orchestrated by type 2 cytokines. In this mini-review, we discuss and summarize the recent evidence linking type 2 immune responses and intestinal homeostasis, as well as how the failure of this regulating pathway may lead to IBD (Figure 1).

Immune cells in type 2 immune responses and IBD

IBD is characterized by activation of T cell-derived pro-inflammatory cytokines, such as IL-17 [8], TNF α [9], and IFN- γ [10,11], as well as malfunction of peripheral- and tissue-specific regulatory T cells [12,13]. T_H1 and T_H 17 immune responses, dominated by the production of IL-12, IL-23, and

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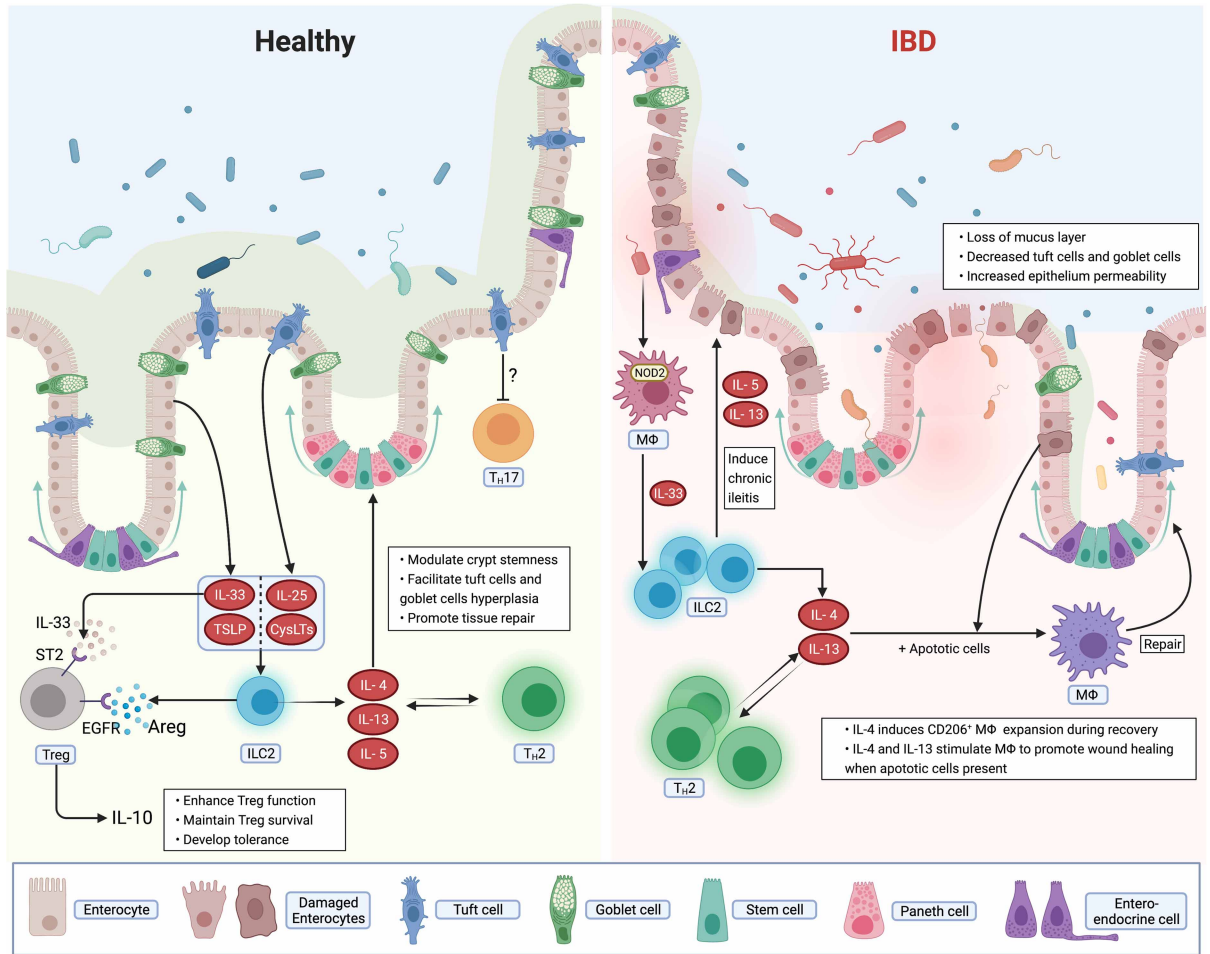


Figure 1. Type 2 immune responses in the intestinal mucosa from healthy and inflammatory bowel disease (IBD).

Type 2 immune responses mediated by T_H2 and ILC2 during homeostatic conditions, inflammation and tissue repair: (a) At steady-state conditions, type 2 cytokines produced by either ILC2 or T_H2 orchestrate epithelial homeostasis. Type 2 immune responses can promote crypt stemness and the epithelial cell differentiation towards the tuft and/or goblet cell lineage. This process is crucial to maintain a healthy mucus layer and eventually the integrity of the intestinal barrier. Epithelial cells derived alarmins (IL-25, CysLTs, IL-33, and TSLP) activate ILC2 promoting the establishment of tolerogenic immune responses. (b) In the inflamed intestine during IBD, ILC2, and T_H2 accumulate in inflamed lesions. Damaged epithelium permeability and loss of the mucus layer might result in commensals bacterial translocation and dissemination. Dissemination of bacteria result in NOD2-dependent activation which results in the production of IL-33. Eventually, ILC2s are activated by IL-33 and in turn produce IL-5 and IL-13, which fuel chronic ileitis. (c) During the process of tissue repair, IL-4 or IL-13 combined with apoptotic cells stimulate macrophages to promote wound healing and tissue remodeling.

IL-17 cytokines, have been well-studied in IBD [8]. However, the role of T_H2 immunity in IBD is not well understood. Here we discuss the potential role of T_H2 and its innate counterpart ILC2 in the aetiology of IBD.

T_H2 in IBD

T_H2 immune responses contribute to tissue repair and damage control, which can be considered as the counterpart to T_H1 immune responses [14]. The activation of transcription factors associated with T_H2 differentiation (e.g. GATA3, STAT6, and c-MAF), represses T_H1 or T_H17 differentiation, thus inhibiting the production of type 1 and type 17 effector cytokines (e.g. IFN γ , IL-1 β , TNF α , and IL-17), which are known to drive IBD pathogenicity [15–17].

Although direct links between T_{H2} immune response and IBD have not been shown, there is emerging research showing indirect links. T_{H2} cells are more abundant in the lamina propria of UC compared with CD patients [2]. Moreover, T_{H2} cells are enriched in CD patients who do not respond to anti-TNF therapy, suggesting that increased T_{H2} is associated with IBD severity [18]. Despite the accumulation of T_{H2} cells in inflamed tissues, it is not known whether T_{H2} cells are beneficial or detrimental. However, it is known that the type 2 cytokine IL-4 promotes macrophage-mediated wound healing and alleviates colitis [14,19]. A similar effect has been observed in studies with IL-13, which together with apoptotic cells, is required in macrophage-dependent pathogen clearance and tissue integrity restoration [20]. Phenome-wide association studies also found that carriers of the R130Q variant links high IL-13 activity with low susceptibility for developing CD [21]. Moreover, sustained CD4T cell-derived IL-13 activity protects mice from experimental colitis [21]. A recent case report suggested that blocking IL-4 activity using the IL-4Ralpha antagonist dupilumab may cause enteritis as a side effect when initially used to treat atopic dermatitis [22]. Thus, T_{H2} immune response may play a protective role in the pathogenesis of IBD.

On the other hand, T_{H2} cells may directly cause intestinal inflammation, as seen by experiments in which IL-4 expressing CD4⁺ T cells from mice with ileitis were sufficient to initiate ileitis upon being adoptively transferred in immunocompromised recipients [23]. In addition, despite the primarily role of T_{H1} immune responses in the establishment of ileitis in SAMP1/YitFc mice, the terminal ileal tissue manifested an elevated type 2 immunity signature (IL-5, IL-13, and GATA3/Tbet ratio) during chronic inflammation compared with the healthy control [23]. T_{H2} responses are also the driving force in an oxazolone-induced colitis murine model, which can be attenuated by suppressing type 2 cytokines [24–26]. Blocking GATA3 with a specific DNase reduces inflammation histologically in a 2,4,6-trinitrobenzene sulfonic acid (TNBS)-induced colitis murine model [27]. In human, UC patients show enhanced type 2 response in their mucosal samples compared with CD [18]. Interestingly, the source of IL-13 may be type 2 natural killer T cells rather than conventional T_{H2} cells [28,29]. Contrasting with the preclinical data, the IL-13 neutralizers tralokinumab and anrukinzumab did not achieve clinically significant responses in UC patients [30,31]. These contradictory findings may reflect the heterogeneous aetiology and immunological response in IBD patients. Future studies might address the source of type 2 cytokines and how their interplay with T_{H2} cells contributes to the disease development.

Type 2 innate lymphoid cells (ILC2) in IBD

Unlike T_{H2} , which responds to antigen-specific activation signals, ILC2 responds to a broader range of stimuli derived from immune cells, epithelial cells, or even neurons [32] and, functions in parasite invasion, allergic reactions, tissue repair, and intestinal homeostasis [33]. ILC2 serves as an initial response before the adaptive immune system resolves the challenge. When challenged, the tuft cell-derived IL-25 activates small intestine resident ILC2s. ILC2 attracts and activates T_{H2} cells by producing the type 2 cytokines IL-4, IL-5, and IL-13 [7,34]. As a result, effector cytokines such as IL-13 promote tissue remodeling by modulating crypt stemness and stimulating goblet cell (GC) and tuft cell hyperplasia in order to facilitate pathogen clearance [34–36].

Besides acting on epithelial cells, ILC2 impacts other immune cells. For example, ILC2 produces and secretes amphiregulin (AREG), which can trigger epidermal growth factor receptor (EGFR) signaling of regulatory T cells (Treg) to consequently enhance their immunosuppressive potential [37–39]. Moreover, similar to negative selection in the thymus, ILC expresses MHC II molecules, which deplete commensal bacteria-specific CD4⁺ T cells [40]. Although this mechanism has been fully demonstrated in ILC3s, studies have demonstrated that MHC II is also expressed on ILC2s [41,42]. Emerging research also highlights the dialog between ILC2 and the enteric nervous system (ENS), which is crucial for establishing homeostasis. Neuromedin U (NMU), a neuropeptide produced by neurons, can be sensed by NMU receptor 1 (NMUR1)-expressing ILC2s, which are restricted to the gut [43]. NMUR1 signaling is sufficient to activate and induce ILC2-derived type 2 immune responses [43]. This neuron-ILC2 axis seems to be relevant to oral tolerance as the immune-suppressive cytokine IL-10 is highly regulated by the NMU-NMUR1 axis [44]. In addition, choline acetyltransferase (ChAT⁺) enteric neurons targets ILC2 through the production of the α -calcitonin gene-related peptide, which inhibits ILC2-induced inflammation and tuft cell expansion [45]. Thus, the cross-talk between ENS and ILC2 may be critical for establishing intestinal immune homeostasis.

In patients diagnosed early with IBD, increased ILC2 (restricted to inflamed tissue), was observed in UC, but not in CD patients, when compared with non-inflamed or healthy specimens [46]. However, such observations were no longer seen in UC and CD patients with at least a 1-year medical history [46]. This observation is in agreement with a recent study showing ILC2 accumulation in CD patients with a dysregulated bacteria-sensing

processes [47]. Regardless whether ILC2 plays a protective or pathogenic role in IBD [48,49], the relationship between ILC2, the innate mucosal barrier, as well as immune and neural modulation, provides new therapeutic targets for IBD treatment.

Intestinal epithelial cells

The intestinal barrier is composed primarily of a single layer of epithelial cells, which produce biochemical molecules that reinforce the physical barrier. The intestinal barrier acts as the first line of defence against possible pathogens; its malfunction may result in pathogen translocation into the lamina propria, triggering a cascade of events leading to IBD [50]. Many intestinal barrier components, such as tuft cells and mucin-producing GCs, are regulated by type 2 immunity.

Tuft cell

Intestinal tuft cells are chemoreceptive cells characterized by the presence of brush-like microvilli projecting to the intestinal lumen. Although tuft cells were first discovered in the 1950s, its critical role in maintaining intestinal homeostasis has only been recently characterized. This knowledge gap is likely due to limited research tools to visualize and investigate tuft cells who are low in abundance both in experimental models and human (e.g. tuft cells only make up 0.4% of the cells in the mouse intestinal epithelium) [51,52]. Recent studies have shown that tuft cells sense luminal cues, such as microbe-derived metabolites [53] and, regulate the invasion of pathogens, such as protists [54], helminth [55], and/or virus [56]. In response to pathogen invasion, tuft cells significantly expand and secrete large amounts of IL-25 and cysteinyl leukotrienes (cysLTs), which subsequently activate ILC2 [54]. In turn, tuft cell expansion required IL-13 and IL-5 signals given the evidence that deletion of these two genes impaired tuft cell hyperplasia in response to *Nippostrongylus brasiliensis* [34]. In the TNF α overexpressing mouse model of colitis, administration of succinate resulted in tuft cell hyperplasia and decreased ileum inflammation [57]. In addition to tuft cell hyperplasia, treated mice demonstrated lower type 17 cytokines and retinoic acid receptor-related orphan receptor gamma-t (ROR γ t+) cells compared with control mice [57]. Banerjee and colleagues reported that CD patients possess tuft cell deficiency in ileal lesions [57]. Tuft cell-associated IBD phenotypes is not restricted to the ileum; a pediatric study reported that patients with acute duodenitis, ulcer, or celiac disease showed significantly lower tuft cells in the duodenum compared with controls.

Inflammation negatively impacts tuft cells. A previous study showed that inflammatory severity inversely correlates with tuft cell count [58]. In mouse, doublecortin like kinase 1 (DCLK1) is a broadly used marker for tuft cell analysis. Conditional knock out mice lacking DCLK1 in intra epithelial cells develops spontaneous colitis from age around 3–4 weeks and this was associated with decreased COX-2 expression and therefore the reduced prostaglandin E2 (PGE2) production, which is a key regulator in colonic epithelial cells proliferation [59]. *Dclk1*^{-/-} mice also showed delayed tissue regeneration following intestinal damage [60]. In agreement with a potential role in tissue regeneration, DCLK1 deficiency reduces the development of intestinal adenomas and limits pro-survival signaling and self-renewal in *Apc*^{Min/+} mice [61,62]. The mechanism by which tuft cells orchestrates tissue remodeling through cross-talk with immune cells requires future investigation. Moreover, whether the reduction in tuft cells in IBD lesions is a protective response or the consequence of IBD remains to be fully understood.

Goblet cells

GCs are epithelial cells with a narrow basal end and wide apical surface. GCs in the small intestine and colon share the same functional property, which is to secrete the mucus-producing mucin. As the main function of the mucus layer is to lubricate the intestinal lumen and to protect it from pathogen invasion, GCs are critical for maintaining the barrier integrity. In agreement, mucin-2 (*Muc2*)-deficient mice show spontaneous colitis [63]. GC hyperplasia is often seen in allergies or helminth infections, wherein the T_H2 immune response is enhanced. ILC2 has been proposed to be a major modulator of GC homeostasis and function [7,34,55]. In experimental colitis, IL-33 induced ILC2 activation, resulting in GC hyperplasia that contributes to epithelial structural restoration [64]. In addition, ILC2-derived IL-13 induces GC hyperplasia in an *in vitro* organoid culture system [65]. However, whether T_H2 and/or ILC2 have direct effects on GC homeostasis and function in IBD patients needs to be determined.

Aberrant GC function has been characterized in UC patients. A proteomic study in active UC patients demonstrated that MUC2 was significantly decreased in both colonic lesions and uninfamed sites [66]. This

phenotype is in line with a systematic review showing that 5 out of 8 human studies during the past 25 years reported decreased MUC2 protein levels in UC patients compared with healthy individuals [67]. The pathogenicity associated with decreased MUC2 was linked to increased bacterial penetration in the inner layer of mucus, which may lead to intestinal inflammation and dysbiosis [68]. Single-cell transcriptional analysis identified inter-crypt goblet cells (icGCs), a functionally distinct GC subpopulation from the canonical crypt-resident GCs [69]. Mucus secreted by icGCs seals the gap between opening crypts, thus preventing microbial penetration into the stem cells niche [69]. Moreover, UC patients demonstrated reduced icGCs and mucus defects [69]. In line with its protective role, icGC-deficient mice develop age-dependent colitis [69]. Whether icGC homeostasis and function is regulated by type 2 immunity, similar to conventional GCs, requires further investigation.

Epithelium-derived type 2 immunity associated cytokines

Cytokines, which modulate intestinal barrier function, are crucial for the development of type 2 immune responses. In the following section, we will discuss classical epithelium-derived cytokines (called intestinal alarmins) that modulate type 2 immunity.

IL-25

IL-25 (also named IL-17E) belongs to the IL-17 family and triggers the heterodimer complex IL-17RA and IL-17RB (IL-25R). IL-25, which is primarily produced and secreted by tuft cells, promotes type 2 responses [70]. The proliferation of IL-25 producing tuft cells is stimulated by IL-4, IL-5, and IL-13 in a positive feedback loop [34,70].

The *IL-25* gene is located within a region reported to confer susceptibility to IBD; however, a study in a small cohort of patients did not show the association between polymorphisms within the *IL-25* and IBD [71]. Studies in larger cohorts of IBD patients are needed to investigate the involvement of polymorphisms in the *IL-25* gene in IBD aetiology.

IL-25 can act on IL25R-expressing ILC2s, CD4⁺ T cells and mesenchymal stem cells (MSC) [72]. IL-25R⁺ CD4⁺ T cells and IL-25R⁺ Lgr5⁺ MSCs are enriched in the inflamed colonic mucosa of CD and UC patients compared with healthy individuals or non-inflamed tissue from IBD patients [72]. However, reductions in circulating IL-25R⁺ CD4⁺ was negatively correlated with inflammation-associated severity [72]. *In vitro* studies showed that IL-25 may prime Lgr5⁺ cells to secrete pro-survival factors involved in the PI3K-Akt pathway to maintain epithelial cells homeostasis [72]. IL-25 was also found to ameliorate experimental TNBS- and dextran sodium sulfate (DSS)-induced colitis by inhibiting T_H17 cells [72–75]. In line with these observations, Caruso and colleagues showed lower IL-25 transcriptional and protein levels in IBD colonic biopsies compared with healthy individuals [76]. Interestingly, IL-25R is expressed by CD14⁺ cells, a critical source of IL-12 in CD mucosa [76]. *In vitro* studies demonstrated that IL-25 signaling in CD14⁺ cells suppresses IBD inflammatory cytokines, such as IL-12 and IL-23 [76]. Furthermore, IL-25 treatment could reduce partially the TNBS-colitis histologic scoring, suggesting that IL-25 may limit inflammation development by reducing IL-12 and IFN γ protein expression and cellular infiltration to the mucosa [76]. IL-25 also shows tumor suppressive properties; blocking IL-25 signaling in a colitis-induced tumor model increases tumor numbers when compared with control mice [77].

Despite the research described above, some studies suggest that IL-25 may aggravate mucosal inflammation. Reynolds et al. [78] demonstrated that IL-25-deficient mice were protected from DSS-induced colitis; however, IL-25-deficient mice showed poorer outcomes compared with wild type control when infection was induced by *Citrobacter rodentium*. Thus, the role of IL-25 in either ameliorating or exacerbating intestinal inflammation may be context dependent.

IL-33

The type 2 cytokine IL-33 belongs to the IL-1 family. In response to intestinal damage or stress, IL-33 is released by necrotic epithelial cells, stromal cells and/or endothelial cells in both mouse and humans [79]. IL-33, a potent driver of T_H2 cells and ILC2 differentiation and function, plays an important role in allergic inflammation, which depends on IL-33 binding to its receptor ST2 (IL1RL1) and IL-1 receptor accessory protein (IL1RAP) [80].

The dual-function of IL-33 has led to several debatable insights. Recently, IL-33 has been reported to delay *C. rodentium* clearance by limiting T_H17 responses and increasing intestinal permeability [81]. The pathogenic

role of IL-33 was also observed in the SAMPl/YitFc (SAMP) mouse model of human CD ileitis [47]. In this study, the IL-33/ST2 pathway promoted NOD2-induced ILC2 expansion in ileitis; furthermore, blocking IL-33 protected SAMP mice [47]. Similar observations were seen in IL-33 deficient mice with DSS-induced colitis [48]. In inflamed colonic biopsies from UC patients, IL-33 was increased compared with inflamed or non-inflamed tissue samples from CD patients, indicating a pathogenic role in UC [82,83]. In contrast with the pathogenic role of IL-33, the protective role of IL-33 may involve the interaction with regulatory T cells. ST2 is one of the top differentially expressed genes in colonic Treg cells (cTreg) compared with Treg cells in mesenteric lymph node (MLN) [84]. Interestingly, cTreg differentiation and function is modulated by IL-33-ST2 signaling [84]. During inflammation, *Rag1*^{-/-} mice receiving the combination of CD45.1⁺ naïve T cells (RB^{hi}) and ST2^{-/-} Treg have been reported to show impaired cTreg compared with RB^{hi} + wild type Treg, thus leading to more severe colitis [84]. Furthermore, IL-33 treatment reduced TNBS-colitis severity [85]. IL-33 can also favor tissue remodeling and induce tuft and GC function [64,65]. The failure to regulate the IL-33-tuft cell axis may be associated with *SPRY2* (Sprouty2, an intracellular signaling regulator). *SPRY2* expression has been shown to be enhanced in both CD and UC patients, resulting in tuft cells and GC inhibition [35]. Collectively, different studies showing both pathogenic and protective roles of IL-33 suggest that IL-33 may be a double-edged sword.

Thymic stromal lymphopoietin (TSLP)

TSLP, which is mainly secreted by epithelial cells and stroma cells [86], is regarded as a potent ILC2 activator. Its role has been extensively studied in dermic allergic and asthma [87]. The role of TSLP in UC patients remains controversial. Tanaka et al. found that TSLP was increased in inflamed lesions compared with non-inflamed tissues from UC patients. Furthermore, they showed that IL-4 and TNF- α may exacerbate TSLP expression, leading to enhanced inflammation [88]. However, others showed that TSLP was reduced in UC patients compared with healthy or non-inflamed tissues [89,90]. Patients receiving anti-IL-13 (tralokinumab) also showed more TSLP transcriptional expression with improved tissue healing compared before treatment [89]. TSLP may trigger ILC2 to produce AREG, which in turn promotes epithelium repair via the AREG/EGFR signaling pathway. In line with this observation, TSLP receptor-deficient mice showed poorer outcomes compared with their wild type control in DSS model [91]. Furthermore, exogenous delivery of TSLP using an engineered lactic acid bacteria ameliorated DSS-induced inflammation by inducing TGF- β , thus reinforcing its immunosuppressive function [92].

Perspectives

- Type 2 immune responses are critical to control pathogen infections and to promote tissue repair whereas breakdown on these mechanisms may lead to IBD (Figure 1)
- The current literature linking type 2 immune responses and IBD is rather inconclusive, as pathogenic and protective roles of type 2 immune responses in IBD has been reported.
- More intensive research on the specific function of type 2 cytokines in specific phases of the disease are urgently needed to better interpret the role of type 2 immune responses in IBD.

Competing Interests

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Author Contribution

X.L. and E.J.V. wrote, revised and edited the manuscript. The figure was prepared by X.L. and revised by E.J.V.

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Abbreviations

AREG, amphiregulin; CD, Crohn's disease; DCLK1, doublecortin like kinase 1; DSS, dextran sodium sulfate; EGFR, epidermal growth factor receptor; ENS, enteric nervous system; GCs, goblet cells; IBD, inflammatory bowel diseases; iGCs, inter-crypt goblet cells; MSC, mesenchymal stem cells; NMU, neuromedin U; NMUR1, NMU receptor 1; ROR γ +, receptor-related orphan receptor gamma-t; T_H2, T helper 2 cells; TNBS, 2,4,6-trinitrobenzene sulfonic acid; Treg, regulatory T cells; TSLP, thymic stromal lymphopoietin; UC, ulcerative colitis.

Reference

- 1 Ananthakrishnan, A.N., Bernstein, C.N., Iliopoulos, D., Macpherson, A., Neurath, M.F., Ali, R.A.R. et al. (2018) Environmental triggers in IBD: a review of progress and evidence. *Nat. Rev. Gastroenterol. Hepatol.* **15**, 39–49 <https://doi.org/10.1038/nrgastro.2017.136>
- 2 Imam, T., Park, S., Kaplan, M.H. and Olson, M.R. (2018) Effector T helper cell subsets in inflammatory bowel diseases. *Front. Immunol.* **9**, 1212 <https://doi.org/10.3389/fimmu.2018.01212>
- 3 Mohammadi, R., Hosseini-Safa, A., Ehsani Ardakani, M.J. and Rostami-Nejad, M. (2015) The relationship between intestinal parasites and some immune-mediated intestinal conditions. *Gastroenterol. Hepatol. Bed. Bench.* **8**, 123–131 PMID: 25926937
- 4 Salgame, P., Yap, G.S. and Gause, W.C. (2013) Effect of helminth-induced immunity on infections with microbial pathogens. *Nat. Immunol.* **14**, 1118–1126 <https://doi.org/10.1038/ni.2736>
- 5 Anthony, R.M., Rutitzky, L.I., Urban, J.F., Stadecker, M.J. and Gause, W.C. (2007) Protective immune mechanisms in helminth infection. *Nat. Rev. Immunol.* **7**, 975–987 <https://doi.org/10.1038/nri2199>
- 6 Zhu, J. (2015) T helper 2 (Th2) cell differentiation, type 2 innate lymphoid cell (ILC2) development and regulation of interleukin-4 (IL-4) and IL-13 production. *Cytokine* **75**, 14–24 <https://doi.org/10.1016/j.cyto.2015.05.010>
- 7 Howitt, M.R., Lavoie, S., Michaud, M., Blum, A.M., Tran, S.V., Weinstock, J.V. et al. (2016) Tuft cells, taste-chemosensory cells, orchestrate parasite type 2 immunity in the gut. *Science* **351**, 1329–1333 <https://doi.org/10.1126/science.aaf1648>
- 8 Moschen, A.R., Tilg, H. and Raine, T. (2019) IL-12, IL-23 and IL-17 in IBD: immunobiology and therapeutic targeting. *Nat. Rev. Gastroenterol. Hepatol.* **16**, 185–196 <https://doi.org/10.1038/s41575-018-0084-8>
- 9 Billmeier, U., Dieterich, W., Neurath, M.F. and Atraya, R. (2016) Molecular mechanism of action of anti-tumor necrosis factor antibodies in inflammatory bowel diseases. *World J. Gastroenterol.* **22**, 9300–9313 <https://doi.org/10.3748/wjg.v22.i42.9300>
- 10 Langer, V., Vivi, E., Regensburger, D., Winkler, T.H., Waldner, M.J., Rath, T. et al. (2019) IFN- γ drives inflammatory bowel disease pathogenesis through VE-cadherin-directed vascular barrier disruption. *J. Clin. Invest.* **129**, 4691–4707 <https://doi.org/10.1172/JCI124884>
- 11 Neurath, M.F. (2014) Cytokines in inflammatory bowel disease. *Nat. Rev. Immunol.* **14**, 329–342 <https://doi.org/10.1038/nri3661>
- 12 Tindemans, I., Joosse, M.E. and Samsom, J.N. (2020) Dissecting the heterogeneity in T-cell mediated inflammation in IBD. *Cells* **9**, 110 <https://doi.org/10.3390/cells9010110>
- 13 Veltkamp, C., Anstaett, M., Wahl, K., Möller, S., Gangl, S., Bachmann, O. et al. (2011) Apoptosis of regulatory T lymphocytes is increased in chronic inflammatory bowel disease and reversed by anti-TNF α treatment. *Gut* **60**, 1345–1353 <https://doi.org/10.1136/gut.2010.217117>
- 14 Gause, W.C., Wynn, T.A. and Allen, J.E. (2013) Type 2 immunity and wound healing: evolutionary refinement of adaptive immunity by helminths. *Nat. Rev. Immunol.* **13**, 607–614 <https://doi.org/10.1038/nri3476>
- 15 Biedermann, T., Röcken, M. and Carballido, J.M. (2004) TH1 and TH2 lymphocyte development and regulation of TH cell-mediated immune responses of the skin. *J. Investig. Dermatol. Symp. Proc.* **9**, 5–14 <https://doi.org/10.1111/j.1087-0024.2004.00829.x>
- 16 Kaiko, G.E., Horvat, J.C., Beagley, K.W. and Hansbro, P.M. (2008) Immunological decision-making: how does the immune system decide to mount a helper T-cell response? *Immunology* **123**, 326–338 <https://doi.org/10.1111/j.1365-2567.2007.02719.x>
- 17 Selin, K.A., Hedin, C.R.H. and Villablanca, E.J. (2021) Immunological networks defining the heterogeneity of inflammatory bowel diseases. *J. Crohns Colitis* **15**, 1093 <https://doi.org/10.1093/ecco-jcc/jjab085>
- 18 Li, J., Ueno, A., Fort Gasia, M., Luider, J., Wang, T., Hirota, C. et al. (2016) Profiles of lamina propria T helper cell subsets discriminate between ulcerative colitis and Crohn's disease. *Inflamm. Bowel Dis.* **22**, 1779–1792 <https://doi.org/10.1097/MIB.0000000000000811>
- 19 Jayme, T.S., Leung, G., Wang, A., Workentine, M.L., Rajeev, S., Shute, A. et al. (2020) Human interleukin-4-treated regulatory macrophages promote epithelial wound healing and reduce colitis in a mouse model. *Sci. Adv.* **6**, eaba4376 <https://doi.org/10.1126/sciadv.aba4376>
- 20 Bosurgi, L., Cao, Y.G., Cabeza-Cabrero, M., Tucci, A., Hughes, L.D., Kong, Y. et al. (2017) Macrophage function in tissue repair and remodeling requires IL-4 or IL-13 with apoptotic cells. *Science* **356**, 1072–1076 <https://doi.org/10.1126/science.aai8132>
- 21 Karmele, E.P., Pasricha, T.S., Ramalingam, T.R., Thompson, R.W., Gieseck, R.L., Knilians, K.J. et al. (2019) Anti-IL-13R α 2 therapy promotes recovery in a murine model of inflammatory bowel disease. *Mucosal Immunol.* **12**, 1174–1186 <https://doi.org/10.1038/s41385-019-0189-6>

- 22 Shimodaira, Y., Takahashi, S. and Iijima, K. (2021) Anti-IL-4R α monoclonal antibody dupilumab mimics ulcerative colitis: a case report. *BMC Gastroenterol.* **21**, 207 <https://doi.org/10.1186/s12876-021-01803-8>
- 23 Bamias, G., Martin, C., Mishina, M., Ross, W.G., Rivera-Nieves, J., Marini, M. et al. (2005) Proinflammatory effects of Th2 cytokines in a murine model of chronic small intestinal inflammation. *Gastroenterology* **128**, 654–666 <https://doi.org/10.1053/j.gastro.2004.11.053>
- 24 Boirivant, M., Fuss, I.J., Chu, A. and Strober, W. (1998) Oxazolone colitis: a murine model of T helper cell type 2 colitis treatable with antibodies to interleukin 4. *J. Exp. Med.* **188**, 1929–1939 <https://doi.org/10.1084/jem.188.10.1929>
- 25 Mf N, B.W. (2016) Oxazolone-induced colitis as a model of Th2 immune responses in the intestinal mucosa. *Methods Mol. Biol.* **1422**, 253–261 https://doi.org/10.1007/978-1-4939-3603-8_23
- 26 Itani, S., Watanabe, T., Nadatani, Y., Sugimura, N., Shimada, S., Takeda, S. et al. (2016) NLRP3 inflammasome has a protective effect against oxazolone-induced colitis: a possible role in ulcerative colitis. *Sci. Rep.* **6**, 39075 <https://doi.org/10.1038/srep39075>
- 27 Popp, V., Gerlach, K., Mott, S., Turowska, A., Garn, H., Atreya, R. et al. (2017) Rectal delivery of a DNzyme that specifically blocks the transcription factor GATA3 and reduces colitis in mice. *Gastroenterology* **152**, 176–192.e5 <https://doi.org/10.1053/j.gastro.2016.09.005>
- 28 Fuss, I.J., Heller, F., Boirivant, M., Leon, F., Yoshida, M., Fichtner-Feigl, S. et al. (2004) Nonclassical CD1d-restricted NK T cells that produce IL-13 characterize an atypical Th2 response in ulcerative colitis. *J. Clin. Invest.* **113**, 1490–1497 <https://doi.org/10.1172/JCI19836>
- 29 Sagami, S., Ueno, Y., Tanaka, S., Fujita, A., Niitsu, H., Hayashi, R. et al. (2017) Choline deficiency causes colonic type II natural killer T (NKT) cell loss and alleviates murine colitis under type I NKT cell deficiency. *PLoS One* **12**, e0169681 <https://doi.org/10.1371/journal.pone.0169681>
- 30 Danese, S., Rudziński, J., Brandt, W., Dupas, J.-L., Peyrin-Biroulet, L., Bouhnik, Y. et al. (2015) Tralokinumab for moderate-to-severe UC: a randomised, double-blind, placebo-controlled, phase IIa study. *Gut* **64**, 243–249 <https://doi.org/10.1136/gutjnl-2014-308004>
- 31 Reinisch, W., Panés, J., Khurana, S., Toth, G., Hua, F., Comer, G.M. et al. (2015) Anrukizumab, an anti-interleukin 13 monoclonal antibody, in active UC: efficacy and safety from a phase IIa randomised multicentre study. *Gut* **64**, 894–900 <https://doi.org/10.1136/gutjnl-2014-308337>
- 32 McGinty, J.W. and von Moltke, J. (2020) A three course menu for ILC and bystander T cell activation. *Curr. Opin. Immunol.* **62**, 15–21 <https://doi.org/10.1016/j.coi.2019.11.005>
- 33 Schulz-Kuhnt, A., Neurath, M.F., Wirtz, S. and Atreya, I. (2021) Innate lymphoid cells as regulators of epithelial integrity: therapeutic implications for inflammatory bowel diseases. *Front. Med.* **8**, 656745 <https://doi.org/10.3389/fmed.2021.656745>
- 34 von Moltke, J., Ji, M., Liang, H.-E. and Locksley, R.M. (2016) Tuft-cell-derived IL-25 regulates an intestinal ILC2–epithelial response circuit. *Nature* **529**, 221–225 <https://doi.org/10.1038/nature16161>
- 35 Schumacher, M.A., Hsieh, J.J., Liu, C.Y., Appel, K.L., Waddell, A., Almohazey, D. et al. (2021) Sprouty2 limits intestinal tuft and goblet cell numbers through GSK3 β -mediated restriction of epithelial IL-33. *Nat. Commun.* **12**, 836 <https://doi.org/10.1038/s41467-021-21113-7>
- 36 Henriksson, J. T., Coursey, T.G., Corry, D.B., De Paiva, C.S. and Pflugfelder, S.C. (2015) IL-13 stimulates proliferation and expression of mucin and immunomodulatory genes in cultured conjunctival goblet cells. *Invest. Ophthalmol. Vis. Sci.* **56**, 4186–4197 <https://doi.org/10.1167/iovs.14-15496>
- 37 Zaiss, D.M.W., van Loosdregt, J., Gorlani, A., Bekker, C.P.J., Gröne, A., Sibilia, M. et al. (2013) Amphiregulin enhances regulatory T cell suppressive function via the epidermal growth factor receptor. *Immunity* **38**, 275–284 <https://doi.org/10.1016/j.immuni.2012.09.023>
- 38 Zaiss, D.M.W., Gause, W.C., Osborne, L.C. and Artis, D. (2015) Emerging functions of amphiregulin in orchestrating immunity, inflammation and tissue repair. *Immunity* **42**, 216–226 <https://doi.org/10.1016/j.immuni.2015.01.020>
- 39 Wang, S., Zhang, Y., Wang, Y., Ye, P., Li, J., Li, H. et al. (2016) Amphiregulin confers regulatory T cell suppressive function and tumor invasion via the EGFR/GSK-3 β /Foxp3 axis. *J. Biol. Chem.* **291**, 21085–21095 <https://doi.org/10.1074/jbc.M116.717892>
- 40 Hepworth, M.R., Monticelli, L.A., Fung, T.C., Ziegler, C.G.K., Grunberg, S., Sinha, R. et al. (2013) Innate lymphoid cells regulate CD4+ T cell responses to intestinal commensal bacteria. *Nature* **498**, 113–117 <https://doi.org/10.1038/nature12240>
- 41 Oliphant, C.J., Hwang, Y.Y., Walker, J.A., Salimi, M., Wong, S.H., Brewer, J.M. et al. (2014) MHCII-mediated dialog between group 2 innate lymphoid cells and CD4(+) T cells potentiates type 2 immunity and promotes parasitic helminth expulsion. *Immunity* **41**, 283–295 <https://doi.org/10.1016/j.immuni.2014.06.016>
- 42 Jiang, M., Liu, H., Li, Z., Wang, J., Zhang, F., Cao, K. et al. (2019) ILC2s induce adaptive Th2-type immunity in acute exacerbation of chronic obstructive pulmonary disease. *Mediators Inflamm.* **2019**, 3140183 <https://doi.org/10.1155/2019/3140183>
- 43 Klose, C.S.N., Mahlaköiv, T., Moeller, J.B., Rankin, L.C., Flamar, A.-L., Kabata, H. et al. (2017) The neuropeptide neuromedin U stimulates innate lymphoid cells and type 2 inflammation. *Nature* **549**, 282–286 <https://doi.org/10.1038/nature23676>
- 44 Bando, J.K., Gilfillan, S., Di Luccia, B., Fachi, J.L., Sécca, C., Cella, M. et al. (2019) ILC2s are the predominant source of intestinal ILC-derived IL-10. *J. Exp. Med.* **217**, e20191520 <https://doi.org/10.1084/jem.20191520>
- 45 Xu, H., Ding, J., Porter, C.B.M., Wallrapp, A., Tabaka, M., Ma, S. et al. (2019) Transcriptional atlas of intestinal immune cells reveals that neuropeptide α -CGRP modulates group 2 innate lymphoid cell responses. *Immunity* **51**, 696–708.e9 <https://doi.org/10.1016/j.immuni.2019.09.004>
- 46 Forkel, M., van Tol, S., Höög, C., Michaëlsson, J., Almer, S. and Mjösberg, J. (2019) Distinct alterations in the composition of mucosal innate lymphoid cells in newly diagnosed and established Crohn's disease and ulcerative colitis. *J. Crohns Colitis* **13**, 67–78 <https://doi.org/10.1093/ecco-jcc/jiy119>
- 47 De Salvo, C., Buela, K.-A., Creyns, B., Corridoni, D., Rana, N., Wargo, H.L. et al. (2021) NOD2 drives early IL-33-dependent expansion of group 2 innate lymphoid cells during Crohn's disease-like ileitis. *J. Clin. Invest.* **131**, e140624 <https://doi.org/10.1172/JCI140624>
- 48 Qiu, X., Qi, C., Li, X., Fang, D. and Fang, M. (2020) IL-33 deficiency protects mice from DSS-induced experimental colitis by suppressing ILC2 and Th17 cell responses. *Inflamm. Res.* **69**, 1111–1122 <https://doi.org/10.1007/s00011-020-01384-4>
- 49 You, Y., Zhang, X., Wang, X., Yue, D., Meng, F., Zhu, J. et al. (2020) ILC2 proliferation by IL-33 stimulation alleviates acute colitis in Rag1 $^{-/-}$ mouse through promoting M2 macrophage polarization. *J. Immunol. Res.* **2020**, 5018975 <https://doi.org/10.1155/2021/9843489>
- 50 Vancamelbeke, M. and Vermeire, S. (2017) The intestinal barrier: a fundamental role in health and disease. *Expert. Rev. Gastroenterol. Hepatol.* **11**, 821–834 <https://doi.org/10.1080/17474124.2017.1343143>
- 51 Gerbe, F., Legraverend, C. and Jay, P. (2012) The intestinal epithelium tuft cells: specification and function. *Cell. Mol. Life Sci.* **69**, 2907–2917 <https://doi.org/10.1007/s00018-012-0984-7>
- 52 Schütz, B., Ruppert, A.-L., Strobel, O., Lazarus, M., Urade, Y., Büchler, M.W., et al. (2019) Distribution pattern and molecular signature of cholinergic tuft cells in human gastro-intestinal and pancreatic-biliary tract. *Sci. Rep.* **9**, 17466 <https://doi.org/10.1038/s41598-019-53997-3>

- 53 Schneider, C., O'Leary, C.E., von Moltke, J., Liang, H.-E., Ang, Q.Y., Turnbaugh, P.J., et al. (2018) A metabolite-triggered tuft cell-ILC2 circuit drives small intestinal remodeling. *Cell* **174**, 271–284.e14 <https://doi.org/10.1016/j.cell.2018.05.014>
- 54 McGinty, J.W., Ting, H.-A., Billipp, T.E., Nadjisombati, M.S., Khan, D.M., Barrett, N.A. et al. (2020) Tuft cell-derived leukotrienes drive rapid anti-helminth immunity in the small intestine but are dispensable for anti-protist immunity. *Immunity* **52**, 528–541.e7 <https://doi.org/10.1016/j.immuni.2020.02.005>
- 55 Gerbe, F., Sidot, E., Smyth, D.J., Ohmoto, M., Matsumoto, I., Dardalhon, V. et al. (2016) Intestinal epithelial tuft cells initiate type 2 mucosal immunity to helminth parasites. *Nature* **529**, 226–230 <https://doi.org/10.1038/nature16527>
- 56 Wilen, C.B., Lee, S., Hsieh, L.L., Orchard, R.C., Desai, C., Hykes, B.L. et al. (2018) Tropism for tuft cells determines immune promotion of norovirus pathogenesis. *Science* **360**, 204–208 <https://doi.org/10.1126/science.aar3799>
- 57 Banerjee, A., Herring, C.A., Chen, B., Kim, H., Simmons, A.J., Southard-Smith, A.N. et al. (2020) Succinate produced by intestinal microbes promotes specification of tuft cells to suppress ileal inflammation. *Gastroenterology* **159**, 2101–2115.e5 <https://doi.org/10.1053/j.gastro.2020.08.029>
- 58 Huh, W.J., Roland, J.T., Asai, M. and Kaji, I. (2020) Distribution of duodenal tuft cells is altered in pediatric patients with acute and chronic enteropathy. *Biomed. Res. Tokyo Jpn.* **41**, 113–118 <https://doi.org/10.2220/biomedres.41.113>
- 59 Yi, J., Bergstrom, K., Fu, J., Shan, X., McDaniel, J.M., McGee, S. et al. (2019) Dclk1 in tuft cells promotes inflammation-driven epithelial restitution and mitigates chronic colitis. *Cell Death Differ.* **26**, 1656–1669 <https://doi.org/10.1038/s41418-018-0237-x>
- 60 Qu, D., Weygant, N., May, R., Chandrakesan, P., Madhoun, M., Ali, N. et al. (2015) Ablation of doublecortin-like kinase 1 in the colonic epithelium exacerbates dextran sulfate sodium-induced colitis. *PLoS One* **10**, e0134212 <https://doi.org/10.1371/journal.pone.0134212>
- 61 Westphalen, C.B., Asfaha, S., Hayakawa, Y., Takemoto, Y., Lukin, D.J., Nuber, A.H. et al. (2014) Long-lived intestinal tuft cells serve as colon cancer-initiating cells. *J. Clin. Invest.* **124**, 1283–1295 <https://doi.org/10.1172/JCI73434>
- 62 Chandrakesan, P., Yao, J., Qu, D., May, R., Weygant, N., Ge, Y. et al. (2017) Dclk1, a tumor stem cell marker, regulates pro-survival signaling and self-renewal of intestinal tumor cells. *Mol. Cancer* **16**, 30 <https://doi.org/10.1186/s12943-017-0594-y>
- 63 Van der Sluis, M., De Koning, B.A.E., De Bruijn, A.C.J.M., Velcich, A., Meijerink, J.P.P., Van Goudoever, J.B. et al. (2006) Muc2-deficient mice spontaneously develop colitis, indicating that MUC2 is critical for colonic protection. *Gastroenterology* **131**, 117–129 <https://doi.org/10.1053/j.gastro.2006.04.020>
- 64 Monticelli, L.A., Osborne, L.C., Noti, M., Tran, S.V., Zaiss, D.M.W. and Artis, D. (2015) IL-33 promotes an innate immune pathway of intestinal tissue protection dependent on amphiregulin–EGFR interactions. *Proc. Natl Acad. Sci. U.S.A.* **112**, 10762–10767 <https://doi.org/10.1073/pnas.1509070112>
- 65 Waddell, A., Vallance, J.E., Hummel, A., Alenghat, T. and Rosen, M.J. (2019) IL-33 Induces murine intestinal goblet cell differentiation indirectly via innate lymphoid cell IL-13 secretion. *J. Immunol.* **202**, 598–607 <https://doi.org/10.4049/jimmunol.1800292>
- 66 van der Post, S., Jabbar, K.S., Birchenough, G., Arike, L., Akhtar, N., Sjoval, H. et al. (2019) Structural weakening of the colonic mucus barrier is an early event in ulcerative colitis pathogenesis. *Gut* **68**, 2142–2151 <https://doi.org/10.1136/gutjnl-2018-317571>
- 67 Bankole, E., Read, E., Curtis, M.A., Neves, J.F. and Garnett, J.A. (2021) The relationship between mucins and ulcerative colitis: a systematic review. *J. Clin. Med.* **10**, 1935 <https://doi.org/10.3390/jcm10091935>
- 68 Johansson, M.E.V., Gustafsson, J.K., Holmén-Larsson, J., Jabbar, K.S., Xia, L., Xu, H. et al. (2014) Bacteria penetrate the normally impenetrable inner colon mucus layer in both murine colitis models and patients with ulcerative colitis. *Gut* **63**, 281–291 <https://doi.org/10.1136/gutjnl-2012-303207>
- 69 Nyström, E.E.L., Martínez-Abad, B., Arike, L., Birchenough, G.M.H., Nonnecke, E.B., Castillo, P.A. et al. (2021) An intercrypt subpopulation of goblet cells is essential for colonic mucus barrier function. *Science* **372**, eabb1590 <https://doi.org/10.1126/science.abb1590>
- 70 Fort, M.M., Cheung, J., Yen, D., Li, J., Zurawski, S.M., Lo, S. et al. (2001) IL-25 induces IL-4, IL-5, and IL-13 and Th2-associated pathologies in vivo. *Immunity* **15**, 985–995 [https://doi.org/10.1016/S1074-7613\(01\)00243-6](https://doi.org/10.1016/S1074-7613(01)00243-6)
- 71 Büning, C., Genschel, J., Weltrich, R., Lochs, H. and Schmidt, H. (2003) The interleukin-25 gene located in the inflammatory bowel disease (IBD) 4 region: no association with inflammatory bowel disease. *Eur. J. Immunogenet.* **30**, 329–333 <https://doi.org/10.1046/j.1365-2370.2003.00411.x>
- 72 Su, J., Xie, C., Fan, Y., Cheng, W., Hu, Y., Huang, Q. et al. (2017) Interleukin-25 enhances the capacity of mesenchymal stem cells to induce intestinal epithelial cell regeneration. *Am. J. Transl. Res.* **9**, 5320–5331 PMID: 29312486
- 73 Cheng, W., Su, J., Hu, Y., Huang, Q., Shi, H., Wang, L. et al. (2017) Interleukin-25 primed mesenchymal stem cells achieve better therapeutic effects on dextran sulfate sodium-induced colitis via inhibiting Th17 immune response and inducing T regulatory cell phenotype. *Am. J. Transl. Res.* **9**, 4149–4160 PMID: 28979689
- 74 Su, J., Chen, T., Ji, X.-Y., Liu, C., Yadav, P.K., Wu, R. et al. (2013) IL-25 downregulates Th1/Th17 immune response in an IL-10-dependent manner in inflammatory bowel disease. *Inflamm. Bowel Dis.* **19**, 720–728 <https://doi.org/10.1097/MIB.0b013e3182802a76>
- 75 Shi, T., Xie, Y., Fu, Y., Zhou, Q., Ma, Z., Ma, J. et al. (2017) The signaling axis of microRNA-31/interleukin-25 regulates Th1/Th17-mediated inflammation response in colitis. *Mucosal Immunol.* **10**, 983–995 <https://doi.org/10.1038/mi.2016.102>
- 76 Caruso, R., Sarra, M., Stolfi, C., Rizzo, A., Fina, D., Fantini, M.C. et al. (2009) Interleukin-25 inhibits interleukin-12 production and Th1 cell-driven inflammation in the gut. *Gastroenterology* **136**, 2270–2279 <https://doi.org/10.1053/j.gastro.2009.02.049>
- 77 Thelen, T.D., Green, R.M. and Ziegler, S.F. (2016) Acute blockade of IL-25 in a colitis associated colon cancer model leads to increased tumor burden. *Sci. Rep.* **6**, 25643 <https://doi.org/10.1038/srep25643>
- 78 Reynolds, J.M., Lee, Y.-H., Shi, Y., Wang, X., Angkasekwinai, P., Nallaparaju, K.C. et al. (2015) Interleukin-17B antagonizes interleukin-25-mediated mucosal inflammation. *Immunity* **42**, 692–703 <https://doi.org/10.1016/j.immuni.2015.03.008>
- 79 Mirchandani, A.S., Salmond, R.J. and Liew, F.Y. (2012) Interleukin-33 and the function of innate lymphoid cells. *Trends Immunol.* **33**, 389–396 <https://doi.org/10.1016/j.it.2012.04.005>
- 80 Griesenauer, B. and Paczesny, S. (2017) The ST2/IL-33 axis in immune cells during inflammatory diseases. *Front. Immunol.* **8**, 475 <https://doi.org/10.3389/fimmu.2017.00475>
- 81 Palmieri, V., Ebel, J.-F., Ngo Thi Phuong, N., Klopffleisch, R., Vu, V.P., Adamczyk, A. et al. (2021) Interleukin-33 signaling exacerbates experimental infectious colitis by enhancing gut permeability and inhibiting protective Th17 immunity. *Mucosal Immunol.* **14**, 923–936 <https://doi.org/10.1038/s41385-021-00386-7>
- 82 Seidelin, J.B., Bjerrum, J.T., Coskun, M., Widjaya, B., Vainer, B. and Nielsen, O.H. (2010) IL-33 is upregulated in colonocytes of ulcerative colitis. *Immunol. Lett.* **128**, 80–85 <https://doi.org/10.1016/j.imlet.2009.11.001>

- 83 Kobori, A., Yagi, Y., Imaeda, H., Ban, H., Bamba, S., Tsujikawa, T. et al. (2010) Interleukin-33 expression is specifically enhanced in inflamed mucosa of ulcerative colitis. *J. Gastroenterol.* **45**, 999–1007 <https://doi.org/10.1007/s00535-010-0245-1>
- 84 Schiering, C., Krausgruber, T., Chomka, A., Fröhlich, A., Adelmann, K., Wohlfert, E.A. et al. (2014) The alarmin IL-33 promotes regulatory T-cell function in the intestine. *Nature* **513**, 564–568 <https://doi.org/10.1038/nature13577>
- 85 Duan, L., Chen, J., Zhang, H., Yang, H., Zhu, P., Xiong, A. et al. (2012) Interleukin-33 ameliorates experimental colitis through promoting Th2/Foxp3 + regulatory T-cell responses in mice. *Mol. Med.* **18**, 753–761 <https://doi.org/10.2119/molmed.2011.00428>
- 86 Ziegler, S.F., Roan, F., Bell, B.D., Stoklasek, T.A., Kitajima, M. and Han, H. (2013) The biology of thymic stromal lymphopoietin (TSLP). *Adv. Pharmacol. San Diego Calif.* **66**, 129–155 <https://doi.org/10.1016/B978-0-12-404717-4.00004-4>
- 87 Verstraete, K., Peelman, F., Braun, H., Lopez, J., Van Rompaey, D., Dansercoer, A., et al. (2017) Structure and antagonism of the receptor complex mediated by human TSLP in allergy and asthma. *Nat. Commun.* **8**, 14937 <https://doi.org/10.1038/ncomms14937>
- 88 Tanaka, J., Saga, K., Kido, M., Nishiura, H., Akamatsu, T., Chiba, T., et al. (2010) Proinflammatory Th2 cytokines induce production of thymic stromal lymphopoietin in human colonic epithelial cells. *Dig. Dis. Sci.* **55**, 1896–1904 <https://doi.org/10.1007/s10620-009-0979-x>
- 89 Whiteoak, S.R., Claridge, A., Balendran, C.A., Harris, R.J., Gwiggner, M., Bondanese, V.P., et al. (2018) MicroRNA-31 targets thymic stromal lymphopoietin in mucosal infiltrated CD4+ T cells: a role in achieving mucosal healing in ulcerative colitis? *Inflamm. Bowel Dis.* **24**, 2377–2385 <https://doi.org/10.1093/ibd/izy213>
- 90 Tahaghoghi-Hajghorbani, S., Ajami, A., Ghorbanalipoor, S., Hosseini-khah, Z., Taghilo, S., Khaje-Enayati, P., et al. (2019) Protective effect of TSLP and IL-33 cytokines in ulcerative colitis. *Autoimmun. Highlights* **10**, 1 <https://doi.org/10.1186/s13317-019-0110-z>
- 91 Taylor, B.C., Zaph, C., Troy, A.E., Du, Y., Guild, K.J., Comeau, M.R., et al. (2009) TSLP regulates intestinal immunity and inflammation in mouse models of helminth infection and colitis. *J. Exp. Med.* **206**, 655–667 <https://doi.org/10.1084/jem.20081499>
- 92 Aubry, C., Michon, C., Chain, F., Chvatchenko, Y., Goffin, L., Zimmerli, S.C., et al. (2015) Protective effect of TSLP delivered at the gut mucosa level by recombinant lactic acid bacteria in DSS-induced colitis mouse model. *Microb. Cell Factories* **14**, 176 <https://doi.org/10.1186/s12934-015-0367-5>