Mining coherent dense subgraphs across massive biological networks for functional discovery

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Received on January 15, 2005; accepted on March 27, 2005

ABSTRACT

Motivation: The rapid accumulation of biological network data translates into an urgent need for computational methods for graph pattern mining. One important problem is to identify recurrent patterns across multiple networks to discover biological modules. However, existing algorithms for frequent pattern mining become very costly in time and space as the pattern sizes and network numbers increase. Currently, no efficient algorithm is available for mining recurrent patterns across large collections of genome-wide networks.

Results: We developed a novel algorithm, CODENSE, to efficiently mine frequent coherent dense subgraphs across large numbers of massive graphs. Compared with previous methods, our approach is scalable in the number and size of the input graphs and adjustable in terms of exact or approximate pattern mining. Applying CODENSE to 39 co-expression networks derived from microarray datasets, we discovered a large number of functionally homogeneous clusters and made functional predictions for 169 uncharacterized yeast genes.

Availability: http://zhoulab.usc.edu/CODENSE/

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1 INTRODUCTION

The recent development of high-throughput technologies provides a range of opportunities to systematically characterize diverse types of biological networks. ‘Network Biology’ has been an emerging field in biology. The variety of biological networks can be classified into two categories: (1) physical networks, which represent physical interactions among molecules, e.g. protein-interaction, protein–DNA interaction and metabolic reactions and (2) conceptual networks, which represent functional associations of molecules derived from genomic data, e.g. co-expression relationships extracted from microarray data and genetic interactions obtained from synthetic lethality experiments. While the physical network data are as yet very limited in size, the large amount of microarray data allows us to infer conceptual functional associations of genes under various conditions for many model organisms, thus providing valuable information to study the functions and the dynamics of biological systems.

Studying the building principles of biological networks could potentially revolutionize our view of biology and disease pathologies (Barabasi and Oltvai, 2004). The popular clustering approach can draw densely connected modules from biological networks, which are often biologically meaningful, e.g. a dense protein interaction subnetwork may correspond to a protein complex (Bader and Hogue, 2003; Spirin and Mirny, 2003), and a dense co-expression network may represent a tight co-expression cluster (Sharan and Shamir, 2000). On account of the noisy nature of high-throughput data, a significant number of spurious edges exist in biological networks, which may lead to the discovery of false patterns. Since biological modules are expected to be active across multiple conditions, we can easily filter out spurious edges by mining frequent patterns in multiple biological networks simultaneously. A straightforward approach is to aggregate these networks together and identify dense subgraphs in the aggregated graph. However, it could result in false dense subgraphs that may not occur frequently in the original networks. Figure 1a illustrates such an example with a cartoon of six graphs. If we simply add these graphs together to construct a summary graph, we may find a dense subgraph comprising vertices a, b, c and d. Unfortunately, this subgraph is neither dense nor frequent in the original graphs.

A potential solution to the false pattern problem is mining frequent subgraphs directly. A subgraph is frequent if it occurs multiple times in a set of graphs. Frequent subgraph discovery, in general, is considered a hard problem. However, biological networks can often be modeled as a special class of graph where each gene occurs once and only once in a graph. That means, our graph has distinct node labels, and we do not have the ‘subgraph isomorphism problem’ which is NP-hard and so...
Fig. 1. (a) Given six graphs with the same vertex set but different edge sets, we construct a summary graph by adding these six graphs together and by deleting edges that occur less than three times in the graphs. The dense subgraph in the summary graph \( \{a, b, c, d\} \) does not actually occur in any original graph. (b) The vertices \( e \) and \( f \) are shared by cliques \( \{a, b, c, d, e, f\} \) and \( \{e, f, h, i\} \); they can be assigned to both cliques only by approaches that are able to detect overlapping dense subgraphs (cliques are the densest subgraphs).

Fig. 2. Shown are six graphs with the same vertex set but different edge sets. The bold subgraph \( \{c-e, c-f, c-h, f-e, f-h, e-g, g-h, h-d, g-d\} \) occurs in three of the six graphs (graphs 1, 3, and 6). However, the vertices/edges of this subgraph may not be tightly associated in their occurrence, because one large component, the subgraph \( \{c-e, c-f, c-h, f-e, f-h, e-h\} \) occurs in every network.

far, constitutes the bottleneck of subgraph frequency counting. We term such graphs relation graphs. Recently, we and others have designed efficient approaches to identify frequent subgraphs across multiple relation networks by decomposing the networks into smaller pieces and applying pattern expansion techniques (Kuramochi and Karypis, 2004; Yan et al., 2005) or by performing frequent set mining with subsequent connectivity checking (Koyuturk et al., 2004). However, these approaches encounter scalability and interpretability issues when being applied to massive biological networks: (1) In both approaches we tested, the time and memory requirements increase exponentially with increasing size of patterns and increasing number of networks. The number of frequent dense subgraphs is explosive when there are very large frequent dense subgraphs, e.g. subgraphs with hundreds of edges. (2) A frequent dense subgraph may not represent a tight association among its nodes. Figure 2 shows a sample network dataset. Vertices \( e, c, f, h, d \) and \( g \) form a frequent dense subgraph. However, biologically it is more interesting to divide this subgraph into two modules, one comprising \( e, c, f \) and \( h \); the other comprising \( h, d, g \) and \( e \) since these two modules have different occurrences throughout this graph set (for details refer to the figure caption). As one can see, frequent dense subgraphs may not capture accurate information for the discovery of biological modules.

In this paper, we address the two aforementioned issues and develop a novel algorithm, called ‘CODENSE’, to mine coherent dense subgraphs, a concept having better interpretability than frequent graph. All edges in a coherent subgraph should exhibit correlated occurrences in the whole graph set. We also term this kind of subgraph ‘network module’. According to the definition of coherent dense subgraph, we are able to distinguish the two modules shown in Figure 2. Moreover, the design of CODENSE can solve the scalability issue. Instead of mining each biological network individually, CODENSE compresses the networks into two metagraphs and performs clustering in these two graphs only. Thus, CODENSE can handle any large number of networks. Using CODENSE, we can successfully identify high-quality network modules within limited time and memory.

As a side product, CODENSE also provides a solution to a graph mining problem—discovery of overlapping graph clusters. It is known that under different conditions, one gene may serve different roles and be involved in different functional groups (Gasch and Eisen, 2002), thus identifying overlapping clusters is important in biological applications. However, most graph clustering algorithms follow the methodology of graph partitioning (Spirin and Mirny, 2003; Van Dongen, 2000); they usually cannot identify overlapping clusters. For example, in Figure 1b, two cliques...
{a, b, c, d, e, f} and {e, f, h, i} share two common vertices {e, f}. If a partition-based method the former clique identified first, the latter clique will be missed. Here, as a component of CODENSE, we designed a novel algorithm, MODES (Mining Overlapping DEEnse Subgraphs), to identify overlapping graph clusters.

As an application example, we used CODENSE to identify frequent co-expression clusters across multiple microarray datasets. A microarray dataset is modeled as an unweighted and undirected network, where each gene is represented by one node and two genes are connected with an edge if they show high expression correlation. A densely connected subgraph in these networks corresponds to a tight co-expression cluster. However, several studies pointed out that clusters derived from a single microarray dataset often include spurious links and may not be functionally homogeneous (Allocco et al., 2004; Clare and King, 2002). A recent study showed that genes co-expressed in multiple datasets tend to have the same functions (Lee et al., 2004). Here, applying our methods to 39 microarray datasets of Saccharomyces cerevisiae, we demonstrated that recurrent expression clusters are very likely to be homogeneous in function and regulation, and can be used to perform large-scale functional annotation of uncharacterized genes.

The remainder of this paper is organized as follows. Section 2 gives the problem formulation. Our algorithms of mining coherent dense subgraphs and overlapping dense subgraphs are examined in Sections 3 and 4, respectively. We give a thorough comparison between our approach and the others in Section 5. The experimental study and biological applications are examined in Section 6. Section 7 concludes our study.

2 PROBLEM FORMULATION

A relation graph set consists of n undirected simple graphs, \( D = \{G_i = (V_i, E_i)\}, i = 1, \ldots, n, E_i \subseteq V \times V \), where a common vertex set \( V \) is shared by the graphs in the set. We denote the vertex set of a graph \( G \) by \( V(G) \) and the edge set by \( E(G) \). Let \( w_i(u, v) \) be the weight of an edge \( e_i(u, v) \) in \( G_i \). For an unweighted graph, \( w_i(u, v) = 1 \) if there is an edge between \( u \) and \( v \), otherwise 0. We choose to illustrate the principles on unweighted and undirected graphs in this paper, although our algorithm should be extendable to weighted and directed graphs.

**Definition 1 (Support).** Given a relation graph dataset, \( D = \{G_1, G_2, \ldots, G_n\} \), where \( G_i = (V, E_i) \), the support of a graph \( g \) is the number of graphs (in \( D \)) where \( g \) is a subgraph, written \( \text{support}(g) \). A graph is frequent if its support is greater than a minimum support threshold.

**Definition 2 (Summary Graph).** Given a relation graph dataset, \( D = \{G_1, G_2, \ldots, G_n\} \), where \( G_i = (V, E_i) \), the summary graph of \( D \) is an unweighted graph \( \hat{G} = (V, \hat{E}) \) where an edge is present if it occurs in more than \( k \) graphs in \( D \), where \( k \) is a user-defined support threshold (see an example in Fig. 1a).

**Definition 3 (Edge Support Vector).** Given a relation graph dataset, \( D = \{G_1, G_2, \ldots, G_n\} \), where \( G_i = (V, E_i) \), the support vector of an edge \( e \), written \( \text{support}(e) \), is of length \( n \) where \( n \) is the number of graphs. The \( i \)-th element of \( \text{support}(e) \) corresponds to the weight of edge \( e \) in the \( i \)-th graph.

The support vector of the edge \( (a, b) \) for the six graphs shown in Figure 1a is \([1, 1, 1, 0, 0, 0]\), while the support vector of the edge \( (b, c) \) is \([0, 0, 0, 1, 1, 1]\). As one can see, edges \((a, b)\) and \((b, c)\) are not correlated in this dataset, though both of them are frequent.

We use a special graph, termed second-order graph (denoted as \( S \)), to illustrate the co-occurrence of edges across all graphs in a relation graph set \( D \). Each edge in \( D \) is transformed into a vertex in \( S \), and two vertices \( u \) and \( v \) in \( S \) will be connected if their corresponding edge support vectors \( \text{support}(u) \) and \( \text{support}(v) \) in \( D \) show high similarity. Depending on whether or not the edges are weighted, the similarity measure could be Euclidean distance or Pearson’s correlation. Figure 3 (Step 3b) shows how to generate a second-order graph from a set of edge support vectors. For example, the Euclidean distance between the support vectors of edges \((c, e)\) and \((c, i)\) is only 1, so we create an edge between the vertices \((c, e)\) and \((c, i)\) in the second-order graph \( S \) shown in Figure 3. In contrast to the second-order graph, we term the original graphs \( G_i \) the first-order graphs. The utilization of second-order graph discussed in this paper is one type of second-order analysis, a concept that has been proposed in our previous publication (Zhou et al., 2005).

**Definition 4 (Second-Order Graph).** Given a relation graph dataset, \( D = \{G_1, G_2, \ldots, G_n\} \), where \( G_i = (V, E_i) \), the second-order graph is an unweighted graph \( S = (V \times V, E_s) \) where the vertex set of \( S \) is the edge set of \( G \), and an edge connects vertices \( u \) and \( v \) if the similarity between the corresponding edge support vectors \( \text{support}(u) \) and \( \text{support}(v) \) is greater than a threshold.

In reality, if \( G_i \) is large and dense, \( S \) will be impractically large. Therefore, to achieve efficiency, in this paper we construct \( S \) each time only for a subgraph of the summary graph \( \hat{G} \).

**Definition 5 (Coherent Graph).** Given a relation graph dataset, \( D = \{G_1, G_2, \ldots, G_n\} \), where \( G_i = (V, E_i) \), a subgraph \( \text{sub}(\hat{G}) \) is coherent if all the edges of \( \text{sub}(\hat{G}) \) have support higher than \( k \) and the second-order graph of \( \text{sub}(\hat{G}) \) is dense.

**Definition 6 (Graph Density).** The density of a graph \( g \), written \( \text{density}(g) \), is \( 2m/[n(n-1)] \) where \( m \) is the number of edges and \( n \) the number of vertices in \( g \).
The problem of mining coherent dense subgraphs is formulated as follows: given a relation graph dataset, \( D = \{G_1, G_2, \ldots, G_n\} \), discover subgraphs \( g \) that satisfy the following two criteria simultaneously: (1) \( g \) is a densely connected subgraph of the summary graph; and (2) \( g \) is a coherent graph. As discussed in Section 1, the coherent dense subgraphs have significant biological interests.

3 CODENSE: MINING COHERENT DENSE SUBGRAPHS

A scalable algorithm for mining coherent dense subgraphs must perform well despite increasing number of graphs and increasing size of patterns. In order to tackle this problem, we investigate the relation between a coherent dense subgraph and the summary graph as well as its second-order graph. We make the following two observations:

(1) If a frequent subgraph is dense, then it must be a dense subgraph in the summary graph. However, the reverse conclusion is not true. A dense subgraph in the summary graph may be neither frequent nor dense in the original dataset (e.g. Fig. 1a).

(2) If a subgraph is coherent (its edges show high correlation in their occurrences across a graph set), then its second-order graph must be dense.

**Fig. 3. CODENSE: discover coherent dense subgraphs across multiple graphs (dense subgraphs are marked bold).**

**Algorithm 1: CODENSE**

1. build a summary graph \( \hat{G} \) across multiple relation graphs \( G_1, G_2, \ldots, G_n \);
2. mine dense summary subgraphs \( \text{sub}(\hat{G}) \) in \( \hat{G} \) using MODES;
   for each dense summary subgraph \( \text{sub}(\hat{G}) \) do
   (3) construct the second-order graph \( S \);
   (4) mine dense subgraphs \( \text{sub}(S) \) in \( S \) using MODES;
   (5) for each dense subgraph \( \text{sub}(S) \) do
      * convert \( \text{sub}(S) \) into the first-order graph \( G \);
      * mine dense subgraphs \( \text{sub}(G) \) in \( G \) using MODES;
      * output \( \text{sub}(G) \);

These two observations provide a clue to mining coherent dense subgraphs with reasonable computational cost. According to Observation 1, each frequent dense subgraph is a subgraph of a dense summary graph. We can start from the summary graph and mine dense summary subgraphs first. Once it is done, we can single out coherent subgraphs from...
dense summary subgraphs by mining their corresponding second-order graphs.

Our CODENSE algorithm consists of five steps, as outlined in Algorithm 1 and illustrated in Figure 3. In Steps 2, 4 and 5, a novel algorithm to discover overlapping dense subgraphs, called ‘MODES’, is employed. We will describe the design of the MODES algorithm in Section 4.

In Step 1, CODENSE builds a summary graph by eliminating infrequent edges.

In Step 2, CODENSE identifies dense subgraphs (possibly overlapping) in the summary graph. Although the dense subgraphs in the summary graph may not be truly frequently occurring in the original graphs, they do serve as a superset of potential frequent dense subgraphs in the original graphs. We start with this superset to refine the search results.

In Step 3, CODENSE builds a second-order graph for each dense summary subgraph.

In Step 4, CODENSE identifies dense subgraphs in the second-order graph. The high connectivity among vertices in the second-order graph indicates that the corresponding edges show high similarity in their occurrences across the $n$ original graphs.

In Step 5, CODENSE discovers the real coherent dense subgraphs. Although a dense subgraph sub($S$) found in Step 4 guarantees the co-occurrence of its edges across the $n$ relation graphs, those edges may no longer form a densely connected graph in the original summary graph. To eliminate such cases, we convert the vertices in sub($S$) back to edges and then apply the MODES algorithm to identify dense subgraphs. The resulting subgraphs by construction will satisfy the criteria for coherent dense subgraphs: (1) they are dense subgraphs and all of their edges occur frequently; and (2) their edges are highly correlated in their occurrences across the $n$ relation graphs.

In comparison with previous frequent graph mining algorithms, our algorithm may not show a significant advantage when the number of relation graphs $n$ is small. However, when $n$ is large, CODENSE will achieve significant time and memory efficiency since it works on two graphs only: summary graph and second-order graph instead of the $n$ original graphs.

In Step 3 by transforming all edges directly into the second-order graph with further steps unchanged. The clustering of the summary graph serves the purpose of restricting the second-order graph to a reasonable size to avoid excessive computation.

### 4 MODES: MINING OVERLAPPING DENSE SUBGRAPHS

The overlapping dense subgraph mining algorithm, called MODES, is frequently used in CODENSE. In this section, we present the details of its design. MODES is developed based on HCS (Mining Highly Connected Subgraphs) (Hartuv and Shamir, 2000), with two new features: (1) MODES is more efficient in identifying dense subgraphs; and more importantly, (2) MODES can discover overlapping subgraphs.

It is computationally intractable to enumerate all dense subgraphs in a large graph. Usually, a large graph is first clustered hierarchically and the dense clusters are singled out. HCS is this kind of algorithm. It recursively partitions the graph into two subgraphs until its minimum cut is no less than half of its vertex set size.

**Definition 7 (Minimum Cut).** Given a graph $G$, an edge cut is a set of edges $E_c$ such that $E(G) - E_c$ is disconnected. A minimum cut is the smallest set in all edge cuts.

There are two issues remaining in HCS. First, HCS cannot identify overlapping dense subgraphs because of its nature of graph-partitioning. Second, the above recursive partitioning process is time consuming. The fastest deterministic minimum cut algorithm in practice has time complexity $O(|V||E| + |V|^2 \log |V|)$, where $|V|$ and $|E|$ are the vertex set size and the edge set size of a given graph (Stoer and Wagner, 1997). The minimum cut criterion adopted by HCS favors cutting small sets of nodes from the graph. When applying the algorithm repeatedly to a large graph consisting of more than thousands of vertices, such unbalanced cuts could lead to unexpected high costs. As verified by our experiments, we found HCS often cuts off one node in each iteration, thus having time complexity $O(|V|^2|E| + |V|^3 \log |V|)$.

To avoid the undesirable bias for partitioning out small sets of vertices and to speed up the process, we apply the normalized cut (Shi and Malik, 2000) instead of minimum cut in the initial runs of the HCS algorithm. Normalized cut is able to better balance the sizes of the partitions. When the size of the partitions generated by normalized cut is reasonably small, we proceed with the minimum cut algorithm to identify dense subgraphs. We revert to the use of the minimum cut in the later stage to exploit its power better in clustering without severe effects on computational cost.

In order to identify overlapping dense subgraphs, we designed the following procedure, as outlined in Algorithm 2 and illustrated in Figure 4: (1) We mine dense subgraphs using the above modified HCS algorithm in a given graph $G$. (2) Each discovered dense subgraph sub($G$) is then condensed into a single vertex $v'$; any vertex $v$ that does not belong to sub($G$) will have an edge with $v'$ if $v$ has edge with a vertex in sub($G$). (3) The condensed graph, written $G'$, is then re-clustered using the above modified HCS algorithm. (4) Once the clustering is done, if any newly discovered dense subgraph
sub(G')] may not be dense any more, in order to extract the dense subgraphs, we perform Step 1 again on sub(G') to identify its dense subgraphs. This procedure is repeated until no new dense graph is discovered. To speed up the computation, we require that a new dense subgraph should have <90% overlap with any existing dense subgraph.

The running time of computing normalized cut is determined by the extraction of the second smallest eigenvalue of 
$I - D^{-1/2}WD^{-1/2}$, where $W(i,j)$ is the weight of edge $(i,j), D(i,i) = \sum_j W(i,j)$. Its complexity is $O(n^c), 1 \leq c \leq 3$. ($O(n^3)$ is the upper bound on solving the eigensystem of an $n$-dimension square matrix) (Shi and Malik, 2000). Empirically, the value of $c$ is around 1.5 when the graph is very sparse (Shi and Malik, 2000). Assume a graph $G = (V,E)$ is clustered to $k$ partitions $V_1,V_2,\ldots,V_k$, where $V_1 \cup V_2 \cup \cdots \cup V_k = V$. The total cost of clustering these $k$ partitions is $O(|V_1|^c + |V_2|^c + \ldots + |V_k|^c)$ which is at most $O(|V|^c)$ given $c \geq 1$. For hierarchical recursive clustering, the cost is $O(d|V|^c)$, where $d$ is the largest recursive steps. The value of $d$ is between $O(\log |V|)$ and $O(|V|)$. Since normalized cut prefers balanced partitions, in general, $d$ is far away from $|V|$ but close to $\log |V|$, which means our algorithm MODES can run much faster than the worst case.

MODES may iterate several times in order to discover overlapping dense subgraphs. Each iteration involves a hierarchical clustering. Assume that the maximum number of recursions for a given graph is $r$ (Algorithm 2, MODES). The running time of MODES is $O(r|V|^{2.5})$ for a sparse graph. In practice, we may restrict the recursion depth.

Figure 4 illustrates a clustering example of mining overlapping dense subgraphs. The upper-left graph in the figure is the original graph. Obviously, it has two dense connected subgraphs. The readers can simulate our MODES algorithm to detect these two subgraphs. As demonstrated in this example, MODES is able to discover some dense subgraphs that traditional clustering approaches cannot find.

**Algorithm 2: MODES**

1. mine dense subgraphs, sub(G), in a given graph G using the modified HCS algorithm in which normalized-cut and min-cut are combined for recursive partitions;
2. for each dense subgraph sub(G) do
   - condense sub(G) into a condensed vertex $v'$ in the original graph;
   - denote the condensed graph by $G'$;
3. mine dense subgraphs sub($G'$) from the condensed graph $G'$ using the modified HCS algorithm again;
4. for each dense subgraph sub($G'$) do
   if sub($G'$) contains any condensed vertex then
     (a) restore each condensed component $C$ in sub($G'$) ;
     (b) remove vertices $v$, if $v \in C$ and $v$ is connected to less than $p\% (|V| \text{ sub}(G')) - |V(C)|$ vertices;
   - repeat Steps 1–4 until no new dense subgraph is discovered in sub($G'$);

**5 COMPARISON WITH OTHER METHODS**

Our approach proposed so far simplifies the problem of identifying coherent dense subgraphs across $n$ graphs into a problem of identifying dense subgraphs in two special graphs: the summary graph and the second-order graph. Here, we compare CODENSE with other methods and highlight its major advantages.

*By transforming all necessary information of the $n$ graphs into two graphs, CODENSE achieves significant time and memory efficiency.* Prior frequent graph mining approaches, mostly based on graph pattern expansion technique (Kuramochi and Karypis, 2004; Yan et al., 2005), iteratively extend frequent patterns and check their support. However, these approaches are infeasible in discovering large patterns. The problem of counting the number of distinct maximal frequent subgraphs is #P-complete, thereby providing a strong implication that the problem of mining maximal frequent subgraphs may be NP-hard (Yang, 2004).

CODENSE can mine coherent subgraph patterns which likely represent true network modules. As discussed in the introduction section, frequent patterns may contain sub
components that differ significantly in their support, although this is not an issue for coherent patterns.

**CODENSE can mine both exact and approximate patterns.** We can control the similarity threshold of the edge support vectors for edge construction in the second-order graph as well as the density threshold for the subgraph discovery in the second-order graph. The lower the two thresholds, the higher the degree of approximation in the output of CODENSE. Conversely, if the support vector similarity threshold is set to 100% and the second-order dense subgraph is required to form cliques, the identified subgraphs shall occur exactly in the same graphs.

**CODENSE can be extended to pattern mining on weighted graphs** since the core algorithms, normalized-cut and minimum-cut based clustering, can be applied to both weighted and unweighted graphs.

CODENSE can be further modified and extended to identify more subtle patterns. For example, instead of clustering edges with overall similar edge support vectors, one may use biclustering algorithms (Cheng and Church, 2000) to identify edges showing similar supports in a subset of datasets. Efficient algorithm design for this purpose is currently under development.

6 EXPERIMENTAL STUDY

6.1 Graph modeling and parameter setting

In this study, we use the co-expression networks derived from microarray datasets as a testing system for CODENSE. We integrated 39 yeast microarray datasets, each comprising the expression profiles of 6661 genes in at least 8 experiments, from Stanford Microarray Database and the NCBI Gene Expression Omnibus (details of the dataset are available at http://zhoulab.usc.edu/CODENSE/). The similarity between two genes in one microarray data set is measured by Pearson’s correlation between their expression patterns. We transform the Pearson’s correlation (denoted as r) into another quantity, \[ \sqrt{(n-2)r^2/1-r^2} \], and model this quantity as a t-distribution with \( n-2 \) degrees of freedom, where \( n \) is the number of measurements used in the computation of the Pearson’s correlation. From each microarray data set, we construct a relation network where two genes are connected if the Pearson’s correlation between their expression patterns is significant at \( \alpha = 0.01 \) level.

We construct a summary graph \( \hat{G} \), collecting edges which support at least 6 over the 39 relation networks. MODES is first applied to \( \hat{G} \) to identify subgraphs with density \( \geq d_1 \). For each identified subgraph, we then construct a second-order graph by transforming edges to vertices. Here, we build the edge support vector (of length 39) by computing the Pearson’s correlation between the expression profiles of two genes (two vertices) in each of the 39 datasets (note that this is different from the binary edge support vectors illustrated in Fig. 3). If the Pearson’s correlation between support vectors of two edges is significant at \( \alpha = 0.001 \) level, we connect their transformed vertices with an edge in the second-order graph. We again apply the MODES algorithm to the second-order graph to identify subgraphs with density \( \geq d_2 \). Such identified second-order graph is then transformed back to the summary graph (vertices \( \rightarrow \) edges), and MODES is employed once more to identify subgraphs with density \( \geq d_3 \). In this experiment, we set the three density cutoffs \( d_1 = d_2 = d_3 = 0.4 \). However, they can be adjusted to accommodate specific needs of the users. For example, decreasing \( d_1 \) and \( d_3 \) will favor sparse coherent patterns; and increasing \( d_2 \) will strengthen the co-occurrence of edges across all networks in an identified pattern. In this study, we only focus on the coherent subgraphs with at least four vertices.

6.2 Functional module discovery

We applied CODENSE to discover coherent clusters across the 39 co-expression networks and compare the result with the dense subgraphs generated by MODES on the summary graph.

To quantify the comparison, we assess the clustering quality by determining the percentage of functionally homogeneous clusters among all identified clusters. We used the Gene Ontology (GO) biological process annotation, and consider a cluster to be functionally homogeneous if (1) the functional homogeneity modeled by the hypergeometric distribution (Wu et al., 2002) shall be significant at \( \alpha = 0.01 \); and (2) at least 40% of its member genes with known annotations belong to a specific GO functional category. Using the GeneOntology biological process annotation, we define specific functions to be those associated with GeneOnology nodes that are more than 5 levels below the root.

We found that CODENSE significantly increased the percentage of functionally homogeneous clusters from the MODES results by filtering out larger amounts of noisy genes or noisy clusters. MODES identified 366 clusters, among which 151 (42%) are functionally homogeneous. Starting with the 366 clusters, after the second-order clustering, CODENSE identified 770 clusters that have at least 4 annotated genes. Of these clusters, 76% are functionally homogeneous. This shows a 34% increase of functionally homogeneous clusters compared with the MODES results.

The major performance improvement of CODENSE over MODES is attributed to the power of the second-order clustering in eliminating dense summary subgraphs whose edges do not show co-occurrence across all networks. For example, MODES identified a five-gene clique in the summary graph, \{ MSF1, PHB1, CBP4, ND11, SCO2 \}. However, the five genes come from diverse functional categories, such as ‘protein biosynthesis’, ‘replicative cell aging’ and ‘mitochondrial electron transport’. In fact, although all edges of this clique occur in at east 6 networks, their co-occurrence is not significant across the 39 networks (Fig. 5a). The second-order...
clustering can filter out such pseudoclusters, thus providing more reliable results.

Moreover, we found instances in which the second-order clustering can remove the noisy genes in a functionally diverse cluster so that its subcluster(s) becomes functionally homogeneous. For example, in a cluster containing 54 genes that are associated with diverse functional categories, such as ‘organic acid metabolism’, ‘carboxylic acid metabolism’ and ‘amino acid and derivative metabolism’, none of these categories is significantly overpresented in this cluster. After the second-order clustering, a coherent subcluster emerges with seven annotated genes, which is dominated by the function ‘organic acid metabolism’ ($P$-value = $2.18 \times 10^{-5}$).

### 6.3 Functional annotation

The large number of functionally homogeneous clusters identified by CODENSE provides a solid foundation for functional annotation of uncharacterized genes. For those clusters containing unknown genes, if the most dominating GO functional category is significantly overrepresented (Bonferroni corrected hypergeometric $P$-value <0.01), we annotated the unknown genes with that function. To assess the prediction accuracy of our method, we employed a ‘leave-one-out’ approach by masking a known gene to be unknown and assign its function based on the remaining known genes in the cluster. We consider a prediction to be correct, if the lowest common ancestor of the predicted and known functional categories of that gene is five levels below the root in the GO hierarchy. That is, the predicted and the known categories will merge into the same category at least at the level 6 of the GO hierarchy. Note that the annotated yeast genes encompass 160 functional categories at the level 6 of the GO hierarchy. We have assigned functions to 448 known genes and achieved a prediction accuracy of 50%.

By applying this approach to unknown genes, we made a functional prediction for 169 genes, covering a wide range of functional categories from cellular protein metabolism, protein biosynthesis, ribosome biogenesis, nucleobase, nucleoside, nucleotide and nucleic acid metabolism, cellular biosynthesis, etc. Figure 5b illustrates an example of our predictions, in which the uncharacterized gene ASC1 is predicted to be involved in ‘protein biosynthesis’, because all of the remaining five genes in the same subgraph participate in that biological process. The comprehensive prediction results are available at http://zhoulab.usc.edu/CODENSE/. Many of our predictions are supported by experimental studies in the literature. For example, we predicted RRP15 to participate in ‘ribosome biogenesis’. According to a recent study (De et al., 2005), this gene is involved in pre-rRNA processing. We assigned the function ‘protein biosynthesis’ to YMR116C; and two studies showed that it is involved in translation regulation (Chantrel et al., 1998) and control (Gerbasi et al., 2004). We predicted QRI5 to be involved in ‘protein biosynthesis’; QRI5 has been shown to participate in a common regulatory process together with MSS51 (Simon et al., 1992) and the GO annotation of MSS51 is ‘positive regulation of translation and protein biosynthesis’.

## 7 CONCLUSIONS

We developed a novel algorithm, CODENSE, to efficiently mine coherent dense subgraphs across massive biological networks. In comparison with previous approaches, CODENSE is scalable in the number and the size of the networks to mine, adjustable in terms of exact or approximate coherent pattern mining and extendable to weighted and directed networks. It provides an efficient tool for the identification of network modules and for the functional discovery in the ever increasing biological networks. The method can integrate heterogeneous network data (e.g. protein interaction network, genetic interaction network and co-expression networks) to reveal consistent biological signals. The discovered network modules can be used in a variety of biological applications, e.g. predict the functions of unknown genes, construct the transcription modules and infer the potential protein assembly mechanisms.

## ACKNOWLEDGEMENTS

We thank Min Xu for the helpful discussions and Ming-Chih J. Kao for proof reading of the manuscript. The work of H.H.
is partially supported by the nih grant 1P50CA112952. The work of X.J.Z. is supported by the usc faculty set up grant and the NIH grant 5R01GM067243. The work of X.Y. and J.H. is supported by NSF IIS-02-091999.

REFERENCES


