Structural bioinformatics

Porter: a new, accurate server for protein secondary structure prediction

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ABSTRACT

Summary: Porter is a new system for protein secondary structure prediction in three classes. Porter relies on bidirectional recurrent neural networks with shortcut connections, accurate coding of input profiles obtained from multiple sequence alignments, second stage filtering by recurrent neural networks, incorporation of long range information and large-scale ensembles of predictors. Porter’s accuracy, tested by rigorous 5-fold cross-validation on a large set of proteins, exceeds 79%, significantly above a copy of the state-of-the-art SSpro server, better than any system published to date.

Availability: Porter is available as a public web server at http://distill.ucd.ie/porter/

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Protein secondary structure (SS) prediction is an important stage for the prediction of protein structure and function. Accurate SS information has been shown to improve the sensitivity of threading methods (e.g. Jones, 1999b) and is at the core of most ab initio methods (e.g. see Bradley et al., 2003) for the prediction of protein structure. Virtually all modern methods for protein SS prediction are based on machine learning techniques (Jones, 1999a; Pollastri et al., 2002), and exploit evolutionary information in the form of profiles extracted from alignments of multiple homologous sequences (MSAs). The progress of these methods over the last 10 years has been slow, but steady, and is due to numerous factors: the ever-increasing size of training sets; more sensitive methods for the detection of homologues, such as PSI-BLAST (Altschul et al., 1997); the use of ensembles of multiple predictors trained independently, sometimes tens of them (Petersen et al., 2000); more sophisticated machine learning techniques (e.g. Pollastri et al., 2002).

We have developed Porter, a new server for protein SS prediction. Porter is based on two layers of Bidirectional Recurrent Neural Networks (BRNN) and is an evolution of SSpro (Pollastri et al., 2002), one of the most accurate public servers to date (Rost and Eyrich, 2001; Lesk et al., 2001). The novel elements of Porter are accurate coding of input profiles obtained from MSA, second stage filtering by recurrent neural networks, incorporation of long-range information, large-scale ensembles of predictors and larger training sets.

Datasets. Porter is trained on the December 2003 25% pdbs select list. After processing by DSSP (Kabsch and Sander, 1983) the set contains 2171 proteins and 344,653 amino acids. We assign eight DSSP classes as follows: H, G, I → Helix; E, B → Strand; S, T, . → Coil. This assignment is known to be ‘hard’ and had been adopted at CASP (Lesk et al., 2001). More lenient assignments generally lead to higher performances. Profiles obtained from MSA have been shown to improve significantly SS prediction performances (starting from Rost and Sander, 1993). In Porter, we use MSA extracted from the NR database as available on March 3, 2004, containing over 1.4 million sequences. Redundancy in the database was first reduced at a 98% threshold, leading to 1.05 million sequences finally. The alignments were generated by three runs of PSI-BLAST (Altschul et al., 1997).

Input coding. In Porter, the input at each residue is coded as a letter out of an alphabet of 25. Beside the 20 standard amino acids, B, U, X, Z and . (gap) are considered. The input presented to the networks is the frequency of each of the 24 non-gap symbols, plus the total frequency of gaps in each column of the alignment. This input coding scheme is richer than the 20-letter scheme adopted in SSpro (Pollastri et al., 2002).

Output filtering, incorporation of long-range information. We adopt a filtering network as for example in Rost and Sander (1993), but we augment the input to this network by the predictions of the first-stage network averaged over multiple contiguous windows, i.e. if \( \sigma_j = (\sigma_j, \beta_j, \gamma_j) \) are the outputs in position \( j \) of the first stage network corresponding to the estimated probabilities of helix, strand and coil given the inputs, the input to the second stage network in position \( j \) is the array \( I_j \):

\[
I_j = \left( \sigma_j, \sum_{h=k-w}^{k+w} \sigma_h, \ldots, \sum_{h=k-p}^{k+p} \sigma_h \right),
\]

where \( k_f = j + f(2w + 1) \), \( 2w + 1 \) is the size of the window over which first-stage predictions are averaged and \( 2p + 1 \) is the number of windows considered. In Porter \( w = 7 \) and \( p = 7 \), i.e. predictions at 225 contiguous residues are considered by the filtering network.

Large-scale ensembles. Five two-stage BRNN models are trained independently and ensemble averaged to build Porter. Differences among models are introduced by two factors: stochastic elements in the training protocol, such as different initial weights of the networks and different shuffling of the examples; different architecture and size of the models. In particular, we resorted to BRNN architectures with shortcuts (Baldi et al., 1999). In these, connections along the forward and backward hidden chains span more than...
one-residue intervals, creating shorter paths between inputs and outputs. Averaging the five models’ outputs leads to improvements in the range of 1–1.5% over single models. In Petersen et al. (2000), a slight improvement in the prediction accuracy was obtained by ‘brute ensembling’ of several tens of different models trained independently. Here, we adopted a less expensive technique: a copy of each of the five models is saved at regular intervals during training. The training protocol (similar to that described by Pollastri et al., 2002) guarantees that differences during training are non-trivial. In Porter we build an ensemble of 45 such copies (9 for each of the 5 models).

Results and conclusions. We measured the performances of each incremental improvement separately, by a 5-fold cross-validation procedure. The percentages of correctly classified residues (Q3), helices and strands (Qα, Qβ), and Matthews’ correlation coefficients for helices and strands (Cα, Cβ) by all systems are shown in Table 1. Q3 differences >0.07% are statistically significant. An exact copy of SSpro, retrained (SSproR) and incremental improvements leading to Porter

<table>
<thead>
<tr>
<th>Method</th>
<th>Q3 (%)</th>
<th>Qα (%)</th>
<th>Qβ (%)</th>
<th>Cα (%)</th>
<th>Cβ (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>SSpro 2.0</td>
<td>78.13</td>
<td>82.4</td>
<td>66.2</td>
<td>75.2</td>
<td>63.4</td>
</tr>
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<td>68.8</td>
<td>74.6</td>
<td>64.7</td>
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<tr>
<td>SSproRs</td>
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<td>82.0</td>
<td>68.4</td>
<td>74.7</td>
<td>65.1</td>
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<tr>
<td>+25 sym</td>
<td>78.54</td>
<td>81.9</td>
<td>68.6</td>
<td>74.8</td>
<td>65.4</td>
</tr>
<tr>
<td>+Filter</td>
<td>78.89</td>
<td>82.4</td>
<td>69.2</td>
<td>75.3</td>
<td>66.2</td>
</tr>
<tr>
<td>Porter</td>
<td>79.01</td>
<td>82.2</td>
<td>69.4</td>
<td>75.6</td>
<td>66.4</td>
</tr>
</tbody>
</table>

Results for all systems except SSpro 2.0 are measured in 5-fold cross-validation. Differences >0.07% are statistically significant. SSproRs, shortcut models; 25 sym, 25 input symbols.

Table 1. Overall Q3, Qα, Qβ, Cα, and Cβ for SSpro 2.0 in Pollastri et al. (2002), SSpro retrained (SSproR) and incremental improvements.

We also tested Porter on the EVA (Rost and Eyrich, 2001) common set 2, as available in November 2004, containing 134 proteins. To ensure a fair comparison, we retrained Porter from scratch, after having excluded from its training set all sequences with >25% similarity to any sequence in common2. On this set, Porter achieves SOV = 72.0% and Q3 = 76.8%, better by at least 1.2 and 1.9%, respectively, than all the other servers evaluated.

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REFERENCES