ART: a next-generation sequencing read simulator

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ABSTRACT

Summary: ART is a set of simulation tools that generate synthetic next-generation sequencing reads. This functionality is essential for testing and benchmarking tools for next-generation sequencing data analysis including read alignment, de novo assembly and genetic variation discovery. ART generates simulated sequencing reads by emulating the sequencing process with built-in, technology-specific read error models and base quality value parameters parameterized empirically in large sequencing datasets. We currently support all three major commercial next-generation sequencing platforms: Roche’s 454, Illumina’s Solexa and Applied Biosystems’ SOLiD. ART also allows the flexibility to use customized read error model parameters and quality profiles.

Availability: Both source and binary software packages are available at http://www.niehs.nih.gov/research/resources/software/art

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Supplementary information: Supplementary data are available at Bioinformatics online.

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1 INTRODUCTION

In the past few years, high-throughput next-generation sequencing technologies have effectively replaced earlier data types for genome-wide studies measuring gene expression changes and discovering genomic/epigenetic variations, and many tools were developed for analyzing such datasets. Simulated data is indispensable for guiding tool development and evaluating tool performance, and therefore it is essential to develop simulation software that can produce next-generation sequencing reads that captures the most essential characteristics of real data. Currently available read simulation programs include Wgsim from the Samtools package (Li et al., 2009) for generating Illumina sequencing reads, MetaSim (Richter et al., 2008) for simulating metagenomic data, Mason (http://seqan.de/projects/mason.html) for both Illumina and 454 reads, SimSeq (https://github.com/jstjohn/SimSeq) for Illumina reads and FlowSim (Balzer et al., 2010) for 454 reads. Although these programs work well in their domain, there is a need for a read simulation program that can deal with all major sequencing platforms, and generate sequence reads with both substitution and insertion–deletion (INDEL) errors, as appropriate for the error modes of each specific platform.

As a general simulator, our ART software was initially developed for simulation studies helping to design data collection modalities of the 1000 Genomes Project (Durbin et al., 2010). ART has been subsequently used by many users worldwide to facilitate sequencing software development. ART takes a set of DNA sequences (representing e.g. a reference genome), and generates ‘synthetic’ sequencing reads in a way that mimics the technology-specific sequencing process. ART comes with a set of technology-specific read error profiles, but it can also take user-supplied profiles to generate sequencing data with customized read length and error characteristics. ART can report simulated reads in the standard SAM alignment format and UCSC BED files.

2 FEATURES AND METHODS

ART simulates both single-end and paired-end sequencing reads of the three main commercial next-generation sequencing platforms: 454, Illumina and SOLiD. The built-in read length and read error profiles were derived from large sets of actual real sequencing data (see Supplementary Material). ART supports all three types of common sequencing errors: base substitutions, insertions and deletions.

2.1 Illumina read simulation

Illumina sequencing by synthesis is a base-by-base sequencing technology using a reversible terminator-based method, enabling detection of single bases as they are incorporated into growing DNA strands complementary to the template (Bentley, 2006). Since this technology reads out one base at a time, the main error mode is substitution rather than insertion or deletion. The probability of a substitution error is determined by the base quality score associated with the called base. The distribution of base quality scores is position-dependent: the mean quality score decreases as the distance from the associated with the called base. The distribution of base quality scores is position-dependent: the mean quality score decreases as the distance from the
Table 1. ART simulation speed. Speed measured for generating $10^8$ read coverage of human chromosome 17, for 454, Illumina, and SOLiD technology-specific parameters

<table>
<thead>
<tr>
<th>Platform</th>
<th>Read length</th>
<th>Single</th>
<th>Paired</th>
<th>Single</th>
<th>Paired</th>
</tr>
</thead>
<tbody>
<tr>
<td>454</td>
<td>Varied</td>
<td>491</td>
<td>676</td>
<td>7,049</td>
<td>10,490</td>
</tr>
<tr>
<td>Illumina</td>
<td>50 bp</td>
<td>290</td>
<td>300</td>
<td>55,997</td>
<td>54,130</td>
</tr>
<tr>
<td>SOLiD</td>
<td>33 bp</td>
<td>728</td>
<td>696</td>
<td>33,798</td>
<td>33,870</td>
</tr>
</tbody>
</table>

2.2 454 read simulation

Roche/454 sequencing is a pyrosequencing technology that tests for the presence of each of the four DNA nucleotides (T, A, C, G) in a cyclical fashion. All consecutive bases within a homopolymer run are incorporated within a single cycle, and the read-out is an intensity signal that is proportional with the number of incorporated bases (Margulies et al., 2005). The dominant error mode is base over- or under-call, resulting in INDEL type errors. While sequencing error rate only slightly increases with the number of flow cycles, the error rate increases dramatically with the frequency of long homopolymer runs. Accordingly, ART models the 454 sequencing error profile with homopolymer length-dependent over-call (insertion) and under-call (deletion) error distributions, and models base quality profiles as homopolymer length-dependent first-order Markov chains. ART uses an empirical distribution of 454 read lengths. By default, ART generates 454 reads with built-in distributions derived for the 454 GS FLX sequencer model.

2.3 SOLiD read simulation

Applied Biosystems’ SOLiD sequencing technology is based on ligation of oligonucleotides. It uses four fluorescent color dyes to encode the 16 different dinucleotides, each dye encoding four dinucleotides. SOLiD performs double interrogation of each base by combining the four-dye encoding scheme with a sequencing assay that samples every base (http://www.appliedbiosystems.com/absite/us/en/home/applications-technologies/solid-next-generation-sequencing.html). Different from either 454 or Illumina technology, the SOLiD base caller reports nucleotide transition color codes, rather than nucleotide sequences. Accordingly, ART also generates nucleotide transition codes or ‘color-space’ reads. For paired-end reads, a Gaussian distribution is used to model the distribution of DNA fragment sizes. The built-in empirical error profiles of SOLiD reads were derived from the read data generated at Applied Biosystems. ART provides an option to tune sequencing error rates with a linear scaling factor.

2.4 Performance

To test ART’s speed, we used human chromosome 17 as reference, and generated reads representing $10^8$ coverage for each of the three sequencing platforms. The test was performed on a desktop computer with Intel Xeon 2.93 GHz CPU, running a Linux operating system. This procedure took <12 min (Table 1), with Illumina reads being the fastest and SOLiD reads the slowest.

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REFERENCES


