piPipes: a set of pipelines for piRNA and transposon analysis via small RNA-seq, RNA-seq, degradome- and CAGE-seq, ChIP-seq and genomic DNA sequencing

Bo W. Han1,2,3,†, Wei Wang1,2,3,4,†, Phillip D. Zamore1,2,3,* and Zhiping Weng3,4,*

1RNA Therapeutics Institute, 2Howard Hughes Medical Institute, 3Department of Biochemistry & Molecular Pharmacology and 4Program in Bioinformatics and Integrative Biology, University of Massachusetts Medical School, 368 Plantation Street, Worcester, MA 01605, USA

Associate Editor: Ivo Hofacker

ABSTRACT
Motivation: PIWI-interacting RNAs (piRNAs), 23–36 nt small silencing RNAs, repress transposon expression in the metazoan germ line, thereby protecting the genome. Although high-throughput sequencing has made it possible to examine the genome and transcriptome at unprecedented resolution, extracting useful information from gigabytes of sequencing data still requires substantial computational skills. Additionally, researchers may analyze and interpret the same data differently, generating results that are difficult to reconcile. To address these issues, we developed a coordinated set of pipelines, ‘piPipes’, to analyze piRNA and transposon-derived RNAs from a variety of high-throughput sequencing libraries, including small RNA, RNA, degradome or 7-methyl guanosine cap analysis of gene expression (CAGE), chromatin immunoprecipitation (ChIP) and genomic DNA-seq. piPipes can also produce figures and tables suitable for publication. By facilitating data analysis, piPipes provides an opportunity to standardize computational methods in the piRNA field.

Supplementary information: Supplementary information, including flowcharts and example figures for each pipeline, are available at Bioinformatics online.

Availability and implementation: piPipes is implemented in Bash, C++, Python, Perl and R. piPipes is free, open-source software distributed under the GPLv3 license and is available at http://bowhan.github.io/piPipes/.

Contact: Phillip.Zamore@umassmed.edu or Zhiping.Weng@umassmed.edu

Supplementary information: Supplementary data are available at Bioinformatics online.

Received on April 25, 2014; revised on August 19, 2014; accepted on September 28, 2014

1 INTRODUCTION
piRNAs, a class of 23–36 nt long small silencing RNAs, suppress transposon expression in the metazoan germ line and, in some animals, the adjacent gonadal somatic cells (Luteijn and Ketting, 2013). By preventing transposition, the piRNA pathway ensures that genetic information passes faithfully to the next generation. Disruption of the piRNA pathway typically leads to transposon mobilization, double-stranded DNA breaks and sterility. High-throughput sequencing technologies have been widely deployed in the study of piRNAs. Small RNA-seq reveals the identity and abundance of piRNAs (Brennecke et al., 2007); RNA-seq detects and quantifies mRNA and transposon transcripts (Reuter et al., 2011); degradome-seq (also termed RACE-seq) detects the cleavage products of PIWI-proteins guided by piRNAs (Reuter et al., 2011); chromatin immunoprecipitation (ChIP)-seq detects chromatin modifications directed by piRNAs or transcription factor-binding events that regulate piRNA precursor or target transcription (Sienski et al., 2012); and genomic DNA sequencing detects new transposition events caused by transposons that escape piRNA repression or background differences between experimental strains and the assembled genome (Khurana et al., 2011; Sienski et al., 2012). Correctly extracting biological knowledge from such voluminous data requires significant computational expertise and effort. The repetitive nature of transposon sequences lays another layer of complexity. Moreover, different laboratories use diverse methods to analyze and interpret data (e.g. the way of treating reads that map to multiple locations in a reference genome). To provide a standardized set of tools to analyze these diverse data types, we developed piPipes, a collection of integrated pipelines for small RNA-seq, RNA-seq, degradome- and cap analysis of gene expression-seq (CAGE-seq), ChIP-seq and genome-seq analyses.

2 METHODS
piPipes comprises five pipelines designed to analyze small RNA-seq, RNA-seq, degradome- and CAGE-seq, ChIP-seq or genome-seq data. The small RNA-seq pipeline reports the abundance, length distribution, nucleotide composition and 5'-to-5' distance (‘Ping-Pong’ signature) of piRNAs assigned to genomic annotations, including individual transposon families and piRNA clusters, the initial sources of piRNA precursors. The RNA-seq pipeline reports the normal abundance of transscripts from both genes and transposons. The degradome-seq pipeline offers methods to identify piRNA-directed cleavage products. This pipeline can also be used to analyze any long RNA sequencing method designed to define RNA 5' ends, e.g. CAGE-seq. The ChIP-seq pipeline uses the widely used peak-calling algorithm MACS2 (Zhang et al., 2008), focusing on piRNA clusters and transposons. The genome-seq pipeline
Fig. 1. Gallery of piPipes Figures (A) Barplot representing length distribution of *Drosophila* w1 ovary small RNAs assigned to sense (blue) and antisense (red) strands of transposons. (B) Scatterplot comparing w1 to *auh*H^N2Q^C^4^ Drosophila ovary RNA-seq reads assigned to mRNA (NM; red), non-coding RNA (NR; green) and transposons (blue). (C) Metagene plot of H3K9me3 ChIP-seq of piRNA clusters from flies in which piwi mRNA was depleted by double-stranded RNA-triggered RNA driven by a triple Gal4 driver (SRX215630). (D) Circos plot representing the locations of, from the periphery to the center, cytological position, piRNA clusters, SV discovered by TEMP (tiles), retroSeq (tiles) and VariationHunter (links) using genomic sequencing of 2–4-day-old ovaries.
detects novel transposition events as well as structural variation. Supplementary Figure S1 illustrates the general piPipes workflow, using the small RNA-seq pipeline as an example. First, all reads aligned to ribosomal RNA (rRNA) sequences are removed. The remaining reads are then mapped to microRNA (miRNA) hairpin sequences to quantify the abundance and 5' and 3'-end heterogeneity of mature miRNAs. Reads that do not match rRNAs or miRNAs are then mapped to the reference genome. piPipes next assigns reads to different genomic features (e.g., transposons, piRNA clusters and genes) by their coordinates. To achieve maximal speed, piPipes parallelizes this step on multiple threads using ParaFly software from the Trinity package (Grabherr et al., 2011). For the reads assigned to each genomic feature, piPipes draws publication-quality graphs of length distribution and nucleotide composition, as well as the distance between the 5' ends of two small RNAs from opposite strands of the same locus, a standard method for detecting piRNA 'Ping-Pong' amplification or siRNA phasing (Fig. 1A and Supplementary Fig. S1B).

Furthermore, piPipes generates a table that summarizes the number of unique and multiple mappers counted as species (distinct sequences) or reads. The RNA-seq pipeline also starts with rRNA removal. The remaining reads are then mapped to the genome using STAR (Dobin et al., 2013). piPipes quantifies transcription abundance from genomic alignment by both Cufflinks (Trapnell et al., 2010) and HTSeq-count ( Anders et al., 2014). In addition, direct mapping of the reads to the transcriptome is performed using Bowtie2 followed by eXpress quantification (Roberts et al., 2014). piPipes next assigns reads to different genomic features using ParaFly software from the Trinity package (Grabherr et al., 2011). piPipes calls peaks using MACS2 (Zhang et al., 2010) and TEMP (Zhuang et al., 2014), to discover transposon insertion, element polymorphism in populations. piPipes also performs differential analysis on genomic transcripts. In the dual-sample mode, the ChiP-seq pipeline uses MACS2 to identify differentially enriched loci (Supplementary Fig. S4).

ACKNOWLEDGEMENTS

The authors thank the members of the Zamore and Weng laboratories for helpful discussions, and Jia Xu, Jui-Hung Hung and Soo Lee for their initial work.

Funding: This work was funded by National Institutes of Health grants [GM62862 and GM65236] to P.D.Z. and [U41HG007000] to Z.W.

Conflicts of interest: none declared.

REFERENCES


Zhang,H. et al. (2013) RCircos: an R package for Circos 2D track plots. BMC Bioinformatics, 14, 244.
