Graphical algorithm for integration of genetic and biological data: proof of principle using psoriasis as a model

Lam C. Tsoi1,*, James T. Elder2,3 and Goncalo R. Abecasis1,*

1Department of Biostatistics, Center for Statistical Genetics, University of Michigan, Ann Arbor, MI, USA, 2Department of Dermatology, University of Michigan, Ann Arbor, MI, USA, and 3Ann Arbor Veterans Affairs Hospital, Ann Arbor, MI, USA

*To whom correspondence should be addressed.
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Abstract

Motivation: Pathway analysis to reveal biological mechanisms for results from genetic association studies have great potential to better understand complex traits with major human disease impact. However, current approaches have not been optimized to maximize statistical power to identify enriched functions/pathways, especially when the genetic data derives from studies using platforms (e.g. Immunochip and Metabochip) customized to have pre-selected markers from previously identified top-rank loci. We present here a novel approach, called Minimum distance-based Enrichment Analysis for Genetic Association (MEAGA), with the potential to address both of these important concerns.

Results: MEAGA performs enrichment analysis using graphical algorithms to identify sub-graphs among genes and measure their closeness in interaction database. It also incorporates a statistic summarizing the numbers and total distances of the sub-graphs, depicting the overlap between observed genetic signals and defined function/pathway gene-sets. MEAGA uses sampling technique to approximate empirical and multiple testing-corrected \( P \)-values. We show in simulation studies that MEAGA is more powerful compared to count-based strategies in identifying disease-associated functions/pathways, and the increase in power is influenced by the shortest distances among associated genes in the interactome. We applied MEAGA to the results of a meta-analysis of psoriasis using Immunochip datasets, and showed that associated genes are significantly enriched in immune-related functions and closer with each other in the protein–protein interaction network.

Availability and implementation: http://genome.sph.umich.edu/wiki/MEAGA
Contact: tsoi.teen@gmail.com or goncalo@umich.edu

Supplementary information: Supplementary data are available at Bioinformatics online.

1 Introduction

In the past decade, association studies have been used to investigate genetic architecture of complex traits and diseases (McCarth et al. 2008). Studies ranging from genotyping-based genome-wide association studies (GWAS; WTCCC 2007; Yang et al. 2010) to large-scale meta-analyses of GWAS (Liu et al. 2013; Teslovich et al. 2010), to the more recent next-generation sequencing-based association analysis (Tang et al. 2013; Zhan et al. 2013), have facilitated the identification of genetic susceptibility loci for different traits and diseases. However, identification of the underlying biological mechanisms from these powerful datasets has not always been straightforward.

To better understand these underlying biological mechanisms, it has been asked whether the identified loci contain genes enriched in
certain gene-sets such as genes annotated with biological functions [e.g. Gene Ontology: GO (Ashburner et al. 2000)] or pathways [e.g. KEGG (Kanehisa et al. 2012)]. It would also be possible to investigate if their encoding proteins could physically interact to form functional units such as heterodimeric cytokines (e.g. IL23A and IL12B as sub-units of cytokine IL23) or transcription factor complexes (e.g. p53). While advances in genotyping technology have greatly increased the power of available datasets, the development of optimal strategies to bridge between genetic and functional results remains a major theoretical and experimental goal of genetics research.

Depending on the genotyping platform being used, loci densities and genomic distribution of the markers in the association analysis might vary. One of the challenges in enrichment testing is to select the proper set of background markers/genes as “null” observations to approximate its overlap against the gene-set in question. Previous studies have proposed different strategies to approximate the null observations: ALIGATOR (Holmans et al. 2009) randomly samples markers from the original platform to mimic its empirical marker distribution; INRICH (Lee et al. 2011) exhibits a sampling strategy using genomic interval as unit, and the sampled intervals are constrained to have similar marker and gene densities as those from the input intervals (i.e. susceptibility loci). Each of the sampled null observations from the above approaches would be used to construct empirical distributions for the numbers of overlap with functional gene-sets, and used to estimate the statistical significance of the observed data.

Both of the above count-based approaches enumerate the number of associated genes overlapping with each gene-set and work well (Holmans et al. 2009; Sklar et al. 2011) in identifying biological functions or pathways if the platform was designed in genome-wide scale (i.e. most common genetic markers are well-tagged by the genotyped markers). Recently, cost-effective genotyping platforms, such as the Immunochip (Cortes and Brown 2011; Parkes et al. 2013) and MetaboChip (Voight et al. 2012), containing common and low-frequency variants have been designed to facilitate the identification of novel association signals (Beecham et al. 2013; Ellinghaus et al. 2013; Tsoi et al. 2012) and with the goal to fine-map established loci (Gong et al. 2013; Voight et al. 2012). These platforms contain dense markers in the top-ranked loci from previous studies (e.g. studies in autoimmune diseases for Immunochip; studies in metabolic and atherosclerotic/cardiovascular traits/diseases for MetaboChip). Performing functional enrichment analysis on the signals identified using these customized platforms poses analytical challenges, as the pre-selected markers are enriched in potential or known susceptibility loci. The potential to identify the correct underlying biological mechanisms/pathways can be revealed only if sufficient gene-sets-overlapping genes from the significant loci are identified to provide enough statistical power. Moreover, as different studies have identified interactions between genes in disease susceptibility loci (Cotsapas et al. 2011; Rossin et al. 2011), a gene-set enrichment approach that incorporates independent information from biological interaction data could thus increase the power to identify functions or pathways crucial to the disease.

We present here a novel approach, called Minimum distance-based Enrichment Analysis for Genetic Association (MEAGA), to perform functional/pathway enrichment test for results from association studies. Instead of only enumerating the overlap between genes in the associated regions and genes annotated in a gene-set (i.e. count-based; Holmans et al. 2009; Lee et al. 2011), we use graphical algorithm and incorporates a statistic to measure the closeness between the overlapping genes in Steiner Tree(s) identified in a user-defined interaction database. Here we demonstrate in simulation studies that MEAGA is more powerful than a count-based approach when protein-coding genes in associated regions tend to be closer with each other in an interaction network. We also applied MEAGA to the results of a meta-analysis of psoriasis, a common inflammatory and hyperproliferative skin disease, using Immunochip datasets (Tsoi et al. 2012). Although the original study did not identify any enriched biological functions among the identified loci, MEAGA is able to reveal associated genes are closer with each other in the protein–protein interaction (PPI) network and are also significantly enriched in immune-related functions.

2 Methods

2.1 Overview

MEAGA tests the hypothesis that genes from the susceptibility loci in the trait/disease-associated function/pathway are closer with each other in the biological interactome than any other genes. The overview of MEAGA is illustrated in Fig. 1. MEAGA takes the markers used in the association analysis as input. Users will pre-specify the

Fig. 1. The workflow overview of MEAGA. MEAGA performs enrichment analysis on association results using gene-sets obtained from functional/pathway databases. For each gene-set being tested, MEAGA identifies overlapping genes in association signals, and constructs Steiner Tree(s) in the biological interactome (e.g. PPI database) using Kou’s algorithm. Kou’s algorithm requires the shortest distances and paths between every gene-pair being computed among the overlapping genes. As illustrated in the bottom right figure, we could use genes present only in the gene-set (red lines in the bottom-right figure) or all genes (black lines) to compute shortest paths in the interactome. MEAGA then constructs Steiner trees and computes statistic $S_T$. MEAGA uses sampling technique to estimate empirical and multiple-testing corrected P-values.
association signals (presumably the most significant marker in each of the significant loci) and annotate the tagged genes for each marker (e.g. using linkage disequilibrium blocks or genomic distance). For each functional gene-set being tested, MEAGA first identifies the overlapping genes from the signals, then uses graphical algorithms to construct sub-graph(s) with minimum distance(s) in the interactome. MEAGA next computes a statistic ($S_T$) summarizing the number of overlapping genes and the overall shortest distance(s) of the sub-graph(s). MEAGA uses a sampling strategy to approximate the null distribution of $S$ and compute empirical and multiple testing-corrected $P$-values. There is no restriction for the type of interaction data [e.g. co-expression network (Langfelder and Horvath 2008); text-mining-derived networks (Franceschini et al. 2013); PPI network (Chatr-Aryamontri et al. 2013)] being used in MEAGA. Throughout this study, we used PPI data as this provides the experimentally verified information and it is well-known that proteins often work together for many biological functions (Cotsapas et al. 2011).

2.2 Data

We obtained the functional and pathway annotation data from the GO (Ashburner et al. 2000), KEGG (Kanehisa et al. 2012) and Reactome (Croft et al. 2013) databases (latest versions downloaded on May 7, 2013). Since the GO database has a directed acyclic graph (DAG) layout, in which each term is defined by one or more less-specific terms in the same domain, we processed the GO’s gene-to-GO file so we also annotated each gene with the ‘ancestral’ terms of its annotations in the DAG of GO database. Altogether, we obtained over 1.5 million gene to function/pathway connections from 18,987 genes and 19,383 functions/pathways. We downloaded and examined PPI data from three different sources: BioGrid (Chatr-Aryamontri et al. 2013), HPRD (Keshava Prasad et al. 2009) and STRING (Franceschini et al. 2013) (Supplementary Materials).

2.3 MEAGA

2.3.1 Sub-graphs with minimum distances

We first denote $G'$ as the overlapping genes between a gene-set and genes in the susceptibility loci ($G$). By assuming genes involved in similar biological mechanisms associated with the traits or diseases would tend to be closer with each other in the interaction network (see Section 3), we use graphical distance as metric. Specifically, from the interactome network, we identify Steiner trees ($T'$) among genes in $G'$; each Steiner tree is a connected and undirected acyclic graph with minimum total length. Note that a Steiner tree would allow other genes in the interactome to act as intermediate nodes when there is no direct connections between genes in $G'$. Therefore, if $G'$ contains all the genes in the interactome Steiner tree problem would become the well-studied minimum spanning tree problem (Kruskal 1956; Prim 1957). We use the heuristic Kou’s algorithm (Kou et al. 1981), which has been applied to other biological problems using Steiner tree technique (Richards et al. 2010; Zheng and Lu 2007), to identify $T'$.

2.3.2 Algorithm

MEAGA uses Kou’s algorithm to identify Steiner tree(s) among genes in $G'$ in the interactome for each of the function/pathway being tested. It is possible that not all genes in $G'$ are connected in the interactome, so there could be more than one Steiner tree identified. Also, any gene in $G'$ not present in the interactome would be treated as an isolated Steiner tree. We developed a statistic which summarizes the number of genes in $G'$ ($|G'|$), the number of identified Steiner trees ($|T'|$), and the distances for all $T'$:

$$S_T = \frac{|G'|}{|T'|} \sum_{i} m_i w_i$$

where $m_i$ is the number of genes from $G'$ in Steiner tree $i$, and $w_i$ is the weight inversely proportional to the total distance ($d_i$) in tree $i$. Two Steiner trees could have the same $m$ but different total distances if there are different numbers of intermediate nodes in the interactome coming from genes not in $G'$; similarly, two Steiner trees could have same total distance but contain different $m$ values. Therefore, the shortest possible distance for a tree with size $m$ could be used as a reference to normalize $d$. For Steiner tree with more than one gene, we set $w_i = a_i/d_{0i}$, where $a_i$ is the shortest possible distance for tree with size $m_i$, and for genes in $G'$ isolated in the interactome, we set $w_i = 1/D_{max}$, where $D_{max}$ is a penalty score that equals the most distant length among all the pairwise shortest paths between genes in the interactome. Note that $\sum_{i} m_i |T'|$ in $S_T$ is the average number of genes in $G'$ among $T'$, and $w_i$ is a $m_i$-depending weight. If all genes in $G'$ are isolated in the interactome, $S_T$ would equal $|G'|/D_{max}$, which is proportional to $|G'|$. Therefore, count-based functional enrichment approach is a special case of MEAGA if the interactome does not provide any information among genes in $G'$ (see Section 3 from simulation studies).

Variables such as gene length, linkage disequilibrium structure, and minor allele frequency of the variant could affect the results of the functional enrichment test. To compute the empirical $P$-values for each gene set effectively, we use a sampling strategy adopted from previous study (Holmans et al. 2009). Briefly, MEAGA first randomly samples markers used in the association analysis until the number of sampled genes equal $|G|$ (user would be able to define the marker-to-genе region assignment, see below). It then applies the algorithm described above to identify gene-set overlapping genes among the sampled genes, and computes the $S$ statistic using the identified Steiner tree(s). MEAGA repeats the sampling procedure $N$ times to construct null distribution of $S$, and the empirical $P$-value is computed as the proportion of samples in $N$ with statistic $S$ as large as the $S_T$ of the observed data. Multiple testing is then performed by sampling from the $N$ samples a random gene set as ‘observed data’, and then computes its empirical $P$-value among all tested function/pathway gene-sets based on comparing its statistic with those obtained from bootstrapping the $N$ samples. This procedure is performed $M$ times, and the multiple testing-corrected $p$-value is computed as the proportion of the $M$ samples with minimum empirical $p$-value (across the function/pathway gene sets) less than or equal to that from the observed data.

2.3.3 Implementation

We implemented MEAGA in Python, and used the graphical features available in the package NetworkX (Hagberg et al. 2008). Statistical computations were implemented in R (http://www.R-project.org). The construction of Steiner trees and the computation of the statistic $S$ for each sampling step could be time consuming, and MEAGA supports multiprocessing for these procedures using the multiprocessing module in Python. The Kou’s algorithm requires the shortest distances and their paths to be first computed between all $G'$. For effective performance, we pre-computed all shortest paths between genes in each function/pathway gene-set and stored them in a database to be readily retrieved when performing the Kou’s algorithm. Figure 1 demonstrates that we could use genes present only in
2.4 Simulation and application to association results

To evaluate the performance of MEAGA, and to compare the results with the count-based approach, we performed simulation to estimate the statistical power evaluated under the same type I error rate for each approach. We simulated null functions/pathways by selecting genes via randomly drawing markers successively from the set of all markers used in a real study (see below). The true associated functions/pathways were simulated similarly, except some of the genes were selected based on sampling the markers from the trait-susceptibility loci. We simulated shortest distances between the genes in the interactome, and varied the differences in distances for genes from the associated loci versus genes that are not. Statistical power is defined by the proportion of true associated functions identified to be significant. We used the 112 243 markers from a meta-analysis for psoriasis (2 Immunochip and 3 GWAS datasets; totally 10 388 psoriasis and 22 806 control samples) in the simulation to mimic real data (Tsoi et al. 2012).

Finally, we used MEAGA to perform functional/pathway enrichment test for the best single nucleotide polymorphisms identified in the 39 psoriasis independent loci. We used an interval of ± 100 kb to define the tagging genes for each marker (Tsoi et al. 2012). Since functionally related genes could be from the same locus, multiple-counting this overlap would give rise to false positive results (Holmans et al. 2009; Lee et al. 2011). Therefore, we restricted our analysis to functions/pathways with genes in G’ all coming from different loci.

3 Results

3.1 Simulation results

Figure 2 illustrates the results of the simulation studies under different approaches (count-based versus MEAGA), simulation settings (i.e. different numbers of total functions, trait associated functions, and genes in the functions), and weighting schemes used. We varied the distances of the Steiner trees identified from genes in association signals, using those obtained from randomly sampled genes as reference. As described above, the distance of a Steiner tree is inversely proportional to its weight wj; the magnification scale ‘x’ in Figure 2 is the relative weight set for associated genes referencing the non-associated genes in the platform. For example, ‘2x’ in Figure 2 represents the distances for Steiner trees from randomly selected genes in the platform are set to be two times those than those for associated genes.

Our results show that when the distances from association signals are relatively closer with each other in the interactome (i.e. 1.3x to 4x), MEAGA could obtain higher statistical power in identifying the true associated functions when comparing with count-based approach. The gain in statistical power increases when the relative weight of the Steiner trees increases for the associated genes. If the interactome does not provide any information (i.e. 1x), the count-based approach would become a special case for the MEAGA algorithm, and we illustrated in Figure 2 (black and grey lines) that the count-based approach has identical statistical power with MEAGA under this situation. We also simulated a ‘counter intuitive’ situation where the associated genes are further away with each other in the interactome when comparing with the non-associated genes (0.8x in Fig. 2). The results show MEAGA only loses a small proportion of its power (purple in Fig. 2) when compared with the count-based approach. Section 3.2.1 illustrates that in reality the associated genes tend to be closer with each other in the interactome.

As expected, an increase in the number of gene-set overlapping genes (i.e. |G’| in the x-axis of Fig. 2) would increase the statistical power in the identification of the true functions for all the scenarios being tested. The results also suggest the total number of genes in the functions (top and middle panels in Fig. 2) and the number of functions being tested (top and bottom panels in Fig. 2) both have effects on the statistical power. For example, when performing enrichment test for 500 small-sized (~20 genes) functions/pathways (top panel in Fig. 2), we would gain the maximum power using different approaches if we have 10 overlapping genes from the association loci; however, testing for medium-sized (~50 genes) functions/pathways (middle panel in Fig. 2) would only have less than 20% statistical power when using the count-based approach or if the interactome does not provide any information for associated genes, but we could achieve at least 80% statistical power if the total distances of Steiner trees from associated genes in the interactome are at least twofold shorter than those from non-associated genes. We also tested the performance of MEAGA when the gene interactions from the network are permuted to different degrees (Supplementary Fig. S1). While the results illustrate that the statistical power of accurately identifying associated functions/pathways decreases when the network is permuted to a greater degree, MEAGA would still perform as well as the count-based approach for a complete random network. In other words, when the interactome becomes...
3.2 Application to meta-analysis results
3.2.1 Gene–gene connections in the interactome
Before performing functional enrichment testing on the meta-analysis results of psoriasis (Tsai et al. 2012), we evaluated the connection for genes identified in the associated loci. We sampled 10 000 random gene-sets from markers used in the meta-analysis, and calculated the proportion of gene pairs that are connected in the interactome as assessed by BioGrid (Fig. 3 upper panel). We found that this proportion for genes identified in the psoriasis susceptibility loci is among the top 5.5% (vertical line) of those from the randomly sampled gene-sets. We then normalized the proportion values by the total distances of the Steiner trees, and the normalized value of genes from the association loci is among the top 2.6% of the randomly sampled gene-sets (lower panel vertical line). In concordance with previous studies (Cotsapas et al. 2011; Rossin et al. 2011) suggesting that proteins encoded by genes in association loci identified in autoimmune diseases tend to be physically interact (via direct interaction or 2° of separation), our results show the first time that using the network topology (in terms of the distances of Steiner trees) of the interactome could enhance the resolution for identifying closely interacting genes from the associated loci in an autoimmune disease. The results further justify the assumption used in MEAGA of using information from the interactome (PPI in this case) for enrichment analysis to identify underlying biological functions/pathways. Performing the above analyses on PPI obtained from HPRD and STRING yielded very similar results (Supplementary Fig. S2).

3.2.2 MEAGA highlights immune-related functions/pathways for psoriasis
To perform enrichment testing on the best signals identified in the meta-analysis, we restricted our analysis to functions/pathways with at least 5 and at most 500 genes, and functions/pathways overlapping with at least two genes from the association signals. After limiting to the overlapping genes (\(G^7\)) coming only from different loci, we tested 743 functions/pathways using MEAGA. We first evaluated the impact of using different interaction data in the interactome on the results. The empirical \(P\)-values computed using only genes present in functions/pathways (x-axis) versus using all genes (y-axis) in the interactome are positively correlated (Spearman correlation = 0.88), and using only genes present in the functions/pathways tend to give stronger signals in the enrichment analysis (Supplementary Fig. S3). On further investigation, we realized that using all genes in the interactome to construct Steiner trees would tend to be less informative because the human PPI network is suggested to behave like a scale-free network (Barabasi and Oltvai 2004), with hub proteins lessening the average distances from one protein to the other. Therefore, using all interactions could introduce noise and thus MEAGA would behave like the count-based approach (Spearman correlation = 0.94).

Using the genes present only in functions/pathways to construct the Steiner trees in the interactome specified by BioGrid, we identified 28 significantly enriched functions/pathways using the \(P_{\text{adj}} \leq 0.1\) criteria. Seven of these were identified as significant using ALIGATOR, whereas INRICH identified only one significant function (Supplementary Materials and Table S1). The top five results are shown in Table 1 (results from MEAGA using other PPI sources are illustrated in Supplementary Table S2). Sensitivity analysis using different parameters such as minimum/maximum number of genes allowed in the functions/pathways still yielded similar results. Immune-related functions such as ‘regulation of leukocyte mediated cytotoxicity’, ‘positive regulation of lymphocyte differentiation’ and ‘regulation of myeloid cell differentiation’ are significantly enriched among the genes from the psoriasis susceptibility loci. MEAGA also successfully reveals those biological functions (e.g. ‘regulation of response to biotic stimulus’) which contain three important genes forming protein complexes and involved in the IL-23 signaling pathways (i.e. IL23A, IL23R, IL12B) (Nair et al. 2009). Figure 4 demonstrates the Steiner trees identified in one of the enriched functions. While previous studies have suggested that TRAF3BP2 is one of the best candidate genes in the psoriasis susceptibility locus 6q21 (Ellinghaus et al. 2010), MEAGA shows that FYN from the same locus is annotated in one of the enriched functions. In close proximity with genes from other susceptibility loci in the interactome. In fact, a recent study suggests FYN might be involved in the immunity in psoriasis, and could be a potential drug candidate (Manzinger and Kemeny 2013). Our results illustrate that MEAGA not only could identify enriched functions and interactions among associated loci, but also could prioritize the genes in the loci based on the functional and interaction data.

We also tested MEAGA for all the psoriasis susceptibility loci using markers from a GWAS study as background (Nair et al. 2009), and identified 95 significant functions highlighting different specialized immune-related pathways among the genes in the disease loci (Supplementary Table S3). We acknowledge that this analysis could be biased as the markers in the Immunochip datasets tend to be related to immune mechanisms, but it provides an overview of the biological functions or pathways that the genes from the psoriasis loci involved, when comparing with the enrichment in the whole genome in general.

4 Discussion
We present a novel functional enrichment method called MEAGA in this study. MEAGA uses graphical algorithm to identify significant
of genes annotated in the function. Full results shown in the Supplementary Materials. We used PPI obtained from BioGrid in this analysis.

sential for making valid and novel biological inferences (Chatr-

immune-related functions potentially involved in the pathogenesis

we also applied MEAGA to the results of a meta-analysis to identify

have high coherency in the interactome. We showed in the simula-

biological functions containing genes from susceptibility loci which

Fig. 4. The networks represent the Steiner trees identified in the enriched

function ‘regulation of response to biotic stimulus’. The edges are the PPI,

and the nodes are the protein-coding genes in the function. Genes in dark

grey colour are from the psoriasis susceptibility loci

Table 1. Top 5 significant enriched functions identified by MEAGA for genes in psoriasis susceptibility loci

| Significant functions | |G'| |T'| P   | P_{adj} | G'          |
|-----------------------|---------|------------|------|-------|------------|
| Regulation of response to biotic stimulus (87) | 7       | 2          | 5.0 \times 10^{-5} | 2.6 \times 10^{-2} | DDX58, ELMO1, FYN, IL12B, IL23A, IL23R, TNAIP3 |
| Regulation of transcription factor import into nucleus (67) | 4       | 2          | 1.0 \times 10^{-4} | 4.0 \times 10^{-2} | DDX58, IL12B, IL23A, NFkBIA |
| Regulation of leukocyte-mediated cytotoxicity (33) | 6       | 3          | 1.5 \times 10^{-4} | 5.5 \times 10^{-2} | ICAM1, IL12B, IL23A, IL23R, NOS2, STAT5A |
| Regulation of cell killing (36) | 6       | 3          | 1.5 \times 10^{-4} | 5.5 \times 10^{-2} | ICAM1, IL12B, IL23A, IL23R, NOS2, STAT5A |
| Positive regulation of lymphocyte differentiation (65) | 5       | 2          | 2.0 \times 10^{-4} | 7.0 \times 10^{-2} | IL12B, IL23A, IL23R, IL4, STAT5A |

For illustration purpose, we restricted the significant results to those functions/pathways with at most 100 genes. Numbers in the bracket represent the number of genes annotated in the function. Full results shown in the Supplementary Materials. We used PPI obtained from BioGrid in this analysis.

above when using linkage disequilibrium-based intervals to define gene region for each associated marker (results not shown).

We have used graph theory techniques in this study, and we show that the systems biology approach could facilitate the understanding of biology for complex disease. Biological systems possess network properties and often function together, and there have already been several interesting published studies using network approaches to model biological units using genome-scale data such as gene expression or somatic mutation data (Alcaraz et al. 2014; Hofree et al. 2013; Ulitsky et al. 2010; Vandin et al. 2011). We believe advanced and efficient tools for providing biological inference of genetic data for complex traits could be based on these methodological foundations, and we have provided Supplementary Table S4 to illustrate how existing network algorithms which were applied to other genome-scale data could be useful for future studies of complex traits.

With the recent advancements in the technological and methodological aspects for using next-generation sequencing to perform genetic association tests, we envision that MEAGA may prove valuable in providing biological inferences for these studies. This is particularly so for cost-effective targeted sequencing-based studies (Tang et al. 2013; Zhan et al. 2013) when the investigated regions are pre-selected to have strong/suggestive association, integrating independent information would enhance the statistical power in identifying the underlying biological mechanisms of the trait/disease being studied.

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References

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