

p53 Antibodies in the Sera of Patients with Various Types of Cancer: A Review¹

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Abstract

p53 antibodies (p53-Abs) were discovered 20 years ago during the course of tumor-associated antigens screening. The discovery of p53 mutation and accumulation of p53 in human tumors shed new light on the p53 humoral response. In the present review, we have compiled more than 130 papers published in this specific field since 1992. We demonstrate that p53-Abs are found predominantly in human cancer patients with a specificity of 96%. Such antibodies are predominantly associated with p53 gene missense mutations and p53 accumulation in the tumor, but the sensitivity of such detection is only 30%. It has been demonstrated that this immune response is due to a self-immunization process linked to the strong immunogenicity of the p53 protein. The clinical value of these antibodies remains subject to debate, but consistent results have been observed in breast, colon, oral, and gastric cancers, in which they have been associated with high-grade tumors and poor survival. The finding of p53-Abs in the sera of individuals who are at high risk of cancer, such as exposed workers or heavy smokers, indicates that they have promising potential in the early detection of cancer.

Introduction

The discovery of the p53 protein was the culmination of two types of studies: (a) the very well known virological approach, with the binding of p53 protein to oncoviral proteins (1–5); and (b) the more discrete serological approach with the study of TAAs³ (6, 7). In 1979, DeLeo *et al.* (7) showed that the humoral response of mice to some methylcholanthrene-induced tumor cells such as MethA was directed to the p53 protein. This protein was found to accumulate in tumor cells of different origin but was undetectable in normal cells. It was also found that animals bearing SV40 tumors elicited an immune response specific for p53 (1, 5, 6). In 1982, Crawford *et al.* (8) first described antibodies against human p53 protein in 9% of breast cancer patient sera. No significant clinical correlation was reported, and, at that time, no information was available concerning mutations of the p53 gene. Caron de Fromentel *et al.* (8) later found that such antibodies were present in sera of children with a wide variety of cancers. The average frequency was 12%, but this figure increased to 20% in Burkitt's lymphoma.

Those studies, performed in the early 1980s, were virtually ignored for more than 10 years because of a lack of interest in p53 during that period. In the early 1990s, it was discovered that the p53 gene is the most common target for molecular alteration in every type of human cancer (10). This provoked considerable interest in the study of the p53 protein and its function in normal and transformed cells. It also

led to the rediscovery of this humoral response, which had been found in cancer patients. Since 1992, more than 150 papers have been published on p53-Abs. This review will summarize this literature and focus on the future applications of this assay.

For a comprehensive view of the p53 discovery, the reader is referred to the excellent review published by Crawford (11). More reviews on p53 have recently been published (12–15).

The p53 Protein

The tumor suppressor p53 is a phosphoprotein barely detectable in the nucleus of normal cells (16). On cellular stress, particularly that induced by DNA damage, p53 can arrest cell cycle progression, thus, allowing the DNA to be repaired (17) or it can lead to apoptosis (18). These functions are achieved, in part, by the transactivational properties of p53, which activate a series of genes involved in cell cycle regulation. In cancer cells that bear a mutant p53, this protein is no longer able to control cell proliferation, which results in inefficient DNA repair and the emergence of genetically unstable cells (12–15). The most common changes of p53 in human cancers are point missense mutations within the coding sequences of the gene (10, 19). Such mutations are found in all of the major histogenetic groups, including cancers of the colon, stomach, breast, lung, brain, and esophagus (20). It is estimated that p53 mutations is the most frequent genetic event in human cancers and accounts for more than 50% of cases. More than 90% of the point mutations reported thus far are clustered between exons 4 and 10 and are localized in the DNA binding domain of the p53 protein (21). One of the most striking features of the inactive mutant p53 protein is its increased stability (half-life of several hours compared with 20 min for wild-type p53) and its accumulation in the nucleus of neoplastic cells. Positive immunostaining is usually indicative of abnormalities of the p53 gene and its product, but it is highly dependent on the type of p53 mutation for review (22, 23).

The Specificity of p53 Antibodies in Cancer Patients

Dosage of p53 Antibodies. The initial work on p53-Abs used either immunoprecipitation or Western blot as the detection method (8, 9). Later, several ELISAs were developed to handle large numbers of specimens (24, 25), and some of them are now commercially available. The diversity of all of these assays could account for the variation in the frequency of p53-Abs observed in the literature. One of the most important parameters seems to be the antigen used for these assays. It has been shown that p53-Ab recognized immunodominant epitopes localized in the NH₂ and COOH termini of the protein (see below). It is thus essential to use the entire p53 protein as antigen. Several attempts to develop an ELISA with synthetic peptides corresponding to these immunodominant epitopes have been unsuccessful, because they lead to a high level of false-negative results (26).

p53 is heavily phosphorylated at the NH₂ and COOH termini. Such phosphorylation can have an important influence on the reactivity of p53-Abs toward the protein, which suggests that p53 expressed in mammalian cell is a better antigen than those expressed in *Escherichia coli*. Recently, it has been shown that IgA p53-Abs are found

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³ The abbreviations used are: p53-Ab, p53 antibody; TAA, tumor-associated antigen; LSH, loop-sheet-helix (motif); SCLC, small cell lung cancer; NSCLC, non-SCLC; ADC, adenocarcinoma.

as the only isotype in some patients with head and neck cancers. The fact that a majority of ELISAs use a secondary antibody specific for IgG may eventually lead to false-negative sera.⁴

p53-Abs have also been found in other body fluids such as the ascites of women with ovarian cancer, the pleural effusions of patients with pancreatic, colon, and lung tumors, and the saliva of patients with oral cancer (27–29). Such antibodies are correlated with serum p53-Abs (27, 28).

p53 Antibodies and p53 Mutation Frequency. Eighty serological analyses of p53 status in 18 cancer types that were published before August 1999 have been compiled (Table 1). Control experiments were performed in 36 studies, involving 2404 individuals who were either healthy or had nonmalignant diseases. Studies involving 9489 patients with various types of cancer were analyzed. Statistical analysis demonstrated that p53 antibodies are a specific marker of patients with neoplasia ($P < 10^{-4}$; Table 2). An analysis of p53-Abs in each type of cancer also showed a significant correlation with malignant diseases for most types of cancer except for testicular carcinoma, melanoma, hepatoma, and glioma, in which the frequency of p53-Abs was very similar to that of the control group (Table 2). Fig. 1 shows the correlation between the frequency of p53-Abs and the frequency of p53 mutations published in the literature. There is a very strong correlation between the two rates, again arguing that p53 mutations are involved in the appearance of these antibodies. Apart from the healthy control, it is striking to observe that three cancers, well-known to be devoid of p53 mutations, hepatoma (30), testicular carcinoma (31, 32), and melanoma (33, 34), are also negative for p53-Abs. However, cancers such as esophageal carcinoma and oral squamous cell carcinoma, which have a high rate of p53 mutations, also have a high frequency of p53-Abs. The only exception is glioma, with a very low rate of p53-Abs (35, 36) despite a high frequency of p53 mutations (37). In a complete study involving both molecular and serological analyses, Rainov *et al.* (35) found p53 mutations in 24 of 60 glioblastomas, but none of these patients had p53-Abs. Several explanations can account for the lack of a p53 immune response in this type of cancer. It is possible that the immune privilege of the brain does not allow the induction of this humoral response. It is also possible that there is inefficient antigen presentation if p53 cannot cross the brain barrier, or if no immune response can be elicited in the brain. Furthermore, we should keep in mind that, in one series, 70% of the patients were treated with dexamethasone before serum collection (35), which can induce perturbation in the immune response. In the other studies, such information was not available.

Relationship between p53 Antibodies, p53 Mutations, and p53 Accumulation

There is generally a very good correlation between the presence of p53-Abs and p53 accumulation and/or mutation in the tumor. Winter *et al.* (38) have shown that only missense mutations can lead to a p53 humoral response. As shown in Table 1, this finding was confirmed by numerous studies.

Only a subset of patients with p53 mutations will have p53 antibodies in their tumor. As shown in Fig. 1, about 20–40% of patients with a p53 mutation will have p53-Abs in their sera. These results are not due to a lack of sensitivity of the current methods of detection but to a real absence of p53-Abs. It had been suggested that only p53 mutations that are localized in exons 5 and 6 with an altered protein conformation and that bind to hsp 70 are associated with p53-Abs (39, 40). These analyses, performed on a small number of patients, were

not confirmed in larger series of patients. Indeed, a compilation of all of the serological analyses performed in conjunction with molecular analyses indicates that the repartition of p53 mutations in patients with p53-Abs is similar to that in patients without antibodies (Fig. 2). Several studies have shown that, despite similar types of cancer, identical p53 mutation, and p53 accumulation, some patients could be either positive or negative for p53-Abs (41–44). These observations demonstrate that other factors contribute to this humoral response. Examination of p53-Abs during follow-up of patients during therapy indicates that the level of p53-Abs can vary depending on the tumor burden (see below). Follow-up of patients who were devoid of p53-Abs at the time of diagnosis failed to detect any *de novo* antibody production after therapy failure or relapse despite p53 accumulation in the tumor and a long follow-up. This suggests that the capacity to elicit this humoral response is linked to the biological background of the patients. It is possible that, for an identical mutation, the immune response is dependent on the specific combination of MHC class I and II molecules expressed by each individual.

There may be several explanations for the presence of p53-Abs in tumors with a wild-type p53. Technical failure cannot be neglected and is difficult to assess, nor can we exclude the possibility that the tumor is composed of heterogeneous tissue and that the fragment analyzed does not bear p53 alteration. We should also keep in mind that serological analysis is a global assay that does not depend on sampling. An undetectable metastasis with p53 alteration might be associated with a p53-negative primary tumor. As discussed below, the mechanism that leads to the formation of these antibodies is poorly understood. Accumulation may be an important component in the development of this immune response, but we cannot exclude the fact that modified processing of the mutated protein can also lead to such a response.

Specificity of p53 Antibodies

As stated above, mutant p53 accumulation is an important component of this humoral response. Thus, it is possible that such antibodies are specific to mutant p53 or at least to a mutant p53 conformation. Several works have shown that these antibodies recognize both wild-type and mutant p53 (38, 45, 46). Using a set of overlapping synthetic peptides corresponding to human p53, precise mapping of the epitopes of p53 protein recognized by p53-Abs was performed (47). This method is very powerful and very rapid for the analysis of a large number of sample. Nevertheless, we should bear in mind that it only works with antibodies that recognize small linear epitopes. Analysis of more than 200 sera from various types of cancer unambiguously demonstrated that these antibodies recognize immunodominant epitopes localized in the NH₂ terminus and, to a lesser extent, in the COOH terminus of human p53 (26, 47, 48). Only a few antibodies recognize the central region of the p53 protein that harbors the mutations. We know that the identification of such epitopes is not due to a technical bias based on the use of short synthetic peptides because immunoprecipitation and immunoblot analysis of truncated p53 have led to the same observation (45, 48).

Such a finding is totally in accordance with the work performed in mice on the production of p53 monoclonal antibodies. Immunization of mice with murine, xenopus, or human wild-type p53 led to the production of monoclonal antibodies directed to linear epitopes localized in the NH₂ and COOH termini of p53 (49–52). The analysis of mouse sera immunized with these proteins indicates that it is due to a specific immune response of the mouse toward this region of the protein and not to a bias in the selection of the hybridoma (49). Monoclonal antibodies specific for the central region of the protein

⁴ M. Tavassoli and T. Soussi. Expression of p53 in oral squamous cell carcinoma is associated with the presence of p53 autoantibodies in sera and saliva, manuscript in preparation.

Table 1 Frequency of p53-Abs in various types of cancer: a survey of the literature, 1979–1999

Methodology	Positive	Total	Comment	Reference
Control studies ^a				
IP ^b	0	164	Healthy women	(8)
IP	1	88	Children with nonmalignant disease	(9)
Wb	0	51	Healthy control	(38)
Wb	0	67	Acute and chronic liver diseases	(110)
ELISA ^c	2	206	Healthy control	(24)
Wb	1	73	Healthy women	(111)
ELISA	1	76	Healthy control	(65)
ELISA	0	49	Nonmalignant lung disease	(112)
ELISA	0	17	Healthy control	(113)
ELISA	0	15	Healthy control	(69)
Wb	0	48	Healthy control	(70)
ELISA + Wb	3	330	Healthy control	(66)
ELISA	0	40	Healthy control	(84)
ELISA	2	36	Nonmalignant liver disease	(84)
ELISA	4	63	Healthy control	(92)
ELISA	0	41	Healthy control (<i>n</i> = 29) Nonmalignant lung disease (<i>n</i> = 12)	(114)
ELISA + Wb + IP	0	30	Healthy control	(91)
ELISA + Wb + IP	1	19	Nonmalignant esophageal disease	(91)
Wb	0	24	Healthy control (<i>n</i> = 10) Nonmalignant lung disease (<i>n</i> = 14)	(76)
ELISA	0	195	Healthy control	(115)
ELISA	0	20	Healthy control	(93)
ELISA	0	9	Gastric polyps	(71)
ELISA	0	40	Nonmalignant liver disease	(116)
ELISA	0	10	Healthy control	(85)
ELISA	1	69	Healthy women	(117)
ELISA	0	140	Chronic hepatitis C	(118)
ELISA	0	41	Healthy control	(119)
ELISA	2	50	Healthy control	(120)
Wb	0	10	Healthy control	(121)
ELISA + Wb	0	11	Healthy control (<i>n</i> = 6) + HBV-infected patients (<i>n</i> = 5)	(122)
ELISA + Wb	0	60	Healthy control	(123)
ELISA + Wb + IP	0	50	Healthy control	(124)
ELISA + IP	0	51	Healthy women	(26)
ELISA	0	24	Nonmalignant digestive disease	(82)
ELISA	0	9	Healthy control	(125)
ELISA	0	41	Healthy control	(81)
ELISA	0	17	Chronic inflammatory diseases of head and neck	(81)
Wb	0	50	Healthy control	(79)
ELISA	17 ^d	70	Nonmalignant oral diseases	(126)
Breast				
IP	14	155	Association with high grade	(8)
IP	14	122	Association with high grade	(127)
Wb	7	60	Association with specific p53 mutation binding to hsp70	(39)
IP + Wb	15	100	Association with high grade and with tumor negative for steroid hormone receptor	(45)
ELISA	3	105		(128)
ELISA	12	93		(47)
ELISA	10	290	Association with tumor negative for steroid hormone receptor	(24)
Wb	45	176	Association with p53 accumulation in the tumor	(111)
ELISA	48	182	Association with high histological grade and p53 accumulation in the tumor	(65)
IF	21	50	Association with good survival	(68)
ELISA	42	353	Association with short survival	(64)
ELISA + Wb	15	165	Association with short survival	(66)
ELISA	2	12		(120)
ELISA	9	61	Association with tumor size	(129)
ELISA	39	82	No clinical correlation detected	(67)
Lung				
Wb	6	46	Correlation with missense mutation and p53 accumulation	(38)
ELISA	10	42		(47)
ELISA	6	73		(24)
ELISA	10	42		(48)
ELISA	16	136		(112)
ELISA + IP + Wb	9	107	Association with p53 accumulation in the tumor	(130)
ELISA	18	67	NSCLC; association with better survival after radiotherapy	(131)
ELISA	13	62	NSCLC; association with p53 accumulation in the tumor	(114)
ELISA + Wb	38	188	NSCLC; association with advanced stage and p53 accumulation in the tumor; no association with survival	(132)
Wb	9	111	NSCLC; association with poor survival	(76)
Wb	1	14	SCLC	(76)
ELISA	19	84	NSCLC; association with poor survival, especially for SCC	(75)
ELISA	17	140	NSCLC; association with p53 accumulation in the tumor and short survival for SCC only	(77)
Wb	27	170	SCLC; no association with clinical parameters	(79)
ELISA	54	231	SCLC	(78)
ELISA	20	97	SCLC; association with poor prognosis for limited-stage SCLC patients	(78)
Colon cancer				
IP	2	16		(127)
ELISA	13	82		(24)
ELISA	63	249	Association with poor differentiation and short survival	(73)
ELISA	59	184	Association with p53 accumulation and short survival	(72)
ELISA	42	235		(115)
ELISA	10	41	Association with p53 accumulation in the tumor	(85)

Table 1 *Continued*

Methodology	Positive	Total	Comment	Reference
ELISA	10	42		(120)
Wb	32	47		(121)
IP + Wb + ELISA	9	65		(124)
ELISA	14	54	Association with p53 accumulation in the tumor	(82)
ELISA	53	229	No clinical correlation detected	(74)
Lymphoma				
IP	14	119	High prevalence in Burkitt ¹ lymphoma and T-cell lymphoma	(9)
ELISA	3	115		(24)
ELISA	2	14	From patients with Sjögren syndrome	(133)
Gastric cancer				
ELISA	23	120	Association with p53 accumulation in the tumor and short survival	(69)
Wb	61	501	Association with large tumor, poor differentiation and short survival	(70)
ELISA	8	25	Association with poor differentiation, high stage, and poor survival	(71)
ELISA	13	81	No clinical correlation detected	(134)
Oral cancer				
Wb	7	14		(135)
ELISA	24	70	Leukoplakia	(92)
ELISA	15	50	Association with poor differentiation and p53 accumulation in the tumor	(92)
ELISA	32	74	No clinical correlation detected	(136)
ELISA	18	82		(137)
ELISA	23	117	Association with p53 accumulation in the tumor	(119)
ELISA	15	80	Association with p53 accumulation in the tumor and short survival	(80)
ELISA	9	39		(125)
ELISA	39	143	Association with therapy failure	(81)
Wb	7	30	Associated with tumors with p53 complexed to Hsp70	(40)
ELISA	37	97		(126)
ELISA	11	30	Patient with recurrence	(126)
ELISA	47	177	Association with tumor size	(138)
ELISA	32	73	Association with p53 accumulation in the tumor	(139)
Esophageal cancer				
ELISA + Wb + IP	11	33	Esophageal cancer, association with p53 accumulation in the tumor	(91)
ELISA + Wb + IP	3	36	Barrett's metaplasia	(91)
ELISA	6	20		(93)
IP + Wb + ELISA	16	65	Association with p53 accumulation in the tumor and p53 mutation in the core domain	(44)
ELISA	16	63	Association with p53 mutation in the tumor; no relationship with clinical parameter	(140)
ELISA	33	57	Not done	(141)
Liver cancer (HCC)				
Wb	20	80	European patients	(110)
			Advanced disease	
ELISA	9	130	Association with p53 accumulation in the tumor and better survival	(84)
ELISA	5	39		(142)
ELISA	28	86	Association with poor overall survival	(116)
ELISA	3	7		(118)
ELISA + Wb	14	38	Association with p53 accumulation in the tumor	(122)
Ovarian cancer				
ELISA	10	72		(128)
ELISA	11	86		(24)
ELISA	12	33	Association with poor survival	(113)
ELISA	3	17	Benign ovarian tumors	(113)
ELISA	15	40		(86)
ELISA	18	86	No clinical correlation detected	(143)
ELISA	8	30		(28)
ELISA	11	38	Association with poor histological differentiation and p53 accumulation in the tumor	(117)
ELISA	10	30		(144)
ELISA	41	174	Association with poor histological differentiation and short survival	(145)
ELISA + IP	4	46		(26)
Pancreatic cancer				
ELISA	2	46		(128)
ELISA	14	73		(47)
ELISA	2	46		(24)
ELISA + Wb	5	78		(146)
ELISA	8	29		(147)
ELISA + Wb	5	96	Chronic pancreatitis	(123)
ELISA + Wb	23	145	Association with high grading (grade III), short survival, and p53 accumulation in the tumor	(123)
ELISA	6	33	1/19 acute pancreatitis 4/33 chronic pancreatitis	(148)
Prostate cancer				
ELISA	2	83		(128)
ELISA	2	65		(24)
Thyroid cancer				
ELISA	4	108		(47)
Bladder cancer				
ELISA	8	29		(47)
Leukemia				
ELISA	4	88		(47)
ELISA	1	107		(24)
Wb	1	50	HTLV-1 asymptomatic carrier	(149)
Wb	2	50	ATL	(149)
Wb	3	50	HAM/TSP	(149)
ELISA	3	83	Association with p53 mutation	(150)

Table 1 *Continued*

Methodology	Positive	Total	Comment	Reference
Testicular cancer				
ELISA	0	144		(24)
Hepatoma				
ELISA	1	150		(24)
Melanoma				
ELISA	0	58		(24)
Multiple myeloma				
ELISA	5	165		(24)
ELISA	18	119		(151)
ELISA	0	80		(150)
Glioma				
ELISA	2	12		(36)
ELISA	0	70		(35)
ELISA	4	107		^b
Endometrial cancer				
ELISA	2	10		(144)

^a We have included all individuals both healthy and with nonmalignant disease.

^b IP, immunoprecipitation; WB, Western blot; HTLV-1, human T-cell lymphotropic virus, type 1; ATL, adult T-cell leukemia; HAM/TSP, HTLV-associated myelopathy/tropical spastic paraparesis; IF, immunofluorescence; HCC, hepatocellular carcinoma.

^c ELISA, 20% of the assays were homemade, whereas 80% were commercial (2 main distributors).

^d This single study represented 50% of positive cases in healthy controls.

^e P. Murray, T. Soussi, and M. Tavassoli. Serum p53 antibodies: predictors of survival in small-cell lung cancer?, manuscript in preparation.

^f Unpublished observations.

were obtained only after a special immunization or selection procedure (53, 54).

Taken together (a) the presence of immunodominant epitopes outside the hot spot region of the p53 mutation; (b) the correlation between p53 accumulation (and p53 gene mutation) in tumor cells and p53-Abs responses; (c) the similarity of humoral responses in patients independent of the cancer type; and (d) the similarity of antigenic site profiles in patients and hyperimmunized animals—all suggest that p53 accumulation is the major component of the humoral response in patients with cancer. This accumulation could lead to a self-immunization process culminating in the appearance of p53-Abs. As stated above, the level of p53 proteins in a normal organism is very low, which suggests very weak (if any) tolerance to endogenous p53. Isotyping of p53-Abs has shown that they correspond mainly to IgG1 and IgG2 subclasses, although some patients exhibit a predominant IgA response (25). Some patients also had IgM, although none had p53 IgM as the only isotype. No IgG3 or IgG4 was detected. Again,

this result strengthens the hypothesis of an active humoral response to p53.

It is not clear whether p53 mutation is really required for the production of p53-Abs or whether the sole accumulation of p53 protein can lead to this humoral response. This question is difficult to answer because there is no normal situation of wild-type p53 accumulation in humans that could be used to test this hypothesis. On the other hand, such a situation occurs in animals. p53-Abs have been discovered in the sera of animals that bear tumors induced by SV40 (1, 5). In such tumors, wild-type p53 is stabilized through its interaction with SV40 large T antigen. These observations indicate that p53 accumulation alone is the main component of this autoimmunization.

The mechanism by which p53 is presented to the immune system is unknown. It is possible that it is released after cell necrosis, but thus far, p53 has not been found reproducibly in human sera (see below; 55, 56). Either the p53 protein is very rapidly eliminated from sera, or else other mechanisms of p53 presentation are involved.

X-ray crystallography of human p53 was an important step in the understanding of the structure of this protein. The central region (amino acids 102–292) was crystallized in the form of a protein-DNA complex (57). This core region has been shown to include the following motifs: (a) two antiparallel β sheets composed of four and five β -strands, respectively. These two sheets form a kind of compact sandwich that holds the other elements; (b) a LSH containing three β -strands, an α -helix, and the L1 loop; (c) an L2 loop containing a small helix; and (d) an L3 loop composed mainly of turns. It is quite remarkable to note the striking correspondence between these various structural elements and the four evolutionarily conserved blocks (II to V). The LSH motif and the L3 helix are involved in direct DNA interaction (LSH with the major groove and L3 with the minor groove). The L2 loop is presumed to provide stabilization by associating with the L3 loop. These two loops are held together by a zinc atom tetraordinated to Cys176 and His179 on the L2 loop and to Cys278 and Cys242 on the L3 loop.

Furthermore, the conformational changes in p53 were dissected by a new battery of monoclonal antibodies directed against the central region of the protein (53). None of these antibodies was able to recognize native, wild-type p53. On the other hand, regardless of the location of their epitopes, they were all able to recognize p53 mutants that had undergone conformational changes (53). This result indicates

Table 2 *p53-Ab frequency in various types of cancer: statistical evaluation*

Status	Frequency of p53-Abs	P^a
Healthy	35/2404 ^b	
Esophageal cancer	85/274	$<10^{-4}$
Oral cancer	309/1062	$<10^{-4}$
Bladder cancer	8/29	$<10^{-4}$
Colon cancer	307/1244	$<10^{-4}$
HCC ^c	82/387	$<10^{-4}$
Ovarian cancer	140/635	$<10^{-4}$
Lung cancer	219/1282	$<10^{-4}$
Breast cancer	296/2006	$<10^{-4}$
Gastric cancer	105/727	$<10^{-4}$
Pancreatic cancer	60/650	$<10^{-4}$
Multiple myeloma	23/364	$<10^{-4}$
Lymphoma	19/248	$<10^{-4}$
Leukemia	14/428	0.005
Glioma	6/144	0.03
Prostate cancer	4/148	NS
Testicular cancer	0/144	NA
Melanoma	0/58	NA
Hepatoma	0/150	NA
Total cancers	1600/9489	$<10^{-4}$

^a Correlation between cancer patients and healthy individuals were tested by the χ^2 test. The levels of significance were set at $P < 0.05$.

^b As shown in Table 1, one study contributed to 17 (50%) positives in healthy controls. All of the statistical analyses including this study were performed emphasizing the strength of these trends. If this study is omitted (18 positive controls), prostate carcinoma reaches significance with a P of 0.015.

^c HCC, hepatocellular carcinoma; NS, not significant; NA, not applicable.

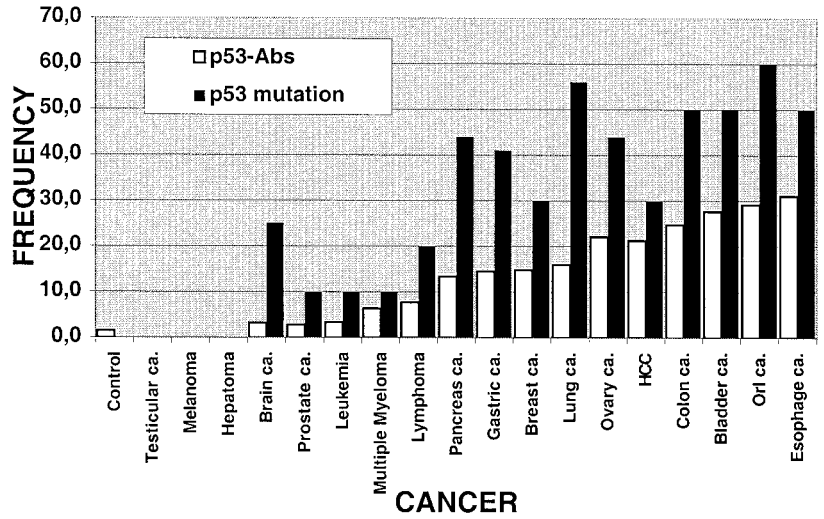


Fig. 1. Relationship between the frequency of p53 mutation and p53 antibodies in various types of cancer. The frequency of p53 mutations is taken from the literature, and the frequency of p53-Abs is taken from Table 1.

that the central region of wild-type p53 protein has a very compact structure that is held in place by the two antiparallel B sheets. Such conformation and the observation that this region is highly hydrophobic explain its poor immunogenicity.

A recent study (58) indicates that critical residues of epitopes recognized by several anti-p53 monoclonal antibodies correspond to key residues of p53 involved in interaction of the mdm2 protein with the NH₂ terminus of the p53 protein. This can explain why such a region is at the surface of the molecule. It should also be mentioned that the p53 protein, whatever its phylogenetic origin, is an incredibly potent antigen. Immunization of mice with p53 requires only a small amount of protein, and adjuvant is not necessary to obtain a high titer of antibodies.⁵

Recently, several p53 homologues have been identified (59–62). Although, the conservation of the sequence is only partial, it is conceivable that p53-Abs could either cross-react with such proteins or were produced toward one of the homologues. Extensive assays of sera with or without p53-Abs indicate that most sera are specific for p53. Nevertheless, specific antibodies toward p73 have been detected.⁶

p53 Antibodies and Clinical Parameters

Numerous studies have attempted to evaluate the clinical value of p53-Abs (Table 1). As for p53 mutation and p53 immunohistochemical analysis, these studies have reported contradictory results. Before examining them, one should bear in mind one question that has not been assessed thus far, *i.e.*, the role of such antibodies in the neoplastic process. This question has never been thoroughly discussed or experimentally tested. During our lifetime, it is quite possible that we experience several unknown precancers that arise after a genetic variation in an oncogene or a tumor suppressor gene. Such preneoplastic cancers could be quickly eliminated either because the mechanisms that control cellular proliferation have been able to overcome these tumoral cells or because they have been eliminated from the organism through various surveillance mechanisms including the immune system. It is plausible that early p53 accumulation, such as that seen in lung or oral cancer, can lead to the production of a humoral and cellular response that participates in the elimination of the tumor. Such a hypothesis is supported by the work of Roth *et al.* (63).

Immunization with canarypox virus recombinants that expressed human or murine p53 protected BALB/c mice from a challenge with a highly tumorigenic mouse fibroblast tumor cell line that express high levels of mutant p53. This tumor protection was equally effective regardless of whether wild-type or mutant p53 was used for the immunization, which indicated that the immunological response was not dependent on any particular p53 mutation and that immunization with this live virus vaccine works effectively against mutant p53 protein expressed in a tumor cell (63). Although there is no formal proof that a natural response toward p53 can protect from precancerous tumor, it is not possible to exclude such a hypothesis. However, it is quite possible that such an immune response is totally neutral toward the organism and is simply the consequence of self-immunization toward a self-protein. In that case, any clinical correlation with p53-Abs would be due to the presence of p53 inactivation via mutation.

In breast cancer, several studies indicate that p53-Abs are found in patients with tumors that have high grades and/or that are negative for steroid hormone receptors (8, 24, 45, 64, 65), two clinical parameters already known to be associated with p53 mutations and bad prognosis. Two studies, on 353 and 165 patients, found an association between p53-Abs and short survival (64, 66), whereas one study (82 patients) did not find any association (67), and another study (50 patients) found an association with good survival (68). In gastric cancer, three of four studies found an association between p53-Abs and poorly differentiated tumors and short survival (69–71). In colon cancer, two of three studies also found an association between p53-Abs and short survival (72–74). In lung cancer, as for p53 mutations, controversies exist concerning the clinical value of p53-Abs (Table 1). In NSCLC,

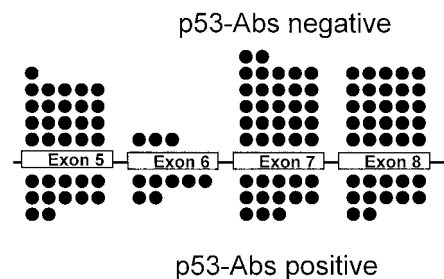


Fig. 2. Distribution of p53 mutations and p53-Abs in patients in whom both serological and molecular analyses were performed.

⁵ Y. Legros and T. Soussi, unpublished observation; D. Lane, personal communication.
⁶ O. Tominaga, K. Unzal, and T. Soussi. p73 antibodies are found in the sera of patients with various types of cancer, manuscript in preparation.

Table 3 Frequency of p53-Abs in high-risk individuals: a survey of the literature 1979–1999

Methodology	Selected Individuals/ Possible Carcinogen	Clinical Status at the Time of p53-Ab Detection	Description	p53 Alteration	Type of Study (Reference)
ELISA, IP, WB	Heavy smokers/tobacco residues	No detectable malignant or premalignant lesion	Two individuals with p53-Abs Patient 90: no follow-up; died 8 months later from lung cancer Patient 37: effective follow-up; developed a lung cancer 2 years later; tumor was detected before clinical manifestation of the cancer; patient still alive after therapy	ND ^a IHC+ ^b	Prospective with follow-up (48, 88)
ELISA, IP, WB	Factory workers in industry/vinyl chloride	ND	Patient 24 with ASL: p53-Abs 10 yr before diagnosis Patient 23 with ASL: p53-Abs 3 mo before diagnosis Patient 19 with Raynaud's syndrome: p53-Abs 4 yr before diagnosis Worker 22 without diagnosed cancer: p53-Abs 4 yr before assay Worker 21 without diagnosed cancer: p53-Abs 6 yr before assay	ND ND ND ND ND	Retrospective study (42)
ELISA, IP, WB	Heavy smokers with COPD/tobacco residue	ND	Patient 12 with lung cancer; p53-Abs 7 mo before diagnosis Patient 10 with lung cancer; p53-Abs 6 mo before diagnosis Patient 8 with breast cancer; p53-Abs 7 mo before diagnosis Patient 11 with prostate cancer; p53-Abs 11 mo before diagnosis	ND Mutant p53 IHC+ IHC+	Retrospective study (90)
ELISA	Betel nut chewer/tobacco and copper in betel nut	Leukoplakia	Exposed individuals: 4/63 ^c Premalignant (leukoplakia): 15/50 Cancer: 24/70 No follow-up of these patients	ND	Prospective study (92)
ELISA, IP, WB	Barrett's metaplasia	Barrett's metaplasia	Patient 88 with Barrett's esophagus; p53-Abs 1 mo before detection of ADC Patient 2 with ADC; p53-Abs 1 mo before diagnosis	ND Mutant p53	Retrospective study (91)

^a ND, not done; ASL, angiosarcoma of the liver; IHC, immunohistochemistry.

^b IHC +, p53 accumulation in the tumor.

^c These four normal subjects were heavy consumers of tobacco and betel quid.

p53-Abs seem to be associated with poor survival, especially in squamous cell carcinoma (75–77), whereas in SCLC, the studies are very divergent (78, 79). In oral cancer, two studies have also demonstrated an association between p53-Abs and short survival (80, 81).

Taken together, in all of these studies, there is a trend toward an association between p53-Abs and tumors with poor differentiation, a feature already observed with p53 mutations. The value of p53-Abs in terms of survival is promising, but additional studies are necessary before this can be clearly established.

p53 and Follow-Up of Patients during Therapy

Because p53 accumulation is the main trigger of this humoral response, it was of interest to examine the behavior of these p53-Abs during therapy to see whether there was a relationship between tumor disappearance and a decrease in p53-Abs. Several studies have addressed this question in various types of cancer (24, 28, 82–87). Such studies can only be performed using a quantitative assay, but this has not been taken into account in many reports. Using immunoprecipitation and two different ELISA formats, Zalcman *et al.* (83) showed that there is a good correlation between the specific evolution of the p53-Abs titer and the response to therapy in patients with lung cancer. A similar situation was described in colorectal (82) and ovarian cancer (24). In other studies, clinical data were not available. In several patients, the disappearance of p53-Abs was very rapid, nearly as rapid as the half-life of human IgG (83, 88). Several arguments demonstrate the specificity of p53-Abs variation during therapy: (a) there is no

variation in total serum immunoglobulins; (b) there is no variation in the amount of antibodies directed toward other antigens; and (c) a decrease of p53-Abs can occur in patients who have been treated by surgery without any chemo- or radiotherapy. All of these observations indicate that constant stimulation of the immune system is necessary to maintain a high level of p53-Abs. Removing the tumor would prevent such stimulation.

In breast cancer, it is possible to detect the reappearance of p53-Abs 2 years after initial therapy. This increase in p53-Abs has been detected 3 months before the detection of a relapse (5). Thus, in such tumor types, p53-Abs could be a useful tool for controlling the response to therapy and for monitoring certain early relapses before they are clinically detectable. As indicated above, several studies have demonstrated that sera that score negative at the time of diagnosis never turn positive during follow-up (82, 83, 86).

p53 Antibodies and Populations at High Risk of Cancer

As demonstrated in the previous section, p53 accumulation is the major component in the appearance of these p53-Abs. These antibodies are usually IgG indicating a secondary response after a prolonged immunization before the diagnosis of the disease (25). All of the studies described up until now used sera taken at the time of diagnosis before any treatment. Thus, it is reasonable to presume that such p53-Abs could be used as an early indicator of p53 mutations in tumors in which such alterations occur early during tumoral progression (Table 3). One good model for testing this hypothesis is that of

Table 4 Quantification of p53 in human sera

Assay	Patients	Amount of p53	Comment	References
ELISA ^a /PAb240	71 controls	0.37 ± 0.44 ng/ml	No statistical difference between controls and cancers	(97)
	40 colon cancers	0.31 ± 0.46 ng/ml		
	12 polyps	0.84 ± 0.44 ng/ml		
ELISA ^a /PAb240	58 controls	0.31 ± 0.41 ng/ml	No statistical difference between controls and cancers	(95)
	23 lung cancers	0.55 ± 1.16 ng/ml		
ELISA	100 controls	<0.04 ng/ml	No p53 protein	(55)
	13 nonmalignant lung diseases	<0.04 ng/ml		
	114 patients with lung cancers	<0.04 ng/ml ^b		
ELISA ^a /PAb240	47 controls	0.55 ng/ml	Statistical difference between controls and cancer, <i>P</i> < 0.02	(96)
	61 "normal" with history of colon cancer	0.09 ng/ml		
	54 adenomas	0.44 ng/ml		
	22 carcinomas	0.55 ng/ml		
ELISA ^a /PAb1801	10 controls	0.29 ng/ml	No statistical difference between controls and cancer	(94)
	36 asbestos controls	0.61 ± 0.33 ng/ml		
	27 lung cancers	0.33 ± 0.07 ng/ml		
ELISA ^a /PAb1801	No controls			(93)
	15 esophageal cancer	0.6–8.6 ng/ml		
ELISA	800 cancer patients	Negative		(56)
ELISA	67 lung cancer patients	Negative		(38)
HPLC ^c	144 controls	0.22–0.6 mg/ml	Should be taken with caution	(98)
	184 colon cancers	1.8–5.6 mg/ml		

^a All of these ELISA were performed with the same commercial kit using a specific monoclonal antibody as a capture antibody.

^b Two sera were shown to be false positive after checking for heterophilic antibodies.

^c HPLC, high-performance liquid chromatography.

lung cancer and heavy smokers. It is well established that p53 accumulation is an early event in lung cancer and that such cancer is strongly associated with tobacco smoking. Indeed, in 1994, p53-Abs were found in two heavy smoker who were negative for any detectable lung cancer (48). One of the patients could not be followed but died 8 months later from a rapidly growing lung tumor. The second patient, PT37, was placed under surveillance, with regular assay for p53-Abs and thoracic X-rays. Two years later, lung cancer was detected in this patient before any clinical manifestations of the disease (88). The patient showed good response to therapy that paralleled the total disappearance of p53-Abs (83, 88). Since 1996, this patient has been tumor-free, and a recent check-up indicated that neither the tumor nor p53-Abs had reappeared. To our knowledge, this is the only prospective study that addressed the importance of p53-Abs in individuals at high risk for cancer, and that used such assays for clinical management of the patient. Since that work, several studies have demonstrated that p53-Abs can be found in the sera of high-risk individuals.

Angiosarcoma of the liver is an extremely rare cancer in humans; it is found in individuals, including workers in several types of industries, who have been exposed to several carcinogens such as vinyl chloride. p53-Abs were detected in the sera of individuals several years before the diagnosis of the tumors (Table 3; Ref. 42). This work is of importance because it is known that p53 mutations are frequent in individuals exposed to various carcinogens, and such mutations usually occur early in the transforming process (89). Therefore, this assay could be useful for early identification of cancer in individuals occupationally exposed to carcinogens. Similarly, p53-Abs have been detected in the sera of patients with chronic obstructive pulmonary disease and in heavy smokers (Table 3; Ref. 90).

There exist certain clinical situations in which nonmalignant lesions can predate their progression toward cancer. This is the case in Barrett's esophagus. The histopathological sequence for (Barrett's) metaplasia, which develops—as a consequence of chronic reflux—to dysplasia and then to carcinoma is well established for these tumors. In Barrett's esophagus, a variety of molecular changes have been characterized and correlated with tumor initiation and progression. Mutations and accumulation of p53 are found mainly in the transition from low- to high-grade dysplasia and are associated with an in-

creased risk of cancer. The finding of p53-Abs in patients with Barrett's esophagus may be promising if confirmed in a larger population because it may predate clinical diagnosis of esophageal ADC (91). A similar situation occurs in individuals with premalignant oral lesions (leukoplakia) due to tobacco or betel nut chewing. Such individuals are at high risk of developing oral cancer (5–10%). p53-Abs have been found at high frequency in patients with premalignant and malignant lesions, which suggests that such antibodies could be used for early detection of cancer (Table 3; Ref.92). Unfortunately, no follow-up has been performed on these patients. Due to the high frequency of this type of cancer in countries such as India or Pakistan, this kind of diagnosis could be of importance. The recent discovery that p53-Abs can be found in saliva indicates that easy screening could be organized to verify the value of these antibodies (27).

p53 Protein in Sera

This question continues to be a subject of debate with highly divergent results (Table 4; Refs. 55, 56, 93–98). It should be pointed out that most assays use a commercial ELISA kit that was developed for the detection of p53 protein in cell or tumor extracts but that has not been fully verified on serum samples. Sera have always been tested undiluted, which can lead to high background and false-positives. Indeed, in carefully controlled experiments, Levesque *et al.* (55) demonstrated that some false-positive sera were caused either by the presence of human antibodies with broad antispecies specificities that can cross-react with some antibodies used in the assay or by other nonspecific reactants that interfere with the assay.

Due to the lack of reliability of the various assays used, serum p53 protein should not be considered as valid as long as the protein has not been formally identified in sera using a reliable assay.

Antibodies toward Other Oncogenes and Tumor Suppressor Genes

Only a few published studies have addressed this question. Antibodies to ras (99), c-myc (100), L-myc (101), c-myc (102), mdm2 (103) or HER2-neu (104, 105) have been detected in sera of patients with various types of cancers.

The presence of antibodies to HER-2/neu were detected in 12

(11%) of 107 breast cancer patients *versus* none (0%) of 200 normal controls (104). The presence of antibodies to HER-2/neu was also correlated with the accumulation of HER-2/neu protein in the patient's primary tumor. Such accumulation of HER-2/neu protein is well known to be associated with a poor prognosis. No information concerning the association of such antibodies with clinical data has been published thus far.

The paucity of these data compared with p53 suggests that p53 immunogenicity is a rather specific situation related to the striking immunogenicity of the p53 protein.

This field is advancing, especially with the use of new methodology, such as SEREX, that enables identification of specific antibodies associated with gene overexpression in tumors (106, 107). Such methodology can shed light on new tumor antigens, which could lead to the discovery of new cancer genes, repeating the history of the p53 with the identification of the protein prior to cloning the gene.

p53 Antibodies: Future Directions

Although several authors have questioned the specificity of p53-Abs, a review of the literature (Tables 1 and 2) clearly demonstrates the specificity of this serological analysis because such antibodies are truly rare in the normal population. It is possible to estimate that the specificity of this assay attains 95%. Such specificity is supported by our knowledge of p53 that accumulates specifically in the nucleus of tumor cells after gene mutation. Among the various TAAs that have been analyzed over the years, the production of p53-Abs is surely the best studied and most clearly explains humoral response. One of the disadvantages of this assay is its lack of sensitivity inasmuch as only 20–40% of patients with p53 mutations will develop p53-Abs. This lack of sensitivity totally precludes the use of the assay to evaluate p53 alteration in human tumor. Nevertheless, if we estimate that there are 8 million patients with various types of cancer throughout the world, and 50% of them have a mutation in their p53 gene, then we can deduce that about 1 million of these patients have p53-Abs.

There exist several situations in which p53-Abs could have clinical utility. The first is in the monitoring of sera during therapy. Only prospective studies on various types of cancer in which relapses occur several months or years after treatment will enable us to validate this assay. The use of standardized assays that have been validated for quantitative analysis should help in such studies.

The second situation concerns p53-Abs in high-risk individuals. One of the challenges of the next millennium is the early detection of tumors using highly sensitive assays with gene probes specific for tumor genetic alterations (108). Such approaches are still under development and remain costly. I do believe that there is still room for serological assays of tumors markers such as those described in the present review. In developing countries, there is an increased burden of tumors due to carcinogen exposure. This is the result of an increase in cigarette consumption, higher pollution caused by political laxism, uncontrolled industrial development, and the absence of regulation in waste evacuation. It is possible that p53 mutations in cancer related to such exposure are high (109). The use of a low-cost assay for the detection of p53-Abs could be of public health benefit in such countries.

Over the past 10 years, a tremendous amount of work has been performed on p53. Such an effort, both in academic and private laboratories has never been made for any other single gene thus far. This work has led to the development of numerous clinical studies including gene therapy protocols, with the aim of achieving more efficient cure of the disease.

Finally, let me dedicate this review to Patient PT37 (88). I think that this patient is the first to have benefited from significant improvement,

and, indeed, whose very life has been saved, thanks to the knowledge we have gained about p53, particularly from the TAA studies published 20 years ago (7).

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REFERENCES

1. Melero, J. A., Stitt, D. T., Mangel, W. F., and Carroll, R. B. Identification of new polypeptide species (48–55K) immunoprecipitable by antiserum to purified large T antigen and present in simian virus 40-infected and transformed cells. *J. Virol.*, 93: 466–480, 1979.
2. Linzer, D. I. H., and Levine, A. J. Characterization of a 54K dalton cellular SV40 tumor antigen present in SV40-transformed cells and uninfected embryonal carcinoma cells. *Cell*, 17: 43–52, 1979.
3. Linzer, D. I., Maltzman, W., and Levine, A. J. The SV40 A gene product is required for the production of a 54,000 MW cellular tumor antigen. *Virology*, 98: 308–318, 1979.
4. Lane, D. P., and Crawford, L. V. T antigen is bound to a host protein in SV40-transformed cells. *Nature (Lond.)*, 278: 261–263, 1979.
5. Kress, M., May, E., Cassingena, R., and May, P. Simian Virus 40-transformed cells express new species of proteins precipitable by anti-simian virus 40 serum. *J. Virol.*, 31: 472–483, 1979.
6. Rotter, V., Witte, O. N., Coffman, R., and Baltimore, D. Abelson murine leukemia virus-induced tumors elicit antibodies against a host cell protein, p50. *J. Virol.*, 36: 547–555, 1980.
7. De Leo, A. B., Jay, G., Appella, E., Dubois, G. C., Law, L. W., and Old, L. J. Detection of a transformation-related antigen in chemically induced sarcomas and other transformed cells of the mouse. *Proc. Natl. Acad. Sci. USA*, 76: 2420–2424, 1979.
8. Crawford, L. V., Pim, D. C., and Bulbrook, R. D. Detection of antibodies against the cellular protein p53 in sera from patients with breast cancer. *Int. J. Cancer*, 30: 403–408, 1982.
9. Caron de Fromental, C., May-Levin, F., Mouriesse, H., Lemerle, J., Chandrasekaran, K., and May, P. Presence of circulating antibodies against cellular protein p53 in a notable proportion of children with B-cell lymphoma. *Int. J. Cancer*, 39: 185–189, 1987.
10. Caron de Fromental, C., and Soussi, T. *TP53* tumor suppressor gene: a model for investigating human mutagenesis. *Genes Chromosomes Cancer*, 4: 1–15, 1992.
11. Crawford, L. The 53,000-dalton cellular protein and its role in transformation. *Int. Rev. Exp. Pathol.*, 25: 1–50, 1983.
12. Levine, A. J. p53, the cellular gatekeeper for growth and division. *Cell*, 88: 323–331, 1997.
13. Oren, M., and Rotter, V. Introduction: p53—the first twenty years. *Cell. Mol. Life Sci.*, 55: 9–11, 1999.
14. Giaccia, A. J., and Kastan, M. B. The complexity of p53 modulation: emerging patterns from divergent signals. *Genes Dev.*, 12: 2973–2983, 1998.
15. Prives, C. Signaling to p53: breaking the MDM2-p53 circuit. *Cell*, 95: 5–8, 1998.
16. Benchimol, S., Pim, D., and Crawford, L. Radioimmunoassay of the cellular protein p53 in mouse and human cell lines. *EMBO J.*, 1: 1055–1062, 1982.
17. Kastan, M. B., Onyekwere, O., Sidransky, D., Vogelstein, B., and Craig, R. W. Participation of p53 protein in the cellular response to DNA damage. *Cancer Res.*, 51: 6304–6311, 1991.
18. Yonish-Rouach, E., Resnitzky, D., Lotem, J., Sachs, L., Kimchi, A., and Oren, M. Wild-type p53 induces apoptosis of myeloid leukaemic cells that is inhibited by interleukin-6. *Nature (Lond.)*, 352: 345–347, 1991.
19. Soussi, T., Legros, Y., Lubin, R., Ory, K., and Schlichtholz, B. Multifactorial analysis of p53 alteration in human cancer—a review. *Int. J. Cancer*, 57: 1–9, 1994.
20. Greenblatt, M. S., Bennett, W. P., Hollstein, M., and Harris, C. C. Mutations in the p53 tumor suppressor gene: clues to cancer etiology and molecular pathogenesis. *Cancer Res.*, 54: 4855–4878, 1994.
21. Bérout, C., and Soussi, T. p53 gene mutation: software and database. *Nucleic Acids Res.*, 26: 200–204, 1998.
22. Casey, G., Lopez, M. E., Ramos, J. C., Plummer, S. J., Arboleda, M. J., Shaughnessy, M., Karlan, B., and Slamon, D. J. DNA sequence analysis of exons 2 through 11 and immunohistochemical staining are required to detect all known p53 alterations in human malignancies. *Oncogene*, 13: 1971–1981, 1996.
23. Dowell, S. P., Wilson, P. O. G., Derias, N. W., Lane, D. P., and Hall, P. A. Clinical utility of the immunocytochemical detection of p53 protein in cytological specimens. *Cancer Res.*, 54: 2914–2918, 1994.
24. Angelopoulou, K., Diamandis, E. P., Sutherland, D. J. A., Kellen, J. A., and Bunting, P. S. Prevalence of serum antibodies against the p53 tumor suppressor gene protein in various cancers. *Int. J. Cancer*, 58: 480–487, 1994.
25. Lubin, R., Schlichtholz, B., Teillaud, J. L., Garay, E., Bussel, A., Wild, C., and Soussi, T. p53 antibodies in patients with various types of cancer: assay, identification and characterization. *Clin. Cancer Res.*, 1: 1463–1469, 1995.
26. Vennegoor, C., Nijman, H. W., Drijfhout, J. W., Vernie, L., Verstraeten, R. A., vonMensdorffPouilly, S., Hilgers, J., Verheijen, R. H. M., Kast, W. M., Melief, C. J. M., and Kenemans, P. Autoantibodies to p53 in ovarian cancer patients and healthy women: a comparison between whole p53 protein and 18-mer peptides for screening purposes. *Cancer Lett.*, 116: 93–101, 1997.

27. Tavassoli, M., Brunel, N., Maher, R., Johnson, N. W., and Soussi, T. p53 antibodies in the saliva of patients with squamous cell carcinoma of the oral cavity. *Int. J. Cancer*, **78**: 390–391, 1998.
28. Angelopoulou, K., and Diamandis, E. P. Detection of the *TP53* tumour suppressor gene product and p53 auto-antibodies in the ascites of women with ovarian cancer. *Eur. J. Cancer*, **33**: 115–121, 1997.
29. Munker, R., Stotzer, O., Darsow, M., Classen, S., Lebeau, A., and Wilmanns, W. Autoantibodies against p53 are not increased in human ascites and pleural effusions. *Cancer Immunol. Immunother.*, **42**: 200–201, 1996.
30. Puisieux, A., Galvin, K., Troalen, F., Bressac, B., Marçais, C., Galun, E., Ponchel, F., Yacicier, C., Ji, J. W., and Ozturk, M. Retinoblastoma and p53 tumor suppressor genes in human hepatoma cell lines. *FASEB J.*, **7**: 1407–1413, 1993.
31. Fleischhacker, M., Strohmeier, T., Imai, Y., Slamon, D. J., and Koefler, H. P. Mutations of the p53 gene are not detectable in human testicular tumors. *Mod. Pathol.*, **7**: 435–439, 1994.
32. Peng, H. Q., Hogg, D., Malkin, D., Bailey, D., Gallie, B. L., Bulbul, M., Jewett, M., Buchanan, J., and Goss, P. E. Mutations of the p53 gene do not occur in testis cancer. *Cancer Res.*, **53**: 3574–3578, 1993.
33. Lubbe, J., Reichel, M., Burg, G., and Kleihues, P. Absence of p53 gene mutations in cutaneous melanoma. *J. Invest. Dermatol.*, **102**: 819–821, 1994.
34. Luca, M., Lenzi, R., Lee-Jackson, D., Gutman, M., Fidler, I. J., and Bar-Eli, M. p53 mutations are infrequent and do not correlate with the metastatic potential of human melanoma cells. *Int. J. Oncol.*, **3**: 19–22, 1993.
35. Rainov, N. G., Dobberstein, K. U., Fittkau, M., Bahn, H., Holzhausen, H. J., Gantchev, L., and Burkert, W. Absence of p53 antibodies in sera from glioma patients. *Clin. Cancer Res.*, **1**: 775–781, 1995.
36. Weller, M., Bornemann, A., Stander, M., Schabet, M., Dichgans, J., and Meyer-mann, R. Humoral immune response to p53 in malignant glioma. *J. Neurol.*, **245**: 169–172, 1998.
37. Ohgaki, H., Eibl, R. H., Schwab, M., Reichel, M. B., Mariani, L., Gehring, M., Petersen, I., Holl, T., Wiestler, O. D., and Kleihues, P. Mutations of the p53 tumor suppressor gene in neoplasms of the human nervous system. *Mol. Carcinog.*, **8**: 74–80, 1993.
38. Winter, S. F., Minna, J. D., Johnson, B. E., Takahashi, T., Gazdar, A. F., and Carbone, D. P. Development of antibodies against p53 in lung cancer patients appears to be dependent on the type of p53 mutation. *Cancer Res.*, **52**: 4168–4174, 1992.
39. Davidoff, A. M., Iglehart, J. D., and Marks, J. R. Immune response to p53 is dependent upon p53/HSP70 complexes in breast cancers. *Proc. Natl. Acad. Sci. USA*, **89**: 3439–3442, 1992.
40. Kaur, J., Srivastava, A., and Ralhan, R. Serum p53 antibodies in patients with oral lesions: Correlation with p53/HSP70 complexes. *Int. J. Cancer*, **74**: 609–613, 1997.
41. Winter, S. F., Sekido, Y., Minna, J. D., McIntire, D., Johnson, B. E., Gazdar, A. F., and Carbone, D. P. Antibodies against autologous tumor cell proteins in patients with small-cell lung cancer—association with improved survival. *J. Natl. Cancer Inst.*, **85**: 2012–2018, 1993.
42. Trivers, G. E., Cawley, H. L., Debenedetti, V. M. G., Hollstein, M., Marion, M. J., Bennett, W. P., Hoover, M. L., Prives, C. C., Tamburro, C. C., and Harris, C. C. Anti-p53 antibodies in sera of workers occupationally exposed to vinyl chloride. *J. Natl. Cancer Inst.*, **87**: 1400–1407, 1995.
43. Hammel, P., LeroyViard, K., Chaumette, M. T., Villaudy, J., Falzone, M. C., Rouillard, D., Hamelin, R., Boissier, B., and Remvikos, Y. Correlations between p53-protein accumulation, serum antibodies and gene mutation in colorectal cancer. *Int. J. Cancer*, **81**: 712–718, 1999.
44. von Brevern, M. C., Hollstein, M. C., Cawley, H. M., De Benedetti, V. M. G., Bennett, W. P., Liang, L., He, A. G., Zhu, S. M., Tursz, T., Janin, N., and Trivers, G. E. Circulating anti-p53 antibodies in esophageal cancer patients are found predominantly in individuals with p53 core domain mutations in their tumors. *Cancer Res.*, **56**: 4917–4921, 1996.
45. Schlichtholz, B., Legros, Y., Gillet, D., Gaillard, C., Marty, M., Lane, D., Calvo, F., and Soussi, T. The immune response to p53 in breast cancer patients is directed against immunodominant epitopes unrelated to the mutational hot spot. *Cancer Res.*, **52**: 6380–6384, 1992.
46. Labrecque, S., Naor, N., Thomson, D., and Matlashewski, G. Analysis of the anti-p53 antibody response in cancer patients. *Cancer Res.*, **53**: 3468–3471, 1993.
47. Lubin, R., Schlichtholz, B., Bengoufa, D., Zalcman, G., Tredaniel, J., Hirsch, A., Caron de Fromental, C., Preudhomme, C., Fenaux, P., Fournier, G., Mangin, P., Laurent-Puig, P., Pelletier, G., Schlumberger, M., Desgrandchamps, F., Leduc, A., Peyrat, J. P., Janin, N., Bressac, B., and Soussi, T. Analysis of p53 antibodies in patients with various cancers define B-Cell epitopes of human p53: distribution on primary structure and exposure on protein surface. *Cancer Res.*, **53**: 5872–5876, 1993.
48. Schlichtholz, B., Tredaniel, J., Lubin, R., Zalcman, G., Hirsch, A., and Soussi, T. Analyses of p53 antibodies in sera of patients with lung carcinoma define immunodominant regions in the p53 protein. *Br. J. Cancer*, **69**: 809–816, 1994.
49. Legros, Y., Lafon, C., and Soussi, T. Linear antigenic sites defined by the B-cell response to human p53 are localized predominantly in the amino- and carboxy-termini of the protein. *Oncogene*, **9**: 2071–2076, 1994.
50. Hardy-Bessard, A. C., Garay, E., Lacroinque, V., Legros, Y., Demarquay, C., Houque, A., Portefaix, J. M., Granier, C., and Soussi, T. Regulation of the specific DNA binding activity of *Xenopus laevis* p53: evidence for conserved regulation through the carboxy-terminus of the protein. *Oncogene*, **16**: 883–890, 1998.
51. Vojtesek, B., Bartek, J., Midgley, C. A., and Lane, D. P. An immunohistochemical analysis of the human nuclear phosphoprotein-p53: new monoclonal antibodies and epitope mapping using recombinant-p53. *J. Immunol. Methods*, **151**: 237–244, 1992.
52. Bartek, J., Bartkova, J., Lukas, J., Staskova, Z., Vojtesek, B., and Lane, D. P. Immunohistochemical analysis of the p53 oncoprotein on paraffin sections using a series of novel monoclonal antibodies. *J. Pathol.*, **169**: 27–34, 1993.
53. Legros, Y., Meyer, A., Ory, K., and Soussi, T. Mutations in p53 produce a common conformational effect that can be detected with a panel of monoclonal antibodies directed toward the central part of the p53 protein. *Oncogene*, **9**: 3689–3694, 1994.
54. Vojtesek, B., Dolezalova, H., Lauerova, L., Svitakova, M., Havlik, P., Kovarik, J., Midgley, C. A., and Lane, D. P. Conformational changes in p53 analysed using new antibodies to the core DNA binding domain of the protein. *Oncogene*, **10**: 389–393, 1995.
55. Levesque, M. A., Dcosta, M., and Diamandis, E. P. p53 protein is absent from the serum of patients with lung cancer. *Br. J. Cancer*, **74**: 1434–1440, 1996.
56. Hassapoglidou, S., Diamandis, E. P., and Sutherland, D. J. A. Quantification of p53 protein in tumor cell lines, breast tissue extracts and serum with time-resolved immunofluorometry. *Oncogene*, **8**: 1501–1509, 1993.
57. Cho, Y. J., Gorina, S., Jeffrey, P. D., and Pavletich, N. P. Crystal structure of a p53 tumor suppressor DNA complex: understanding tumorigenic mutations. *Science (Washington DC)*, **265**: 346–355, 1994.
58. Portefaix, J. M., Thébault, S., Bourgain-Guglielmetti, F., Del Rio, M., Granier, M., Mani, J. C., Teulon-Navaro, I., Nicolas, M., Soussi, T., and Pau, B. Critical residues of epitopes recognized by several anti-p53 monoclonal antibodies correspond to key residues of p53 involved in interaction with the mdm2 protein. *J. Immunol. Methods*, in press, 2000.
59. Kaghad, M., Bonnet, H., Yang, A., Creancier, L., Biscan, J. C., Valent, A., Minty, A., Chalou, P., Lelias, J. M., Dumont, X., Ferrara, P., McKeon, F., and Caput, D. Monoallelically expressed gene related to p53 at 1p36, a region frequently deleted in neuroblastoma and other human cancers. *Cell*, **90**: 809–819, 1997.
60. Osada, M., Ohba, M., Kawahara, C., Ishioka, C., Kanamaru, R., Katoh, I., Ikawa, Y., Nimura, Y., Nakagawara, A., Obinata, M., and Ikawa, S. Cloning and functional analysis of human p51, which structurally and functionally resembles p53. *Nat. Med.*, **4**: 839–843, 1998.
61. Trink, B., Okami, K., Wu, L., Sriuranpong, V., Jen, J., and Sidransky, D. A new human p53 homologue. *Nat. Med.*, **4**: 747–748, 1998.
62. Schmale, H., and Bamberger, C. A novel protein with strong homology to the tumor suppressor p53. *Oncogene*, **15**: 1363–1367, 1997.
63. Roth, J., Dittmer, D., Rea, D., Tartaglia, J., Paoletti, E., and Levine, A. J. p53 as a target for cancer vaccines: recombinant canarypox virus vectors expressing p53 protect mice against lethal tumor cell challenge. *Proc. Natl. Acad. Sci. USA*, **93**: 4781–4786, 1996.
64. Peyrat, J. P., Bonnetterre, J., Lubin, R., Vanlemmens, L., Fournier, J., and Soussi, T. Prognostic significance of circulating p53 antibodies in patients undergoing surgery for locoregional breast cancer. *Lancet*, **345**: 621–622, 1995.
65. Mudenda, B., Green, J. A., Green, B., Jenkins, J. R., Robertson, L., Tarunina, M., and Leinster, S. J. The relationship between serum p53 autoantibodies and characteristics of human breast cancer. *Br. J. Cancer*, **69**: 1115–1119, 1994.
66. Lenner, P., Wiklund, F., Emdin, S. O., Amerlov, C., Eklund, C., Hallmans, G., Zentgraf, H., and Dillner, J. Serum antibodies against p53 in relation to cancer risk and prognosis in breast cancer: a population-based epidemiological study. *Br. J. Cancer*, **79**: 927–932, 1999.
67. Willsher, P. C., Pinder, S. E., Robertson, L., Nicholson, R. I., Ellis, I. O., Bell, J. A., Blamey, R. W., Green, J. A., and Robertson, J. F. R. The significance of p53 autoantibodies in the serum of patients with breast cancer. *Anticancer Res.*, **16**: 927–930, 1996.
68. Porzolt, F., M., S., Hoher, D., Mucbe, D., Gaus, W., and Montenarh, M. Biological relevance of auto-antibodies against p53 in patients with metastatic breast cancer. *Onkologie*, **17**: 402–408, 1994.
69. Maehara, Y., Kakeji, Y., Watanabe, A., Baba, H., Kusumoto, H., Kohnoe, S., and Sugimachi, K. Clinical implications of serum anti-p53 antibodies for patients with gastric carcinoma. *Cancer (Phila.)*, **85**: 302–308, 1999.
70. Wu, C. W., Lin, Y. Y., Chen, G. D., Chi, C. W., Carbone, D. P., and Chen, J. Y. Serum anti-p53 antibodies in gastric adenocarcinoma patients are associated with poor prognosis, lymph node metastasis and poorly differentiated nuclear grade. *Br. J. Cancer*, **80**: 483–488, 1999.
71. Shiota, G., Ishida, M., Noguchi, N., Takano, Y., Oyama, K., Okubo, M., Katayama, S., Harada, K., Hori, K., Ashida, K., Kishimoto, Y., Hosoda, A., Suou, T., Ito, H., and Kawasaki, H. Clinical significance of serum p53 antibody in patients with gastric cancer. *Res. Commun. Mol. Pathol. Pharmacol.*, **99**: 41–51, 1998.
72. Kressner, U., Glimelius, B., Bergstrom, R., Pahlman, L., Larsson, A., and Lindmark, G. Increased serum p53 antibody levels indicate poor prognosis in patients with colorectal cancer. *Br. J. Cancer*, **77**: 1848–1851, 1998.
73. Houbiers, J. G. A., Vanderburg, S. H., Vandewatering, L. M. G., Tollenaar, R. A. E. M., Brand, A., Vandevelde, C. J. H., and Melief, C. J. M. Antibodies against p53 are associated with poor prognosis of colorectal cancer. *Br. J. Cancer*, **72**: 637–641, 1995.
74. Angelopoulou, K., Stratis, M., and Diamandis, E. P. Humoral immune response against p53 protein in patients with colorectal carcinoma. *Int. J. Cancer*, **70**: 46–51, 1997.
75. Laudanski, J., Burzykowski, T., Niklinska, W., Chyczewski, K., Furman, M., and Niklinski, J. Prognostic value of serum p53 antibodies in patients with resected non-small cell lung cancer. *Lung Cancer*, **22**: 191–200, 1998.
76. Lai, C.-L., Tsai, C.-M., Tsai, T.-T., Ing-Tiau Kuo, B., Chang, K.-T., Fu, H.-T., Perng, R.-P., and Chen, J.-Y. Presence of serum anti-p53 antibodies is associated with pleural effusion and poor prognosis in lung cancer patients. *Clin. Cancer Res.*, **4**: 3025–3030, 1998.
77. Komiya, T., Hirashima, T., Takada, M., Masuda, N., Yasumitsu, T., Nakagawa, K., Hosono, Y., Kikui, M., Tsuji, S., Fukuoka, M., and Kawase, I. Prognostic signifi-

- cance of serum p53 antibodies in squamous cell carcinoma of the lung. *Anticancer Res.*, 17: 3721–3724, 1997.
78. Zalcman, G., Trédaniel, G., Schlichtholz, B., Urban, T., Milleron, B., Lubin, R., Meignin, V., Couderc, J. L., Hirsch, A., and Soussi, T. Prognostic significance of serum p53 antibody in patients with limited-stage small cell lung cancer. *Int. J. Cancer*, 89: 81–86, 2000.
 79. Rosenfeld, M. R., Malats, N., Schramm, L., Graus, F., Cardenal, F., Vinolas, N., Rosell, R., Tora, M., Real, F. X., Posner, J. B., and Dalmau, J. Serum anti-p53 antibodies and prognosis of patients with small-cell lung cancer. *J. Natl. Cancer Inst.*, 89: 381–385, 1997.
 80. Bourhis, J., Lubin, R., Roche, B., Koscielny, S., Bosq, J., Dubois, I., Talbot, M., Marandas, P., Schwaab, G., Wibault, P., Luboinski, B., Eschwege, F., and Soussi, T. Analysis of p53 serum antibodies in patients with head and neck squamous cell carcinoma. *J. Natl. Cancer Inst.*, 88: 1228–1233, 1996.
 81. Werner, J. A., Gottschlich, S., Folz, B. J., Goeroegh, T., Lippert, B. M., Maass, J. D., and Rudert, H. p53 serum antibodies as prognostic indicator in head and neck cancer. *Cancer Immunol. Immunother.*, 44: 112–116, 1997.
 82. Hammel, P., Boissier, B., Chaumette, M. T., Piedbois, P., Rotman, N., Kouyoumdjian, J. C., Lubin, R., Delchier, J. C., and Soussi, T. Detection and monitoring of serum p53 antibodies in patients with colorectal cancer. *Gut*, 40: 356–361, 1997.
 83. Zalcman, G., Schlichtholz, B., Trédaniel, J., Urban, T., Lubin, R., Dubois, I., Milleron, B., Hirsh, A., and Soussi, T. Monitoring of p53 auto antibodies in lung cancer during therapy: relationship to response to treatment. *Clin. Cancer Res.*, 4: 1359–1366, 1998.
 84. Saffroy, R., Lelong, J. C., Azoulay, D., Salvucci, M., Reynes, M., Bismuth, H., Debuire, B., and Lemoine, A. Clinical significance of circulating anti-p53 antibodies in European patients with hepatocellular carcinoma. *Br. J. Cancer*, 79: 604–610, 1999.
 85. Polge, A., Bourgaux, J. F., Bancel, E., Pignodel, C., Boyer, J. C., Poirey, S., DeBormier, B. M., Balmes, J. L., and Bali, J. P. p53 and follow-up of colorectal adenocarcinomas. *Dig. Dis. Sci.*, 43: 1434–1442, 1998.
 86. Gadducci, A., Ferdeghini, M., Buttitta, F., Cosio, S., Fanucchi, A., Annicchiarico, C., and Genazzani, A. R. Serum anti-p53 antibodies in the follow-up of patients with advanced ovarian carcinoma. *Anticancer Res.*, 18: 3763–3766, 1998.
 87. Shimada, H., Arima, M., Nakajima, K., Koide, Y., Okazumi, S., Matsubara, H., Miyazawa, Y., Takeda, A., Hayashi, H., Yoshida, T., Ochiai, T., and Isono, K. "Detection of serum p53 antibodies in mucosal esophageal cancer and negative conversion after treatment" (Letter). *Am. J. Gastroenterol.*, 93: 1388–1389, 1998.
 88. Lubin, R., Zalcman, G., Bouchet, L., Trédaniel, J., Legros, Y., Cazals, D., Hirsh, A., and Soussi, T. Serum p53 antibodies as early markers of lung cancer. *Nat. Med.*, 1: 701–702, 1995.
 89. Bennett, W. P., Colby, T. V., Travis, W. D., Borkowski, A., Jones, R. T., Lane, D. P., Metcalf, R. A., Samet, J. M., Takeshima, Y., Gu, J. R., Vahakangas, K. H., Soini, Y., Paaikko, P., Welsh, J. A., Trump, B. F., and Harris, C. C. p53 protein accumulates frequently in early bronchial neoplasia. *Cancer Res.*, 53: 4817–4822, 1993.
 90. Trivers, G. E., De Benedetti, V. M. G., Cawley, H. L., Caron, G., Harrington, A. M., Bennet, W. P., Jett, J. R., Colby, T. V., Tazelaar, H., Pairolo, P., Miller, R. D., and Harris, C. C. Anti-p53 antibodies in sera from patients with chronic obstructive pulmonary disease can predate a diagnosis of cancer. *Clin. Cancer Res.*, 2: 1767–1775, 1996.
 91. Cawley, H. M., Meltzer, S. J., De Benedetti, V. M., Hollstein, M. C., Muehlbauer, K. R., Liang, L., Bennett, W. P., Souza, R. F., Greenwald, B. D., Cottrell, J., Salabas, A., Bartsch, H., and Trivers, G. E. Anti-p53 antibodies in patients with Barrett's esophagus or esophageal carcinoma can predate cancer diagnosis. *Gastroenterology*, 115: 19–27, 1998.
 92. Ralhan, R., Nath, N., Agarwal, S., Mathur, M., Wasylyk, B., and Shukla, N. K. Circulating p53 antibodies as early markers of oral cancer: correlation with p53 alterations. *Clin. Cancer Res.*, 4: 2147–2152, 1998.
 93. Parashar, K., and Sobti, R. C. A study on p53 protein and anti-p53 antibodies in the sera of patients with oesophageal cancer. *Mutat. Res. Fundam. Mol. Mech. Mutat.*, 422: 271–277, 1998.
 94. Partanen, R., Koskinen, H., Oksa, P., Hemminki, K., Carney, W., Smith, S., and Brandtrauf, P. Serum oncoproteins in asbestosis patients. *Clin. Chem.*, 41: 1844–1847, 1995.
 95. Luo, J. C., Zehab, R., Anttila, S., Ridana, M., Husgafvel-Pursiainen, K., Vainio, H., Carney, W., DeVivo, I., Milling, C., and Brandt-Rauf, P. W. Detection of serum p53 protein in lung cancer patients. *J. Occup. Med.*, 36: 155–160, 1994.
 96. Luo, J. C., Neugot, A. I., Garbowski, G., Forde, K. A., Treat, M., Smith, S., Carney, W. P., and Brandt-Rauf, P. W. Levels of p53 antigen in the plasma of patients with adenomas and carcinomas of the colon. *Cancer Lett.*, 8: 235–240, 1995.
 97. Wong, S. K., Seowchoen, F., Leong, A. P. K., Ho, Y. H., and Aw, S. E. Mutant plasma p53 protein levels: prognostication in colorectal carcinoma. *Br. J. Surg.*, 84: 664–665, 1997.
 98. Zusman, I., Zusman, R., Sandler, B., Korol, D., Gurevich, P., Smirnov, P., Bass, D., Or, A., Shani, A., Idelevich, E., Huszar, M., Tendler, Y., Klein, T., Shezen, E., and Glick, J. HPLC determination of serum levels of soluble p53 antigen as a new method for colon cancer detection, and its clinical implication: clinical study. *Int. J. Oncol.*, 8: 1249–1256, 1996.
 99. Takahashi, M., Chen, W., Byrd, D. R., Disis, M. L., Huseby, E. S., Qin, H., McCahill, L., Nelson, H., Shimada, H., Okuno, K., Yasutomi, M., Peace, D. J., and Cheever, M. A. Antibody to ras proteins in patients with colon cancer. *Clin. Cancer Res.*, 1: 1071–1077, 1995.
 100. Sorokine, I., Ben-Mahrez, K., Bracone, A., Thierry, D., Ishii, S., Imamoto, F., and Kohiyama, M. Presence of circulating anti-*c-myc* oncogene product antibodies in human sera. *Int. J. Cancer*, 47: 665–669, 1991.
 101. Yamamoto, A., Shimizu, E., Ogura, T., and Sone, S. Detection of auto-antibodies against *L-myc* oncogene products in sera from lung cancer patients. *Int. J. Cancer*, 69: 283–289, 1996.
 102. Ben-Mahrez, K., Thierry, D., Sorokine, I., Danna-Muller, A., and Kohiyama, M. Detection of circulating antibodies against c-myc protein in cancer patient sera. *Br. J. Cancer*, 57: 529–534, 1988.
 103. Selter, H., Amela-Neuschwander, S., Villena-Heinsen, C., and Montenarh, M. Antibodies against murine double minute-2 (mdm2) in sera of patients with various gynaecological diseases. *Cancer Lett.*, 96: 111–115, 1995.
 104. Disis, M. L., Pupa, S. M., Gralow, J. R., Dittadi, R., Menard, S., and Cheever, M. A. High-titer HER-2/neu protein-specific antibody can be detected in patients with early-stage breast cancer. *J. Clin. Oncol.*, 15: 3363–3367, 1997.
 105. Ward, R. L., Hawkins, N. J., Coomber, D., and Disis, M. L. Antibody immunity to the HER-2/neu oncogenic protein in patients with colorectal cancer. *Hum. Immunol.*, 60: 510–515, 1999.
 106. Chen, Y. T., Scanlan, M. J., Sahin, U., Tureci, O., Gure, A. O., Tsang, S., Williamson, B., Stockert, E., Pfreundschuh, M., and Old, L. J. A testicular antigen aberrantly expressed in human cancers detected by autologous antibody screening. *Proc. Natl. Acad. Sci. USA*, 94: 1914–1918, 1997.
 107. Tureci, O., Sahin, U., and Pfreundschuh, M. Serological analysis of human tumor antigens: molecular definition and implications. *Mol. Med. Today*, 3: 342–349, 1997.
 108. Mao, L., Lee, J. S., Kurie, J. M., Fan, Y. H., Lippman, S. M., Lee, J. J., Ro, J. Y., Broxson, A., Yu, R., Morice, R. C., Kemp, B. L., Khuri, F. R., Walsh, G. L., Hittelman, W. N., and Hong, W. K. Clonal genetic alterations in the lungs of current and former smokers. *J. Natl. Cancer Inst.*, 89: 857–862, 1997.
 109. Rusin, M., Butkiewicz, D., Malusecka, E., Zborek, A., Harasim, J., Czyzewski, K., Bennett, W. P., Shields, P. G., Weston, A., Welsh, J. A., Krzyzowska-Gruca, S., Chorazy, M., and Harris, C. C. Molecular epidemiological study of non-small-cell lung cancer from an environmentally polluted region of Poland. *Br. J. Cancer*, 80: 1445–1452, 1999.
 110. Volkmann, M., Muller, M., Hofmann, W. J., Meyer, M., Hagemstein, J., Rath, U., Kommerell, B., Zentgraf, H., and Galle, P. R. The humoral immune response to p53 in patients with hepatocellular carcinoma is specific for malignancy and independent of the α -fetoprotein status. *Hepatology*, 18: 559–565, 1993.
 111. Green, J. A., Mudenda, B., Jenkins, J., Leinster, S. J., Tarunina, M., Green, B., and Robertson, L. Serum p53 auto-antibodies: incidence in familial breast cancer. *Eur. J. Cancer*, 30A: 580–584, 1994.
 112. Wild, C. P., Ridana, M., Anttila, S., Lubin, R., Soussi, T., Husgafvel-Fursiainen, K., and Vainio, H. p53 antibodies in the sera of lung cancer patients: comparison with p53 mutation in the tumour tissue. *Int. J. Cancer*, 64: 176–181, 1995.
 113. Mayerhofer, K., Tempfer, C., Kucera, E., Hefler, L., Zeisler, H., Kainz, C., Zeilinger, R., and Sliutz, G. Humoral p53 antibody response is a prognostic parameter in ovarian cancer. *Anticancer Res.*, 19: 875–878, 1999.
 114. Iizasa, T., Fujisawa, T., Saitoh, Y., Hiroshima, K., and Ohwada, H. Serum anti-p53 autoantibodies in primary resected non-small-cell lung carcinoma. *Cancer Immunol. Immunother.*, 46: 345–349, 1998.
 115. Hallak, R., Mueller, J., Lotter, O., Gansauge, S., Gansauge, F., Jumma, M. E., Montenarh, M., Safi, F., and Beger, H. p53 genetic alterations, protein expression and autoantibodies in human colorectal carcinoma: a comparative study. *Int. J. Oncol.*, 12: 785–791, 1998.
 116. Shiota, G., Kishimoto, Y., Suyama, A., Okubo, M., Katayama, S., Harada, K., Ishida, M., Hori, K., Suou, T., and Kawasaki, H. Prognostic significance of serum anti-p53 antibody in patients with hepatocellular carcinoma. *J. Hepatol.*, 27: 661–668, 1997.
 117. Green, J. A., Roberston, L. J., Campbell, I. R., and Jenkins, J. Expression of the p53 gene and presence of serum autoantibodies in ovarian cancer: correlation with differentiation. *Cancer Detect. Prev.*, 19: 151–155, 1995.
 118. Raedle, J., Roth, W. K., Oremek, G., Caspary, W. F., and Zeuzem, S. α -fetoprotein and p53 autoantibodies in patients with chronic hepatitis C. *Dig. Dis. Sci.*, 40: 2587–2594, 1995.
 119. Maass, J. D., Gottschlich, S., Lippert, B. M., Niemann, A. M., Goeroegh, T., and Werner, J. A. Antibodies in response to the cellular protein p53 in patients with squamous cell carcinoma of the head and neck. *Laryngo-Rhino-Otol.*, 75: 53–56, 1996.
 120. Coomber, D., Hawkins, N. J., Clark, M., Meagher, A., and Ward, R. L. Characterisation and clinicopathological correlates of serum anti-p53 antibodies in breast and colon cancer. *J. Cancer Res. Clin. Oncol.*, 122: 757–762, 1996.
 121. Shibata, Y., Kotanogi, H., Andoh, H., Koyama, K., Itoh, H., and Kudo, S. Detection of circulating anti p53 antibodies in patients with colorectal carcinoma and the antibody's relation to clinical factors. *Dis. Colon Rectum*, 39: 1269–1274, 1996.
 122. Ryder, S. D., Rizzi, P. M., Volkmann, M., Metivier, E., Pereira, L. M. M. B., Galle, P. R., Naoumov, N. V., Zentgraf, H., and Williams, R. Use of specific ELISA for the detection of antibodies directed against p53 protein in patients with hepatocellular carcinoma. *J. Clin. Pathol.*, 49: 295–299, 1996.
 123. Gansauge, S., Gansauge, F., Negri, G., Galle, P., Muller, J., Nussler, A. K., Poch, B., and Beger, H. G. The role of anti-p53-autoantibodies in pancreatic disorders. *Int. J. Pancreatol.*, 19: 171–178, 1996.
 124. Stoeger, Z., Evron, E., Goland, S., Shani, A., Wolkowicz, R., Goldfinger, N., Rotter, V., and Fogel, M. Anti-p53 antibodies in colon cancer patients. *Ann. NY. Acad. Sci.*, 815: 496–498, 1997.
 125. Friedrich, R. E., BartelFriedrich, S., Plambeck, K., Bahlo, M., and Klapdor, R. p53 auto-antibodies in the sera of patients with oral squamous cell carcinoma. *Anticancer Res.*, 17: 3183–3184, 1997.

126. Wollenberg, B., Jan, N. V., Pitzke, P., Reiter, W., and Stieber, P. Anti-p53 antibodies in serum of smokers and head and neck cancer patients. *Anticancer Res.*, *17*: 413–418, 1997.
127. Crawford, L. V., Pim, D. C., and Lamb, P. The cellular protein p53 in human tumors. *Mol. Biol. Med.*, *2*: 261–272, 1984.
128. Angelopoulou, K., and Diamandis, E. P. Autoantibodies against the p53 tumor suppressor gene product quantified in cancer patients serum with time-resolved immunofluorometry. *The Cancer J.*, *6*: 315–321, 1993.
129. Regidor, P. A., Regidor, M., Callies, R., and Schindler, A. E. Detection of p53 auto-antibodies in the sera of breast cancer patients with new recurrence using an ELISA assay. *Eur. J. Gynaecol. Oncol.*, *17*: 192–199, 1996.
130. Guinee, D. G., Travis, W. D., Trivers, G. E., Debenedetti, V. M. G., Cawley, H., Welsh, J. A., Bennett, W. P., Jett, J., Colby, T. V., Tazelaar, H., Abbondanzo, S. L., Pairolero, P., Trastek, V., Caporaso, N. E., Liotta, L. A., and Harris, C. C. Gender comparisons in human lung cancer: analysis of p53 mutations, anti-p53 serum antibodies and C-erbB-2 expression. *Carcinogenesis (Lond.)*, *16*: 993–1002, 1995.
131. Bergqvist, M., Brattstrom, D., Larsson, A., Holmertz, J., Hesselius, P., Rosenberg, L., Wagenius, G., and Brodin, O. p53 auto-antibodies in non-small cell lung cancer patients can predict increased life expectancy after radiotherapy. *Anticancer Res.*, *18*: 1999–2002, 1998.
132. Mitsudomi, T., Suzuki, S., Yatabe, Y., Nishio, M., Kuwabara, M., Gotoh, K., Hatoaka, S., Shinoda, M., Suyama, M., Ogawa, M., Takahashi, T., and Ariyoshi, Y. Clinical implications of p53 autoantibodies in the sera of patients with non-small-cell lung cancer. *J. Natl. Cancer Inst.*, *90*: 1563–1568, 1998.
133. Royer, B., CazalsHatem, D., Sibilia, J., Agbalika, F., Cayuela, J. M., Soussi, T., Maloisel, F., Clauvel, J. P., Brouet, J. C., and Mariette, X. Lymphomas in patients with Sjögren's syndrome are marginal zone B-cell neoplasms, arise in diverse extranodal and nodal sites, and are not associated with viruses. *Blood*, *90*: 766–775, 1997.
134. Nakajima, K., Suzuki, T., Shimada, H., Hayashi, H., Takeda, A., and Ochiai, T. Detection of preoperative serum anti-p53 antibodies in gastric cancer. *Tumor Biol.*, *20*: 147–152, 1999.
135. Cough, M. J., Koch, M., Brennan, J. A., and Sidransky, D. The humoral response to oncoprotein p53 in head and neck cancer. *Head Neck*, *16*: 495, 1994.
136. Lavieille, J. P., Righini, C., Reyt, E., Brambilla, C., and Riva, C. Implications of p53 alterations and anti-p53 antibody response in head and neck squamous cell carcinomas. *Oral Oncol.*, *34*: 84–92, 1998.
137. Maass, J. D., Gottschlich, S., Goeroegh, T., Lippert, B. M., and Werner, J. A. Head and neck cancer and p53-immunogenicity. *Anticancer Res.*, *17*: 2873–2874, 1997.
138. Maass, J. D., Gottschlich, S., Niemann, A. M., Goeroegh, T., Lippert, B. M., and Werner, J. A. Serum p53-antibodies in head and neck cancer. *In: J. A. Werner, B. M. Lippert, and H. H. Lippert (eds.), Head and Neck Cancer: Advances in Basic Research*, Vol. 1114, pp. 351–355. London: Elsevier, 1996.
139. Lavieille, J. P., Lubin, R., Soussi, T., Reyt, E., Brambilla, C., and Riva, C. Analysis of p53 antibody response in patients with squamous cell carcinoma of the head and neck. *Anticancer Res.*, *16*: 2385–2388, 1996.
140. Collet, B., Raoul, J. L., LeBerre, N., Heresbach, D., Meritte, H., Quillien, V., and DeCertaïnes, J. D. Serum anti-p53 antibodies in patients with squamous cell carcinoma of the esophagus: comparison with p53 alterations and lymph node invasion. *Int. J. Oncol.*, *11*: 617–621, 1997.
141. Shimada, H., Nakajima, K., Ochiai, T., Koide, Y., Okazumi, S. I., Matsubara, H., Takeda, A., Miyazawa, Y., Arima, M., and Isono, K. Detection of serum p53 antibodies in patients with esophageal squamous cell carcinoma: correlation with clinicopathologic features and tumor markers. *Oncol. Rep.*, *5*: 871–874, 1998.
142. Edis, C., Kahler, C., Klotz, W., Herold, M., Feichtinger, H., Konigsreiner, A., Margreiter, R., Jäschke, W., and Vogel, W. A comparison between α -fetoprotein and p53 antibodies in the diagnosis of hepatocellular carcinoma. *Transplant. Proc.*, *30*: 780–781, 1998.
143. Gadducci, A., Ferdeghini, M., Buttitta, F., Cosio, S., Fanucchi, A., Annicchiarico, C., Galetti, O., Bevilacqua, G., and Genazzani, A. R. Assessment of the prognostic relevance of serum anti-p53 antibodies in epithelial ovarian cancer. *Gynecol. Oncol.*, *72*: 76–81, 1999.
144. Gadducci, A., Ferdeghini, M., Buttitta, F., Fanucchi, A., Annicchiarico, C., Prontera, C., Bevilacqua, G., and Genazzani, A. R. Preoperative serum antibodies against the p53 protein in patients with ovarian and endometrial cancer. *Anticancer Res.*, *16*: 3519–3523, 1996.
145. Angelopoulou, K., Rosen, B., Stratis, M., Yu, H., Solomou, M., and Diamandis, E. P. Circulating antibodies against p53 protein in patients with ovarian carcinoma: correlation with clinicopathologic features and survival. *Cancer (Phila.)*, *78*: 2146–2152, 1996.
146. Marxsen, J., Schmiegel, W., Roder, C., Harder, R., Juhl, H., Hennebruns, D., Kremer, B., and Kalthoff, H. Detection of the anti-p53 antibody response in malignant and benign pancreatic disease. *Br. J. Cancer*, *70*: 1031–1034, 1994.
147. Laurent-Puig, P., Lubin, R., Semhouncloux, S., Pelletier, G., Fourre, C., Ducreux, M., Briantais, M. J., Buffet, C., and Soussi, T. Antibodies against p53 protein in serum of patients with benign or malignant pancreatic and biliary diseases. *Gut*, *36*: 455–458, 1995.
148. Raedle, J., Oremek, G., Welker, M., Roth, W. K., Caspary, W. F., and Zeuzem, S. p53 autoantibodies in patients with pancreatitis and pancreatic carcinoma. *Pancreas*, *13*: 241–246, 1996.
149. Chen, Y. M. A., Chen, S. H., Fu, C. Y., Chen, J. Y., and Osame, M. Antibody reactivities to tumor-suppressor protein p53 and HTLV-I Tof Rex and Tax in HTLV-I-infected people with differing clinical status. *Int. J. Cancer*, *71*: 196–202, 1997.
150. Preudhomme, C., Vanrumbeke, M., Detourmignies, L., Facon, T., Lepelley, P., Soussi, T., and Fenaux, P. Very low incidence of p53 antibodies in adult non-Hodgkin's lymphoma and multiple myeloma. *Br. J. Haematol.*, *100*: 184–186, 1998.
151. Pulkki, K., Ollikainen, H., Rajamäki, A., and Pelliniemi, T. T. serum p53 oncoprotein and p53 antibodies as prognostic factor in multiple myeloma (MM). *Blood*, *86*: 64a, 1995.