

In Vivo and *in Vitro* Prolactin Secretion by Transplanted Rat "Mammotropic" Pituitary Tumors*

P. K. TALWALKER,† A. RATNER, AND J. MEITES

(Department of Physiology, Michigan State University, East Lansing, Michigan)

SUMMARY

The functional, transplanted, autonomous rat pituitary "mammotropic" tumor (Furth, MtT.F₄) was compared with normal rat pituitary for capacity to elaborate prolactin *in vivo* and *in vitro*. The prolactin content of transplanted anterior pituitary (AP) and MtT.F₄, as measured by the intradermal pigeon crop technic, was 1.6 and 4.8 per cent, respectively, of that present in the AP *in situ*. Considerable amounts of prolactin were detected in the blood plasma of rats bearing an MtT.F₄, the levels being higher in rats with larger than in rats with smaller tumors. Prolactin activity could not be detected in the blood plasma of normally cycling, estrogen-treated, pregnant, or lactating rats, or in rats bearing a single AP transplant underneath the kidney capsule. The amount of prolactin released into the medium by MtT.F₄ tissue during 3 days of culture was 8–31 times greater than that present initially, indicating active synthesis and release of prolactin by the tumor. These results show that the MtT.F₄, like nontumorous AP transplants removed from hypothalamic inhibition, synthesizes prolactin and releases it at a rapid rate, leaving little in the tissue itself.

The pioneer work of Furth (7) has led to the identification and isolation of a variety of transplantable hormone-secreting rat and mouse pituitary tumors. One of these is a chromophobic, autonomous rat pituitary "mammotropic" tumor, strain F₄ (MtT.F₄), originally induced by estrogen treatment (6, 9). Following transplantation in the rat this tumor appears to elaborate prolactin, STH, and ACTH, but no FSH-LH or TSH (2, 3, 7, 9, 21).

Animal cell cultures rarely perform differentiated functions characteristic of the tissue of origin; this has been particularly true of cultures of endocrine tissue. However, in our laboratory (13, 14) as well as others (11, 15) it has recently been observed that cultures of rat anterior pituitary (AP) actively release prolactin into the medium, whereas release of all other hormones is markedly diminished. The purpose of this study was to test the capacity of the MtT.F₄ to secrete prolactin *in vivo* and *in vitro* as compared with that of normal rat AP. A report of some of the present work has previously been published in abstract form (19).

MATERIALS AND METHODS

Animals.—Mature, virgin female, highly inbred Fischer¹ rats of the CDF strain, 3–4 months old, were used in this

* Published with the approval of the Director of the Michigan Agricultural Experiment Station as journal article No. 3365.

This investigation was supported in part by NIH grant AM 4784-04, the Michigan Cancer Foundation, and an Institutional Research grant from the American Cancer Society.

† Present address: Research Division, Pharmaceutical Dept., CIBA Limited, Basel, Switzerland.

¹ Charles River Breeding Laboratories, Brookline, Mass.

Received for publication April 25, 1964.

study. The rats were maintained in a temperature-controlled (75 ± 1° F.) and artificially illuminated (14 hr/day) room. They were fed ad libitum on Wayne Lab Blox pellets supplemented with canned (Dash) dog food. White King Squabs, 5–8 weeks old, were used for all prolactin assays.

Prolactin assay.—The sensitive intradermal pigeon-crop method was utilized. The test material was injected over one side of the crop-sac, whereas standard NIH prolactin was injected over the other side of the crop-sac in the same pigeons. Prolactin activity was expressed as IU/100 mg of tissue (wet weight) or IU/100 ml plasma. The details of the assay procedure have been described elsewhere (14).

Transplantation and assays of MtT.F₄ and AP.—Serial transplantation of MtT.F₄ was carried out in our laboratory. The tumor transplants were made by giving injections to rats subcutaneously, in the back of the neck, of 0.1 ml. tumor mince in an equal volume of medium 199 (pH 7.4) containing streptomycin² (100 µg/ml) and penicillin G potassium² (100 U/ml). In some rats injections were made at two sites, resulting in the formation of two tumors. The tumors were palpable within 4–5 weeks. When the tumor reached 1–2 cm. in diameter it was removed, and a portion of it was used for cultures and for histological examination; another part of it was weighed, homogenized with physiological saline, and assayed for prolactin activity. In a total, six tumors (passage 41) from six rats were assayed separately for prolactin activity.

² Nutritional Biochemicals Corporation, Cleveland, Ohio.

TABLE 1
PROLACTIN ASSAYS OF FRESH AP, AP TRANSPLANTS,
AND MtT.F₄ TRANSPLANTS

Tissue	No. donor rats	No. assays	Total no. pigeons	Prolactin IU/100 mg. tissue (wet weight)	Comparative prolactin content (per cent)
Fresh AP	16	4	18	1.2 ± 0.07*	100
AP transplant†	20	2	8	0.02	1.6
MtT.F ₄ transplant	6	6	30	0.06 ± 0.01	4.8

AP = anterior pituitary; MtT.F₄ = mammatropic pituitary tumor, strain F₄.

* Mean ± standard error of mean.

† A single anterior pituitary was transplanted underneath the kidney capsule of each rat. Ten days later the transplant was removed and assayed for prolactin activity.

Intact rats were given implants underneath the kidney capsule of one AP from each of the donor rats of the same strain and approximately of the same age group. Ten days later the AP transplants were removed, trimmed of adhering tissue under a dissecting microscope, pooled from ten rats each, weighed, homogenized with saline, and assayed for prolactin activity. Some of the AP transplants were used for histological examination. The AP from normally cycling rats was also removed, homogenized with saline, and assayed for prolactin activity.

Collection of rat blood and assay for prolactin activity.—Rats were anesthetized with ether, and blood was collected in a heparinized syringe from the posterior abdominal artery. After centrifugation the plasma was dialyzed, lyophilized, and assayed for prolactin activity. Rats in different states were used as blood donors, as indicated in Table 2.

Cultures of MtT.F₄.—A modification of the watch glass technic (5) was used, in which sterile, plastic (3.5 × 1 cm.) Petri dishes³ were employed. Platforms were formed from stainless steel mesh rectangles (1 × 2.3 cm.), bent at each end to make a platform 4 mm. high. These were placed in Petri dishes with 3 ml. of culture medium 199⁴ containing 2 U insulin,⁵ 100 U penicillin G potassium, and 100 μg streptomycin/ml. For some of the cultures calf, rabbit, or horse serum,⁶ or calf serum and chicken embryo extract,⁷ was added to medium 199, as shown in Table 3.

The MtT.F₄ was removed and placed in a Petri dish (10 × 1.5 cm.) containing a few drops of medium. The tumor was cut into small pieces each about 2–3 mm. in diameter, and two such explants were placed on a strip of lens paper (1.5 × 3 cm.), which was then placed on top of the platform inside the Petri dish. The tumor explants were cultured for 3 or 6 days at 35° C. in 95 per cent O₂–5 per cent CO₂ atmosphere. Six MtT.F₄ were used to carry out a total of 148 cultures. At the end of the culture period medium from eight to ten culture dishes was pooled, dialyzed, lyophilized, and assayed for

prolactin activity. The explants were fixed in Bouins fluid, sectioned at 6 μ, and stained with hematoxylin and eosin. The details of the culture procedure have been described elsewhere (14).

To determine whether prolactin activity could be detected in medium without MtT.F₄, medium 199 alone or medium 199 containing 20 per cent calf, rabbit, or horse serum was incubated for 3 days. At termination the medium was dialyzed, lyophilized, and assayed for prolactin activity. Also, pregnant rat mammary gland and pigeon crop gland were cultured separately for 3 days and the medium was similarly treated and assayed for prolactin. In another experiment, standard NIH ovine prolactin was added to medium 199 with or without 20 per cent calf serum, and incubated for 3 days to ascertain the effects of incubation alone on prolactin activity.

RESULTS

The average prolactin content (Table 1) of MtT.F₄ and AP transplants was found to be 0.06 and 0.02 IU/100 mg of tissue (wet weight), respectively, as compared with 1.2 IU/100 mg of fresh AP. This represents 4.8 and 1.6 per cent, respectively, of the content of prolactin present in fresh AP.

No prolactin activity was detected in blood plasma (Table 2) of 3- to 4-month-old mature, female, cycling rats, female rats given injections daily of 10 μg. of estradiol for 10 days, 11- to 12-day pregnant rats, 10-day post-partum lactating rats, or rats with a single AP transplant of 10 days' duration. However, considerable amounts of prolactin could be detected in the blood plasma of rats bearing an MtT.F₄. The plasma prolactin activity was found to be higher in rats bearing large as compared with rats with smaller tumors.

Explants of MtT.F₄ actively released prolactin into the culture medium (Table 3). The amount of prolactin released during 3 days of culture (0.5–1.9 IU/100 mg MtT.F₄) was 8–31 times more than the initial content of prolactin in MtT.F₄ (0.06 IU/100 mg MtT.F₄). Prolactin was also detected in the medium from monolayer cultures of MtT.F₄ incubated for 22 days (unpublished). Neither incubation of the medium alone for 3 days without MtT.F₄ nor medium from cultures of rat mammary gland or pigeon crop gland contained prolactin. Prolactin potency was not altered when standard NIH ovine prolactin was incubated for 3 days in medium 199 with or without 20 per cent calf serum.

Histological examination of the MtT.F₄ explants at the end of culture showed variable degrees of maintenance. The best maintenance of explants, comparable to fresh MtT.F₄, was obtained when 20 per cent calf serum was added to medium 199. The histological appearance of the AP transplants underneath the kidney capsule, 10 days after transplantation, indicated that most of the tissue was viable. These AP transplants were actively secreting prolactin, as indicated by mammary growth in the hosts rats.

DISCUSSION

The prolactin content of MtT.F₄ and AP transplants is very low as compared with that of the AP *in situ*. The

³ Falcon Plastics, Los Angeles, Calif.

⁴ Difco Laboratories, Detroit, Mich.

⁵ Eli Lilly and Co., Indianapolis, Ind.

⁶ Microbiological Associates, Bethesda, Md.

⁷ Cappel Laboratories, West Chester, Penn.

TABLE 2
PROLACTIN ASSAYS OF BLOOD PLASMA FROM RATS IN DIFFERENT STATES

Type of donor rat	No. donor rats	No. assays	Total no. pigeons	Prolactin (IU/100 ml plasma)	Remarks
Mature, female	4	4	16	0	3-4 months old
Estradiol	4	4	16	0	10 μ g., 1 \times 10 days
Pregnant	3	3	12	0	Mid-pregnancy
Lactating	5	5	20	0	10 days post-partum
AP transplant	8	2	10	0	10 days after single AP transplant
MtT.F ₄ transplant	2	2	8	33	Each rat had 2 tumors (av. diam., 2.5 cm.)
MtT.F ₄ transplant	1	1	4	6.2	One tumor (av. diam., 2.4 cm.)
MtT.F ₄ transplant	2	2	8	0.4	Each rat had 2 tumors (av. diam., about 1 cm.)

AP = anterior pituitary.

MtT.F₄ = "Mammotropic" pituitary tumor, strain F₄.

TABLE 3
PROLACTIN ASSAYS OF DIFFERENT MEDIA FROM CULTURES CONTAINING MtT.F₄ TRANSPLANTS

Medium	No. cultures	Days of cultures	No. assays	Total no. pigeons	Prolactin (IU/100 mg MtT.F ₄)	Prolactin in medium as compared with content in fresh MtT.F ₄
"199"	25	1-3	3	12	1.0 \pm 0.2*	\times 16
"199"	10	1-3	1	4	1.9	\times 31
		4-6	1	4	1.4	\times 23
"199" + 20% calf serum	47	1-3	5	20	1.0 \pm 0.20	\times 16
	27	1-3	3	12	1.7 \pm 0.25	\times 29
		4-6	3	12	1.2 \pm 0.22	\times 20
"199" + 20% rabbit serum	9	1-3	1	4	0.54	\times 9
		4-6	1	4	0.51	\times 8
"199" + 20% horse serum	10	1-3	1	4	1.1	\times 18
		4-6	1	4	1.0	\times 16
"199" + 20% calf serum + 5% chicken embryo extract	10	1-3	1	6	1.3	\times 21
	10	1-3	1	6	1.0	\times 16
		4-6	1	6	0.9	\times 15

MtT.F₄ = Mammotropic pituitary tumor, strain F₄.

* Mean \pm standard error of mean.

MtT.F₄ and AP transplants release considerable amounts of prolactin *in vivo*, as indicated by mammary gland stimulation and by blood assay for prolactin in rats with tumors. Therefore, the MtT.F₄ and AP transplant have less capacity to retain (or store) prolactin than the normal AP *in situ*.

Placement of hypothalamic lesions, transection of the pituitary stalk, transplantation of the AP to noncranial sites, administration of certain depressant drugs, and cultures of AP *in vitro* have all demonstrated that the hypothalamus exerts an inhibitory effect on prolactin secretion (13). We have recently provided evidence that the hypothalamus contains a factor(s) which inhibits

prolactin secretion *in vitro* (20). The decreased capacity of the MtT.F₄ or AP transplant to retain prolactin is probably due to removal of hypothalamic inhibition, with a resultant increase in both synthesis and release of this hormone.

Considerable amounts of prolactin were detected in the blood plasma of rats with a MtT.F₄. The high blood prolactin levels may be due in part to the large size of the pituitary tumors (400-1000 times larger than a normal rat AP); it may actually secrete a smaller amount of prolactin per unit weight of pituitary tissue. The MtT.F₄ during 3 days of culture released 0.5-1.9 IU of prolactin per 100 mg. MtT.F₄, whereas under similar conditions

fresh bovine and rat AP released 8–12 IU (Talwalker and Meites, unpublished) and 10–40 IU per 100 mg. of tissue, respectively (13, 14). This indicates that, on a per unit weight basis, the *in vitro* capacity of MtT.F₄ to secrete prolactin is much lower than that of the normal AP. Nevertheless, the MtT.F₄, during 3 days of culture, released 8–31 times more prolactin than was present initially in the tumor, indicating continuous synthesis and release of prolactin. The comparatively lower prolactin release by MtT.F₄ *in vitro*, in contrast to normal AP, cannot be attributed to inactivation of the hormone, since incubation per se of prolactin did not alter its activity. Prolactin was not detectable, by the assay procedure used, in blood plasma from normally cycling, estrogen-treated, pregnant, or lactating rats or in rats with a single AP transplant. Wolthuis (22, 23) reported the presence of prolactin in the blood plasma of rats in similar states by employing a more sensitive prolactin assay involving a luteotropic reaction by the ovaries.

The MtT.F₄ was initially induced by Furth with estrogen (6, 9). It has been observed that direct circulatory connection with the hypothalamus is not necessary for *in vivo* growth of the tumor (6), or for an *in vivo* or *in vitro* (13, 16) prolactin secretory response to estrogen. Estrogen administration *in vivo* has been found to deplete the rat hypothalamus of prolactin-inhibiting activity (Ratner and Meites, in press). Thus, estrogen stimulation may lead to pituitary tumor formation and increased prolactin secretion as a result of direct action of the estrogen on the AP or hypothalamus or on both. It should be noted that transplantation alone of the AP to non-cranial sites in certain strains of mice (1, 4, 8) may lead to pituitary tumor formation. Tumorigenesis has also been observed in pituitary grafts in rats which have been subjected to estrogen treatment (6, 10, 12). Thus, removal of hypothalamic inhibition to the AP may lead to increased prolactin secretion and formation of prolactin-secreting tumors under appropriate experimental conditions.

ACKNOWLEDGMENTS

We express our sincere appreciation to Drs. Jacob Furth and Untae Kim of the Department of Pathology, College of Physicians and Surgeons, Columbia University, New York, New York, for providing us with "mammatropic" pituitary tumors; to the Endocrinology Study Section, NIH, for ovine prolactin; and to T. E. Staley for technical assistance.

REFERENCES

- BARDIN, C. W., AND LEIBERT, A. D. Neoplastic Changes Occurring in Pituitary Isografts in Mice. *Proc. Am. Assoc. Cancer Res.*, **3**:93, 1960.
- BATES, R. W.; CLIFTON, K. H.; AND ANDERSON, E. Prolactin and Thyrotrophin Content of Functional Transplantable Pituitary Tumors. *Proc. Soc. Exp. Biol. Med.*, **93**:525–26, 1956.
- BATES, R. W.; MILKOVIC, S.; AND GARRISON, M. M. Concentration of Prolactin, Growth Hormone and ACTH in Blood and Tumor of Rats with Transplantable Mammatropic Pituitary Tumors. *Endocrinology*, **71**:943–48, 1962.
- BOOT, L. M.; MÜHLBOCK, O.; ROPCKE, G.; AND TENGBERGEN, W. E. Further Investigations on Induction of Mammary Cancer in Mice by Isografts of Hypophyseal Tissue. *Cancer Res.*, **22**:713–27, 1962.
- CHEN, J. M. The Cultivation in Fluid Medium of Organized Liver, Pancreas and Other Tissues of Foetal Rats. *Exp. Cell Res.*, **7**:518–29, 1954.
- CLIFTON, K. H., AND FURTH, J. Changes in Hormone Sensitivity of Pituitary Mammatropes during Progression from Normal to Autonomous. *Cancer Res.*, **21**:913–20, 1961.
- FURTH, J. Vistas in the Etiology and Pathogenesis of Tumors. *Fed. Proc.*, **20**:865–73, 1961.
- GARDNER, W. U. Tumors in Transplanted Pituitary Glands in Mice. *Am. Assoc. Cancer Res.*, **3**:113, 1960.
- KIM, U.; FURTH, J.; AND YANNOPOULAS, K. Observations on Hormonal Control of Mammary Cancer. I. Estrogen and Mammatropes. *Cancer Res.*, **31**:233, 1963.
- KULLANDER, S. On Tumor Formation in Gonadal and Hypophyseal Transplants into Anterior Eye Chambers of Gonadectomized Rats. *Cancer Res.*, **20**:1079–82, 1960.
- . Studies on the Growth and Hormone Production of Rat Hypophysis in Tissue Culture. *Acta Endocrinol.*, **43**:147–54, 1963.
- MARTINS, T. Action des hautes doses d'estrine sur l'hypophyse *in situ*, ou greffée dans la chambre antérieure de l'oeil du rat. *Compt. Rend. Soc. Biol.*, **123**:702–4, 1936.
- MEITES, J.; NICOLL, C. S.; AND TALWALKER, P. K. The Central Nervous System and the Secretion and Release of Prolactin. In: A. V. NALBANDOV (ed.), *Advances in Neuroendocrinology*, Chapter 8. Urbana: University of Illinois Press, 1963.
- NICOLL, C. S., AND MEITES, J. Prolactin Secretion *in Vitro*: Effects of Thyroid Hormones and Insulin. *Endocrinology*, **72**:544–51, 1963.
- PASTEELS, J. L. Secretion of Prolactin by the Pituitary in Tissue Culture. *Compt. Rend. Séances Acad. Sci.*, **253**:2140–42, 1961.
- RATNER, A.; TALWALKER, P. K.; AND MEITES, J. Effect of Estrogen Administrations *in Vivo* on Prolactin Release by Rat Pituitary *in Vitro*. *Proc. Soc. Exp. Biol. Med.*, **112**:12–15, 1963.
- TAKEMOTO, H.; YOKORO, K.; FURTH, J.; AND COHEN, A. I. Adrenotropic Activity of Mammo-somatotropic Tumors in Rats and Mice. *Cancer Res.*, **22**:917–24, 1962.
- TALWALKER, P. K., AND MEITES, J. Mammary Lobulo-alveolar Growth Induced by Anterior Pituitary Hormones in Adrenoovariectomized and Adreno-ovariectomized-hypophysectomized Rats. *Proc. Soc. Exp. Biol. Med.*, **107**:880–83, 1961.
- TALWALKER, P. K.; RATNER, A.; AND MEITES, J. *In Vivo* and *in Vitro* Prolactin Production by Rat "Mammatropic" Pituitary Tumors. *Fed. Proc.*, **21**:196, 1962.
- . *In Vitro* Inhibition of Pituitary Prolactin Synthesis and Release by Hypothalamic Extract. *Am. J. Physiol.*, **205**:213–18, 1963.
- WHERRY, F. E.; TRIGG, L. N.; GRINDELAND, R. E.; AND ANDERSON, E. Identification of the Hormones Secreted by an Autonomous Mammatropic Pituitary Tumor in Rats. *Proc. Soc. Exp. Biol. Med.*, **110**:362–65, 1962.
- WOLTHUIS, O. L. The Effects of Sex Steroid on the Prolactin Content of Hypophyses and Serum in Rats. *Acta Endocrinol.*, **43**:137–46, 1963.
- WOLTHUIS, O. L., AND DE JONGH, S. E. The Prolactin Production of a Pituitary Graft and of the Hypophysis *in situ*. *Acta Endocrinol.*, **43**:271–79, 1963.