

Prognostic Value of Steroid Receptor Determination in Leukemia¹

D. Duval and F. Homo

Institut National de la Santé et de la Recherche Médicale U 7, Physiology and Pharmacology, Hôpital Necker, 161. rue de Sèvres, 75015 Paris, France

Abstract

Determinations of steroid receptors have been used to predict steroid sensitivity in various neoplastic tumors. It appears, however, that simple steroid binding measurements are not sufficient for that purpose in lymphoid tumors. This conclusion is based on a literature survey showing, first, that numerous factors are capable of modulating cellular steroid receptor content; second, that the results of steroid receptor determinations are critically dependent on experimental procedures; and, third, that the correlation between steroid receptor content and sensitivity is not obligatory in animal or human leukemic cells.

Introduction

In all steroid-responsive tissues the initiation of hormonal response appears to be mediated by a specific receptor system in the cytoplasm of the target cells. The main argument supporting this assumption is the close correlation between the affinity of various steroids for the cytoplasmic binding sites and physiological potency (3, 10, 16, 19, 65).

Given the fact that the binding step is an obligatory event in the mechanism of steroid action, attempts have been made to infer the steroid sensitivity of various tissues from the determination of their steroid receptor content. In particular, this procedure has been used in neoplastic tissues such as mammary tumors, prostatic carcinoma, or leukemia. Recent reports, however, suggest that in lymphoid tissue it is not possible to correlate properly cytoplasmic receptor levels with glucocorticoid responses (6, 32, 45).

Therefore, we want to present a critical reevaluation of receptor determination as an index of steroid sensitivity in lymphoid tissue.

Physiological Modulation of Steroid Receptors

Milgrom *et al.* (50) have emphasized the possibility that the hormonal control of target cells could depend not only on variations in the plasma levels of steroids but also on changes in the cellular amount of receptors. They demonstrated that the level of progesterone receptors undergoes an alternative increase and diminution during the estrous cycle and also rises after a single injection of estradiol. Similarly, estradiol in the uterus appears to be able to modulate its own receptors. An injection of estradiol in immature rats induces an initial drop in estradiol receptors

followed by a low replenishment to control and even higher values.

In the case of glucocorticoids, adrenal ablation produces an increase in corticosterone binding sites in rat liver (4, 18) and heart (26). We have recently demonstrated that adrenalectomy in mice produces not only an increase in thymus size and cellularity but also an augmentation of the number of dexamethasone binding sites per cell.² This elevation of receptors appears to be due to a true increase in the number of receptors rather than to an unmasking of sites previously occupied by endogenous hormone (26).² The existence of a possible mechanism of receptor regulation raises the question of such a modulation under conditions when fluctuations of steroid plasma levels are observed, such as circadian rhythms, stress, and even steroid therapy.

Evaluation of receptor amounts in target organs is made on the basis of protein or DNA content or per cell, assuming that the binding sites are uniformly distributed. This assumption, however, could be inaccurate, particularly in the lymphoid tissue where different cell subsets with different origins and different sensitivities to steroids have been characterized (2, 9, 74). Attempts to isolate subpopulations and to measure their steroid receptors have given rise to puzzling results. Schaumburg and Crone (67) showed that in chickens thymus and bursa lymphocytes contain different amounts of glucocorticoid receptors (600 and 1200 sites/cell, respectively). In mice, spleen cell subpopulations rich in B- and T-cells do not bind the same amount of steroids,³ whereas thymocyte subfractions contain similar quantities of binding sites (14). In humans no difference in receptors could be demonstrated between subpopulations of circulating lymphocytes (42).⁴ Examination of thymus cells in children, however, reveals a low number of glucocorticoid receptors as compared to circulating lymphocytes in adults.⁵ Moreover, the hormonal state could deeply influence the repartition of the lymphocyte subpopulations. Shortman and Jackson (69) showed that adrenalectomy or hydrocortisone treatment modulates the percentage of high θ and low θ thymus cells in mouse. Yu *et al.* (79) and Fauci and Dale (17) demonstrated that steroid treatments in humans induce changes in lymphocyte recirculation and homing. Therefore, determination of the steroid receptors should take into account the species, the nature of the subpopulation studied, and the hormonal state of the individual.

Claman (9) and Baxter *et al.* (2) emphasized that in

² D. Duval, H. Dardenne, and F. Homo, submitted for publication.

³ D. Duval, J. P. Dausse, F. Homo, and C. Fournier, submitted for publication.

⁴ F. Homo, D. Duval, C. Thierry, and B. Serrou, submitted for publication.

⁵ F. Homo and D. Duval, manuscript in preparation.

¹ Presented at the John E. Fogarty International Center Conference on Hormones and Cancer, March 29 to 31, 1978, Bethesda, Md.

lymphoid tissue glucocorticoid susceptibility depends on not only the immunological origin of the cell but also on parameters such as the degree of maturation or differentiation and the stage of cell proliferation. Several authors have attempted to follow the ontogeny of glucocorticoid receptors in parallel with the onset of glucocorticoid-inducible functions in embryos and neonates.

According to the organs studied, several patterns of development could be observed. In chick neural retina Koehler and Moscona (37) described a decrease of hydrocortisone receptors during embryonic life, whereas Lippman *et al.* (47) were unable to demonstrate receptor changes during the same period. Similar discrepancies exist for steroid receptor development in rat liver between the results of Feldman (18) and those of Giannopoulos (23). Giannopoulos *et al.* (24) reported a 2-fold decrease of thymus binding sites in rabbits during the neonatal period. However, we recently showed that in mice no changes in steroid receptors occur in spleen or thymus during the neonatal period.² Because of these conflicting results, the pattern of ontogeny of steroid receptors remains essentially obscure and requires additional investigations. Possible changes of steroid binding sites during aging have received little attention, but Roth (63) and Roth and Livingston (64) have described a decrease of the receptor content in both leukocytes and adipocytes with senescence. In addition, in a given cell the level of binding sites fluctuates according to the stage of cell proliferation. Cidowski and Michaels (7) demonstrated in synchronized HeLa cells that the number of glucocorticoid receptors increases during S phase and falls after mitosis. Similarly, mitogen-induced blast transformation of human lymphocytes is associated with an elevation of the receptor content/cell (53, 71).

Finally, the extent of steroid binding to the receptors could be dependent on metabolic parameters or modulated by other factors. Munck and Brinck-Johnsen (51) and Munck *et al.* (52) showed that the binding of cortisol to its cytosolic receptors was virtually abolished in the absence of glucose or under anaerobic conditions. Nielsen *et al.* (54) pointed out that dephosphorylation decreases receptor binding capacity, suggesting that the binding process could be energy dependent. Several authors postulated the existence of factors capable of modulating steroid receptor interactions; these factors include phospholipids (68), steroids (72, 75), and unidentified modulators (11, 12, 55, 66).

The different parameters responsible for *in vivo* and perhaps physiological modulations of steroid binding sites are summarized in Table 1.

Conditions of Steroid Receptor Determination *in Vitro*

Some of the discrepancies in the results of different groups working on the same material can be attributed to the methods of binding site measurement. Indeed, quantitative evaluation of steroid receptors appears to be critically dependent on experimental parameters such as isolation of subcellular fractions, temperature, composition of the incubation medium, and nature of the tracer. In lymphoid tissue it is possible to determine steroid receptors either in cell-free cytosolic extracts of 0–4° or in whole-cell assays

Table 1
Modulation of cellular steroid receptor content

Parameter	Conditions	Refs.
Level of circulating steroid	Steroid injection	50
	Adrenalectomy	4, 18, 26
	Stress	?
	Circadian rhythm	?
	Hormonal therapy	?
Nature of cell sub-population	Immunological origin	13, 14, 38, 42, 67
	Changes of repartition with hormonal state	17, 69, 79
Cell differentiation	Ontogenesis	18, 23, 24, 37, 47
	Aging	63, 64
	Cell cycle	7
	Blast transformation	53, 71
Metabolic conditions		51, 52, 54
Other factors	Phospholipids	68
	Steroids	72, 75
	Unknown modulators	11, 12, 55, 66

at 37°. The characteristics of dexamethasone binding in human circulating lymphocytes could therefore differ by 1 order of magnitude according to the method used (31, 42). Moreover, in the whole-cell procedure at 4° and 37° we demonstrated that glucocorticoid receptor specificity varies with temperature (15). In contrast to estrogen and androgen receptors, glucocorticoid receptors appear extremely unstable, Kaine *et al.* (34) showed that, during the preparation of thymus cytosol by centrifugation, 20 to 30% of the steroid binding capacity could be lost. Given the fragility of these binding proteins, numerous parameters are capable of affecting their interaction with the ligand, either stabilizing or accelerating receptor decline.

These factors include glycerol, saccharose, cations, EDTA, pH, ionic strength, steroids, and enzymes (34, 39, 59, 68, 72, 75). Sulfhydryl groups appear to play a key role in the binding process, as recently illustrated by Granberg and Ballard (25).

The nature of the tracer used in receptor determination might modify the results. First, triamcinolone acetonide and dexamethasone are more tightly bound by glucocorticoid receptors than are natural steroids (*e.g.*, corticosterone, cortisol) (5, 15). The use of synthetic molecules thus decreases the possibility of steroid receptor dissociation during the experimental procedure. Second, corticosterone or cortisol is known to interact with plasma proteins or with corticosteroid-binding globulin-like binders in the cytoplasm of target tissue (20, 21, 48), therefore leading to a possible overestimation of receptor sites. The existence of a corticosteroid-binding globulin-like protein in human lymphocytes has been reported by Werthamer *et al.* (78) but could not be demonstrated in mouse thymus (15).

Animals used as experimental material are usually adrenalectomized to avoid competition between the endogenous hormone and the exogenous tracer for binding sites. However, in humans the presence of endogenous steroid filling part of the binding sites could lead to an underestimation of the receptors. Until now, attempts to draw an exchange assay between nonradioactive and radioactive

ligands to measure the totality of the cytosolic glucocorticoid receptors have been unsuccessful (18, 26). Therefore, in humans the actual determinations appear to be only estimations of the empty sites. We recently obtained evidence suggesting that an exchange assay could be performed in the whole-cell assay procedure.²

In addition, it was surprising that treatment of intact cells with agents that produce membrane alterations (neuraminidase, phospholipase) or modify the properties of the membrane (concanavalin A, iodoacetamide) induces a diminution of steroid receptor levels (8, 27-29, 49, 58).

Relationship between Steroid Sensitivity and Glucocorticoid Receptor Levels in Normal and Neoplastic Lymphoid Cells

Suggestions that steroid sensitivity could be correlated with the levels of cellular glucocorticoid receptors arose essentially from experiments performed on neoplastic cell lines *in vitro*. Numerous workers reported a defect of steroid receptors in resistant clones as compared to their steroid-sensitive counterpart. In S₄₉IA lymphoma cells or in P1798 lymphosarcoma cells, for example, resistant cells contain 30 to 50% of the binding sites measured in sensitive ones (2, 30, 33, 35, 36, 62, 76). In contrast to these tissue cultures, where steroid sensitivity is easily appreciated following cell lysis, growth, or differentiation inhibitions, steroid susceptibility is more difficult to determine in human leukemia. Usually, antileukemic therapy includes steroid (prednisolone) and other cytotoxic drugs. It is therefore difficult to ascertain the efficiency of one among these agents. Moreover some patients who initially respond to steroid therapy do not respond at a later date, suggesting that cells initially sensitive become resistant or are replaced by resistant cells (35, 36). Despite these difficulties Lippman *et al.* (43, 44, 46, 47) presented evidence that clinical sensitivity to steroids could be associated with elevated levels of steroid binding sites in peripheral cells from patients with acute lymphoblastic leukemia or acute myeloblastic leukemia.

This fascinating correlation between receptor content and steroid sensitivity has recently been questioned, particularly by Sibley and Tompkins (70). These authors developed corticoreistant variants from the S₄₉IA-sensitive cell line. Among these variants the majority are defective in receptors, but some of them contain receptors indistinguishable from those of the sensitive control.

From the experiments performed in various systems, several points become apparent. First, neoplastic tissues often contain the same receptor contents as normal tissues (41). Second, elevation of receptor content does not induce steroid hypersensitivity (71), and, third, glucocorticoid resistance is not always associated with receptor defect (13, 14, 40, 73).

Furthermore, attempts to associate the steroid binding capacity and the clinical state in human neoplastic lymphoid tissue have given only moderate (22) or negative results (6, 31, 32, 77).⁶ This situation is somewhat comparable to

that occurring in breast tumors, in which all tumors that contain estrogen-binding proteins do not necessarily respond to additive or ablative hormone therapy. The conclusion is that, although receptors appear to be indispensable for sensitivity, sensitivity is not governed by receptors alone but depends on many other steps in the mechanism of action of glucocorticoids. Therefore, receptor determination in human leukemic cells is not sufficient to preclude steroid sensitivity.

References

- Baxter, J.D., and Harris, A. W. Mechanism of Glucocorticoid Action: General Features with Reference to Steroid-mediated Immunosuppression. *Transplant. Proc.*, **7**: 55-65, 1975.
- Bacter, J. D., Harris, A. W., Tompkins, G. M., and Cohn, M. Glucocorticoid Receptors in Lymphoma Cells in Culture: Relationship to Glucocorticoid Killing Activity. *Science*, **171**: 189-191, 1971.
- Baxter, J. D., and Tompkins, G. M. The Relationship between Glucocorticoid Binding and Tyrosine Aminotransferase Induction in Hepatoma Tissue Culture Cells. *Proc. Natl. Acad. Sci. U. S.*, **65**: 709-715, 1970.
- Beato, M., Kalimi, M., Beato, W., and Feigelson, P. Interaction of Glucocorticoids with Rat Liver Nuclei: Effect of Adrenalectomy and Cortisol Administration. *Endocrinology*, **94**: 377-387, 1974.
- Bell, P. A., and Munck, A. Steroid Binding Properties and Stabilization of Cytoplasmic Glucocorticoid Receptors from Rat Thymus Cells. *Biochem. J.*, **136**: 97-107, 1973.
- Bird, C. C., Waddel, A. W., Robertson, A. M. G. Currie, A. R., Steel, C. M., and Evans, J. Cytoplasmic Receptor Levels and Glucocorticoid Response in Human Lymphoblastoid Cell Lines. *Brit. J. Cancer*, **33**: 700-707, 1976.
- Cidrowski, J. A., and Michaels, G. A. Alterations in Glucocorticoid Binding Site Number during the Cell Cycle in HeLa Cells. *Nature*, **266**: 643-645, 1977.
- Cidrowski, J. A., and Munck, A. Concanavalin A Induced Glucocorticoid Resistance in Rat Thymus Cells: Decreased Cytoplasmic and Nuclear Receptor Binding of Dexamethasone. *J. Steroid Biochem.*, **7**: 1141-1145, 1976.
- Claman, H. N. Corticosteroids and Lymphoid Cells. *New Engl. J. Med.*, **287**: 388-397, 1972.
- Dausse, J. P., Duval, D., Meyer, P., Gagnault, J. C., Marchandeau, C., and Raynaud, J. P. The Relationship between Corticosteroid Structure and Effects upon Thymocytes. *Mol. Pharmacol.*, **13**: 948-953, 1977.
- Defer, N., Dastugue, B., and Kruh, J. Rat Liver Chromatin Non-histone Proteins and Glucocorticoid Binding. *Biochimie*, **56**: 1549-1557, 1974.
- Defer, N., Dastugue, B., Kruh, J., Defort, J. J., Beck, G., and Beck, J. P. Comparison of Glucocorticoid Cytoplasmic Receptors from Liver, Zajdela Hepatoma and HTC Cells. *Federation European Biochem. Soc. Letters*, **45**: 179-183, 1974.
- Duval, D., Dardenne, M., Dausse, J. P., and Homo, F. Glucocorticoid Receptors in Corticosteroid and Corticoreistant Thymocyte Subpopulations. II. Studies with Hydrocortisone Treated Mice. *Biochim. Biophys. Acta*, **496**: 312-320, 1977.
- Duval, D., Dausse, J. P., and Dardenne, M. Glucocorticoid Receptors in Corticosteroid and Corticoreistant Thymocyte Subpopulations. I. Characterization of Glucocorticoid Receptors and Isolation of a Corticoreistant Subpopulation. *Biochim. Biophys. Acta*, **451**: 82-91, 1976.
- Duval, D., and Simon, J. Temperature Dependent Changes in Specificity of Glucocorticoid Receptors in Mouse Thymocytes. *In*: M. K. Agarwal (ed.), *Multiple Molecular Forms of Steroid Hormone Receptor*, pp. 229-243. Amsterdam: Elsevier.
- Edelman, I. S. Mechanisms of Action of Steroid. *J. Steroid Biochem.*, **6**: 147-159, 1975.
- Fauci, A. S., and Dale, D. C. The Effect of Hydrocortisone on the Kinetics of Normal Human Lymphocytes. *Blood*, **46**: 235-241, 1975.
- Feldman, D. Ontogeny of Rat Hepatic Glucocorticoid Receptors. *Endocrinology*, **95**: 1219-1227, 1974.
- Feldman, D., Funder, J. W., and Edelman, I. S. Subcellular Mechanisms in the Action of Adrenal Steroids. *Am. J. Med.*, **53**: 545-560, 1972.
- Feldman, D., Funder, J. W., and Edelman, I. S. Evidence for a New Class of Corticosterone Receptors in the Rat Kidney. *Endocrinology*, **92**: 1429-1441, 1973.
- Funder, J. W., Feldman, D., and Edelman, I. S. The Roles of Plasma Binding and Receptor Specificity in the Mineralocorticoid Action of Aldosterone. *Endocrinology*, **92**: 994-1004, 1973.
- Gailani, S., Minowada, J., Silvernail, P., Nussbaum, A., Kaiser, N., Rosen, F., and Shimaoka, K. Specific Glucocorticoid Binding in Human Hemopoietic Cell Lines and Neoplastic Tissue. *Cancer Res.*, **33**: 2653-2657, 1973.

⁶ G. Crabtree, K. Smith, and A. Munck, submitted for publication.

23. Gianopoulos, G. Ontogeny of Glucocorticoid Receptors in Rat Liver. *J. Biol. Chem.*, **250**: 5847-5851, 1975.
24. Giannopoulos, G., Hassan, Z., and Solomon, S. Glucocorticoid Receptors in Fetal and Adult Tissues. *J. Biol. Chem.*, **249**: 2424-2427, 1974.
25. Granberg, J. P., and Ballard, P. L. The Role of Sulphydryl Groups in the Binding of Glucocorticoids by Cytoplasmic Receptors of Lung and Other Mammalian Tissues. *Endocrinology*, **100**: 1160-1168, 1977.
26. Gregory, M. C., Duval, D., and Meyer, P. Changes in Cardiac and Hepatic Glucocorticoid Receptors after Adrenalectomy. *Clin. Sci. Mol. Med.*, **51**: 487-493, 1976.
27. Harrison, R. W., Fairfield, S., and Orth, D. N. Evidence for Glucocorticoid Transport through the Target Cell Membrane. *Biochem. Biophys. Res. Commun.*, **61**: 1262-1267, 1974.
28. Harrison, R. W., Fairfield, S., and Orth, D. N. Evidence for Glucocorticoid Transport into AtT-20/D-1 Cells. *Biochemistry*, **14**: 1304-1307, 1975.
29. Harrison, R. W., Fairfield, S., and Orth, D. N. The Effect of Cell Membrane Alteration on Glucocorticoid Uptake by AtT-20/D-1 Target Cells. *Biochim. Biophys. Acta*, **486**: 357-365, 1977.
30. Hollander, N., and Chiu, Y. W. *In Vitro* Binding of Cortisol $1.2 \text{ }^3\text{H}$ by a Substance in the Supernatant Fraction of P 1798 Mouse Lymphosarcoma. *Biochem. Biophys. Res. Commun.*, **25**: 291-297, 1966.
31. Homo, F., Duval, D., and Meyer, P. Etude de la Liaison de la Dexaméthasone Tritiée dans les Lymphocytes de Sujets Normaux et Leucémiques. *Compt. Rend.*, **280**: 91-94, 1975.
32. Homo, F., Duval, D., Meyer, P., Belas, F., Debre, P., and Binet, J. L. Chronic Lymphatic Leukemia. Cellular Effects of Glucocorticoids *in Vitro*. *Brit. J. Haematol.*, in press, 1978.
33. Honma, Y., Kasukabe, T., Okabe, J., and Hozumi, M. Glucocorticoid Binding and Mechanism of Resistance in Some Clones of Mouse Myeloid Leukemic Cells Resistant to Induction. *J. Cellular Physiol.*, **93**: 227-236, 1977.
34. Kaine, J. L., Nielsen, C. J., and Pratt, W. B. The Kinetics of Specific Glucocorticoid Binding in Rat Thymus Cortisol: Evidence for the Existence of Multiple Binding States. *Mol. Pharmacol.*, **11**: 578-587, 1975.
35. Kaiser, N., Milholland, R. J., and Rosen, F. Glucocorticoid Receptors and Mechanism of Resistance in the Cortisol-sensitive and -Resistant Lines of Lymphosarcoma P1798. *Cancer Res.*, **34**: 621-626, 1974.
36. Kirkpatrick, A. F., Milholland, R. J., and Rosen, F. Stereospecific Glucocorticoid Binding to Subcellular Fractions of the Sensitive and Resistant Lymphosarcoma P 1798. *Nature New Biol.*, **232**: 216-218, 1971.
37. Koehler, D. E., and Moscona, A. A. Corticosteroid Receptors in the Neural Retina and Other Tissues of the Chick Embryo. *Arch. Biochem. Biophys.*, **170**: 102-113, 1975.
38. Konior Yarbro, G. S., Lippman, M. E., Johnson, G. E., and Leventhal, B. G. Glucocorticoid Receptors in Subpopulations of Childhood Acute Lymphocytic Leukemia. *Cancer Res.*, **37**: 2688-2695, 1971.
39. Krieger, N. S., Middlebrook, J. L., and Aranow, L. Effect of Salt on Reversibility of Glucocorticoid Receptor Binding. *J. Steroid Biochem.*, **7**: 395-399, 1976.
40. Krystosek, A., and Sachs, L. Steroid Hormone Receptors and the Differentiation of Myeloid Leukemic Cells. *J. Cellular Physiol.*, **92**: 245-352, 1977.
41. Leinen, J. G., Wittliff, J. L., Kostyu, J. A., and Brown, R. C. Glucocorticoid-binding Components in an Irradiation-induced Thymoma of the C57BL/6J Mouse. *Cancer Res.*, **34**: 2779-2783, 1974.
42. Lippman, M. E., and Barr, R. Glucocorticoid Receptors in Purified Subpopulations of Human Peripheral Blood Lymphocytes. *J. Immunol.*, **118**: 1977-1981, 1977.
43. Lippman, M. E., Halterman, R., Leventhal, B., Perry, S., and Thompson, E. B. Glucocorticoid Binding Proteins in Human Acute Lymphoblastic Leukemic Blast Cells. *J. Clin. Invest.*, **52**: 1715-1725, 1973.
44. Lippman, M. E., Halterman, R., Perry, S., Leventhal, B., and Thompson, E. B. Glucocorticoid Binding Proteins in Human Leukaemic Lymphoblasts. *Nature New Biol.*, **242**: 157-158, 1973.
45. Lippman, M. E., Perry, S., and Thompson, E. B. Cytoplasmic Glucocorticoid-binding Proteins in Glucocorticoid-unresponsive Human and Mouse Leukemic Cell Lines. *Cancer Res.*, **34**: 1572-1576, 1974.
46. Lippman, M. E., Perry, S., and Thompson, E. B. Glucocorticoid Binding Proteins in Myeloblasts of Acute Myelogenous Leukemia. *Am. J. Med.*, **59**: 224-227, 1975.
47. Lippman, M. E., Wiggert, B. O., Chader, G. J., and Thompson, E. B. Glucocorticoid Receptors. Characteristics, Specificity, and Ontogenesis in the Embryonic Chick Neural Retina. *J. Biol. Chem.*, **249**: 5916-5917, 1974.
48. Litwack, J., and Singer, S. Subcellular Actions of Glucocorticoid. *In: Biochemical Actions of Hormones*, Vol. 2, Chap. 5, pp. 113-163, New York: Academic Press, 1972.
49. Milgrom, E., Atger, M., and Baulieu, E. E. Studies on Estrogen Entry into Uterine Cells and on Estradiol Receptor Complex Attachment to the Nucleus. Is the Entry of Estrogen into Uterine Cells a Protein Mediated Process? *Biochim. Biophys. Acta*, **320**: 267-283, 1973.
50. Milgrom, E., Luu Thi, M., and Baulieu, E. E. Control Mechanisms of Steroid Hormone Receptors in the Reproductive Tract. *In: Karolinska Symposia on Research Methods in Reproductive Endocrinology. Sixth Symposium: Protein Synthesis in Reproductive Tissue*, pp. 380-403, 1973.
51. Munck, A., and Brinck-Johnsen, T. Specific and Nonspecific Physicochemical Interactions of Glucocorticoids and Related Steroids with Rat Thymus Cells *In Vitro*. *J. Biol. Chem.*, **243**: 5556-5565, 1968.
52. Munck, A., Wira, C., Young, D. A., Mosher, K. M., Hallahan, C., and Bell, P. A. Glucocorticoid Receptor Complexes and the Earliest Steps in the Action of Glucocorticoids on Thymus Cells. *J. Steroid Biochem.*, **3**: 567-578, 1972.
53. Neifeld, J. P., Lippman, M. E., and Tormey, D. C. Steroid Hormone Receptors in Normal Human Lymphocytes. Induction of Glucocorticoid Receptor Activity by Phytohemagglutinin Stimulation. *J. Biol. Chem.*, **252**: 2972-2977, 1977.
54. Nielsen, C. J., Sando, J. S., and Pratt, W. B. Evidence That Dephosphorylation Inactivates Glucocorticoid Receptors. *Proc. Natl. Acad. Sci. U. S. A.*, **74**: 1398-1402, 1977.
55. Nielsen, C. J., Sando, J. S., Vogel, W. M., and Pratt, W. B. Glucocorticoid Receptor Inactivation under Cell-free Conditions. *J. Biol. Chem.*, **252**: 7568-7578, 1977.
56. Norman, M. R., and Thompson, E. B. Characterization of a Glucocorticoid-sensitive Human Lymphoid Cell Line. *Cancer Res.*, **37**: 3785-3791, 1977.
57. Notides, A. C., Hamilton, D. E., and Rudolph, J. H. The Action of a Human Uterine Protease on the Estrogen Receptor. *Endocrinology*, **93**: 210-216, 1973.
58. Picard, F., Homo, F., and Duval, D. Influence of Enzyme Treatment on Dexamethasone Binding in Isolated Mouse Thymocytes. *J. Steroid Biochem.*, in press, 1978.
59. Rafestin-Oblin, M. E., Michaud, A., Claire, M., and Corvol, P. Dramatic Protective Effect of Ligand against Thermal Degradation on Minerals and Glucocorticoid Receptors of Rat Kidney. *J. Steroid Biochem.*, **8**: 19-23, 1977.
60. Rao, G. S., Schulze-Hagen, K., Rao, H. J., and Brauer, H. Kinetics of Steroid Transport through Cell Membranes: Comparison of the Uptake of Cortisol by Isolated Rat Liver Cells with Binding of Cortisol to Rat Liver Cytosol. *J. Steroid Biochem.*, **7**: 1123-1129, 1976.
61. Rochefort, H., and Baulieu, E. E. Effect of KCl, CaCl₂, Temperature and Estradiol on the Uterine Cytosol Receptor of Estradiol. *Biochimie*, **53**: 893-907, 1971.
62. Rosenau, W., Baxter, J. D., Rousseau, G. G., and Tomkins, G. M. Mechanism of Resistance to Steroids: Glucocorticoid Receptor Defect in Lymphoma Cells. *Nature New Biol.*, **237**: 20-24, 1972.
63. Roth, G. S. Reduced Glucocorticoid Responsiveness and Receptor Concentration in Splenic Leukocytes of Senescent Rats. *Biochim. Biophys. Acta*, **399**: 145-156, 1975.
64. Roth, G. S., and Livingston, J. N. Reductions in Glucocorticoid Inhibition of Glucose Oxidation and Presumptive Glucocorticoid Receptor Content in Rat Adipocytes during Aging. *Endocrinology*, **99**: 831-839, 1976.
65. Rousseau, G. G., and Schmit, J. P. Structure Activity Relationships for Glucocorticoids. I. Determination of Receptor Binding and Biological Activity. *J. Steroid Biochem.*, **8**: 911-919, 1977.
66. Sando, J. J., Nielsen, C. J., and Pratt, W. B. Reactivation of Thymocyte Glucocorticoid Receptors in a Cell-free System. *J. Biol. Chem.*, **252**: 7579-7582, 1977.
67. Schaumburg, B. P., and Crone, M. Binding of Corticosterone of Thymus Cells, Bursa Cells and Blood Lymphocytes from the Chicken. *Biochim. Biophys. Acta*, **237**: 494-501, 1971.
68. Schulte, H. F., Nielsen, C. J., Sando, J. J., and Pratt, W. B. Evidence for a Phospholipid Requirement in the Specific Binding of Glucocorticoids to Receptors of Fibroblasts and Thymic Lymphocytes. *J. Biol. Chem.*, **251**: 2279-2289, 1976.
69. Shortman, K., and Jackson, H. The Differentiation of Lymphocytes. I. Proliferation Kinetics and Interrelationships of Subpopulations of Mouse Thymus Cells. *Cellular Immunol.*, **12**: 230-246, 1974.
70. Sibley, C. H., and Tomkins, G. H. Mechanisms of Steroid Resistance. *Cell*, **2**: 221-227, 1974.
71. Smith, K. A., Crabtree, G. R., Kennedy, S. J., and Munck, A. U. Glucocorticoid Receptors and Glucocorticoid Sensitivity of Mitogen Stimulated and Unstimulated Human Lymphocytes. *Nature*, **267**: 523-525, 1977.
72. Suthers, H. B., Pressley, L. A., and Funder, J. W. Glucocorticoid Receptors: Evidence for a Second, Non Glucocorticoid Binding Site. *Endocrinology*, **99**: 260-269, 1976.
73. Thompson, E. B., Aviv, D., and Lippman, M. E. Variants of HTC Cells with Low Tyrosine Aminotransferase Inducibility and Apparent Normal Glucocorticoid Receptors. *Endocrinology*, **100**: 406-419, 1977.
74. Thompson, E. B., and Lippman, M. E. Mechanisms of Action of Glucocorticoid. *Metab. Clin. Exptl.*, **23**: 159-202, 1974.
75. Tindall, D. J., Cunningham, G. R., and Means, A. R. 5 α -Dihydrotestosterone Binding to Androgen-binding Proteins. Effects of Testosterone

- and Other Compounds *in Vivo* and *in Vitro* on the Number of Binding Sites Measured. *J. Biol. Chem.*, 253: 166-169, 1978.
76. Turnell, R. W., and Burton, A. F. Glucocorticoid Receptors and Lymphocytolysis in Normal and Neoplastic Lymphocytes. *Mol. Cellular Biochem.* 9: 175-189, 1975.
77. Waddell, A. W., Bird, C. C., and Currie, A. R. The Role of Cytoplasmic Glucocorticoid Receptors in the Cytolysis of Human Lymphoblastoid Cells by Methylprednisolone. *Biochem. Soc. Trans.*, 5: 719-721, 1977.
78. Werthamer, S., Samuels, A. J., and Amaral, L. Identification and Partial Purification of "Transcortin"-like Protein within Human Lymphocytes. *J. Biol. Chem.*, 248: 6398-6407, 1973.
79. Yu, D. T., Clements, P. J., Paulus, H. E., Peter, B. J., Levy, R., and Barnett, E. V. Human Lymphocyte Subpopulations: Effects of Corticosteroids. *J. Clin. Invest.*, 53: 565-571, 1974.