Rapid cooling preserves the ischaemic myocardium against mitochondrial damage and left ventricular dysfunction

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Aims We investigated whether rapid cooling instituted by total liquid ventilation (TLV) improves cardiac and mitochondrial function in rabbits submitted to ischaemia-reperfusion.

Methods and results Rabbits were chronically instrumented with a coronary artery occluder and myocardial ultrasonic crystals for assessment of segment length-shortening. Two weeks later they were re-anaesthetized and underwent either a normothermic 30-min coronary artery occlusion (CAO) (Control group, n = 7) or a comparable CAO with cooling initiated by a 10-min hypothermic TLV and maintained by a cold blanket placed on the skin. Cooling was initiated after 5 or 15 min of CAO (Hypo-TLV and Hypo-TLV15 groups, n = 6 and 5, respectively). A last group underwent normothermic TLV during CAO (Normo-TLV group, n = 6). Wall motion was measured in the conscious state over three days of reperfusion before infarct size evaluation and histology. Additional experiments were done for myocardial sampling in anaesthetized rabbits for mitochondrial studies. The Hypo-TLV procedure induced a rapid decrease in myocardial temperature to 32–34°C. Throughout reperfusion, segment length-shortening was significantly increased in Hypo-TLV and Hypo-TLV15 vs. Control and Normo-TLV (15.1 ± 3.3%, 16.4 ± 2.3%, 1.8 ± 0.6%, and 1.1 ± 0.8% at 72 h, respectively). Infarct sizes were also considerably attenuated in Hypo-TLV and Hypo-TLV15 vs. Control and Normo-TLV (4 ± 1%, 11 ± 5%, 39 ± 2%, and 42 ± 5% infarction of risk zones, respectively). Mitochondrial function in myocardial samples obtained at the end of ischaemia or after 10 min of reperfusion was improved by Hypo-TLV with respect to ADP-stimulated respiration and calcium-induced opening of mitochondrial permeability transition pores (mPTP). Calcium concentration opening mPTP was, e.g., increased at the end of ischaemia in the risk zone in Hypo-TLV vs. Control (157 ± 12 vs. 86 ± 12 μM). Histology and electron microscopy also revealed better preservation of lungs and of cardiomyocyte ultrastructure in Hypo-TLV when compared with Control.

Conclusion Institution of rapid cooling by TLV during ischaemia reduces infarct size as well as other sequelae of ischaemia, such as post-ischaemic contractile and mitochondrial dysfunction.

KEYWORDS
Cooling; Contractile function; Mitochondria; Infarction; Total liquid ventilation

1. Introduction
Among the numerous experimental strategies that have been proposed to reduce the size of myocardial infarct, one of the most potent is cooling the heart to 32–34°C during ischaemia.¹⁻³ At these temperatures, the heart beats normally and no external support of the circulation is required. The cooling rates that can be achieved by cooling the skin⁴ or using intravascular thermodes¹ are however too slow to be optimally clinically effective. To exert optimal protection, a cooling strategy should aim to lower temperature to the desired target very fast after the onset of coronary artery occlusion (CAO) to effectively shorten the normothermic ischaemic time. Total liquid ventilation (TLV) with temperature-controlled perfluorocarbons has been proposed for rapid cooling,⁵⁻⁷ as these liquids have a high thermal conductivity and can use the lungs as heat...
exchangers while still maintaining gas exchange (high solubility for $O_2$ and $CO_2$).\textsuperscript{5,8,9} In a previous report, we demonstrated that a left atrial temperature of 32°C could be achieved within 5 min using TLV in anaesthetized rabbits.\textsuperscript{7} This was associated with a dramatic decrease in infarct size when hypothermic TLV (Hypo-TLV) was performed during a 30-min CAO. It remains, however, unknown whether rabbits can recover after Hypo-TLV and whether it would protect the myocardium against regional contractile dysfunction. This latter endpoint must be determined as it is well known that infarct size reduction does not predict full left ventricular contractile recovery, i.e. myocardium could be salvaged but still remain dysfunctional.\textsuperscript{10,11}

Moreover, the exact mechanism by which mild hypothermia protects the heart against ischaemia remains unknown and justifies further investigation. Reduction in ATP consumption,\textsuperscript{12} intracellular acidosis, and $Na^+$ and $Ca^{2+}$ overload\textsuperscript{13} would be likely components of a mechanism resulting in the reduction of myocardial energy utilization\textsuperscript{14} and inhibition of intracellular processes associated with cell necrosis. Preserved mitochondrial function has also been proposed to protect isolated hypothermic hearts against ischaemia,\textsuperscript{15} but this was investigated following only ex vivo 17°C deep hypothermia. The effect of moderate 32–34°C hypothermia on ischaemia-induced mitochondrial dysfunction has not been studied despite the observation that this organelle, and in particular the mitochondrial permeability transition pore (mPTP), is possibly the end-effector of cardioprotection.\textsuperscript{16–19} Such investigations would indeed provide new mechanistic insights to determine whether TLV-induced hypothermia, a cardioprotective manoeuvre not related to preconditioning or post-conditioning, would also protect mitochondria against mPTP opening.

The main goal of the present study was, therefore, to investigate whether rapid cooling induced by TLV improves regional left ventricular function and protects mitochondria in rabbits submitted to lethal ischaemia. Accordingly, we first investigated left ventricular recovery in a well-established model of myocardial infarction in chronically instrumented conscious rabbits. Secondly, we performed additional investigations on mitochondria from ischaemic myocardium of rabbits submitted to CAO. Mitochondrial structure and function were studied by evaluating oxygen consumption, calcium-induced opening of mPTP, and morphology.

2. Methods

Animal instrumentation and ensuing experiments were conducted in accordance with French official regulations after approval by the local ethical committee. The investigation conforms to the Guide for the Care and Use of Laboratory Animals published by the US National Institutes of Health. All experiments were performed in male New Zealand rabbits (2.5–3.0 kg).

2.1 Instrumentation

Rabbits were anaesthetized with a mixture of tiletamine (25 mg/kg iv) and zolazepam (25 mg/kg iv), intubated, and mechanically ventilated with oxygen. Maintenance of anaesthesia was done with pentobarbital and inhaled 2% isoflurane. A left thoracotomy was performed under sterile conditions and a pneumatic occluder was implanted around a major branch of the left coronary artery (marginal artery).\textsuperscript{10,11} A pair of 1-mm ultrasonic piezoelectric crystals was inserted within the left ventricular wall in the perfusion territory of the instrumented coronary artery. The chest was closed in layers. The occluder tubing and the crystal wires were exteriorized between the scapulae. During the post-operative period, rabbits received buprenorphine (0.02 mg/kg/12 h sc, 3 days) and spiramycine (60 000 IU/kg/day im, 5 days). Rabbits recovered for a minimum of 10 days after surgery before inclusion in the study.

2.2 Experimental protocol

Baseline haemodynamic parameters were recorded in the conscious state in all rabbits after surgery. They were then re-anaesthetized using thiopentone (20 mg/kg iv) and intubated for conventional mechanical ventilation. As shown in Figure 1A, rabbits were divided into four groups. All underwent 30 min of CAO to induce infarction. Reperfusion was then initiated and continued for 3 days in the conscious state. The first group (Control) was not subjected to any other treatment. In the second group (Hypo-TLV), cooling was started at the fifth minute of CAO using the combination of 10 min of hypothermic TLV and 25 min of surface cooling with ice-filled cold blankets initiated at the same time as TLV. In the third group (Hypo-TLV\textsubscript{15}), a similar cooling protocol was induced at the 15th min of CAO using hypothermic TLV for 10 min combined with 15 min of surface cooling. The last group (Normo-TLV) was treated with 10 min of normothermic TLV initiated in the fifth min of CAO. In the Hypo-TLV, Hypo-TLV\textsubscript{15}, and Normo-TLV groups, TLV episodes were performed using a mixture of perfluoroobutyltrifluorohydrocarbons and perfluoropropyltetrahydropyran (RM101, Miten, Milano, Italy).\textsuperscript{7} Rabbits were switched to liquid ventilation by filling the lung with 20 mL of perfluorocarbon and then connecting the endotracheal tube to the liquid ventilator. The ventilator was set to a tidal volume of 15 mL/kg body weight and 5 breaths/min. For each breath, the ventilator pumped into and out of the lungs the tidal volume of liquids. This protocol has previously been demonstrated to maintain normal blood gases.\textsuperscript{7} The perfluorocarbon mixture was bubbled with 100% $O_2$. The temperature of the heat exchanger was set to either 15°C in Hypo-TLV and Hypo-TLV\textsubscript{15} or 39°C in the Normo-TLV group. At the end of the TLV procedure, the liquid was aspirated from the lung (approximately 20 mL of liquid) and resumption of conventional gas ventilation was permitted. In the Hypo-TLV and Hypo-TLV\textsubscript{15} groups, warm-up was initiated at the onset of reperfusion using infra-red lamps and a heating pad until the return of normothermia as monitored by the rectal temperature probe. Rabbits were allowed to wake-up and breathe spontaneously as soon as possible after initiation of reperfusion. They were kept in a cage with supplemental oxygen for 24–48 h.

In order to measure myocardial temperature during the TLV procedure, additional rabbits were anaesthetized and mechanically ventilated. A left lateral thoracotomy was performed and two thermal probes were implanted within the left ventricular wall for measurement of myocardial temperatures in ischaemic and non-ischaemic territories during a subsequent CAO. After the institution of CAO, the chest was rapidly closed in layers, and rabbits were randomly divided into Control, Hypo-TLV, or Normo-TLV groups, as described above. In the Hypo-TLV group, the cooling procedure was started at the fifth minute of ischaemia. Rectal and myocardial temperatures were continuously monitored throughout the 30 min of CAO (Figure 2, upper panel).

Finally, in order to avoid blood-sampling in rabbits in which haemodynamics were investigated, we performed blood-gas analyses (ABL77, Radiometer Medical ApS, Brønshøj, Denmark) in additional rabbits submitted to Control or Hypo-TLV procedures ($n = 4$ in each condition). Blood-gas values were corrected for the actual body temperature.
2.4 Post-mortem analyses and histology

After completion of three days of reperfusion, the chronically instrumented rabbits were euthanized using pentobarbital (60 mg/kg iv) followed by potassium chloride. The hearts were excised and the coronary artery was ligated at the occluder site. The ascending aorta was cannulated and perfused retrogradely with 100 mM sucrose, 100 mM KCl, 10 mM HEPES, 5 mM KH2PO4, pH = 7.4) containing mitochondria (0.4 mg protein/ml). Substrate-respiration rate (state 4 oxygen consumption) and ATP synthesis (state 3) were investigated by the addition of 5 mM pyruvate/malate and 300 μM ADP, respectively. The corresponding respiration control ratio (state 3/state 4) was calculated. In the mitochondrial samples submitted to both ischaemia and reperfusion, ATP concentration was measured using an ATP determination kit (FluoProbes®, Interchim, Montluçon, France).

In other experiments, the ability of mitochondria to retain Ca2+ before exogenously induced mPTP opening was monitored, as previously described.20 Briefly, cardiac mitochondria (1 mg protein/mL) energized with 5 mM pyruvate/malate were incubated in the respiration buffer including 1 μM of the Ca2+ green-N fluorescent probe. The reaction was started by the addition of successive 10 μM Ca2+ pulses. After each addition, a rapid uptake was observed followed by a dynamic steady state corresponding to the equilibrium between influx and efflux of Ca2+. When sufficient Ca2+ loading was
2.6 Electron microscopy

In order to further investigate myocardial integrity following hypothermia, experiments were performed in additional Hypo-TLV or Control anesthetized rabbits. After opening the chest, a major branch of the left coronary artery (marginal artery) was occluded for 30 min. The chest was immediately closed and rabbits were subjected to either no procedure (Control) or to cooling with Hypo-TLV initiated in the fifth min of CAO. Just before completion of the 30-min CAO, the chest was reopened in order to reperfuse the ligated artery. At 15 s after the onset of reperfusion, hearts were excised and perfused-fixed through the aorta with 2.5% glutaraldehyde. A 5 × 5 mm sample of myocardium was excised from both the ischaemic and non-ischaemic areas. Each sample was cut into 1 mm³ tissue blocks which were embedded in Epon for standard electron microscopy. Five blocks from each area were randomly selected for analysis.

2.7 Statistical analysis

Values are expressed as means ± SEM. Comparisons were made using either a one- or two-way analysis of variance followed by a Student’s t-test with Bonferroni correction. Significant differences were determined at P < 0.05.

3. Results

As shown in Figure 2 (lower panel) myocardial and rectal temperatures were rapidly decreased in Hypo-TLV when compared with Control and Normo-TLV (n = 5 in each group, P < 0.05). Myocardial temperature in the ischaemic territory decreased by an average of −6.3 ± 0.2°C at the 15th min of CAO in the Hypo-TLV group.

During conventional mechanical ventilation, blood-gases were within normal ranges in Control hearts (e.g. pH 7.4 ± 0.1, pCO₂ 41 ± 4 mmHg, and pO₂ 531 ± 30 mmHg after 30 min of follow-up). Blood-gas values were also not altered at the end of the Hypo-TLV cooling procedure (pH 7.4 ± 0.1, pCO₂ 32 ± 3 mmHg, and pO₂ 597 ± 14 mmHg) and 1 h later (pH 7.4 ± 0.1, pCO₂ 33 ± 3 mmHg, and pO₂ 538 ± 59 mmHg).

Twenty-four rabbits underwent the complete protocol in the chronic study (seven, six, five, and six rabbits in Control, Hypo-TLV, Hypo-TLV₁₅, and Normo-TLV groups, respectively). As shown in Table 1, haemodynamic parameters and rectal temperature were not significantly different among groups at baseline. During CAO, those parameters were similar between groups except heart rate and rectal temperature that were significantly reduced after the onset of cooling in Hypo-TLV and Hypo-TLV₁₅. Conversely, segment length-shortening was significantly increased throughout reperfusion in Hypo-TLV and Hypo-TLV₁₅ when compared with Control. As shown in Figure 1B, this parameter recovered to 95% of its corresponding baseline value after 72 h of reperfusion in these groups. Sizes of risk regions were not significantly different among groups averaging 32 ± 3%, 35 ± 4%, 34 ± 2%, and 36 ± 4% of the left ventricle in Control, Hypo-TLV, Hypo-TLV₁₅, and Normo-TLV groups, respectively. As shown in Figure 1C, infarct size was significantly reduced in Hypo-TLV and Hypo-TLV₁₅ when compared with Control and Normo-TLV (4 ± 1%, 11 ± 5%, 39 ± 2%, and 42 ± 5% of region at risk, respectively). Infarction between crystals assessed by histology was also significantly reduced in the Hypo-TLV (1 ± 1%) and Hypo-TLV₁₅ (3 ± 2%) groups when compared with Control (45 ± 9%) and Normo-TLV (36 ± 6%). In all lungs from the Control group, histology demonstrated foci of minor to severe congestion (Figure 1D, panel a). In the Hypo-TLV group, three rabbits had normal lungs (Figure 1D, panel b) and three others had only minor congestion.

In 10 other rabbits mitochondria were isolated from myocardium obtained at the end of the 30-min CAO (n = 5 in both Control and Hypo-TLV groups). In Control normothermic hearts, mitochondrial dysfunction in the ischaemic territory was demonstrated by a −41% decrease in ATP synthesis (state 3 oxygen consumption) compared with the non-ischaemic zone (447 ± 30 vs. 763 ± 63 nmoles O₂/min/mg of protein, respectively, P < 0.05). In Hypo-TLV hearts (n = 5), state 3 oxygen consumption was decreased by only −25% in ischaemic vs. non-ischaemic zones (631 ± 53 vs. 841 ±
However, substrate-respiration rate (state 4 oxygen consumption) was not altered in the ischaemic zone in either group (data not shown). As illustrated in Figure 3C, the respiratory control ratio (state 3/state 4 oxygen consumption) was therefore significantly decreased by 42% in the ischaemic zone in the Control group. In contrast, the respiratory control ratio was significantly less depressed in Hypo-TLV (−21%). As shown in Figure 3D, Ca2⁺ concentration required to open mPTP was significantly reduced by −49% in the ischaemic vs. non-ischaemic zones in the Control group (86 ± 12 vs. 170 ± 10 μM, respectively). In contrast, hypothermia protected mitochondria against mPTP opening as Ca2⁺ concentration required to open mPTP was not significantly different in the ischaemic and non-ischaemic zones in the Hypo-TLV group (157 ± 12 vs. 188 ± 14 μM, respectively).

Ten rabbits were also included for mitochondrial investigations after 30-min CAO followed by 10 min of reperfusion (n = 5 in both Control and Hypo-TLV groups). The respiratory control ratio (state 3/state 4) was significantly decreased...
by −58% in the reperfused when compared with non-ischaemic zone in the Control group, as shown in Figure 3E. In contrast, the respiratory control ratio was significantly less decreased in Hypo-TLV (−30%). The increase in respiratory control ratio was again related to an alteration in state 3 oxygen consumption as state 4 was unchanged (data not shown). Interestingly, ATP concentrations in myocardium subjected to ischaemia-reperfusion was also significantly improved in Hypo-TLV hearts compared with Control (5.90 ± 1.63 and 0.65 ± 0.17 µM/g of tissue, respectively). As shown in Figure 3F, Ca$^{2+}$ concentration required to open mPTP was significantly reduced by −68% in the territory undergoing ischaemia-reperfusion compared with the non-ischaemic zone in the Control group (57 ± 8 vs. 174 ± 17 µM, respectively). In contrast, it was only reduced by −37% in the Hypo-TLV group (117 ± 16 vs. 183 ± 12 µM, respectively).

Finally, six other rabbits underwent myocardial sampling for electron microscopy (n = 3 in both Control and Hypo-TLV groups). In each heart, we analysed five samples issued from the ischaemic territory and five samples from the normally perfused myocardium (total number of blocks = 60). In all samples from Control and Hypo-TLV rabbits, cellular and extracellular architecture of normally perfused myocardium was intact with normal mitochondria, lack of intracellular oedema (Figure 4A), and normal microvessels (Figure 4B). In all samples of ischaemic myocardium from Control hearts we observed marked intracellular oedema (Figure 4C) with wide-spread irreversible ischaemic damage to the mitochondria characterized by membrane rupture and amorphous densities and (Figure 4E). In the Hypo-TLV group, intracellular oedema was more modest (Figure 4D) and mitochondrial damage was characterized by minor loss of mitochondrial cristae without amorphous densities (Figure 4F). Finally, we observed endothelial cell necrosis in Control samples (Figure 4G) but not in Hypo-TLV hearts (Figure 4H).

4. Discussion

The present study demonstrates that institution of rapid cooling by a brief episode of Hypo-TLV during ischaemia not only reduces infarct size as previously reported\cite{2,3,21-23} but importantly also abolishes post-ischaemic regional contractile dysfunction. Interestingly, our results also demonstrate that hypothermia protects the myocardium against cellular damage and calcium-induced opening of mPTP at the end of the ischaemic period. To our knowledge this is the first study to investigate the functional recovery following myocardial infarction and cooling in a 3-day recovery model and to compare mitochondrial function following in vivo normothermic and hypothermic ischaemia (32–34°C).

Our results document that a short period of hypothermic TLV can be used to induce rapid cooling that can be maintained by less-invasive surface cooling. Importantly, the infarct size reduction elicited by Hypo-TLV was not related to a chemical effect of the perfluorocarbon mixture since infarct size was not altered with normothermic TLV. This result was expected as the cardioprotective effect of

| Table 1 Haemodynamic parameters and rectal temperature in chronically instrumented rabbits |
|----------------------------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|
|                                 | Baseline        | CAO 15 min      | 25 min          | Reperfusion 1 h | 2 h             | 3 h             | 24 h            | 48 h            | 72 h            |
| Heart rate (beats/ min)         |                 |                 |                 |                 |                 |                 |                 |                 |                 |
| Control                         | 234 ± 11        | 248 ± 16        | 242 ± 15        | 234 ± 13        | 239 ± 10        | 246 ± 9         | 242 ± 14        | 242 ± 12        | 243 ± 13        |
| Hypo-TLV                        | 243 ± 11        | 177 ± 13*       | 200 ± 7*        | 243 ± 9         | 248 ± 11        | 244 ± 15        | 251 ± 8         | 240 ± 13        | 244 ± 15        |
| Hypo-TLVs                        | 240 ± 10        | 228 ± 17        | 187 ± 13*       | 211 ± 16        | 252 ± 17        | 242 ± 13        | 220 ± 15        | 227 ± 19        | 245 ± 30        |
| Normo-TLV                       | 234 ± 9         | 241 ± 15        | 238 ± 16        | 230 ± 17        | 250 ± 12        | 257 ± 10        | 241 ± 18        | 243 ± 13        |                 |
| Mean arterial blood pressure (mmHg) |                 |                 |                 |                 |                 |                 |                 |                 |                 |
| Control                         | 84 ± 2          | 68 ± 8          | 68 ± 8          | 73 ± 8          |                 |                 |                 |                 |                 |
| Hypo-TLV                        | 84 ± 4          | 73 ± 6          | 66 ± 4          | 69 ± 5          |                 |                 |                 |                 |                 |
| Hypo-TLVs                        | 76 ± 2          | 73 ± 3          | 70 ± 4          | 67 ± 5          |                 |                 |                 |                 |                 |
| Normo-TLV                       | 84 ± 3          | 74 ± 2          | 67 ± 5          | 68 ± 6          |                 |                 |                 |                 |                 |
| Segment length at the end of diastole (mm) |                 |                 |                 |                 |                 |                 |                 |                 |                 |
| Control                         | 7.5 ± 1.0       | 7.7 ± 1.1       | 7.2 ± 1.2       | 7.8 ± 1.1       | 7.9 ± 1.1       | 7.8 ± 1.1       | 7.9 ± 1.1       | 7.8 ± 1.1       | 8.0 ± 1.1       |
| Hypo-TLV                        | 6.8 ± 1.1       | 7.5 ± 1.2       | 7.4 ± 1.1       | 6.9 ± 1.0       | 6.8 ± 1.0       | 6.8 ± 1.1       | 6.7 ± 1.1       | 6.8 ± 1.1       | 6.7 ± 1.1       |
| Hypo-TLVs                        | 7.7 ± 0.9       | 8.2 ± 1.0       | 8.2 ± 1.0       | 7.8 ± 1.0       | 7.9 ± 0.9       | 7.9 ± 0.9       | 8.3 ± 1.1       | 7.9 ± 1.0       | 7.5 ± 0.9       |
| Normo-TLV                       | 7.1 ± 1.1       | 7.2 ± 0.9       | 7.3 ± 1.0       | 7.1 ± 1.0       | 7.2 ± 1.0       | 7.2 ± 1.0       | 7.4 ± 1.0       | 7.2 ± 1.0       | 7.0 ± 1.0       |
| Segment length systolic shortening (%) |                 |                 |                 |                 |                 |                 |                 |                 |                 |
| Control                         | 15.2 ± 2.3      | 2.1 ± 0.3       | −1.4 ± 0.4      | 0.9 ± 0.6       | 0.9 ± 0.7       | 0.4 ± 0.8       | 0.4 ± 1.0       | 0.6 ± 0.8       | 1.8 ± 0.6       |
| Hypo-TLV                        | 15.8 ± 2.7      | −0.9 ± 0.4      | −1.3 ± 0.3      | 8.3 ± 1.8       | 8.8 ± 1.7       | 11.1 ± 2.2*     | 12.5 ± 2.0*     | 15.3 ± 4.0*     | 15.1 ± 3.3*     |
| Hypo-TLVs                        | 18.3 ± 2.7      | −1.0 ± 0.7      | −0.9 ± 0.7      | 10.2 ± 3.4*     | 10.5 ± 3.8*     | 11.4 ± 2.5*     | 15.5 ± 1.9*     | 16.6 ± 2.1*     | 16.4 ± 2.3*     |
| Normo-TLV                       | 14.9 ± 3.1      | −2.1 ± 0.5      | −1.7 ± 0.5      | 0.9 ± 1.1       | 0.7 ± 1.1       | 0.3 ± 0.8       | 1.0 ± 1.4       | 1.1 ± 1.0       | 1.1 ± 0.8       |
| Rectal temperature (°C)         |                 |                 |                 |                 |                 |                 |                 |                 |                 |
| Control                         | 39.1 ± 0.3      | 38.8 ± 0.3      | 38.6 ± 0.3      | 39.0 ± 0.4      | 38.9 ± 0.8      | 39.0 ± 0.9      |                 |                 |                 |
| Hypo-TLV                        | 39.2 ± 0.1      | 36.3 ± 0.5*     | 34.8 ± 0.6*     | 36.2 ± 0.6*     | 38.1 ± 0.3      | 38.7 ± 0.3      |                 |                 |                 |
| Hypo-TLVs                        | 38.7 ± 0.2      | 38.3 ± 0.2      | 35.5 ± 0.7*     | 36.1 ± 0.3*     | 38.0 ± 0.6      | 38.3 ± 0.3      |                 |                 |                 |
| Normo-TLV                       | 39.3 ± 0.2      | 38.9 ± 0.2      | 38.7 ± 0.2      | 38.3 ± 0.3      | 38.5 ± 0.2      | 38.8 ± 0.2      |                 |                 |                 |

CAO, coronary artery occlusion; Hypo-TLV, hypothermic total liquid ventilation; Hypo-TLVs, Hypo-TLV instituted at the fifteenth min of CAO; Normo-TLV, normothermic total liquid ventilation.

*P < 0.05 vs. Control.
cooling during ischaemia has already been demonstrated in rabbits,\textsuperscript{2,3} dogs,\textsuperscript{21} pigs,\textsuperscript{22} and rats.\textsuperscript{23} The protection is dependent on the depth of cooling and maximal protection is reached at 32°C.\textsuperscript{3} In the present study, myocardial salvage was observed when cooling was initiated either at the fifth or fifteenth min of ischaemia. In our previous report in open-chest anaesthetized rabbits, Hypo-TLV did not reduce infarct size if instituted at the 25th min of a 30 min ischaemia.\textsuperscript{7} Importantly, we further observed in the present report a rapid and potent recovery of post-ischaemic regional contractility in Hypo-TLV and Hypo-TLV\textsubscript{15} groups when compared with normothermic intervention.

Figure 4  Electron microscopic analysis of myocardium samples fixed immediately after the onset of reperfusion. (A, B) Normal ultrastructure of myocardium in the non-ischaemic area in both Control (A, ×12 500) and Hypo-TLV (B, ×8000) groups. (C, D) More marked intracellular oedema in the ischaemic area in the Control (C, ×6300) than in the Hypo-TLV group (D, ×6300). (E) In the ischaemic area in the Control group (E), there is widespread irreversible ischaemic damage to the mitochondria characterized by amorphous densities (small arrows) (×12 500). (F) In the ischaemic area in the Hypo-TLV group, the mitochondrial damage is characterized by minor loss of mitochondrial cristae in the ischaemic area (large arrows, ×12 500). (G) Damage of the microvessels (V) showing endothelial cell necrosis in the ischaemic area in the Control group (×12 500). (H) The microvessels (V) are normal in the ischaemic area in the Hypo-TLV group (×12 500).
The recovery was evident as early as 30 min after the onset of reperfusion and virtually complete after 3 days. Conversely, ischaemic preconditioning that can induce an ultimate infarct size reduction as potent as that observed with Hypo-TLV failed to improve regional contractility during the first hours of reperfusion in similarly instrumented rabbits. These data suggest that in addition to its infarct-sparing effect, cooling during ischaemia exerts a more potent protective effect than preconditioning against early post-ischaemic dysfunction. It has also been reported that a modest decrease in myocardial temperature exerts a positive inotropic and oxygen-saving effect in in situ dog hearts at 34°C. The proposed mechanisms include an increased Ca^{2+} sensitivity of myofilament proteins and improved Ca^{2+}-activated force generation.

In addition to infarct size reduction and improvement in post-ischaemic left ventricular function, we further demonstrated excellent preservation of cardiomyocyte and blood vessel ultrastructure in rabbits subjected to Hypo-TLV compared with Control. This effect was observed by electron microscopy in hearts fixed immediately after the onset of reperfusion, again suggesting that the beneficial effect of cooling occurs during ischaemia. This however does not definitely exclude any reperfusion injury in these conditions. We also demonstrate that hypothermia protects mitochondria by preserving respiratory control ratio and ATP synthesis and reducing mitochondrial Ca^{2+} sensitivity to mPTP opening. Mitochondrial preservation by hypothermia is an important finding since opening of mPTP is a well-known trigger of cell death following myocardial ischaemia. Preconditioning and post-conditioning exert, at least in part, their cardioprotective effect at reperfusion through inhibition of mPTP opening. The beneficial effect of cooling was unexpectedly observed in mitochondria from 'non-reperfused' as well as 'reperfused' myocardium. In 'non-reperfused' conditions, Ca^{2+} concentration required to open mPTP was importantly not significantly decreased in hypothermic hearts (−16%), whereas we observed a −49% drop in normothermic hearts. In hearts subjected to ischaemia-reperfusion, a significant decrease was observed in both hypothermic (−37%) and Control (−68%) hearts, although the decrease was greater in the latter. Hence reperfusion enhanced mitochondrial damage in both normothermic and hypothermic hearts again suggesting that hypothermia protects during ischaemia rather than against reperfusion injury. The exact mechanism by which hypothermia inhibits mPTP opening was not investigated. Inhibition of ischaemia-induced calcium overload could be involved since it was previously reported in isolated guinea pig hearts that hypothermia to 17°C abolished mitochondrial rise of calcium concentration during ischaemia. Interestingly, hypothermia also altered reactive oxygen species production in that study. However, we did not properly demonstrate that mitochondrial protection was a direct effect of hypothermia as one would expect any treatment against ischaemia to protect mitochondria directly or indirectly.

One of our important conclusions is also that brief TLV is safe. We and others have already demonstrated that haemodynamics and gas exchange were not altered during TLV in rabbits, lambs, and cats. However, our preliminary experiments demonstrated an increased mortality in rabbits subjected to prolonged periods of Hypo-TLV, i.e. >30 min. The cause of these deaths was not pinpointed but that problem was not seen in rabbits exposed to shorter periods of Hypo-TLV as in the present report. To our knowledge this is the first study demonstrating that short TLV is compatible with rapid resumption of spontaneous breathing without major hypoxia and with minimal lung trauma. Indeed, our rabbits resumed spontaneous breathing only a few hours after the end of TLV. They breathed normally and were maintained in a closed cage with air supplemented with oxygen during the initial 24–48 h of their recovery. This was to prevent any possibility of hypoxia since atelectasis may occur following TLV. Later, rabbits were kept in conventional cages and still breathed normally without hypoxia. Lung histology also demonstrated that Hypo-TLV did not exert a deleterious effect. We even observed protection against pulmonary oedema and congestion in the Hypo-TLV group, further demonstrating the potency of the protection against cardiac mechanical dysfunction.

A recent editorial emphasized the clinical relevance of inducing cooling by breathing chilled liquids. The present report further proposes a strategy that would be easy to manage by adding a short period of Hypo-TLV to conventional surface cooling. The translation would be easier in patients who are already intubated and mechanically ventilated, e.g. patients with atherosclerotic vascular disease and post-operative myocardial infarction following high-risk surgery for whom immediate coronary revascularization is not possible. Similarly, Hypo-TLV might be useful in myocardial ischaemia secondary to profound blood loss or hypovolaemia. Whether conscious patients awaiting revascularization for acute ST-segment elevation myocardial infarction would benefit from Hypo-TLV and the attendant delay would depend on the projected time before the proposed intervention and reperfusion could be accomplished. However, as rapid cooling virtually stops infarct extension, the time needed for anaesthesia and intubation might not represent a serious limitation. Liquid ventilation could also be a strategy to induce cooling as an aid to resuscitation. In the hours following cardiac arrest and resumption of spontaneous circulation, therapeutic hypothermia is indeed already recommended by international guidelines. The method of rapid induction described here would likely increase its effectiveness.

In conclusion, rapid cooling instituted by a brief episode of hypothermic TLV in rabbits abolished most consequences of ischaemia and resulted in potent infarct size reduction, early and complete contractile recovery, protection against myocardial cellular lesions, and prevention of mitochondrial dysfunction.

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