Activities and expression pattern of the carbohydrate sulfotransferase GlcNAc6ST-3 (I-GlcNAc6ST): functional implications

Jin Kyu Lee, Annette Bistrup, Annemieke van Zante, and Steven D. Rosen

Department of Anatomy and Program in Immunology, University of California, San Francisco, CA 94143-0452, USA

Received on June 30, 2002; revised on September 18, 2002; accepted on September 25, 2002

In recent years, a family of five GlcNAc-6-O-sulfotransferases, called the GlcNAc6STs, has been molecularly cloned. One of these, GlcNAc6ST-2 (originally named HEC-GlcNAc6ST or LSST), shows a very restricted expression at the mRNA level in high endothelial cells (HECs) of lymph nodes high endothelial venules (HEVs). This enzyme has been shown to be involved in elaborating the 6-sulfo sLex structure on a set of mucin-like acceptors within HECs, thus providing a critical recognition determinant for L-selectin during the process of lymphocyte homing to lymph nodes. Limited information has been available about the closely related sulfotransferase known as GlcNAc6ST-3 (I-GlcNAc6ST). Here, employing transfection experiments with a series of glycoprotein acceptors, we report that this sulfotransferase has a marked preference for sulfating O-linked sugars of mucin-type acceptors, whereas other sulfotransferases in the family (GlcNAc6ST-1, GlcNAc6ST-2) and a Gal-6-O-sulfotransferase exhibit strong activity on both mucin-type acceptors and glycoproteins with predominantly N-linked chains. PCR analysis of cDNAs derived from a panel of tissues and purified cell populations confirms the strong expression of GlcNAc6ST-3 in gut-associated tissues and extends the expression to include lymphocytes. In contrast to GlcNAc6ST-2, GlcNAc6ST-3 transcripts are present minimally, if at all, in HECs; moreover, this enzyme is not able to generate the 6-sulfo sLex epitope in transfected cells. These latter findings argue that GlcNAc6ST-3 is not involved in generating HEV-expressed ligands for L-selectin.

Key words: GlcNAc-6-O-sulfotransferase/homing/ L-selectin/mucin/sulfation

Introduction

Carbohydrate sulfation and protein sulfation are important extracellular modifications that contribute to a variety of biological recognition events at the cell surface and in the extracellular matrix (reviewed in Hooper et al., 1996; Bowman and Bertozzi, 1999; Fukuda et al., 2001). Just as the kinases that mediate the diverse phosphorylation events within the cytosol have received great attention in recent years, the Golgi-associated sulfotransferases that impart specific sulfation modifications are now stimulating considerable interest. Over the last several years, approximately 30 Golgi-associated sulfotransferases have been identified at the molecular level in humans (reviewed in Habuchi, 2000; Hemmerich and Rosen, 2000; Fukuda et al., 2001). Several subfamilies have been delineated that show enhanced sequence conservation and related catalytic activities. One of these subfamilies, known as the GlcNAc-6-O-sulfotransferases (GlcNAc6STs), catalyze the transfer of sulfate from 3′-phosphoadenosine 5′-phosphosulfate to the C-6 position of GlcNAc (Fukuda et al., 2001). The nomenclature for the five members of this group is diverse, reflecting the participation of numerous groups in the discovery and characterization of these enzymes (Table I). We employ the GlcNAc6ST nomenclature, which designates the enzymes in order of their cloning.

GlcNAc6ST-1 was the first member to be identified at the molecular level (Uchimura et al., 1998a,b). It is expressed in a wide variety of human tissues, as judged by northern analysis and the frequency of expressed sequence tags (ESTs) in the NCBI human database. By contrast, GlcNAc6ST-2, also known as HEC-GlcNAc6ST (Bistrup et al., 1999) or L-selectin ligand sulfotransferase (LSST) (Hiraoka et al., 1999), shows a very narrow expression pattern in high endothelial cells (HECs) of high endothelial venules (HEVs) of secondary lymphoid organs. Additionally, it is ectopically expressed in certain adenocarcinomas (Seko et al., 2002; Uchimura et al., 2002). GlcNAc6ST-2 is essential to the process of L-selectin-mediated lymphocyte homing by elaborating a critical GlcNAc-6-sulfate modification on the HEV-expressed ligands for L-selectin (Hammerich et al., 2001a). This modification is found in the context of 6-sulfo sLex (Sia\(^{[\alpha-L-Fuc1\rightarrow3\alpha-Gal\beta1\rightarrow4}\)GlcNAc) on O-linked chains borne by a series of sialomucins, including CD34, podocalyxin, MadCAM-1, and GlyCAM-1 (Hammerich et al., 1995; Mitsuoka et al., 1998; Yeh et al., 2001).

GlcNAc6ST-2 is also essential for the formation of the epitope of MECA-79, a monoclonal antibody that stains HEV and blocks L-selectin-dependent adherence of lymphocytes (Berg et al., 1991; Hammerich et al., 1994, 2001a; Yeh et al., 2001). The closest relatives of GlcNAc6ST-2 are GlcNAc6ST-3 and GlcNAc6ST-5, also known as I-GlcNAc6ST (Lee et al., 1999) and C-GlcNAc6ST (Akama et al., 2000; Hammerich et al., 2001b), respectively,
because of their prominent expression in the intestine and cornea, respectively. These three sulfotransferases map to the same region of chromosome 16 and show the highest degree of sequence homology within the GlcNAc6ST subfamily (Hemmerich et al., 2001b). Mutations in the gene for GlcNAc6ST-5 underlie macular corneal dystrophy in humans, which is attributable to the normal role of this enzyme in transferring sulfate to C-6 of GlcNAc within keratan sulfate (Akama et al., 2000, 2001). The remaining member of the family, GlcNAc6ST-4, like GlcNAc6ST-1, shows a broad expression pattern in human tissues (Bhakta et al., 2000; Kitagawa et al., 2000; Uchimura et al., 2000).

This enzyme is also reported to be a chondroitin 6-O-sulfotransferase that transfers sulfate to C-6 of GalNAc (Kitagawa et al., 2000).

The present study focuses on GlcNAc6ST-3 because of the limited functional and biochemical information available about this enzyme. In comparing GlcNAc6ST-3 with GlcNAc6ST-1 and GlcNAc6ST-2, we find striking differences with respect to the acceptor specificities and the sulfated products that can be formed. Moreover, we have expanded the survey of tissue expression for GlcNAc6ST-3 by using a more extensive panel of tissue cDNAs and including isolated cell populations. The pertinence of our findings to L-selectin ligand synthesis is discussed.

**Results**

*Sulfation of different glycoprotein acceptors*

As already reviewed, GlcNAc6ST-2, a close relative of GlcNAc6ST-3, has been established to impart L-selectin ligand activity to several mucins (Bistrup et al., 1999; Hiraoka et al., 1999; Hemmerich et al., 2001a; Yeh et al., 2001). Therefore, we wanted to compare these two enzymes in their ability to sulfate O-linked and N-linked chains. We chose a series of glycoproteins with predominantly O-linked chains or N-linked chains as acceptors. The former consisted of mucin scaffolds (CD34, GlyCAM-1, and MAdCAM-1) that serve as L-selectin ligands and as acceptors. The latter consisted of mucin O-linked chains (CD34, GlyCAM-1, and MAdCAM-1) that serve as L-selectin ligands and as acceptors. We chose a series of glycoproteins with predominantly O-linked chains or N-linked chains as acceptors. We included in the comparison another member of the GlcNAc6ST family (GlcNAc6ST-1) and an example of a Gal-6-O-sulfotransferase, that is, KSGal6ST (Fukuta et al., 1997).

**Table 1. The family of GlcNAc-6-O-sulfotransferases**

<table>
<thead>
<tr>
<th>Systematic name</th>
<th>Other designations</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>GlcNAc6ST-1</td>
<td>GlcNAc6ST, GST-2, CHST2</td>
<td>Uchimura et al., 1998a,b; Li and Tedder, 1999</td>
</tr>
<tr>
<td>GlcNAc6ST-2</td>
<td>HEC-GlcNAc6ST, LSST, GST-3, CHST4</td>
<td>Bistrup et al., 1999; Hiraoka et al., 1999</td>
</tr>
<tr>
<td>GlcNAc6ST-3</td>
<td>I-GlcNAc6ST, GST-4α, CHST5</td>
<td>Lee et al., 1999</td>
</tr>
<tr>
<td>GlcNAc6ST-4</td>
<td>C6ST-2, GST-5, CHST7</td>
<td>Bhakta et al., 2000; Kitagawa et al., 2000; Uchimura et al., 2000</td>
</tr>
<tr>
<td>GlcNAc6ST-5</td>
<td>C-GlcNAc6ST, GST-4β, CHST6</td>
<td>Akama et al., 2000; Hemmerich et al., 2001b</td>
</tr>
</tbody>
</table>

**Fig. 1.** Sulfation of various glycoproteins by GlcNAc6STs and KSGal6ST. COS cells were transfected with combinations of plasmids encoding GlyCAM-IgG, CD34-IgG, MAdCAM-IgG, NCAM-IgG, ICAM-IgG, Fractalkine-IgG or IgG, and each sulfotransferase as indicated. Transfected cells were cultured in the presence of [35S] sulfate. The recombinant IgG chimeric proteins were purified from the conditioned medium by protein A. (A) The purified proteins were analyzed by SDS-PAGE followed by autoradiography. (B) The proteins were visualized by Coomassie staining.
Characterization of a GlcNAc-6-O-sulfotransferase

Fig. 2. N-glycanase treatment of glycoproteins sulfated by GlcNAc6STs and KSGal6ST. The purified glycoproteins sulfated by GlcNAc6STs and KSGal6ST (as described in Materials and methods and Figure 1) were incubated in the presence (+) or absence (−) of N-glycanase and analyzed by SDS-PAGE and autoradiography.

Fig. 3. Dependency of GlcNAc6ST-3 on the core 2 structure. CHO cells were transfected with combinations of plasmid encoding CD34/IgG, MAδCAM/IgG, or vector only (Mock), and GlcNAc6ST-3. A plasmid encoding core 2 GlcNAcT-I was either present (+) or absent (−) during transfection. Transfected cells were cultured in the presence of [35S] sulfate. The recombinant IgG chimeric proteins were purified from the conditioned medium by protein A. The purified proteins were analyzed by SDS–PAGE followed by autoradiography.
bead immunoselection (see Materials and methods). In parallel, T and B cells (CD3⁺ and CD19⁺ cells, respectively) were isolated from tonsillar cell suspensions using fluorescence-activated cell sorting. Semi-quantitative PCR was performed on cDNAs generated from these specimens. As shown in Figure 5B, we observed strong expression of GlcNAc6ST-2 in the isolated HECs, consistent with our previous observations (Bistrup et al., 1999). Interestingly, low levels of GlcNAc6ST-2 transcripts were also detected in the cDNAs derived from the populations of B and T cells, which were estimated to be >99% pure by flow cytometry analysis. A converse expression pattern was observed for GlcNAc6ST-3 (Figure 5A): B and T cells showed significant expression, whereas only a trace level was detectable in HECs. This minimal expression was very likely attributable to contamination of the HECs with adherent lymphocytes. We also found only barely detectable levels of GlcNAc6ST-3 transcripts in HECs isolated from mouse lymph node or Peyer’s patches (data not shown).

Generation of the 6-sulfo sLex epitope

Previous work has shown that both GlcNAc6ST-1 and GlcNAc6ST-2 can generate the 6-sulfo sLex epitope in transfected cells (Bistrup et al., 1999; Kimura et al., 1999; Kanamori et al., 2002; Uchimura et al., 2002). This determinant can be detected by two monoclonal antibodies of nearly identical specificity: G72 (which does not require fucose for activity) and G152 (which exhibits an absolute dependency on fucosylation) (Mitsuoka et al., 1998). We determined the ability of GlcNAc6ST-3 to produce the 6-sulfo sLex on the surface of transfected cells. To provide for the generation of sLex on a core 2 branch, we utilized CHO cells (known as CHO/FTVII/Core2GlcNAcT) that stably expressed Core2GlcNAcT-I and fucosyltransferase VII activities (Bistrup et al., 1999). We transiently transfected these cells with a cDNA encoding CD34 and a cDNA for GlcNAc6ST-2 or GlcNAc6ST-3. Using flow cytometry with the G152 monoclonal antibody, we probed for the 6-sulfo sLex epitope on the cell surface. As shown in Figure 6A, the CHO/FTVII/C2GnT cells transfected with GlcNAc6ST-2 were very strongly positive for the G152 epitope. In contrast, transfection with GlcNAc6ST-3 did not result in G152 staining above background. To confirm that GlcNAc6ST-3 did, in fact, cause increased sulfation of CD34, we labeled CHO cells with [35S]-SO₄ after transfection with a CD34 cDNA and a cDNA for GlcNAc6ST-2 or GlcNAc6ST-3. Lysates were analyzed by SDS–PAGE and autoradiography. Transfection with either sulfotransferase resulted in enhanced sulfation of CD34 relative to that in control cells (not shown). However, only GlcNAc6ST-2 could generate the 6-sulfo sLex determinant on CD34, as detected by immunoprecipitation with the G72 monoclonal antibody (Figure 6B).
distinct with respect to sequence homology and activities from the heparan sulfate GlcNSO$_3$-6-O-sulfotransferase (HS6ST) subfamily of three enzymes that are responsible for addition of sulfate to the 6-position of GlcN or GlcNAc in heparan sulfate chains (reviewed in Fukuda et al., 2001). It is strongly suspected that the existence of multiple sulfotransferases for a given modification underlies higher-order specificities, involving the elaboration of sulfate modifications within the context of different oligosaccharides (Bowman and Bertozzi, 1999; Fukuda et al., 2001). Further acceptor specificity may derive from features of the protein scaffold. An emerging view is that different sulfotransferases acting in concert with specific glycosyltransferases provide for a huge diversity of sulfate-based determinants (“sulfotopes”) that enable a broad range of biological recognition events (Bowman and Bertozzi, 1999).

Among the members of the GlcNAc6ST subfamily, two members have been extensively characterized with respect to their function, as reviewed in the Introduction. GlcNAc6ST-2 is strongly implicated as one of the contributing enzymes in the synthesis of the 6-sulfo sLex determinant on L-selectin ligands, as well as the formation of the MECA-79 epitope carried by these ligands. Akama et al. (2000, 2001) have demonstrated the involvement of GlcNAc6ST-5 in the elaboration of the glycosaminoglycan chains of keratan sulfate. Although GlcNAc6ST-3 is highly homologous to GlcNAc6ST-5, and it is likely that their genes arose by gene duplication, GlcNAc6ST-3 is not able to generate highly sulfated keratan sulfate (Akama et al., 2001). Also, when we compared the tissue distributions of GlcNAc6ST-3 and GlcNAc6ST-5, notable differences were found. The former was strongly expressed in tissues of the gut, whereas the latter was not. A commonality was their expression in peripheral blood mononuclear cells and brain.

To address the issue of the acceptor specificity for GlcNAc6ST-3, we analyzed the ability of several members of the GlcNAc6ST subfamily to sulfate N-linked versus O-linked chains in a series of glycoproteins. Compared to its closest relatives in the GlcNAc6ST family, GlcNAc6ST-3 was much more selective for O-linked chains of mucin-type acceptors with minimal activity in sulfating glycoproteins with predominantly N-linked glycosylation. No such preference was found for GlcNAc6ST-2, despite the fact that the functionally relevant HEV-expressed acceptors for this enzyme are mucin-type glycoproteins (Puri et al., 1995; Hemmerich et al., 2001a). As expected, the sulfation of O-linked chains by GlcNAc6ST-3 within CHO cells required the presence of the core 2 branch. Our findings are compatible with the recent biochemical analyses in which cell-free sulfotransferase assays were performed with a series of recombinantly expressed GlcNAc6STs and defined oligosaccharides acceptors (Bowman et al., 2001; Seko et al., 2002; Uchimura et al., 2002). GlcNAc6ST-1 and GlcNAc6ST-2 were able to sulfate GlcNAc on a core 2 acceptor (Bowman et al., 2001; Uchimura et al., 2002), but both were also active on GlcNAc-Man structures found in N-Linked chains. In contrast, GlcNAc6ST-3 was found to act on the core 2 acceptor but exhibited very limited activity on GlcNAc-Man structures, consistent with our observation that this enzyme preferred mucin-type over N-linked acceptors in

**Discussion**

The GlcNAc-6-sulfate modification is found on keratan sulfate and heparan sulfate glycosaminoglycans, as well as on smaller glycans of N- and O-linked carbohydrate chains in various glycoproteins. Among the mucins that carry this modification are respiratory mucins of cystic fibrosis patents (Lo-Guidice et al., 1994), HEV-expressed ligands for L-selectin (Hemmerich et al., 1995; Mitsuoka et al., 1998), and human colon carcinoma mucins (Capon et al., 1997).

GlcNAc-6-O-sulfotransferase activities have been demonstrated in extracts from a variety of tissues and tumors (Carter et al., 1988; Goso and Hotta, 1993; Spiro et al., 1996; Bowman et al., 1998; Nakazawa et al., 1998; Hasegawa et al., 2000; Seko et al., 1998; Delmotte et al., 2001). Underlying these activities are members of the GlcNAc6ST subfamily of carbohydrate sulfotransferases, of which five have been cloned to date. This subfamily is

**Figure 6.** Generation of 6-sulfo sLex by GlcNAc6ST-2 but not GlcNAc6ST-3. (A) CHO/FTVII/Core2GlcNAcT-I cells were transfected with cDNAs encoding CD34 and either GlcNAc6ST-2 or GlcNAc6ST-3. Cells were stained with the G152 monoclonal antibody and analyzed by flow cytometry. Histogram shows staining for the transfections with GlcNAc6ST-2 (bold) or GlcNAc6ST-3 (stippled), or staining with the isotype control antibody for the GlcNAc6ST-2 transfected cells (solid). (B) CHO cells were transfected with cDNAs encoding FTVII, core 2, GlcNAcT-I, full-length CD34, and either GlcNAc6ST-2 or GlcNAc6ST-3 or vector alone (mock). Transfected cells were cultured in the presence of [35S]sulfate, and lysates were prepared. Equal amounts of protein were subjected to immuno-precipitation with the G72 monoclonal antibody. The precipitates were analyzed by SDS–PAGE.

Downloaded from https://academic.oup.com/glycob/article-abstract/13/4/245/565740 by guest on 22 January 2019
cotransfection assays. Our findings with respect to the acceptor preference for GlcNAc6ST-2 appear to be at variance with those of Hiraoka et al. (1999) who studied the mouse homolog of this enzyme. They, like us, found that this enzyme could direct the sulfation of IgG chimeras of GlyCAM-1, MadCAM-1, and CD34, but they observed no detectable activity above background on NCAM-1/IgG. Whether this discrepancy is attributable to the species difference or to technical issues (e.g., level of enzyme or acceptor glycoprotein) remains to be determined.

Our previous analysis of GlcNAc6ST-2 gene–targeted mice has established a clear role for this enzyme in the elaboration of HEV ligands for L-selectin and in the generation of the MECA-79 epitope on these ligands (Hemmerich et al., 2001a). However, the phenotype of the null mice was only partial in that L-selectin ligands and MECA-79 reactivity were still present in HEVs, albeit to a greatly reduced extent and restricted to the abluminal aspects of the HEVs. Also, we have observed a reduced but still substantial level of sulfation in HEV-expressed ligands from GlcNAc6ST-2 null mice (Van Zante and Rosen, unpublished data). One of the candidates for these residual activities has been GlcNAc6ST-3. Two of our present findings render this possibility remote. First, we found only a trace level of GlcNAc6ST-3 transcripts in isolated HECs, which was very likely attributable to contaminating lymphocytes in the HEC preparation. Second, our transfection experiments indicate that GlcNAc6ST-3 is unable to direct the synthesis of the 6-sulfo sLex determinant, a critical structure for L-selectin ligands. A more likely candidate to account for the residual ligand and MECA-79 reactivity in GlcNAc6ST-2 null mice is GlcNAc6ST-1, which is known to be expressed in HEVs (Uchimura et al., 1998a). This enzyme can participate in the generation of functional L-selectin ligands, the 6-sulfo sLex determinant, and the MECA-79 epitope (Kimura et al., 1999; Kanamori et al., 2002; Uchimura et al., 2002). It should be noted that GlcNAc6ST-1 is also implicated in the generation of the 6-sulfo sLex determinant in the mouse embryo (Fan et al., 1999).

We found evidence for GlcNAc6ST-2 expression in B and T cells, although at a markedly lower level than in HECs (Figure 5B). This low level of expression likely explains the failure to detect this transcript by in situ hybridization in the lymphocyte-rich regions of lymph nodes (Bistrup et al., 1999; Hiraoka et al., 1999). Ohmori et al. (2000) reported the presence of transcripts for GlcNAc6ST-2 in the human lymphoid leukemia line Nal4. However, our results provide the first direct evidence of its presence in primary lymphocytes. The enzyme may account for the occurrence of the 6-sulfo sLex epitope on subpopulations of lymphocytes (Kannagi and Kanamori, 1999).

A novel finding of the present study is the expression of GlcNAc6ST-3 in mononuclear cells. This was established in sets of cDNAs derived by three independent means (Figures 4A, C, and 5A). The preference of GlcNAc6ST-3 to sulfate mucin acceptors focuses attention on leukocyte cell surface molecules with mucin domains. In fact, several such molecules (CD43, CD44, and CD45) are reported to be sulfated (Giordanengo et al., 1995; Maiti et al., 1998; Brown et al., 2001), although the nature of the sulfated moieties has not been reported. In the case of CD44, ligand binding activity (i.e., hyaluronic acid binding) depends on its sulfation (Maiti et al., 1998; Brown et al., 2001). PSGL-1, another sialomucin, is broadly distributed on leukocytes (reviewed in McEver and Cummings, 1997) and carries tyrosine sulfation. On NK subpopulations, PSGL-1 is further decorated in a cell type-specific manner with the PEN5 epitope, which is thought to involve a GlcNAc-6-sulfate modification (Andre et al., 2000). This epitope is of functional interest because it is implicated in L-selectin ligand activity on these cells. The potential involvement of GlcNAc6ST-3 in modulating the function of cell surface mucins on leukocytes deserves further attention.

The present study confirms and extends the original finding that GlcNAc6ST-3 is strongly expressed in the gastrointestinal tract. Studies by Seko et al. (2000, 2002) indicate that GlcNAc6ST-3 corresponds to the GlcNAc-6-O-sulfotransferase activity detected in normal human colon mucosa. In view of the preference of GlcNAc6ST-3 for mucin-type acceptors, demonstrated herein, a role in the modification of gut-associated mucins should be considered. A variety of secreted and membrane-bound mucins are found in the gut (reviewed in Kim and Gum, 1995). Normal functions and pathological roles ascribed to mucins are varied, including mechanical protection, lubrication, facilitating metastasis of carcinoma cells, and providing attachment sites for microbes. Most recently, a gene targeting approach has implicated the Muc2 mucin in the suppression of colorectal cancer (Veleich et al., 2002). Mucins exhibit an enormous diversity of sulfated O-linked chains (Lo-Guidice et al., 1994; Capon et al., 1997). Mucin function undoubtedly will depend on these sulfation modifications whether through influence on overall physicochemical properties or via sulfotopes involved in specific recognition events. Further work on the contribution of GlcNAc6ST-3 to the structure and function of gut-associated mucins under both normal and pathological circumstances is clearly warranted.

Materials and methods

**Sulfation of defined glycoproteins by GlcNAc6STs**

cDNAs encoding Fc chimeras were constructed by amplifying the coding sequence corresponding to the entire coding region of GlyCAM-1 or the extracellular domains of human CD34 and human fractalkine by PCR and cloning the resulting fragments into the plG1 vector to yield fusion proteins with the hinge, CH2 and CH3 regions of human IgG1 at the C-terminus (Simmons, 1993). COS cells were transfected with vectors encoding one of these fusion proteins or the Fc alone plus a cDNA for one of the following sulfotransferases: KSGal6ST (pCDNA3.1), GlcNAc6ST-1 (pCDNA3.1), GlcNAc6ST-2 (pCDNA1.1), GlcNAc6ST-3 (pCDNA3.1), or the empty vector (pCDNA3.1). COS cells were transfected at 80% confluency using Lipofectamine (Life Technologies) in Opti-MEM (Life Technologies, Carlsbad, CA). Cells were grown for 6 h after transfection in Opti-MEM, then cultured for 72 h in serum-free medium (Dulbecco’s modified Eagle’s medium, Gibco BRL,
Carlsbad, CA) supplemented with Na$_2^{35}$S$\cdot$SO$_4$ (0.25 mCi/ml, 1400 Ci/mmol ICN). The recombinant fusion proteins were isolated from the conditioned medium by affinity binding on protein A–agarose (Watson et al., 1990). The proteins were separated by 10% SDS–PAGE and were visualized by Coomassie staining and autoradiography.

For the N-glycanase experiments, Fc-fusion proteins were produced as above. Each 2–5 µl of protein A–agarose-bound protein was boiled for 2 min in 20 mM sodium phosphate, pH 7.5, 50 mM ethylenediamine tetra-acetic acid (EDTA), 0.02% sodium azide, 0.5% SDS, and 5% β-mercaptoethanol. As a control, 25 µg of α1-acid glycoprotein was subjected to N-glycanase treatment. The denatured samples were treated with or without 10 U/ml of recombinant Peptide-N-Glycosidase F (Glyko, Novato, CA) in the presence of 20 mM sodium phosphate, pH 7.5, 50 mM EDTA, 0.02% sodium azide, 1% Nondiet P-40 (Sigma, St. Louis, MO) and 1% β-mercaptoethanol in a volume of 30 µl for 16 h at 37°C. The resulting samples were analyzed by SDS–PAGE as mentioned.

For the experiments to determine the presence of the 6-sulfo sLex determinant, CHO cells were transfected with cDNAs encoding full-length CD34, Core2GlcNAcT-I (pCDNA1.1), FTVI (pCDNA3.1), and either GlcNAc6ST-2 or -3 or the empty vector (pCDNA3.1). The transfected cells were cultured for 3 days in the presence of Na$_2^{35}$S$\cdot$SO$_4$. Whole cell lysates (0.1% Triton-X 100 in PBS) were prepared and equalized for protein content. Equal aliquots of each sample were separated by 10% SDS–PAGE and visualized by Coomassie blue staining and autoradiography. In parallel, equal aliquots were incubated with 8 µg G72 monoclonal antibody immobilized on Protein A–agarose (Zymed, San Francisco, CA). Bound proteins were separated by 10% SDS–PAGE and visualized by Coomassie staining and autoradiography. To determine the presence of the 6-sulfo sLex determinant by flow cytometry, CHO cells stably expressing Core2GlcNAcT-I and FTVI were transiently transfected by Lipofectamine as previously described (Bistrup et al., 1998) or a mouse IgM (Pharmingen, San Diego, CA) isotype control.

For the core 2 dependency experiments, CHO cells were transfected with a cDNA for GlcNAc6ST-3 (or the empty vector), together with a plasmid encoding one of a series of the Ig fusion proteins (CD34/IgG, ICAM/IgG NCAM/IgG or fractalkine/IgG) with or without a plasmid encoding Core2GlcNAcT-I (pCDNA1.1). Transfection methods and analysis of the recombinant fusion proteins were as already described for the COS cell transfections.

Reverse transcriptase PCR experiments

Primers for each GlcNAc6ST were designed to avoid cross-reactivity among the GlcNAc6ST cDNAs. The following primers were used:

- GlcNAc6ST-2: 5’-ATAAAGCTTGTGGATTGTTCAGGGCATTTCCAGGTAGACAGAT-3’ and 5’-AAGTGCGACTAGTCTTGCTGC-3’, which amplify a 467-bp fragment;
- GlcNAc6ST-3: 5’-AACATCACCCACGGTGCCGGGATCGGCAA-3’ and 5’-GTCAGCCGATGGCCACAGTGAAGCTGAATGTG-3’, which amplify a 257-bp fragment;
- GlcNAc6ST-5: 5’-GGTAGGATGATGCCAAAGTGAGCCGTTCA-3’ and 5’-TCCATAACCACATCCCACGGA-TCTGGACCT-3’, which amplify a 262-bp fragment;
- HPRT: 5’-CCTGCTGATTACATCAGAACGCTG-3’ and 5’-TCCACACTTCGTTGGCTCCT-3’, which amplify a 300-bp fragment;
- G3PDH: 5’-TGAAGTCGAGTCAACGGATTTGT-3’ and 5’-CATGTGGCCCATGAGTCCACC-3’, which amplify a 983-bp fragment.

Each PCR reaction was carried out in a total volume of 10 µl of 1× Klen Taq buffer (Clontech, Palo Alto, CA) containing 500 µM dNTPs, 400 nM primers, 0.2 µl Klen Taq Advantage DNA polymerase mix (Clontech), and twofold serially diluted cDNA as template. Thermocycling conditions were as follows: 1 cycle of 3 min at 94°C, 30 s at 65°C (60°C for HPRT), 1 min at 68°C and 35 cycles (33 cycles for HPRT, 22 cycles for G3PDH) of 30 s at 94°C, 1 min at 6°C (60°C for HPRT), 1 min at 68°C, followed by 5 min at 68°C. The resulting amplified DNA was electrophoresed and visualized with ethidium bromide.

To verify the specificity of the PCR reactions, each of three primer pairs was tested against all three GlcNAc6ST cDNAs (20 ng of each cDNAs per reaction). Only the homologous combinations yielded PCR products of the predicted size. In addition the following PCR reactions were verified by directly sequencing the PCR product isolated from the agarose gel: GlcNAc6ST-3 product from CD19+ cells (B cells) cDNA; GlcNAc6ST-2 product from tonsillar B cell cDNA; and GlcNAc6ST-2 product from tonsillar T cell cDNA.

Tissue expression of GlcNAc6ST-3 and GlcNAc6ST-5 were determined using the Rapid-Scan Gene Expression Panel Human-24 (Origene Technologies, Rockville, MD). According to the manufacturer’s description, poly A+ RNA was used to synthesize first-strand cDNA, employing oligo(dT) primers. The amount of first-strand cDNAs from each tissue was normalized to contain an equivalent concentration of β-actin reverse transcripts. The cDNA pools from the 24 human tissues were diluted in water at four different concentrations in steps of 10-fold dilutions with the lowest concentration at approximately 1 pg/ml. Fragments were amplified by PCR using the primers for GlcNAc6ST-3 already given. PCR conditions were as follows: 1 cycle of 3 min at 94°C, 30 s at 68°C, 1 min at 68°C and 35 cycles of 30 s at 94°C, 30 s at 68°C, 1 min at 68°C, followed by 5 min at 68°C.

cDNAs derived from isolated purified populations of peripheral blood mononuclear cells were obtained from Clontech (K1428-1). Mononuclear cells were purified from peripheral blood on a Percoll gradient. Populations of CD4+ T cells, CD8+ T cells, and CD19+ cells (B cells) were obtained by immunomagnetic separation using
antibody conjugated Dynabeads (Dynal, Lake Success, NY). According to the manufacturer, the purity of the populations was > 95%, as evaluated by immunostaining the preparations. PCR was performed as described.

HECs were purified from human tonsils by immunomagnetic selection with MECA-79 by a modification of a previously described procedure (Girard and Springer, 1995; Sassetti et al., 2000). Surgical specimens were digested with a combination of collagenase A (Boehringer-Mannheim, Indianapolis, IN) and dispase I (Roche Diagnostics, Indianapolis, IN), and the resulting cell suspension (2 x 10^8) was incubated with 10 µg MECA-79 in 1 ml staining buffer (phosphate buffered saline [PBS] containing 1% bovine serum albumin) at 4°C for 20 min. Cells were collected by centrifugation, washed with staining buffer, and incubated with 10 µg of biotinylated mouse anti-rat IgM (Caltag, Burlingame, CA) at 4°C for 20 min. Finally, cells were collected, washed, and incubated with 40 µl of streptavidin-conjugated beads (Dynal) at 4°C for 20 min. MECA-79 positive cells were selected using a Dynal cell separation magnet and extensive washing as directed by the manufacturer. The purity of the resulting HEC preparation was 95% as determined by microscopic examination of cellular morphology with the major contaminant being adherent lymphocytes.

To purify T and B lymphocytes, tonsillar lymphocytes were prepared by mincing surgical specimens of human tonsil and flushing the loose lymphocytes through a 100 µm cell strainer with cold RPMI 1640 medium. The filtered cells were pelleted for 5 min at 1000 rpm and washed with PBS. T and B lymphocytes were purified by sorting after incubation with anti-CD3-FITC (Pharmingen) or anti-CD19-PE (CalTag). The purity of the sorted populations was > 99%. Total RNA was isolated from each purified cell population (3 x 10^6 T cells and 1 x 10^7 B cells) by lysis and extraction with RNAzol (Tel-Test, Friendswood, TX). To avoid genomic DNA contamination, the purified RNA was digested with RNase-free Dnase I (Gibco BRL) followed by ethanol precipitation. First strand cDNA was synthesized from 5 µg of total RNA primed with random hexamers using AMV reverse transcriptase (Gibco BRL). PCR reactions were carried out as described.

Acknowledgments

We thank Dr. Geoffrey Kanas of Northwestern Medical School for the CHO/FTVII/Core2GlcNAcT-I cells. We thank David Simmons of the ICRF, Oxford, Great Britain, for pIG plasmids encoding the Fc chimeras of NCAM-1 and ICAM-1. We thank Dr. John Lowe of the University of Michigan for the FTVII cDNA, Dr. Minoru Fukuda (Burnham Institute) for the core2GlcNAcT-I cDNA, and Dr. Sherman Fong of Genentech for the MadCAM-1 cDNA. Dr. Reiji Kannagi of the Aichi Cancer Center in Nagoya, Japan, kindly provided us with the G72 and G152 monoclonal antibodies. We acknowledge many helpful conversations with Dr. Stefan Hemmerich. The research was supported by grants to S.D.R. from the NIH (R01GM57541 and R37GM23547). J.K.L. was supported by a postdoctoral fellowship from the National Arthritis Foundation.

A.B. was supported by a postdoctoral fellowship from the American Heart Foundation (0020106Y).

Abbreviations

CHO, Chinese hamster ovary; EDTA, ethylenediamine tetra-acetic acid; EST, expressed sequence tag; FTVII, fucosyltransferase VII; HEC, high endothelial cell; HEV, high endothelial venule; LSST, L-selectin ligand sulfotransferase; PBS, phosphate buffered saline; PCR, polymerase chain reaction; SDS-PAGE, sodium dodecyl sulfate-polyacrylamide gel electrophoresis; sLex, sialyl Lewis x.

References


Uchimura, K., Muramatsu, H., Kannane, T., Ogawa, H., Yamakawa, T., Fan, Q.W., Mitsuoka, C., Kannagi, R., Habuchi, O., Yokoyama, I., and others. (1998b) Human N-acetylgalactosamine-6-O-sulfotransferase


