

# The Atherogenic Lipoprotein Profile Associated With Obesity and Insulin Resistance Is Largely Attributable to Intra-Abdominal Fat

Delma J. Nieves, Miriam Cnop, Barbara Retzlaff, Carolyn E. Walden, John D. Brunzell, Robert H. Knopp, and Steven E. Kahn

Obesity and insulin resistance are both associated with an atherogenic lipoprotein profile. We examined the effect of insulin sensitivity and central adiposity on lipoproteins in 196 individuals (75 men and 121 women) with an average age of 52.7 years. Subjects were subdivided into three groups based on BMI and their insulin sensitivity index ( $S_I$ ): lean insulin sensitive ( $n = 65$ ), lean insulin resistant ( $n = 73$ ), and obese insulin resistant ( $n = 58$ ). This categorization revealed that both obesity and insulin resistance determined the lipoprotein profile. In addition, the insulin-resistant groups had increased central adiposity. Increasing intra-abdominal fat (IAF) area, quantified by computed tomography scan and decreasing  $S_I$ , were important determinants of an atherogenic profile, marked by increased triglycerides, LDL cholesterol, and apolipoprotein B and decreased HDL cholesterol and LDL buoyancy (Rf). Density gradient ultracentrifugation (DGUC) revealed that in subjects who had more IAF and were more insulin resistant, the cholesterol content was increased in VLDL, intermediate-density lipoprotein (IDL), and dense LDL fractions whereas it was reduced in HDL fractions. Multiple linear regression analysis of the relation between the cholesterol content of each DGUC fraction as the dependent variable and IAF and  $S_I$  as independent variables revealed that the cholesterol concentration in the fractions corresponding to VLDL, IDL, dense LDL, and HDL was associated with IAF, and that  $S_I$  additionally contributed independently to VLDL, but not to IDL, LDL, or HDL. Thus an atherogenic lipoprotein profile appears to be the result primarily of an increase in IAF, perhaps via insulin resistance. *Diabetes* 52:172–179, 2003

From the Division of Metabolism, Endocrinology and Nutrition, Department of Medicine, Veterans Affairs Puget Sound Health Care System and Harborview Medical Center, University of Washington, Seattle, Washington.

Address correspondence and reprint requests to Steven E. Kahn, ChB, VA Puget Sound Health Care System (151), 1660 S. Columbian Way, Seattle, WA 98108. E-mail: skahn@u.washington.edu.

Received for publication 10 May 2002 and accepted in revised form 2 October 2002.

D.J.N. and M.C. contributed equally to this work.

Current address for C.E.W. is College of Medicine Research Office, University of Arizona, Tucson, AZ.

CT, computed tomography; DGUC, density gradient ultracentrifugation; IAF, intra-abdominal fat; IDL, intermediate-density lipoprotein; LIR, lean insulin resistant; LIS, lean insulin sensitive; OIR, obese insulin resistant; Rf, LDL relative flotation; SCF, subcutaneous fat;  $S_I$ , insulin sensitivity index; WHR, waist-to-hip ratio.

Obesity has been clearly demonstrated to be associated with insulin resistance (1–4). Insulin resistance in turn has been found to be linked with many of the conditions typically associated with obesity (5–7) and other conditions that appear to be less typically associated with simple obesity (8). These effects have generally been observed across sex (9,10), ethnic (11–13), and glucose tolerance categories (9,14–19).

In the evaluation of obesity, it has become apparent that it is not only the magnitude of the increase in fat mass, but also the site of distribution that is an important determinant of the development of insulin resistance and the conditions typically associated with obesity. Although a proportion of the variance in insulin sensitivity appears to be related to a central distribution of body fat, it has been debated whether this association is determined primarily by the accumulation of fat in the intra-abdominal or subcutaneous depots (16,20–24). We recently examined this issue in a large cohort of subjects who had central fat distribution, as determined by computed tomography (CT) scan and related to insulin sensitivity (10). In this cross-sectional analysis, we found that intra-abdominal fat (IAF) was a more important determinant of insulin sensitivity than was subcutaneous fat (SCF), whereas SCF was the more critical variable associated with leptin levels. In an important finding, we also observed that some individuals that were considered lean based on their BMI had increased amounts of IAF and were insulin resistant, suggesting that the relation between IAF fat and insulin sensitivity is a continuum that is not influenced by BMI.

The accumulation of central fat and presence of insulin resistance have both been associated with the dyslipidemia seen in the metabolic syndrome (25). Thus an increase in the waist-to-hip ratio (WHR) has been found to be associated with small dense LDL particles (26), as has been an increase in IAF mass (27,28) or insulin sensitivity (29,30). This pattern of central fat distribution has also been associated with an increase in VLDL and intermediate-density lipoprotein (IDL) (26,31) and a decrease in HDL<sub>2</sub> cholesterol (26,32,33). Although these analyses strongly suggest that central fat distribution and/or insulin resistance is an important determinant of adverse changes in lipoprotein profile, we believed that a critical examination of this issue in a large number of apparently healthy

TABLE 1  
Subject characteristics

|  | LIS         | LIR          | OIR            |
|--|-------------|--------------|----------------|
| <i>n</i>   | 65          | 73           | 58             |
| M/F  | 21/44       | 28/45        | 26/32          |
| Age (years)  | 49.1 ± 1.2  | 54.8 ± 1.4†  | 54.1 ± 1.2†    |
| Weight (kg)  | 66.5 ± 1.4  | 70.8 ± 1.2*  | 89.3 ± 1.5‡,   |
| BMI (kg/m <sup>2</sup> )   | 23.3 ± 0.3  | 24.4 ± 0.2†  | 31.0 ± 0.4‡,   |
| WHR  | 0.79 ± 0.01 | 0.82 ± 0.01* | 0.89 ± 0.01‡,  |
| SCF area (cm <sup>2</sup> )  | 128.0 ± 7.9 | 189.5 ± 8.1‡ | 310.0 ± 17.2‡, |
| IAF area (cm <sup>2</sup> )  | 51.0 ± 3.7  | 87.7 ± 4.9‡  | 166.1 ± 9.3‡,  |
| Glucose (mmol/l)   | 5.2 ± 0.05  | 5.3 ± 0.04†  | 5.5 ± 0.05‡,§  |
| Insulin (pmol/l)   | 39.4 ± 1.9  | 60.8 ± 3.6‡  | 82.8 ± 5.3‡,   |
| S <sub>1</sub> [ $\times 10^{-5} \text{ min}^{-1}/(\text{pmol/l})$ ] | 11.18 ± 0.6 | 4.85 ± 0.17‡ | 3.62 ± 0.19‡,§ |

Data are means ± SE. \* $P < 0.05$ , † $P < 0.01$ , ‡ $P < 0.001$  vs. LIS; § $P < 0.05$ , || $P < 0.001$  vs. LIR.

subjects was required to determine the impact of differences in body fat distribution and insulin sensitivity on plasma lipids. Further, we held that an assessment of these associations along with findings from small intervention studies would provide additional insight into the potential sequence of events in the development of an atherogenic lipoprotein profile. Therefore, we measured plasma lipoproteins, including an assessment of cholesterol distribution by density gradient ultracentrifugation (DGUC), in apparently healthy middle-aged men and women who were not receiving treatment for a lipid disorder and in whom we had also quantified insulin sensitivity and IAF. Here we describe the findings of this study and strongly suggest that IAF is a critical determinant of an atherogenic lipoprotein profile, including perturbations in the distribution of cholesterol in the different lipoprotein fractions.

## RESEARCH DESIGN AND METHODS

**Subjects.** The study cohort was comprised of 196 individuals (75 men and 121 women) from the Greater Seattle area who had been recruited by advertisement to participate in a study of the effect of insulin sensitivity on lipoprotein profile alteration after egg consumption (34). Eligible subjects had to be apparently healthy with no history of diabetes or coronary or other vascular disease. In addition, they could not be receiving treatment for lipid disorders or hypertension and had to be free of renal or hepatic disease. At the time of screening, qualifying individuals had to have a fasting plasma glucose  $< 6.4$  mmol/l, LDL cholesterol  $< 190$  mg/dl, and triglycerides  $< 500$  mg/dl. Based on their BMI and insulin sensitivity index (S<sub>1</sub>), subjects were divided a priori into three groups: lean insulin sensitive (LIS), lean insulin resistant (LIR), and obese insulin resistant (OIR), using cutoff points of 27.5 kg/m<sup>2</sup> and  $7.0 \times 10^{-5} \text{ min}/(\text{pmol/l})$ , respectively. The cutoff point for obesity was determined using data from healthy populations and applying criteria set before the redefinition of overweight and obesity (NHANES II), and the cutoff point for insulin sensitivity was based on the highest value for this parameter among a group of apparently healthy obese subjects studied in Seattle (35).

**Characterization of body size and fat distribution.** A number of measures of body fat distribution were quantified. BMI was calculated as weight/height (kg/m<sup>2</sup>). WHR was calculated as waist circumference (cm) divided by hip circumference (cm), using measurements made according to the techniques established by Krotkiewski et al. (36). The waist circumference was measured one-third up the distance between the umbilicus and the xiphoid process. The hip circumference was measured 4 cm below the superior anterior iliac crest.

SCF and IAF areas were quantified by a CT scan at the level of the umbilicus (14). The border of the intra-abdominal cavity was outlined on the CT image, and total fat and IAF areas were quantified by selecting an attenuation range of -250 to -50 Hounsfield units using standard software. The SCF area was calculated by subtracting the IAF area from total fat area. The variability of these measures made by a single observer is 1.5%, and day-to-day variability is  $< 1\%$  (37).

**Insulin sensitivity.** S<sub>1</sub> was determined using the minimal model of glucose kinetics described by Bergman and colleagues (38,39) from the glucose and insulin data obtained during a frequently sampled intravenous glucose toler-

ance test performed after an overnight fast. The test was modified by the injection of tolbutamide, which served to improve parameter identifiability (40). The day-to-day variability of S<sub>1</sub> in our laboratory is 16.9% (41).

**Chemical analyses.** All analyses were performed on blood samples drawn after an overnight fast of at least 12 h. Samples were stored at  $-70^{\circ}\text{C}$  until assayed.

Plasma glucose levels were measured using the glucose oxidase method. Plasma immunoreactive insulin levels were determined using a modification of the double antibody method of Morgan and Lazarow (42).

Plasma triglycerides and total cholesterol were measured by enzymatic analytical chemistry. HDL cholesterol was precipitated using dextran sulfate and measured enzymatically. LDL was calculated according to the Friedewald equation:  $\text{LDL} = \text{TC} - \text{HDL} - (\text{TG}/5)$ , where TC is total cholesterol and TG is triglycerides. Apolipoprotein B was determined by nephelometry (43). Non-equilibrium DGUC was performed with the collection of 38 fractions in which cholesterol was measured (44). The LDL relative flotation (Rf), a measure of LDL particle buoyancy, was determined by dividing the fraction number containing the highest LDL cholesterol concentration by 38, the latter representing the total number of fractions sampled.

**Statistical analyses.** Comparison of demographic and metabolic variables among the LIS, LIR, and OIR groups and of lipoproteins across quartiles of body fat distribution and insulin sensitivity was performed by ANOVA, followed by Fisher's protected least significant differences test, where appropriate. The mean cholesterol differences at each of the 38 DGUC fractions were determined among groups using independent samples *t* test for equality of the means with a 95% confidence interval of the difference. Assessment of the relative contribution of IAF and SCF to S<sub>1</sub>, and of S<sub>1</sub>, IAF, SCF, and BMI to the cholesterol concentration in the individual DGUC fractions, was performed by multiple linear regression analysis. Data are presented as means ± SE unless stated otherwise. Data were considered significant at  $P \leq 0.05$ .

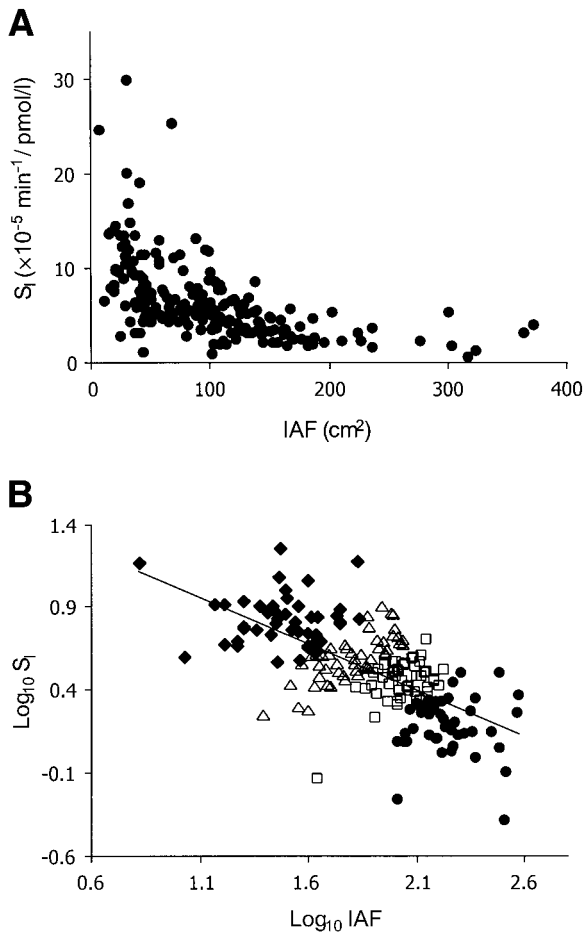
## RESULTS

### BMI, insulin sensitivity, and demographic measures.

As shown in Table 1, the three groups had a similar number of subjects and similar gender distribution, but the LIS group was younger. The two lean groups were fairly well matched for BMI, but differed markedly for S<sub>1</sub>, whereas the two insulin-resistant groups differed with regard to BMI and differed slightly with regard to S<sub>1</sub>. Plasma glucose and insulin levels increased across groups with insulin resistance and obesity.

**Body fat distribution and relationship with insulin sensitivity.** The WHR and abdominal fat areas for the three groups are provided in Table 1. The WHR increased significantly from the LIS group to the LIR and OIR groups. Similarly, SCF and IAF areas increased significantly across the three groups. Thus, although the BMI measurements of the lean groups were similar, the CT scan demonstrated that the LIR group had more abdominal fat than the LIS group.

When all subjects were considered together, SCF and



**FIG. 1.** Relationship between IAF and  $S_1$  in 196 individuals (75 men, 121 women). The two variables are related in a nonlinear manner (A) that is highly correlated ( $r^2 = 0.438$ ,  $P < 0.001$ ), as was demonstrated by log transformation (B). B: The interaction of  $S_1$  and IAF for each individual was determined as the quotient of the two variables; subjects were then subdivided into quartiles ( $n = 49$  per group;  $\blacklozenge, \blacktriangle, \blacksquare, \bullet$ ) based on this interaction. The extreme quartiles represent a group that has the least IAF and is the most insulin sensitive ( $\blacklozenge$ ) and a group that has the most IAF and is the most insulin resistant ( $\bullet$ ).

IAF were linearly related ( $r^2 = 0.192$ ,  $P < 0.001$ ). The relationship between each of these two fat depots and  $S_1$  was curvilinear and best described by a logarithmic function. This relationship was greater for IAF versus  $S_1$  ( $r^2 = 0.438$ ,  $P < 0.001$ ) (Fig. 1) than for SCF versus  $S_1$  ( $r^2 = 0.310$ ,  $P < 0.001$ ). Using multiple linear regression analysis, IAF was found to contribute to this relationship, whereas SCF did not. Further, IAF and  $S_1$  were related in men ( $r^2 = 0.528$ ,  $P < 0.001$ ) and women ( $r^2 = 0.383$ ,  $P < 0.001$ ).

**TABLE 2**  
Lipoprotein measurements

|                           | LIS           | LIR            | OIR              |
|---------------------------|---------------|----------------|------------------|
| <i>n</i>                  | 65            | 73             | 58               |
| Triglycerides (mg/dl)     | 88.5 ± 7.4    | 117.3 ± 7.4*   | 161.4 ± 12.0‡,   |
| Total cholesterol (mg/dl) | 187.9 ± 4.5   | 209.3 ± 3.7‡   | 206.4 ± 4.8†     |
| LDL cholesterol (mg/dl)   | 111.1 ± 3.6   | 132.1 ± 3.0‡   | 128.9 ± 4.2‡     |
| HDL cholesterol (mg/dl)   | 58.7 ± 1.9    | 53.3 ± 1.6*    | 44.8 ± 1.4‡,     |
| Apolipoprotein B (mg/dl)  | 85.0 ± 2.6    | 100.7 ± 2.3‡   | 105.2 ± 3.3‡     |
| Rf                        | 0.288 ± 0.003 | 0.278 ± 0.004* | 0.262 ± 0.004‡,§ |

Data are means ± SE. \* $P < 0.05$ , † $P < 0.01$ , ‡ $P < 0.001$  vs. LIS; § $P < 0.01$ , || $P < 0.001$  vs. LIR.

**Lipoprotein measurements and relation to body fat distribution and insulin sensitivity.** Plasma lipoprotein levels were consistent with a more atherogenic profile in the insulin-resistant subjects, and more so in the obese insulin-resistant individuals (Table 2).

To assess the relationship between body fat distribution and plasma lipoproteins and between  $S_1$  and plasma lipoproteins, we subdivided the cohort into quartiles based on IAF, SCF, BMI, and  $S_1$  measurements. As illustrated in Fig. 2, increasing IAF and decreasing  $S_1$  were both associated with the presence of a more atherogenic lipid profile. The change over quartiles for each lipoprotein variable was significant for both IAF and  $S_1$  ( $P < 0.001$ ). Similarly, there were significant relationships between SCF and the plasma lipoproteins and BMI and the plasma lipoproteins that portended a more atherogenic lipid profile, with the exception that SCF was not related to HDL cholesterol, whereas BMI was not related to LDL cholesterol. The relationships between the different measures of body fat distribution and plasma lipoproteins was greatest for IAF for all measures except HDL cholesterol, where it was slightly greater with BMI than with IAF (ANOVA  $F$  test: 15.82 vs. 13.44).

Because the a priori classification of the subjects into LIS, LIR, and OIR groups based on BMI and  $S_1$  did not eliminate the confounding effect of abdominal obesity, and because IAF and  $S_1$  were highly correlated in a nonlinear manner (Fig. 1A), we calculated the quotient of  $S_1$  and IAF as a composite measure of each individual's value for these two parameters. For comparison purposes, we then divided the study population into four equal-sized groups ( $n = 49$ ) based on the interaction of their individual measures of IAF and  $S_1$  (Fig. 1B). The individuals in the two extreme quartiles differed markedly for this composite measure of IAF and  $S_1$ , which allowed us to compare a group of subjects with the least IAF ( $34.3 \pm 2.0 \text{ cm}^2$ ) who were the most insulin sensitive [ $11.73 \pm 0.71 \times 10^{-5} \text{ min}^{-1}/(\text{pmol/l})$ ] with a group of subjects with the most IAF ( $182.5 \pm 9.7 \text{ cm}^2$ ;  $P < 0.001$ ) who were the most insulin resistant [ $2.86 \pm 0.14 \times 10^{-5} \text{ min}^{-1}/(\text{pmol/l})$ ;  $P < 0.001$ ]. The group that was the most insulin resistant and had the most IAF had a more atherogenic lipoprotein profile than the group that was the most insulin sensitive and had the least IAF. Thus, in the group with the most IAF that was the most insulin resistant, the triglyceride ( $177.8 \pm 13.5$  vs.  $75.8 \pm 4.8 \text{ mg/dl}$ ;  $P < 0.001$ ), total cholesterol ( $210.5 \pm 5.3$  vs.  $182.6 \pm 5.1 \text{ mg/dl}$ ;  $P < 0.001$ ), LDL cholesterol ( $130.1 \pm 4.7$  vs.  $106.0 \pm 4.1 \text{ mg/dl}$ ;  $P < 0.001$ ), and apolipoprotein B ( $107.9 \pm 3.5$  vs.  $79.8 \pm 2.6 \text{ mg/dl}$ ;  $P < 0.001$ ) levels were increased, whereas the HDL cholesterol level was de-

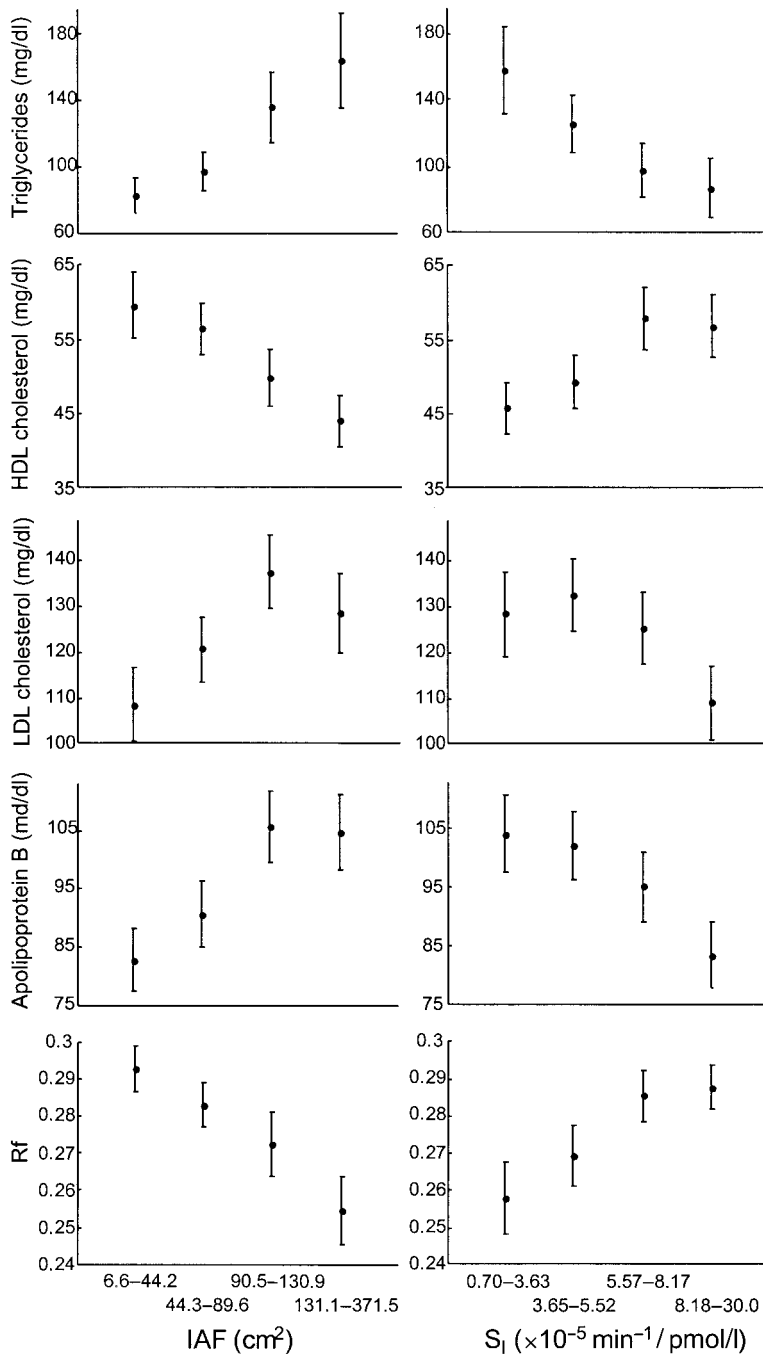
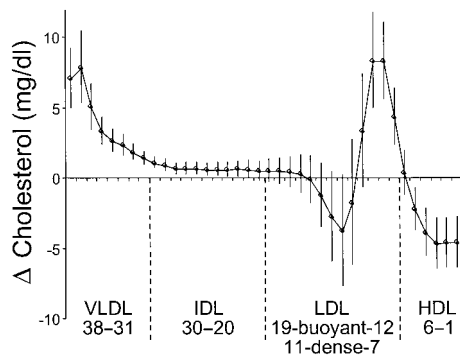


FIG. 2. Relationship between IAF area (*left panels*) and  $S_1$  (*right panels*) and different lipoproteins in 196 individuals. The whole cohort was subdivided into quartiles for IAF area and  $S_1$ . The range for each quartile is listed. Data are means  $\pm$  SE, with 95% confidence interval bars. Increasing IAF area and decreasing  $S_1$  are associated with a more atherogenic profile for each lipoprotein. For each lipoprotein variable, the change across quartiles was significant ( $P < 0.001$ ).

creased ( $44.4 \pm 1.8$  vs.  $60.9 \pm 2.2$  mg/dl;  $P < 0.001$ ) as was the measure of LDL buoyancy distribution ( $R_f$ :  $0.252 \pm 0.005$  vs.  $0.294 \pm 0.003$ ;  $P < 0.001$ ), when compared to the group that had the least IAF and was the most insulin sensitive. These two subject groups also differed markedly for BMI (most IAF, most insulin resistant:  $29.6 \pm 0.6$  kg/m<sup>2</sup> vs. least IAF, most insulin sensitive:  $22.9 \pm 0.3$  kg/m<sup>2</sup>;  $P < 0.001$ ). However, BMI made no additional contribution beyond that of IAF and  $S_1$  in determining the above differences in the lipoproteins between the group that was the most insulin resistant and had the most IAF and the group that was the most insulin sensitive and had the least IAF. **DGUC analysis.** The DGUC analysis allowed us to compare the lipoprotein cholesterol levels encompassing 38 fractions in the two groups that were defined based on the

interaction of IAF and  $S_1$ . When the cholesterol distribution in the most intra-abdominally obese, insulin-resistant group was compared with that in the most intra-abdominally lean, insulin-sensitive group, significant differences were observed in almost all fractions across the density gradient (Fig. 3). The most intra-abdominally obese, insulin-resistant group had more cholesterol in the VLDL, IDL, and dense LDL fractions. In addition, they had less cholesterol in the HDL fractions. Significant differences in the cholesterol content were maintained in fractions 1–4, 7–9, 23, 25, 26, and 29–38 after adjustment for the effect of BMI.

To determine the relative contributions of IAF and  $S_1$  to the cholesterol levels in the DGUC fractions, we performed multiple linear regression analyses for each fraction using data from the whole cohort. In the first model,



**FIG. 3.** Difference plots of the cholesterol concentration in 38 DGUC fractions for intra-abdominally obese, insulin-resistant subjects and intra-abdominally lean, insulin-sensitive subjects ( $n = 49$  per group). The two groups represent the extreme quartiles for the quotient of  $S_1$  and IAF, a composite measure that was determined for each individual (see Fig. 1B). Data are means  $\pm$  SE, with 95% confidence interval bars. The different fractions and their representation of the different cholesterol-containing lipoproteins are illustrated. The intra-abdominally obese, insulin-resistant group had more cholesterol in the VLDL, IDL, and dense LDL fractions and less cholesterol in the HDL fractions.

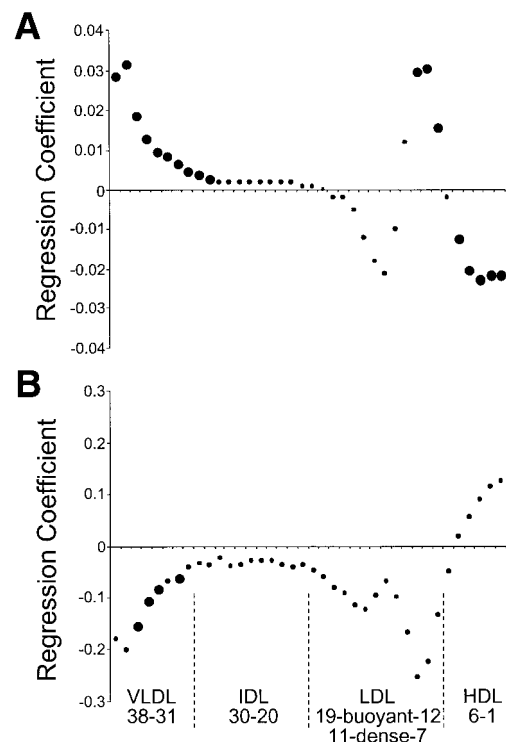
in which IAF and  $S_1$  were the only independent variables and the fractional cholesterol concentration was the dependent variable, the cholesterol concentration in the fractions corresponding to the VLDL, IDL, dense LDL, and HDL fractions was associated with IAF after adjustment for  $S_1$  (Fig. 4A). In this model, after adjustment for IAF,  $S_1$  contributed independently to some of the fractions corresponding to VLDL (fractions 32 and 34–36), but not to those in the IDL, LDL, or HDL ranges (Fig. 4B). In the second model, we included SCF as an independent variable along with IAF and  $S_1$ , with the fractional cholesterol concentration again being the dependent variable. In this model, the cholesterol concentration in the fractions corresponding to VLDL (fractions 31–38), IDL (fractions 29 and 30), buoyant LDL (fractions 12–14), dense LDL (fractions 7–9), and HDL (fractions 1–5) was associated with IAF, whereas  $S_1$  made no additional contribution to any of the fractions. In a third model, we included BMI as an independent variable together with IAF,  $S_1$ , and SCF, and found that the cholesterol concentration in the fractions corresponding to VLDL (fractions 31–38), IDL (fraction 30), and dense LDL (fractions 8–10) was associated with IAF, whereas  $S_1$  again made no additional contribution. Thus IAF, but not  $S_1$ , had independent effects to determine the cholesterol concentration beyond those of SCF and BMI.

**DISCUSSION**

By studying 196 apparently healthy subjects who differed in body habitus and insulin sensitivity, we were able to examine the relationship of these variables to the lipoprotein profile. We found that both obesity and insulin sensitivity are determinants of the lipoprotein profile, with an increased amount of IAF and insulin resistance being associated with a more atherogenic lipid profile: increased triglyceride, total cholesterol, LDL cholesterol, and apolipoprotein B levels and a decreased HDL cholesterol level and Rf (a measure of LDL buoyancy distribution). We were also able to extend the analysis by performing DGUC and examining the effect of differences in IAF and insulin sensitivity on the amount of cholesterol in the different

lipoprotein fractions. This analysis revealed that subjects who had greater amounts of IAF and a lower  $S_1$  had more cholesterol in the VLDL, IDL, and dense LDL fractions but less cholesterol in the HDL fractions. Further, the cholesterol in the VLDL, IDL, dense LDL, and HDL fractions was more strongly associated with IAF than with insulin sensitivity, the relation of the latter two being highly colinear. Insulin sensitivity did contribute to the cholesterol concentration in the VLDL range. When the effect of SCF was also accounted for, we found that IAF was still a determinant of VLDL, IDL, buoyant LDL, dense LDL, and HDL, whereas insulin sensitivity made no contribution to the cholesterol concentration. Finally, when SCF and the role of body size, quantified as BMI, were considered, IAF was still an independent determinant of VLDL, IDL, and dense LDL, whereas insulin sensitivity did not contribute to the cholesterol concentration. Thus it appears that IAF and insulin sensitivity are associated with an atherogenic lipid profile, with the effect of obesity being largely attributable to IAF.

The subjects we studied were recruited from the general population and were classified a priori into three groups based on their BMI and  $S_1$  values. This classification and the subsequent analyses allowed us to make a number of observations about the relationship between obesity and insulin resistance and, in turn, the association of body habitus and insulin sensitivity to plasma lipoproteins. First, it is apparent that BMI cannot be used as a simple means of determining whether an individual is or is not likely to have a lipoprotein profile associated with an



**FIG. 4.** Multiple linear regression analysis of the cholesterol content of each DGUC fraction, with IAF area and  $S_1$  as the independent variables. The regression coefficients for each of the 38 fractions are plotted for IAF (A) and  $S_1$  (B). The different fractions and their representation of the different cholesterol-containing lipoproteins are illustrated. Larger points represent statistically significant relationships determined by multiple linear regression analysis in which the one independent variable has been adjusted for the other ( $P < 0.05$ ).

increased risk of atherosclerosis. Apparently "lean" individuals can be insulin resistant and have increased amounts of IAF, with this characterization being associated with an adverse lipoprotein profile. Second, in insulin-resistant subjects, an increase in body size (based on BMI) is associated with a further change in the lipoprotein profile, specifically an increase in triglycerides and a decrease in both HDL cholesterol and Rf, but no increase in LDL cholesterol, which is consistent with an additional increase in atherogenic risk. This finding is in keeping with the well-recognized role of obesity as an important risk factor for cardiovascular disease (45). Third, IAF and  $S_1$  are highly correlated in a curvilinear manner, and this relationship is continuous irrespective of body size or sex. In turn, the phenotype of intra-abdominal obesity and insulin resistance is associated with a more atherogenic lipid profile. Fourth, SCF is increased in lean subjects who are insulin resistant compared with those who are insulin sensitive, and is further increased in individuals who are obese and insulin resistant. In addition, because SCF and IAF are positively and linearly related, they both increase as  $S_1$  declines. However, IAF is more strongly related to  $S_1$ , and therefore it would appear that IAF is more strongly associated with an adverse lipoprotein profile than is SCF, the latter more likely reflecting total body fat (46).

There has been a great deal of interest in the role of insulin resistance in the pathogenesis of a number of disorders, including type 2 diabetes, hypertension, and dyslipidemia, based on the clear association between insulin resistance and these conditions. A number of different names have been suggested to describe the constellation of associations between insulin resistance and these clinical findings, including syndrome X (6), the insulin resistance syndrome (47), the central adiposity syndrome (48), and the metabolic syndrome (25). In the course of these investigations, the roles of insulin resistance and central body fat distribution have been discussed, but a clear delineation of their roles in determining an atherogenic lipoprotein profile has not been achieved. The present study has provided a rather unique opportunity to further address these associations as it combines sophisticated, discriminatory measures of insulin sensitivity, body fat distribution, and plasma lipoproteins with a large cohort of subjects. Our findings regarding the relationship between IAF and insulin resistance with lipoprotein abnormalities is supported by intervention studies. Caloric restriction, which results in weight loss and decreased IAF, results in an improvement in an atherogenic lipid profile (49–54). However, the changes that follow caloric restriction are compounded somewhat by the fact that insulin sensitivity also improves with weight loss (53,55,56). Similarly, exercise training, which improves the lipoprotein profile, is also associated with reductions in body weight and IAF as well as an increase in insulin sensitivity (37,52,57,58). Both increased IAF and insulin resistance have been associated with increased hepatic lipase activity (33,59,60), leading to small dense LDL and decreased HDL<sub>2</sub> cholesterol levels (61–64); maneuvers to decrease hepatic lipase correct this dyslipidemia (53,65).

Collectively, previous intervention studies and the present cross-sectional analysis in a large cohort in whom discriminatory measures were used for quantification do

provide insight into these relationships. They strongly suggest that an abnormality in body fat distribution leads to the accumulation of intra-abdominal adiposity, which in turn is associated with the development of insulin resistance, followed by dyslipidemia. IAF would therefore be a contributor to an adverse lipoprotein profile and, thus, cardiovascular risk. To confirm whether insulin sensitivity is an intermediate step in IAF's effect or whether IAF has independent effects will require further study.

In summary, using a cross-sectional approach in a large cohort of apparently healthy subjects, we found that IAF appears to be a major determinant of an atherogenic lipid profile. Although insulin resistance is also associated with an adverse lipid profile, it would appear that this association is largely based on the fact that this parameter and IAF are closely correlated. However, it is possible that insulin sensitivity may play an additional contributory role in determining changes in cholesterol content in the VLDL fraction. Finally, these results suggest that approaches aimed at reducing IAF, via lifestyle or medication, would have a beneficial effect on both insulin sensitivity and plasma lipoproteins. Whether a similar relationship between IAF and insulin sensitivity holds for other features of the metabolic syndrome will await further evaluation.

#### ACKNOWLEDGMENTS

This work was supported in part by an American Egg Board grant; National Institutes of Health Grants DK-02456, DK-02654, DK-17047, DK-35816, HL-30086, and RR-37; the Medical Research Service of the Department of Veterans Affairs; and the McMillen Family Trust. D.J.N. was supported by a Medical Student Research Scholarship from the American Federation of Aging Research, and M.C. was supported by a fellowship from the Belgian American Educational Foundation.

We wish to thank the participants in this study who gave of their time. We also wish to acknowledge the nursing efforts of Diane Collins and the staff of the General Clinical Research Center at the University of Washington and the data management effort of Brian Fish. Dr. Santica Marcovina and the staff of the Northwest Lipid Research Laboratory and the Immunoassay Core of the Diabetes Endocrinology Research Center are acknowledged for the performance of the assays.

#### REFERENCES

1. Kolterman OG, Insel J, Saekow M, Olefsky JM: Mechanisms of insulin resistance in human obesity: evidence for receptor and postreceptor defects. *J Clin Invest* 65:1272–1284, 1980
2. Lillioja S, Bogardus C: Obesity and insulin resistance: lessons learned from the Pima Indians. *Diabetes Metab Rev* 4:517–540, 1988
3. Bjorntorp P: Visceral obesity: a "civilization syndrome." *Obes Res* 1:206–222, 1993
4. Peiris AN, Mueller RA, Smith GA, Struve MF, Kissebah AH: Splanchnic insulin metabolism in obesity: influence of body fat distribution. *J Clin Invest* 78:1648–1657, 1986
5. Modan M, Halkin H, Almog S, Lusky A, Eshkol A, Shefi M, Shitrit A, Fuchs Z: Hyperinsulinemia: a link between hypertension obesity and glucose intolerance. *J Clin Invest* 75:809–817, 1985
6. Reaven GM: Role of insulin resistance in human disease. *Diabetes* 37:1595–1607, 1988
7. DeFronzo RA, Ferrannini E: Insulin resistance: a multifaceted syndrome responsible for NIDDM, obesity, hypertension, dyslipidemia, and atherosclerotic cardiovascular disease. *Diabetes Care* 14:173–194, 1991
8. Dunaif A, Segal KR, Futterweit W, Dobrjansky A: Profound peripheral

- insulin resistance, independent of obesity, in polycystic ovary syndrome. *Diabetes* 38:1165–1174, 1989
9. Banerji MA, Lebowitz J, Chaiken RL, Gordon D, Kral JG, Lebowitz HE: Relationship of visceral adipose tissue and glucose disposal is independent of sex in black NIDDM subjects. *Am J Physiol* 273:E425–E432, 1997
  10. Cnop M, Landchild MJ, Vidal J, Havel PJ, Knowles NG, Carr DR, Wang F, Hull RL, Retzlaff BM, Walden CE, Knopp RH, Kahn SE: The concurrent accumulation of intra-abdominal and subcutaneous fat explains the association between insulin resistance and plasma leptin concentrations: distinct metabolic effects of two fat compartments. *Diabetes* 51:1005–1015, 2002
  11. Weinsier RL, Hunter GR, Gower BA, Schutz Y, Darnell BE, Zuckerman PA: Body fat distribution in white and black women: different patterns of intraabdominal and subcutaneous abdominal adipose tissue utilization with weight loss. *Am J Clin Nutr* 74:631–636, 2001
  12. Matsuzawa Y, Shimomura I, Nakamura T, Keno Y, Tokunaga K: Pathophysiology and pathogenesis of visceral fat obesity. *Ann N Y Acad Sci* 748:399–406, 1995
  13. Banerji MA, Faridi N, Atluri R, Chaiken RL, Lebowitz HE: Body composition, visceral fat, leptin, and insulin resistance in Asian Indian men. *J Clin Endocrinol Metab* 84:137–144, 1999
  14. Shuman WP, Newell Morris LL, Leonetti DL, Wahl PW, Mocerri VM, Moss AA, Fujimoto WY: Abnormal body fat distribution detected by computed tomography in diabetic men. *Invest Radiol* 21:483–487, 1986
  15. Fujimoto WY, Bergstrom RW, Leonetti DL, Newell-Morris LL, Shuman WP, Wahl PW: Metabolic and adipose risk factors for NIDDM and coronary disease in third-generation Japanese-American men and women with impaired glucose tolerance. *Diabetologia* 37:524–532, 1994
  16. Carey DG, Jenkins AB, Campbell LV, Freund J, Chisholm DJ: Abdominal fat and insulin resistance in normal and overweight women: direct measurements reveal a strong relationship in subjects at both low and high risk of NIDDM. *Diabetes* 45:633–638, 1996
  17. Pouliot MC, Despres JP, Nadeau A, Moorjani S, Prud'Homme D, Lupien PJ, Tremblay A, Bouchard C: Visceral obesity in men: associations with glucose tolerance, plasma insulin, and lipoprotein levels. *Diabetes* 41:826–834, 1992
  18. Brochu M, Starling RD, Tchernof A, Matthews DE, Garcia-Rubi E, Poehlman ET: Visceral adipose tissue is an independent correlate of glucose disposal in older obese postmenopausal women. *J Clin Endocrinol Metab* 85:2378–2384, 2000
  19. Rendell M, Hulthén UL, Tornquist C, Groop L, Mattiasson I: Relationship between abdominal fat compartments and glucose and lipid metabolism in early postmenopausal women. *J Clin Endocrinol Metab* 86:744–749, 2001
  20. Sparrow D, Borkan GA, Gerzof SG, Wisniewski C, Silbert CK: Relationship of fat distribution to glucose tolerance: results of computed tomography in male participants of the Normative Aging Study. *Diabetes* 35:411–415, 1986
  21. Fujioka S, Matsuzawa Y, Tokunaga K, Tarui S: Contribution of intra-abdominal fat accumulation to the impairment of glucose and lipid metabolism in human obesity. *Metabolism* 36:54–59, 1987
  22. Despres JP, Nadeau A, Tremblay A, Ferland M, Moorjani S, Lupien PJ, Theriault G, Pinault S, Bouchard C: Role of deep abdominal fat in the association between regional adipose tissue distribution and glucose tolerance in obese women. *Diabetes* 38:304–309, 1989
  23. Abate N, Garg A, Peshock RM, Stray-Gundersen J, Grundy SM: Relationships of generalized and regional adiposity to insulin sensitivity in men. *J Clin Invest* 96:88–98, 1995
  24. Goodpaster BH, Thaete FL, Simoneau JA, Kelley DE: Subcutaneous abdominal fat and thigh muscle composition predict insulin sensitivity independently of visceral fat. *Diabetes* 46:1579–1585, 1997
  25. Expert Panel on Detection Evaluation and Treatment of High Blood Cholesterol in Adults: Executive Summary of the Third Report of the National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel III). *JAMA* 285:2486–2497, 2001
  26. Terry RB, Wood PD, Haskell WL, Stefanick ML, Krauss RM: Regional adiposity patterns in relation to lipids, lipoprotein cholesterol, and lipoprotein subfraction mass in men. *J Clin Endocrinol Metab* 68:191–199, 1989
  27. Fujimoto WY, Abbate SL, Kahn SE, Hokanson JE, Brunzell JD: The visceral adiposity syndrome in Japanese-American men. *Obes Res* 2:364–371, 1994
  28. Tchernof A, Lamarche B, Prud'Homme D, Nadeau A, Moorjani S, Labrie F, Lupien PJ, Despres JP: The dense LDL phenotype: association with plasma lipoprotein levels, visceral obesity, and hyperinsulinemia in men. *Diabetes Care* 19:629–637, 1996
  29. Festa A, D'Agostino R Jr, Mykkanen L, Tracy RP, Hales CN, Howard BV, Haffner SM: LDL particle size in relation to insulin, proinsulin, and insulin sensitivity: the Insulin Resistance Atherosclerosis Study. *Diabetes Care* 22:1688–1693, 1999
  30. Reaven GM, Chen YD, Jeppesen J, Maheux P, Krauss RM: Insulin resistance and hyperinsulinemia in individuals with small, dense low density lipoprotein particles. *J Clin Invest* 92:141–146, 1993
  31. Abbasi F, McLaughlin T, Lamendola C, Yeni-Komshian H, Tanaka A, Wang T, Nakajima K, Reaven GM: Fasting remnant lipoprotein cholesterol and triglyceride concentrations are elevated in nondiabetic, insulin-resistant, female volunteers. *J Clin Endocrinol Metab* 84:3903–3906, 1999
  32. Williams PT, Haskell WL, Vranizan KM, Krauss RM: The associations of high-density lipoprotein subclasses with insulin and glucose levels, physical activity, resting heart rate, and regional adiposity in men with coronary artery disease: the Stanford Coronary Risk Intervention Project baseline survey. *Metabolism* 44:106–114, 1995
  33. Carr MC, Hokanson JE, Deeb SS, Purnell JQ, Mitchell ES, Brunzell JD: A hepatic lipase gene promoter polymorphism attenuates the increase in hepatic lipase activity with increasing intra-abdominal fat in women. *Arterioscler Thromb Vasc Biol* 19:2701–2707, 1999
  34. Knopp RH, Retzlaff BM, Walden CE, Wallick S, Fish B, Anderson M, Kahn SE: Effects of insulin resistance and obesity on the LDL response to egg ingestion. *Circulation* 100 (Suppl. 1):I-116, 1999
  35. Kahn SE, Prigeon RL, McCulloch DK, Boyko EJ, Bergman RN, Schwartz MW, Neifing JL, Ward WK, Beard JC, Palmer JP, Porte D Jr: Quantification of the relationship between insulin sensitivity and B-cell function in human subjects: evidence for a hyperbolic function. *Diabetes* 42:1663–1672, 1993
  36. Krotkiewski M, Bjorntorp P, Sjostrom L, Smith U: Impact of obesity on metabolism in men and women: importance of regional adipose tissue distribution. *J Clin Invest* 72:1150–1162, 1983
  37. Schwartz RS, Shuman WP, Larson V, Cain KC, Fellingham GW, Beard JC, Kahn SE, Stratton JR, Cerqueira MD, Abrass IB: The effect of intensive endurance exercise training on body fat distribution in young and older men. *Metabolism* 40:545–551, 1991
  38. Bergman RN, Ider YZ, Bowden CR, Cobelli C: Quantitative estimation of insulin sensitivity. *Am J Physiol* 236:E667–E677, 1979
  39. Beard JC, Bergman RN, Ward WK, Porte D Jr: The insulin sensitivity index in man: correlation between clamp-derived and IVGTT-derived values. *Diabetes* 35:362–369, 1986
  40. Prigeon RL, Kahn SE, Porte D Jr: Reliability of error estimates from the minimal model: implications for measurements in physiological studies. *Am J Physiol* 266:E279–E286, 1994
  41. Abbate SL, Fujimoto WY, Brunzell JD, Kahn SE: Effect of heparin on insulin-glucose interactions measured by the minimal model technique: implications for reproducibility using this method. *Metabolism* 42:353–357, 1993
  42. Morgan DR, Lazarow A: Immunoassay of insulin: two antibody system: plasma insulin levels of normal, subdiabetic, and diabetic rats. *Diabetes* 12:115–126, 1963
  43. Marcovina SM, Albers JJ, Kennedy H, Mei JV, Henderson LO, Hannon WH: International Federation of Clinical Chemistry standardization project for measurements of apolipoproteins A-I and B. IV. Comparability of apolipoprotein B values by use of international reference material. *Clin Chem* 40:586–592, 1994
  44. Purnell JQ, Marcovina SM, Hokanson JE, Kennedy H, Cleary PA, Steffes MW, Brunzell JD: Levels of lipoprotein(a), apolipoprotein B, and lipoprotein cholesterol distribution in IDDM: results from follow-up in the Diabetes Control and Complications Trial. *Diabetes* 44:1218–1226, 1995
  45. National Heart Lung and Blood Institute: Clinical guidelines on the identification, evaluation, and treatment of overweight and obesity in adults: the evidence report. *Obes Res* 6 (Suppl. 2):51S–209S, 1998
  46. Despres J-P, Krauss RM: Obesity and lipoprotein metabolism. In *Handbook of Obesity*. Bray GA, Bouchard C, James WPT, Eds. New York, Marcel Dekker, 1998, p. 651–675
  47. Haffner SM: The insulin resistance syndrome revisited. *Diabetes Care* 19:275–277, 1996
  48. Brunzell JD, Hokanson JE: Dyslipidemia of central obesity and insulin resistance. *Diabetes Care* 22 (Suppl. 3):C10–C13, 1999
  49. Olefsky J, Reaven GM, Farquhar JW: Effects of weight reduction on obesity: studies of lipid and carbohydrate metabolism in normal and hyperlipoproteinemic subjects. *J Clin Invest* 53:64–76, 1974
  50. Sorbris R, Pettersson BG, Nilsson-Ehle P: Effects of weight reduction on plasma lipoproteins and adipose tissue metabolism in obese subjects. *Eur J Clin Invest* 11:491–498, 1981
  51. Colman E, Katzell LL, Rogus E, Coon P, Muller D, Goldberg AP: Weight loss reduces abdominal fat and improves insulin action in middle-aged and older men with impaired glucose tolerance. *Metabolism* 44:1502–1508, 1995

52. Katznel LI, Bleecker ER, Colman EG, Rogus EM, Sorkin JD, Goldberg AP: Effects of weight loss vs. aerobic exercise training on risk factors for coronary disease in healthy, obese, middle-aged and older men: a randomized controlled trial. *JAMA* 274:1915–1921, 1995
53. Purnell JQ, Kahn SE, Albers JJ, Nevin DN, Brunzell JD, Schwartz RS: Effect of weight loss with reduction of intra-abdominal fat on lipid metabolism in older men. *J Clin Endocrinol Metab* 85:977–982, 2000
54. Goldstein DJ: Beneficial health effects of modest weight loss. *Int J Obes Relat Metab Disord* 16:397–415, 1992
55. Franssila-Kallunki A, Rissanen A, Ekstrand A, Ollus A, Groop L: Effects of weight loss on substrate oxidation, energy expenditure, and insulin sensitivity in obese individuals. *Am J Clin Nutr* 55:356–361, 1992
56. Goodpaster BH, Kelley DE, Wing RR, Meier A, Thaete FL: Effects of weight loss on regional fat distribution and insulin sensitivity in obesity. *Diabetes* 48:839–847, 1999
57. Schwartz RS, Cain KC, Shuman WP, Larson V, Stratton JR, Beard JC, Kahn SE, Cerqueira MD, Abrass IB: Effect of intensive endurance training on lipoprotein profiles in young and older men. *Metabolism* 41:649–654, 1992
58. Kahn SE, Larson VG, Beard JC, Cain KC, Fellingham GW, Schwartz RS, Veith RC, Stratton JR, Cerqueira MD, Abrass IB: Effect of exercise on insulin action, glucose tolerance and insulin secretion in aging. *Am J Physiol* 258:E937–E943, 1990
59. Despres JP, Ferland M, Moorjani S, Nadeau A, Tremblay A, Lupien PJ, Theriault G, Bouchard C: Role of hepatic-triglyceride lipase activity in the association between intra-abdominal fat and plasma HDL cholesterol in obese women. *Arteriosclerosis* 9:485–492, 1989
60. Carr MC, Hokanson JE, Zambon A, Deeb SS, Barrett PH, Purnell JQ, Brunzell JD: The contribution of intraabdominal fat to gender differences in hepatic lipase activity and low/high density lipoprotein heterogeneity. *J Clin Endocrinol Metab* 86:2831–2837, 2001
61. Patsch JR, Prasad S, Gotto AM Jr, Patsch W: High density lipoprotein2: relationship of the plasma levels of this lipoprotein species to its composition, to the magnitude of postprandial lipemia, and to the activities of lipoprotein lipase and hepatic lipase. *J Clin Invest* 80:341–347, 1987
62. Kuusi T, Ehnholm C, Viikari J, Harkonen R, Vartiainen E, Puska P, Taskinen MR: Postheparin plasma lipoprotein and hepatic lipase are determinants of hypo- and hyperalphalipoproteinemia. *J Lipid Res* 30: 1117–1126, 1989
63. Zambon A, Austin MA, Brown BG, Hokanson JE, Brunzell JD: Effect of hepatic lipase on LDL in normal men and those with coronary artery disease. *Arterioscler Thromb* 13:147–153, 1993
64. Havel RJ: Genetic underpinnings of LDL size and density: a role for hepatic lipase? *Am J Clin Nutr* 71:1390–1391, 2000
65. Zambon A, Hokanson JE, Brown BG, Brunzell JD: Evidence for a new pathophysiological mechanism for coronary artery disease regression: hepatic lipase-mediated changes in LDL density. *Circulation* 99:1959–1964, 1999