

Lisanne M.M. Gommers,¹ Joost G.J. Hoenderop,¹ René J.M. Bindels,¹ and Jeroen H.F. de Baaij^{1,2}



Hypomagnesemia in Type 2 Diabetes: A Vicious Circle?

Diabetes 2016;65:3–13 | DOI: 10.2337/db15-1028

Over the past decades, hypomagnesemia (serum Mg^{2+} <0.7 mmol/L) has been strongly associated with type 2 diabetes mellitus (T2DM). Patients with hypomagnesemia show a more rapid disease progression and have an increased risk for diabetes complications. Clinical studies demonstrate that T2DM patients with hypomagnesemia have reduced pancreatic β -cell activity and are more insulin resistant. Moreover, dietary Mg^{2+} supplementation for patients with T2DM improves glucose metabolism and insulin sensitivity. Intracellular Mg^{2+} regulates glucokinase, K_{ATP} channels, and L-type Ca^{2+} channels in pancreatic β -cells, preceding insulin secretion. Moreover, insulin receptor autophosphorylation is dependent on intracellular Mg^{2+} concentrations, making Mg^{2+} a direct factor in the development of insulin resistance. Conversely, insulin is an important regulator of Mg^{2+} homeostasis. In the kidney, insulin activates the renal Mg^{2+} channel transient receptor potential melastatin type 6 that determines the final urinary Mg^{2+} excretion. Consequently, patients with T2DM and hypomagnesemia enter a vicious circle in which hypomagnesemia causes insulin resistance and insulin resistance reduces serum Mg^{2+} concentrations. This Perspective provides a systematic overview of the molecular mechanisms underlying the effects of Mg^{2+} on insulin secretion and insulin signaling. In addition to providing a review of current knowledge, we provide novel directions for future research and identify previously neglected contributors to hypomagnesemia in T2DM.

Globally, over 300 million people suffer from type 2 diabetes mellitus (T2DM), and the prevalence is predicted to rise to over 600 million over the next decades (1). T2DM is characterized by a combination of insulin deficiency and insulin resistance. The general pathophysiological concept is that hyperglycemia emerges when endogenous insulin secretion

can no longer match the increased demand owing to insulin resistance (2).

Since the 1940s, it has been reported that T2DM is associated with hypomagnesemia (3,4). Low serum magnesium (Mg^{2+}) levels have been reported in large cohorts of patients with T2DM (5). In T2DM, the prevalence of hypomagnesemia ranges between 14 and 48% compared with between 2.5 and 15% in healthy control subjects (4). Hypomagnesemia is associated with a more rapid, and permanent, decline in renal function in patients with T2DM (6). In addition, epidemiological studies consistently show an inverse relationship between dietary Mg^{2+} intake and risk of developing T2DM (7). Several patient studies have shown beneficial effects of Mg^{2+} supplementation on glucose metabolism and insulin sensitivity (8–10). Recently, Rodríguez-Morán et al. (11) published an excellent overview of the clinical studies addressing the role of Mg^{2+} in T2DM. In our review, we will focus on the molecular mechanisms underlying these clinical observations.

Mg^{2+} is an essential ion for human health, as it is involved in virtually every mechanism in the cell, including energy homeostasis, protein synthesis, and DNA stability (12). Considering these divergent functions, it can be appreciated that serum Mg^{2+} levels are tightly regulated between 0.7 and 1.05 mmol/L in healthy individuals. However, impaired intestinal Mg^{2+} absorption or renal Mg^{2+} wasting can lead to hypomagnesemia. A wide range of genetic and environmental factors can affect the Mg^{2+} -deficient state, which have previously been extensively reviewed (12).

In this review, we address the following questions that are central to the role of hypomagnesemia in T2DM: 1) Does Mg^{2+} regulate insulin secretion? 2) How does Mg^{2+} affect insulin resistance? 3) How does insulin regulate Mg^{2+} homeostasis? Taken together, these questions will aid the understanding of whether hypomagnesemia is a causative factor for or a consequence of T2DM.

¹Department of Physiology, Radboud Institute for Molecular Life Sciences, Radboud University Nijmegen Medical Center, Nijmegen, the Netherlands

²Department of Physiology, Anatomy and Genetics, University of Oxford, Oxford, U.K.
Corresponding author: Jeroen H.F. de Baaij, jeroen.debaaij@radboudumc.nl.

Received 24 July 2015 and accepted 10 September 2015.

© 2016 by the American Diabetes Association. Readers may use this article as long as the work is properly cited, the use is educational and not for profit, and the work is not altered.

INSULIN RESISTANCE

Insulin Sensitivity in Normal Cell Physiology

Increased insulin resistance is the major pathophysiological cause for the development of T2DM. In healthy subjects, insulin increases glycogen production in the liver, lipid synthesis by adipose tissue, and glucose uptake in muscle (5,13–18). Insulin resistance is often the consequence of reduced sensitivity of the insulin receptor that is composed of two insulin-binding α -subunits and two β -subunits. Specifically, upon insulin binding, the tyrosine residues of the β -subunits become autophosphorylated, activating a wide signaling network in the cell (Fig. 1). Depending on the target tissue, direct substrates of the insulin receptor may be recruited to the receptor, of which insulin receptor substrates (IRSs)-1–4 are the most studied. These IRSs, in turn, phosphorylate downstream signaling pathways leading to glucose uptake, glycogenesis, lipid synthesis, and other insulin-dependent actions. Alternatively, the insulin receptor can activate IRS-independent pathways via Src homology 2 domain containing transforming protein causing the activation of mitogen-activated protein kinase signaling and regulation of cell proliferation (16,19). An overview of the main insulin signaling pathways is provided in Fig. 1.

Role of Mg^{2+} in Insulin Sensitivity

Many clinical studies have shown that hypomagnesemia is associated with increased insulin resistance in T2DM

patients (4,20–22). In a cross-sectional study of patients with metabolic syndrome, it was shown that insulin resistance associates with reduced serum Mg^{2+} levels (21). Furthermore, a cohort study of adult black Americans showed that Mg^{2+} deficiency contributes to an insulin-resistant state (20). Similar results were found in healthy human subjects, where induced Mg^{2+} deficiency reduced insulin action and secretion (22). In this segment of the review, we will clarify the association between hypomagnesemia and insulin resistance by focusing on the effects of Mg^{2+} on the insulin receptor activity and downstream signaling events.

Insulin Receptor Phosphorylation

It is widely accepted that Mg^{2+} is essential for autophosphorylation of the β -subunits of the insulin receptor. The crystal structure of the insulin receptor tyrosine kinase shows that two Mg^{2+} ions can bind to the tyrosine kinase domain (23). The role of this Mg^{2+} binding has been shown by in vitro studies using isolated insulin receptors. Here, Mg^{2+} enhances tyrosine kinase activity by increasing the receptor's affinity for ATP (24,25). Indeed, rats with hypomagnesemia have reduced levels of insulin receptor phosphorylation, mimicking a state of insulin resistance (26,27). In contrast, increased insulin receptor phosphorylation was shown in liver tissue of rats fed Mg^{2+} -deficient diets for 11 weeks (28). However, the value of this study can be questioned because insulin

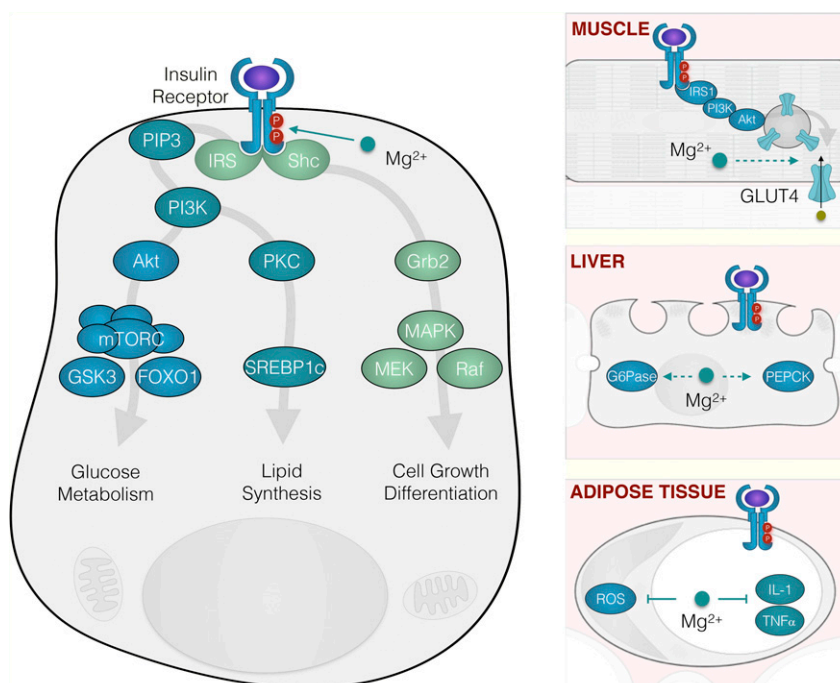


Figure 1— Mg^{2+} affects insulin sensitivity. Mg^{2+} regulates the insulin signaling pathway by increasing the affinity of the insulin receptor tyrosine kinase for ATP. Consequently, hypomagnesemia is associated with a reduced activity of all downstream pathways. In the muscle, Mg^{2+} therefore regulates the membrane trafficking of GLUT4. In the liver, Mg^{2+} is an important regulator of enzymes in gluconeogenesis, including G6Pase and PEPCK. In adipose tissue, Mg^{2+} acts as an anti-inflammatory factor reducing IL-1 and TNF- α secretion. FOXO1, forkhead box class O1; Grb2, growth factor receptor-bound protein 2; GSK3, glycogen synthase kinase 3; MEK/MAPK, mitogen-activated protein kinase kinase; P, phosphorylation; PIP3, phosphatidylinositol 3,4,5 trisphosphate; PI3K, phosphatidylinositol 3-kinase; PKC, protein kinase C; ROS, reactive oxygen species; Shc, Src homology 2 domain containing transforming protein.

phosphorylation was normal in rats with the same serum Mg^{2+} levels at 6 weeks or in muscle tissue. Altogether, Mg^{2+} seems to be an important factor in insulin receptor autophosphorylation. Defective insulin receptor phosphorylation is therefore regarded as the main mechanism by which hypomagnesemia contributes to insulin resistance in T2DM patients.

Glucose Metabolism in the Muscle

Glucose uptake in skeletal muscle accounts for the disposal of ~80% of the dietary glucose load via insulin-dependent glucose uptake using GLUT4 glucose transporters (29). In a recent study in rats with streptozotocin (STZ)-induced diabetes, oral Mg^{2+} supplementation increased GLUT4 expression in the rat muscle and thereby lowered serum glucose levels to the normal range (30). Similar results were obtained in STZ mice treated with Mg^{2+} -rich sea water, showing increased GLUT1 and GLUT4 expression in muscle (31). Although the molecular mechanism is still unknown, these findings suggest that Mg^{2+} regulates glucose uptake in muscle.

Glycogen Synthesis in the Liver

Although many enzymes in the liver require Mg^{2+} for their activity, the role of Mg^{2+} in gluconeogenesis and glycogenesis is poorly studied. The activity of several enzymes involved in gluconeogenesis including glucose-6-phosphatase (G6Pase) and phosphoenolpyruvate carboxykinase (PEPCK) were measured in Mg^{2+} -deficient rats, but only PEPCK activity was increased (32). In the same vein, mice with STZ-induced diabetes showed decreased PEPCK and G6Pase mRNA expression when supplemented with Mg^{2+} -rich deep sea water (31). However, it cannot be excluded that other molecules that are present in the sea water mediate these effects. In contrast, mRNA expression of PEPCK was decreased in Mg^{2+} -deficient rats (33).

Obesity and Inflammatory Aspects

Obesity is a major risk factor for the development of T2DM (16,34). In obese patients with T2DM, adipocytes produce proinflammatory mediators, such as interleukin 1 (IL-1) and tumor necrosis factor- α (TNF- α), and stimulate the production of reactive oxygen species. The inflammatory environment is regarded as an important contributor to insulin resistance and is one of the main reasons that obesity is associated with T2DM (15,16). Among others, chronic inflammation may induce insulin resistance by reducing GLUT4 activity and inhibiting IRS-1 action (reviewed in 35).

Mg^{2+} is an important anti-inflammatory molecule, and therefore hypomagnesemia increases the inflammatory environment in obesity. IL-1 and TNF- α are significantly increased in Mg^{2+} -deficient hamsters and rats (36). Moreover, low serum Mg^{2+} levels are associated with increased levels of TNF- α in obese people without diabetes (37). Additionally, Mg^{2+} deficiency contributes to neutrophil activation and oxidative stress (38). In a community-based cross-sectional study of 488 healthy children, an inverse correlation between serum Mg^{2+} levels and hs-CRP

was demonstrated (39). A recent clinical randomized double-blind placebo-controlled trial of healthy subjects with prediabetes and hypomagnesemia showed reduced hs-CRP levels after Mg^{2+} supplementation for 3 months (40). Given that inflammation and oxidative stress are important factors in the development of insulin resistance (35,41), hypomagnesemia may cause increased insulin resistance.

Novel Perspectives

Although there is extensive evidence that hypomagnesemia reduces insulin receptor phosphorylation and increases chronic inflammation in T2DM patients, the consequences of these signaling events in affected tissues, such as muscle, liver, and adipocytes, have barely been studied. For instance, Ca^{2+} has been shown to increase glucose uptake via GLUT4 in the skeletal muscle (42). Given that Mg^{2+} is an antagonist of Ca^{2+} , GLUT4 membrane trafficking may be reduced in hypomagnesemia. Additionally, some early studies suggest that Mg^{2+} may affect adipokine levels (43). Therefore, future research should be aimed at identifying the effects of hypomagnesemia in target tissues, taking into account the intrinsic effects of Mg^{2+} on gene expression and Ca^{2+} antagonism.

INSULIN SECRETION

Insulin Secretion in Normal Cell Physiology

Acute insulin secretion from pancreatic β -cells is essential to the control of blood glucose homeostasis. Increased blood glucose levels stimulate the influx of glucose in pancreatic β -cells via GLUT2, where it is converted to glucose-6-phosphate (G6P) by glucokinase (44). This enzymatic reaction functions as a glucose sensor to determine the required amount of insulin secretion. G6P is further metabolized by glycolysis to generate ATP, which directly induces closure of K_{ATP} channel Kir6.2 (45). Closure of these channels induces depolarization of the plasma membrane and, consequently, opening of voltage-dependent Ca^{2+} channels (46). The influx of extracellular Ca^{2+} triggers the release of insulin via exocytosis (19) (Fig. 2).

In the early phases of T2DM, insulin release meets the increasing demands by expanding the pancreatic β -cell mass (47). For decades, insulin resistance was thought to be the major cause for T2DM. However, supported by genome-wide association studies, evidence has accumulated that impaired insulin secretion in the pancreatic β -cells is a major contributor to the development of T2DM (48,49).

Role of Mg^{2+} in Insulin Secretion

The clinical evidence for a role of Mg^{2+} in insulin secretion is limited and less well studied than the effects of Mg^{2+} on insulin sensitivity, but several recent clinical studies suggest that T2DM patients with hypomagnesemia display reduced insulin secretion. In individuals without diabetes, decreased serum Mg^{2+} concentrations are associated with a diminished insulin secretion (15). Conversely, HOMA of β -cell activity was negatively correlated with the serum

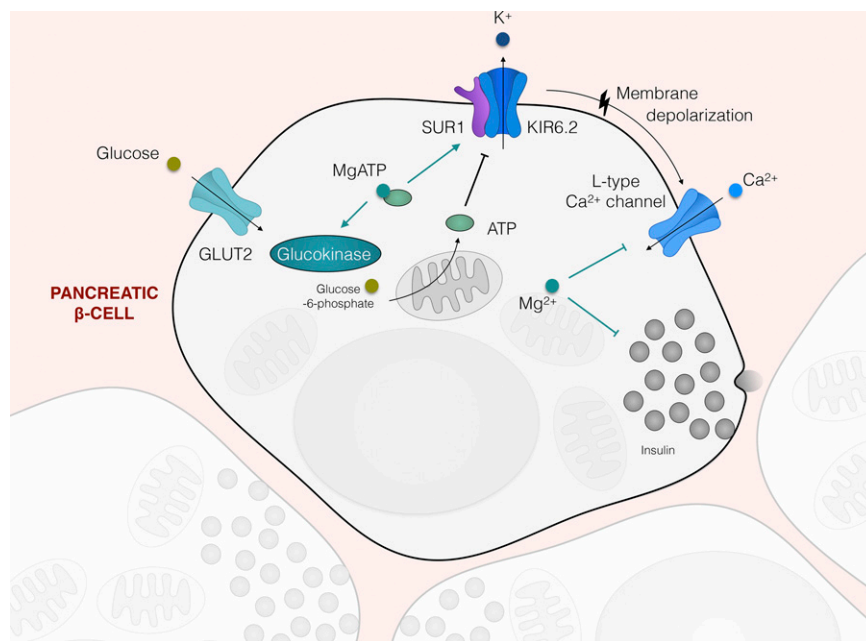


Figure 2— Mg^{2+} regulates insulin secretion in pancreatic β -cells. In pancreatic β -cells, Mg^{2+} directly influences the rate of glucokinase activity by acting as a cofactor for adenine nucleotides. The product of this enzymatic reaction, G6P, is further processed in glycolysis producing ATP. Closure of the K_{ATP} channel is dependent on ATP by its binding to the Kir6.2 subunits. Conversely, $MgATP$ initiates channel opening by binding to the SUR1 subunits of the channel. Importantly, the physiological consequence of channel closure is the depolarization of the membrane, which triggers the influx of Ca^{2+} via the L-type Ca^{2+} channels. This final step initiates insulin vesicle release, which is negatively controlled by Mg^{2+} acting on both Ca^{2+} influx and the L-type Ca^{2+} channels.

Mg^{2+} concentration in a Canadian cohort of patients with T2DM (50). However, this study focused mainly on the lipid metabolism and did not fully explore the effects on insulin secretion. Supplementation of individuals without diabetes with $MgCl_2$ significantly increased β -cell function in a small randomized clinical trial (51). Here, we will examine the molecular mechanisms by which Mg^{2+} improves insulin secretion by showing how Mg^{2+} acts on the main actors involved in insulin secretion.

Glucokinase

After entering the pancreatic β -cells via GLUT2, glucose is converted to G6P by glucokinase. The action of glucokinase depends on $MgATP^{2-}$. Recent studies show that the activity of glucokinase is regulated by $MgATP^{2-}$ at subphysiological concentrations (reported $K_M \sim 0.15$ mmol/L), suggesting that there is always sufficient $MgATP^{2-}$ available in the cell for glucokinase activity (52). However, there may exist a small cooperative effect of $MgATP^{2-}$ on glucose binding (52,53). Mg^{2+} deficiency may, therefore, reduce glucose binding to glucokinase by slowing down metabolism and indirectly reducing insulin secretion.

Glycolysis

G6P is further metabolized via glycolysis and the Krebs cycle, resulting in increased ATP levels. Although many enzymes in this metabolic process use $MgATP$ as a cofactor (54), studies in erythrocytes have shown that glycolysis depends on intracellular Mg^{2+} with a half-maximal

glycolysis flux at 0.03 mmol/L Mg^{2+} . This is well below physiological intracellular Mg^{2+} values, even in Mg^{2+} deficient conditions (55).

K_{ATP} Channel

First described in 1984, K_{ATP} channels are the main regulators of the membrane potential in pancreatic β -cells (56). The K_{ATP} channel in β -cells consists of four Kir6.2 subunits and four regulatory sulfonylurea receptor (SUR)1 subunits, whose activity is controlled by the intracellular ATP-to-ADP ratio. In the presence of Mg^{2+} , it is the balance between $MgATP$, physiologically ranging between 1 and 5 mmol/L, and $MgADP$ levels that determines channel activity. Binding of both $MgATP$ and $MgADP$ to the nucleotide-binding sites of SUR1 subunits results in opening of the channel. By contrast, in the absence of Mg^{2+} , ATP induces closure of the K_{ATP} channel by directly binding to Kir6.2 subunits (57). In high-glucose conditions, increased glycolysis shifts the balance toward ATP, concomitantly leading to reduced $MgADP$ levels inducing channel closure and increased insulin secretion (58). This balance is the therapeutic target of sulfonylurea drugs, which induce channel closure by antagonizing the binding of Mg nucleotides on SUR1 (59). Thus, reduced intracellular Mg^{2+} concentrations, as a result of hypomagnesemia in T2DM, may reduce the $MgATP$ and $MgADP$ levels, favoring inhibition of the K_{ATP} channel and thereby indirectly stimulating insulin secretion. However, the exact effect of hypomagnesemia on intracellular $MgATP$ levels remains to be examined.

L-Type Ca^{2+} Channel

Inhibition of K_{ATP} channels by increased ATP levels results in depolarization of the membrane, activating Ca^{2+} influx via the voltage-dependent L-type Ca^{2+} channel. In the pancreatic β -cell, the main L-type channels are of the $\text{Ca}_v1.2$ and $\text{Ca}_v1.3$ subtypes, of which $\text{Ca}_v1.2$ channels are the main contributors to insulin secretion (60,61). There is a wide range of literature showing that both intracellular and extracellular Mg^{2+} inhibits L-type Ca^{2+} channels (62–64). Although these findings suggest that hypomagnesemia would increase L-type channel activity in the short-term, in the long-term it has been indicated that hypomagnesemia reduces the expression of L-type Ca^{2+} channels and, thus, indirectly diminishes insulin secretion (65). However, it should be noted that these findings rely on measurements performed with cardiomyocytes and have not been confirmed in pancreatic β -cells.

Insulin Vesicle Release

Insulin vesicle release depends on Ca^{2+} binding that initiates exocytosis (45). Mg^{2+} fulfills an essential role in antagonizing Ca^{2+} and thus regulates insulin secretion by pancreatic β -cells. Atwater et al. (66) studied the importance of the Ca^{2+} -to- Mg^{2+} ratio in glucose-induced insulin release in perfused rat pancreas and mouse islets. The results indicated that only a reduction of the physiological Mg^{2+} concentrations stimulated insulin secretion. Conversely, insulin secretion was inhibited only by diminished Ca^{2+} levels. Interestingly, a simultaneous reduction in Ca^{2+} and Mg^{2+} , while maintaining their ratio constant, did not change the insulin release. Mg^{2+} deficiency inevitably changes the Ca^{2+} -to- Mg^{2+} ratio and may therefore affect insulin secretion.

Novel Perspectives

Altogether, the regulation of K_{ATP} channels, L-type Ca^{2+} channels, and vesicle release points toward an inhibitory effect of Mg^{2+} on insulin secretion. These studies suggest that Mg^{2+} deficiency stimulates insulin release by the β -cells. However, a recent clinical study showed a positive correlation between Mg^{2+} concentration and first-phase insulin secretion (15). Why do hypomagnesemic T2DM patients suffer from reduced insulin secretion then? There may be multiple factors contributing to this paradox: 1) many of the studies of K_{ATP} channels and L-type Ca^{2+} channels are executed in cardiac cells and may therefore not be a good representation of the effects of Mg^{2+} in the pancreas; 2) most studies investigate short-term effects of Mg^{2+} on channel activity and may not represent long-term Mg^{2+} deficiency; 3) it has been shown that Mg^{2+} increases insulin synthesis at the transcriptional level (67); 4) all studies have focused on the effects of intracellular Mg^{2+} , leaving the role of extracellular Mg^{2+} unknown; 5) Mg^{2+} is an important regulator of protein synthesis and cell proliferation (68) and may therefore regulate β -cell viability; and 6) Mg^{2+} deficiency may also regulate other factors, including serum K^+ and Ca^{2+} , and thereby indirectly affect insulin signaling. However, it

should be emphasized that the role of Mg^{2+} in insulin secretion has only been examined by a small number of clinical studies. The current clinical evidence on the effect of Mg^{2+} on insulin secretion is too limited to draw firm conclusions on the physiological implications of Mg^{2+} in the pancreatic β -cell. Future clinical and experimental studies are necessary to resolve this question.

Mg^{2+} Homeostasis

Cellular Mg^{2+} Homeostasis

Since protein and DNA synthesis is highly dependent on intracellular Mg^{2+} availability, intracellular Mg^{2+} concentrations are tightly regulated (12). Intracellular Mg^{2+} concentrations are determined by the uptake via dedicated Mg^{2+} channels and transporters, including solute carrier family 41 member 1 (SLC41A1), magnesium transporter 1 (MagT1), and transient receptor potential melastatin type 6 and 7 (TRPM6 and TRPM7). The role of these Mg^{2+} transporters in the establishment of Mg^{2+} homeostasis has been reviewed in detail (12). Several groups have investigated the association between genetic variations in these Mg^{2+} transporters and risk for T2DM (69–71). Until now, a link between T2DM and TRPM7 or MagT1 has not been found. However, single nucleotide polymorphisms (SNPs) in TRPM6 or SLC41A1 have been associated with increased risk for T2DM (69,71). Interestingly, insulin may be an important regulator of cellular Mg^{2+} uptake. Studies in erythrocytes showed that insulin reduces serum Mg^{2+} levels and increases cytosolic Mg^{2+} concentrations (72). Indeed, insulin increases both glucose and Mg^{2+} uptake in pancreatic β -cells and cardiomyocytes, suggesting that glucose and Mg^{2+} homeostasis are linked (73,74). However, the molecular identity of the Mg^{2+} transporters involved in this mechanism has not been identified to date.

Body Mg^{2+} Homeostasis

Mg^{2+} homeostasis in the body is facilitated by the interplay of intestinal absorption, bone Mg^{2+} storage, and renal Mg^{2+} excretion (12). In the kidney, the bulk of filtered Mg^{2+} is reabsorbed passively in the proximal tubule (10–25%) and thick ascending limb of Henle loop (TAL) (60–80%). However, fine-tuning takes place in the distal convoluted tubule (DCT) (5–15%) where transcellular reabsorption determines the final urinary Mg^{2+} excretion, since no reabsorption takes place beyond the DCT (12).

In the DCT, Mg^{2+} is reabsorbed from the pro-urine by TRPM6 channels (12). Patients with TRPM6 mutations suffer from hypomagnesemia with secondary hypocalcemia (Mendelian Inheritance in Man: 602014) due to renal Mg^{2+} wasting (75,76). TRPM6 is highly regulated by dietary Mg^{2+} availability, epidermal growth factor, estrogen, pH, and ATP (77–81). Conversely, vitamin D and parathyroid hormone do not regulate Mg^{2+} reabsorption in the DCT (77). The transepithelial movement of Mg^{2+} is dependent on the electrochemical gradient that is set by Na^+ - K^+ -ATPase activity and depends on the local recycling

of K^+ transport via $K_v1.1$ at the apical membrane and $Kir4.1$ at the basolateral membrane (82–84). Although the basolateral Mg^{2+} extrusion mechanism in the DCT is still under debate, recent publications suggest that this might be facilitated by the $SLC41A1$ Na^+-Mg^{2+} exchanger and is regulated by cyclin M2 (85–87).

Insulin Regulates Mg^{2+} Reabsorption in the Kidney

Hypomagnesemia in T2DM is primarily due to renal Mg^{2+} wasting (88). Insulin extracts from animal pancreas were introduced in medicine in the early 1920s saving the lives of many T2DM patients (89). The first article reporting increased blood Mg^{2+} and Na^+ levels during treatment with impure insulin extracts was published in 1933 (90). However, it was not until the 1960s, when synthetic insulin was available and Mg^{2+} measurements were improved, that it became apparent that insulin regulates Mg^{2+} reabsorption in the kidney (91). Indeed, microperfusion experiments in mouse TAL segments show increased Mg^{2+} permeability after insulin stimulation (92). Moreover, in mouse DCT cells insulin stimulates Mg^{2+} uptake (93). Therefore, we will present an overview of the molecular targets of insulin in the regulation of renal Mg^{2+} transport.

TRPM6

TRPM6 was identified as the molecular target of insulin signaling in 2012 (94) (Fig. 3). Upon insulin binding to the receptor, an intracellular signaling cascade including phosphatidylinositol 3-kinase, Akt, and Rac1 is activated resulting

in increased insertion of TRPM6 in the plasma membrane. Two SNPs in TRPM6 (p.Val1393Ile [rs3750425] and p.Lys1584Glu [rs2274924]) are associated with increased susceptibility for gestational diabetes mellitus (71). Patch clamp analysis showed that these mutations render the channel insensitive to insulin stimulation (94). Follow-up studies could not confirm associations between serum Mg^{2+} values and these two SNPs (69,95). However, these differences may be explained by the dietary Mg^{2+} intake of the subjects, since patients in the original study showed reduced Mg^{2+} intake (<250 mg/day) (71). The mRNA expression of TRPM6 is changed in diabetic rats, although results are contradictory and may depend on the experimental model used (96,97). Whereas some report increased TRPM6 expression (96), others show that TRPM6 is downregulated (97). Given that hypomagnesemia itself also stimulates TRPM6 expression (77,98), it is difficult to distinguish between the effect of hypomagnesemia and T2DM in these studies.

Na^+-Cl^- Cotransporter

In addition to its actions on TRPM6, insulin signaling in DCT increases Na^+ reabsorption via the thiazide-sensitive Na^+-Cl^- cotransporter (NCC) (99–103). Insulin activates an intracellular signaling cascade that includes mTOR complex 2 (mTORC2) and stress-activated protein kinase/oxidative stress responsive kinase (SAPK/OSR1) to increase NCC phosphorylation and activity (99,101,102) (Fig. 3). It has been hypothesized that

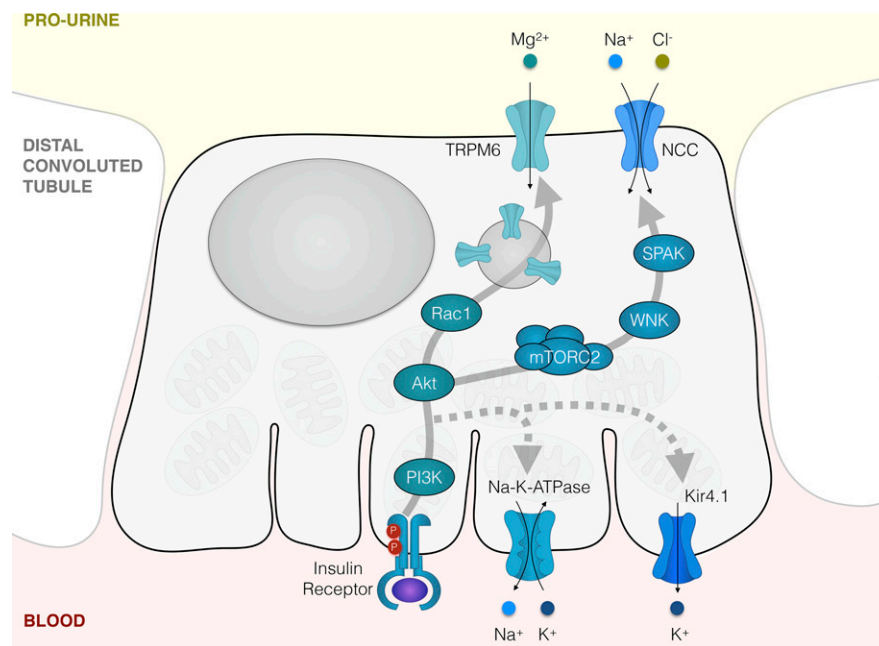


Figure 3—Insulin regulates electrolyte reabsorption in the distal convoluted tubule. Upon binding of insulin to its receptor, an intracellular signaling cascade including phosphatidylinositol 3-kinase (PI3K) and Akt increases the plasma membrane expression of the TRPM6 Mg^{2+} channel and the thiazide-sensitive NCC. Additionally, insulin may stimulate the activity of other channels, such as K^+ channel $Kir4.1$ and the Na^+-K^+ -ATPase. SPAK, STE20/SPS1-related, proline-alanine-rich kinase; WNK, with-no-lysine [K] kinases.

hyperinsulinemia in T2DM causes increased NCC activation and thus renal Na^+ reabsorption, contributing to hypertension that is present in 75% of T2DM patients (104). This hypothesis is supported by studies in Zucker obese rats and db/db mice showing hyperinsulinemia, hypertension, and increased NCC activity (100,101).

Diabetes-DCT Paradox

Central in the theory of insulin-induced hypertension is that the hyperinsulinemic state causes overactivation of the insulin pathway resulting in increased NCC activity (99,101). In contrast, insulin resistance has been proposed to explain the reduced TRPM6 activity and hypomagnesemia in T2DM (94). Until now, it has been widely accepted that Na^+ and Mg^{2+} transport in the DCT go hand in hand, since reduced NCC activity causes hypomagnesemia as shown in patients with Gitelman syndrome and in users of thiazide diuretics (105,106). However, in T2DM, Na^+ and Mg^{2+} reabsorption in DCT may be uncoupled, resulting in increased Na^+ reabsorption and decreased Mg^{2+} reabsorption. Elucidation of the molecular mechanisms of Na^+ and Mg^{2+} cross talk and studies of the insulin resistance of DCT cells in T2DM may contribute to better understanding of the diabetes-DCT paradox.

Kir4.1 and Na^+ - K^+ -ATPase

Reports on diabetic retinopathy suggest that insulin regulates the potassium channel Kir4.1 in the retina (107,108). Given that Kir4.1 is also an important regulator of Na^+ and Mg^{2+} transport in DCT, and patients with mutations in Kir4.1 develop hypomagnesemia and hypokalemia, future studies should include regulation of Kir4.1 in T2DM patients (109,110).

Additionally, insulin has been shown to increase Na^+ - K^+ -ATPase activity in several tissues including heart and muscle (111,112). Na^+ - K^+ -ATPase activity is decreased in animal models of T2DM (reviewed in 113,114). Within the DCT, the membrane expression of the Na^+ - K^+ -ATPase is regulated by TBC1 domain family, member 4 (TBC1D4), which is a substrate of Akt in the insulin pathway (115). Given that the expression of TBC1D4 in the DCT is regulated by dietary Mg^{2+} intake (98), the regulation of the Na^+ - K^+ -ATPase may be changed in hypomagnesemia via TBC1D4.

Other Factors Regulating Mg^{2+} Transport in T2DM

In the complex context of T2DM, many factors may play a role in the development of hypomagnesemia. In contrast to the well-described effects of insulin on renal electrolyte handling, other contributing factors are poorly studied. Therefore, rather than providing a definitive overview of impaired renal regulation of Mg^{2+} transport in T2DM, this part of the article aims to highlight contributors to hypomagnesemia that are underappreciated in current literature and should be subject for further examinations.

Glucose

Initial experiments in 1970s and 1980s showed that glucose increases urinary Mg^{2+} excretion (116,117). However,

the molecular mechanism by which glucose regulates renal Mg^{2+} handling is unknown, and these early experiments do not take into account the action of insulin or diuresis. Recently, the Na^+ -GLUT2 sodium-glucose cotransporter 2 (sGLT2) KO mouse, which is characterized by a marked glucosuria, was shown to have increased urinary Mg^{2+} excretion (118). These findings raise clinical concerns for the use of SGLT2 inhibitors. However, clinical trials with SGLT2 inhibitors have not provided any evidence that Mg^{2+} excretion is changed by SGLT2 inhibitor intake (119,120). Given the substantial glucosuria in T2DM, the physiological role of glucose in Mg^{2+} reabsorption can be questioned.

Hyperfiltration

Increased blood glucose values in T2DM result in hyperfiltration and increased renal urinary flow (121,122). Approximately 10–25% of the filtered Mg^{2+} is reabsorbed by the proximal tubules where water reabsorption precedes Mg^{2+} reabsorption, creating a favorable electrochemical gradient for Mg^{2+} reabsorption. Micropuncture studies have shown that a 1.9 ratio between the concentrations of Mg^{2+} in the tubular fluid and the interstitial fluid is necessary for passive Mg^{2+} reabsorption (123). Consequently, increased urinary volume in T2DM patients will dilute the Mg^{2+} concentration in the pro-urine reducing the transepithelial chemical Mg^{2+} gradient in the proximal tubule. Mg^{2+} reabsorption in TAL and DCT is inversely correlated to urinary flow. Given that increased glomerular filtration results in high urinary flow rates, hyperfiltration may reduce Mg^{2+} reabsorption in T2DM patients.

Oxidative Stress

One of the main contributors to diabetic nephropathy is the oxidative stress in the kidney (124). Interestingly, oxidative stress has been shown to reduce TRPM6 activity (125), and as a result, Mg^{2+} uptake may be diminished in people with diabetes. Previously, methionine sulfoxide reductase B1 (MSRB1) was shown to prevent the effects of oxidative stress on TRPM6 by reducing oxidation of the channel. However, studies of rats with STZ-induced diabetes showed reduced *MsrB1* expression (126). Thus, oxidative stress may contribute to hypomagnesemia in T2DM.

CONCLUSIONS AND PERSPECTIVES

Over the past two decades, there has been a staggering amount of clinical evidence showing a tight association between hypomagnesemia and T2DM. A recent cross-sectional study has shown that hypomagnesemia is associated with an increased risk for complications, including retinopathy, nephropathy, and foot ulcers (127). Importantly, Mg^{2+} supplementation improved insulin sensitivity and metabolic control in a double-blind randomized trial, suggesting that Mg^{2+} is an important factor in the etiology and management of T2DM (9,10). So far, the clinical trials that have been performed using Mg^{2+} supplementation to improve T2DM have mainly focused on general parameters such as blood glucose or HbA_{1c} levels. Therefore,

well-designed, double-blind randomized trials in T2DM patients with hypomagnesemia studying the long-term effects of Mg^{2+} supplementation on T2DM pathophysiology and disease progression are now warranted.

The prevalence of hypomagnesemia in T2DM has been reported to range between 14 and 48%, meaning that millions of people worldwide are affected (4). Nevertheless, serum Mg^{2+} levels are not routinely determined in T2DM patients. Provided that hypomagnesemia is associated with conditions that are often present in T2DM, including hypertension, hypokalemia, and muscle cramps, more clinical attention is necessary to address this problem (12). Patients using widely prescribed drugs such as thiazide diuretics, proton pump inhibitors, and calcineurin inhibitors are especially at risk for developing hypomagnesemia and should be closely monitored (128,129). Additionally, patients with diabetic neuropathy who have episodes of diarrhea may suffer from intestinal malabsorption of Mg^{2+} , which is another risk factor for hypomagnesemia.

Despite the widespread clinical evidence of the association of hypomagnesemia and T2DM, the molecular mechanisms of Mg^{2+} on insulin secretion and insulin resistance are still far from understood. Currently, the strongest line of evidence supports an effect of Mg^{2+} on insulin sensitivity (24–27). Small-scale fundamental studies have shown that Mg^{2+} is essential for insulin receptor phosphorylation, but the effect of Mg^{2+} on downstream targets in the muscle, liver, and adipocytes is largely unknown. The role of Mg^{2+} in the regulation of insulin secretion is more controversial and hampered by the limited number of clinical and experimental studies. In contrast, there have been significant advances demonstrating the important role of insulin in the regulation of Mg^{2+} reabsorption via TRPM6 in the kidney. Insulin resistance reduces renal Mg^{2+} reabsorption resulting in urinary Mg^{2+} wasting. As a consequence, people with diabetes may end up in a vicious circle in which hypomagnesemia enhances insulin resistance and insulin resistance causes hypomagnesemia. However, the picture is still far from complete, and more studies are required to fully understand the complex and dynamic role of Mg^{2+} in T2DM.

Acknowledgments. The authors state their gratitude to Steef Kurstjens (Radboud University Nijmegen Medical Center, Nijmegen, the Netherlands) for the careful reading of the manuscript.

Funding. This work was supported by grants from the Netherlands Organisation for Scientific Research (VICI 016.130.668) and the EURenOmics project from the European Union Seventh Framework Programme (FP7/2007–2013) (agreement no. 305608). J.H.F.d.B. is supported by grants from the Netherlands Organisation for Scientific Research (Rubicon 825.14.021) and the Dutch Kidney Foundation (Nierstichting) (Kolff 140KG17).

Duality of Interest. No potential conflicts of interest relevant to this article were reported.

References

- Olokoba AB, Obateru OA, Olokoba LB. Type 2 diabetes mellitus: a review of current trends. *Oman Med J* 2012;27:269–273
- Bergman M. Pathophysiology of prediabetes and treatment implications for the prevention of type 2 diabetes mellitus. *Endocrine* 2013;43:504–513
- Martin HE, Wertman M. Serum potassium, magnesium, and calcium levels in diabetic acidosis. *J Clin Invest* 1947;26:217–228
- Pham PC, Pham PM, Pham SV, Miller JM, Pham PT. Hypomagnesemia in patients with type 2 diabetes. *Clin J Am Soc Nephrol* 2007;2:366–373
- Chaudhary DP, Sharma R, Bansal DD. Implications of magnesium deficiency in type 2 diabetes: a review. *Biol Trace Elem Res* 2010;134:119–129
- Pham PC, Pham PM, Pham PA, et al. Lower serum magnesium levels are associated with more rapid decline of renal function in patients with diabetes mellitus type 2. *Clin Nephrol* 2005;63:429–436
- Dong JY, Xun P, He K, Qin LQ. Magnesium intake and risk of type 2 diabetes: meta-analysis of prospective cohort studies. *Diabetes Care* 2011;34:2116–2122
- Guerrero-Romero F, Rodríguez-Morán M. Oral magnesium supplementation: an adjuvant alternative to facing the worldwide challenge of type 2 diabetes? *Cir Cir* 2014;82:282–289 [in Spanish]
- Guerrero-Romero F, Simental-Mendía LE, Hernández-Ronquillo G, Rodríguez-Morán M. Oral magnesium supplementation improves glycaemic status in subjects with prediabetes and hypomagnesaemia: a double-blind placebo-controlled randomized trial. *Diabetes Metab* 2015;41:202–207
- Rodríguez-Morán M, Guerrero-Romero F. Oral magnesium supplementation improves insulin sensitivity and metabolic control in type 2 diabetic subjects: a randomized double-blind controlled trial. *Diabetes Care* 2003;26:1147–1152
- Rodríguez-Morán M, Simental Mendía LE, Zambrano Galvan G, Guerrero-Romero F. The role of magnesium in type 2 diabetes: a brief based-clinical review. *Magnes Res* 2011;24:156–162
- de Baaij JHF, Hoenderop JGJ, Bindels RJM. Magnesium in man: implications for health and disease. *Physiol Rev* 2015;95:1–46
- Barbagallo M, Dominguez LJ. Magnesium metabolism in type 2 diabetes mellitus, metabolic syndrome and insulin resistance. *Arch Biochem Biophys* 2007;458:40–47
- Fröjdö S, Vidal H, Pirola L. Alterations of insulin signaling in type 2 diabetes: a review of the current evidence from humans. *Biochim Biophys Acta* 2009;1792:83–92
- Rodríguez-Morán M, Guerrero-Romero F. Insulin secretion is decreased in non-diabetic individuals with hypomagnesaemia. *Diabetes Metab Res Rev* 2011;27:590–596
- Günther T. The biochemical function of Mg^{2+} in insulin secretion, insulin signal transduction and insulin resistance. *Magnes Res* 2010;23:5–18
- Takaya J, Higashino H, Kobayashi Y. Intracellular magnesium and insulin resistance. *Magnes Res* 2004;17:126–136
- Barbagallo M, Dominguez LJ, Galioto A, et al. Role of magnesium in insulin action, diabetes and cardio-metabolic syndrome X. *Mol Aspects Med* 2003;24:39–52
- Guyton AC, Hall JE. *Textbook of Medical Physiology*. 11th ed. Philadelphia, Saunders, 2006
- Humphries S, Kushner H, Falkner B. Low dietary magnesium is associated with insulin resistance in a sample of young, nondiabetic Black Americans. *Am J Hypertens* 1999;12:747–756
- Lima MdeL, Cruz T, Rodrigues LE, et al. Serum and intracellular magnesium deficiency in patients with metabolic syndrome—evidences for its relation to insulin resistance. *Diabetes Res Clin Pract* 2009;83:257–262
- Nadler JL, Buchanan T, Natarajan R, Antonipillai I, Bergman R, Rude R. Magnesium deficiency produces insulin resistance and increased thromboxane synthesis. *Hypertension* 1993;21:1024–1029
- Hubbard SR. Crystal structure of the activated insulin receptor tyrosine kinase in complex with peptide substrate and ATP analog. *EMBO J* 1997;16:5572–5581
- Vicario PP, Bennun A. Separate effects of Mg^{2+} , MgATP, and ATP⁴⁻ on the kinetic mechanism for insulin receptor tyrosine kinase. *Arch Biochem Biophys* 1990;278:99–105

25. Viñals F, Camps M, Testar X, Palacín M, Zorzano A. Effect of cations on the tyrosine kinase activity of the insulin receptor: inhibition by fluoride is magnesium dependent. *Mol Cell Biochem* 1997;171:69–73
26. Paxton R, Ye L. Regulation of heart insulin receptor tyrosine kinase activity by magnesium and spermine. *Mol Cell Biochem* 2005;277:7–17
27. Suárez A, Pulido N, Casla A, Casanova B, Arrieta FJ, Rovira A. Impaired tyrosine-kinase activity of muscle insulin receptors from hypomagnesaemic rats. *Diabetologia* 1995;38:1262–1270
28. Reis MA, Reyes FG, Saad MJ, Velloso LA. Magnesium deficiency modulates the insulin signaling pathway in liver but not muscle of rats. *J Nutr* 2000;130:133–138
29. Biddinger SB, Kahn CR. From mice to men: insights into the insulin resistance syndromes. *Annu Rev Physiol* 2006;68:123–158
30. Solaimani H, Soltani N, Malekzadeh K, et al. Modulation of GLUT4 expression by oral administration of Mg(2+) to control sugar levels in STZ-induced diabetic rats. *Can J Physiol Pharmacol* 2014;92:438–444
31. Ha BG, Park JE, Cho HJ, Shon YH. Stimulatory effects of balanced deep sea water on mitochondrial biogenesis and function. *PLoS One* 2015;10:e0129972
32. McNeill DA, Herbein JH, Ritchey SJ. Hepatic gluconeogenic enzymes, plasma insulin and glucagon response to magnesium deficiency and fasting. *J Nutr* 1982;112:736–743
33. Takaya J, Iharada A, Okihana H, Kaneko K. Down-regulation of hepatic phosphoenolpyruvate carboxykinase expression in magnesium-deficient rats. *Magnes Res* 2012;25:131–139
34. Uysal KT, Wiesbrock SM, Marino MW, Hotamisligil GS. Protection from obesity-induced insulin resistance in mice lacking TNF- α function. *Nature* 1997;389:610–614
35. de Luca C, Olefsky JM. Inflammation and insulin resistance. *FEBS Lett* 2008;582:97–105
36. Weglicki WB, Phillips TM, Freedman AM, Cassidy MM, Dickens BF. Magnesium-deficiency elevates circulating levels of inflammatory cytokines and endothelin. *Mol Cell Biochem* 1992;110:169–173
37. Rodríguez-Morán M, Guerrero-Romero F. Elevated concentrations of TNF- α are related to low serum magnesium levels in obese subjects. *Magnes Res* 2004;17:189–196
38. Bussi re FI, Gueux E, Rock E, et al. Increased phagocytosis and production of reactive oxygen species by neutrophils during magnesium deficiency in rats and inhibition by high magnesium concentration. *Br J Nutr* 2002;87:107–113
39. Rodr guez-Mor n M, Guerrero-Romero F. Serum magnesium and C-reactive protein levels. *Arch Dis Child* 2008;93:676–680
40. Simental-Mendia LE, Rodr guez-Mor n M, Guerrero-Romero F. Oral magnesium supplementation decreases C-reactive protein levels in subjects with prediabetes and hypomagnesaemia: a clinical randomized double-blind placebo-controlled trial. *Arch Med Res* 2014;45:325–330
41. Henriksen EJ, Diamond-Stanic MK, Marchionne EM. Oxidative stress and the etiology of insulin resistance and type 2 diabetes. *Free Radic Biol Med* 2011;51:993–999
42. Li Q, Zhu X, Ishikura S, et al. Ca²⁺ signals promote GLUT4 exocytosis and reduce its endocytosis in muscle cells. *Am J Physiol Endocrinol Metab* 2014;307:E209–E224
43. Takaya J, Yamato F, Higashino H, Kaneko K. Intracellular magnesium and adipokines in umbilical cord plasma and infant birth size. *Pediatr Res* 2007;62:700–703
44. Matschinsky FM. Regulation of pancreatic beta-cell glucokinase: from basics to therapeutics. *Diabetes* 2002;51(Suppl. 3):S394–S404
45. Ashcroft FM, Rorsman P. K(ATP) channels and islet hormone secretion: new insights and controversies. *Nat Rev Endocrinol* 2013;9:660–669
46. Braun M, Ramracheya R, Bengtsson M, et al. Voltage-gated ion channels in human pancreatic beta-cells: electrophysiological characterization and role in insulin secretion. *Diabetes* 2008;57:1618–1628
47. Weir GC, Bonner-Weir S. Five stages of evolving beta-cell dysfunction during progression to diabetes. *Diabetes* 2004;53(Suppl. 3):S16–S21
48. Meier JJ, Bonadonna RC. Role of reduced β -cell mass versus impaired β -cell function in the pathogenesis of type 2 diabetes. *Diabetes Care* 2013;36(Suppl. 2):S113–S119
49. Billings LK, Florez JC. The genetics of type 2 diabetes: what have we learned from GWAS? *Ann N Y Acad Sci* 2010;1212:59–77
50. Randell EW, Mathews M, Gadag V, Zhang H, Sun G. Relationship between serum magnesium values, lipids and anthropometric risk factors. *Atherosclerosis* 2008;196:413–419
51. Guerrero-Romero F, Rodr guez-Mor n M. Magnesium improves the beta-cell function to compensate variation of insulin sensitivity: double-blind, randomized clinical trial. *Eur J Clin Invest* 2011;41:405–410
52. Molnes J, Teigen K, Aukrust I, et al. Binding of ATP at the active site of human pancreatic glucokinase–nucleotide-induced conformational changes with possible implications for its kinetic cooperativity. *FEBS J* 2011;278:2372–2386
53. Storer AC, Cornish-Bowden A. Kinetics of rat liver glucokinase. Co-operative interactions with glucose at physiologically significant concentrations. *Biochem J* 1976;159:7–14
54. Garfinkel L, Garfinkel D. Magnesium regulation of the glycolytic pathway and the enzymes involved. *Magnesium* 1985;4:60–72
55. Laughlin MR, Thompson D. The regulatory role for magnesium in glycolytic flux of the human erythrocyte. *J Biol Chem* 1996;271:28977–28983
56. Ashcroft FM, Harrison DE, Ashcroft SJ. Glucose induces closure of single potassium channels in isolated rat pancreatic beta-cells. *Nature* 1984;312:446–448
57. Ashcroft FM. ATP-sensitive potassium channelopathies: focus on insulin secretion. *J Clin Invest* 2005;115:2047–2058
58. Proks P, de Wet H, Ashcroft FM. Sulfonylureas suppress the stimulatory action of Mg-nucleotides on Kir6.2/SUR1 but not Kir6.2/SUR2A KATP channels: a mechanistic study. *J Gen Physiol* 2014;144:469–486
59. Ashcroft FM. New uses for old drugs: neonatal diabetes and sulphonylureas. *Cell Metab* 2010;11:179–181
60. Schulla V, Renstr m E, Feil R, et al. Impaired insulin secretion and glucose tolerance in beta cell-selective Ca(v)1.2 Ca²⁺ channel null mice. *EMBO J* 2003;22:3844–3854
61. Nitert MD, Nagorny CL, Wendt A, Eliasson L, Mulder H. Cav1.2 rather than Cav1.3 is coupled to glucose-stimulated insulin secretion in INS-1 832/13 cells. *J Mol Endocrinol* 2008;41:1–11
62. Wang M, Berlin JR. Channel phosphorylation and modulation of L-type Ca²⁺ currents by cytosolic Mg²⁺ concentration. *Am J Physiol Cell Physiol* 2006;291:C83–C92
63. Wang M, Tashiro M, Berlin JR. Regulation of L-type calcium current by intracellular magnesium in rat cardiac myocytes. *J Physiol* 2004;555:383–396
64. Zhang J, Berra-Romani R, Sinnegger-Brauns MJ, Striessnig J, Blaustein MP, Matteson DR. Role of Cav1.2 L-type Ca²⁺ channels in vascular tone: effects of nifedipine and Mg²⁺. *Am J Physiol Heart Circ Physiol* 2007;292:H415–H425
65. Shimaoka T, Wang Y, Morishima M, Miyamoto S, Ono K. Hypomagnesemic down-regulation of L-type Ca(2+) channel in cardiomyocyte as an arrhythmogenic substrate in rats. *Pathophysiology* 2015;22:87–93
66. Atwater I, Frankel BJ, Rojas E, Grodsky GM. Beta cell membrane potential and insulin release; role of calcium and calcium:magnesium ratio. *Q J Exp Physiol* 1983;68:233–245
67. Balon TW, Gu JL, Tokuyama Y, Jasman AP, Nadler JL. Magnesium supplementation reduces development of diabetes in a rat model of spontaneous NIDDM. *Am J Physiol* 1995;269:E745–E752
68. Rubin H. The membrane, magnesium, mitosis (MMM) model of cell proliferation control. *Magnes Res* 2005;18:268–274
69. Chan KH, Chacko SA, Song Y, et al. Genetic variations in magnesium-related ion channels may affect diabetes risk among African American and Hispanic American women. *J Nutr* 2015;145:418–424
70. Romero JR, Castonguay AJ, Barton NS, Germer S, Martin M, Zee RY. Gene variation of the transient receptor potential cation channel, subfamily M,

members 6 (TRPM6) and 7 (TRPM7), and type 2 diabetes mellitus: a case-control study. *Transl Res* 2010;156:235–241

71. Song Y, Hsu YH, Niu T, Manson JE, Buring JE, Liu S. Common genetic variants of the ion channel transient receptor potential membrane melastatin 6 and 7 (TRPM6 and TRPM7), magnesium intake, and risk of type 2 diabetes in women. *BMC Med Genet* 2009;10:4

72. Paolisso G, Sgambato S, Passariello N, et al. Insulin induces opposite changes in plasma and erythrocyte magnesium concentrations in normal man. *Diabetologia* 1986;29:644–647

73. Gylfe E. Insulin secretagogues induce Ca(2+)-like changes in cytoplasmic Mg2+ in pancreatic beta-cells. *Biochim Biophys Acta* 1990;1055:82–86

74. Romani AM, Matthews VD, Scarpa A. Parallel stimulation of glucose and Mg(2+) accumulation by insulin in rat hearts and cardiac ventricular myocytes. *Circ Res* 2000;86:326–333

75. Schlingmann KP, Weber S, Peters M, et al. Hypomagnesemia with secondary hypocalcemia is caused by mutations in TRPM6, a new member of the TRPM gene family. *Nat Genet* 2002;31:166–170

76. Walder RY, Landau D, Meyer P, et al. Mutation of TRPM6 causes familial hypomagnesemia with secondary hypocalcemia. *Nat Genet* 2002;31:171–174

77. Groenestege WM, Hoenderop JG, van den Heuvel L, Knoers N, Bindels RJ. The epithelial Mg2+ channel transient receptor potential melastatin 6 is regulated by dietary Mg2+ content and estrogens. *J Am Soc Nephrol* 2006;17:1035–1043

78. Li M, Du J, Jiang J, et al. Molecular determinants of Mg2+ and Ca2+ permeability and pH sensitivity in TRPM6 and TRPM7. *J Biol Chem* 2007;282:25817–25830

79. Cao G, van der Wijst J, van der Kemp A, van Zeeland F, Bindels RJ, Hoenderop JG. Regulation of the epithelial Mg2+ channel TRPM6 by estrogen and the associated repressor protein of estrogen receptor activity (REA). *J Biol Chem* 2009;284:14788–14795

80. Thebault S, Alexander RT, Tiel Groenestege WM, Hoenderop JG, Bindels RJ. EGF increases TRPM6 activity and surface expression. *J Am Soc Nephrol* 2009;20:78–85

81. de Baaij JH, Blanchard MG, Lavrijsen M, Leipziger J, Bindels RJ, Hoenderop JG. P2X4 receptor regulation of transient receptor potential melastatin type 6 (TRPM6) Mg2+ channels. *Pflugers Arch* 2014;466:1941–1952

82. de Baaij JH, Dorresteyn EM, Hennekam EA, et al. Recurrent FXD2 p.Gly41Arg mutation in patients with isolated dominant hypomagnesaemia. *Nephrol Dial Transplant* 2015;30:952–957

83. Glaudemans B, van der Wijst J, Scola RH, et al. A missense mutation in the Kv1.1 voltage-gated potassium channel-encoding gene KCNA1 is linked to human autosomal dominant hypomagnesemia. *J Clin Invest* 2009;119:936–942

84. Reichold M, Zdebek AA, Lieberer E, et al. KCNJ10 gene mutations causing EAST syndrome (epilepsy, ataxia, sensorineural deafness, and tubulopathy) disrupt channel function. *Proc Natl Acad Sci U S A* 2010;107:14490–14495

85. Arjona FJ, de Baaij JH, Schlingmann KP, et al. CNNM2 mutations cause impaired brain development and seizures in patients with hypomagnesemia. *PLoS Genet* 2014;10:e1004267

86. Hurd TW, Otto EA, Mishima E, et al. Mutation of the Mg2+ transporter SLC41A1 results in a nephronophthisis-like phenotype. *J Am Soc Nephrol* 2013;24:967–977

87. Kolisek M, Nestler A, Vormann J, Schweigel-Röntgen M. Human gene SLC41A1 encodes for the Na+/Mg2+ exchanger. *Am J Physiol Cell Physiol* 2012;302:C318–C326

88. McNair P, Christensen MS, Christiansen C, Madsbad S, Transbøl I. Renal hypomagnesaemia in human diabetes mellitus: its relation to glucose homeostasis. *Eur J Clin Invest* 1982;12:81–85

89. Banting FG, Best CH, Collip JB, Campbell WR, Fletcher AA. Pancreatic extracts in the treatment of diabetes mellitus. *Can Med Assoc J* 1922;12:141–146

90. Atchley DW, Loeb RF, Richards DW, Benedict EM, Driscoll ME. On diabetic acidosis: a detailed study of electrolyte balances following the withdrawal and reestablishment of insulin therapy. *J Clin Invest* 1933;12:297–326

91. Aikawa JK. Effect of glucose and insulin on magnesium metabolism in rabbits. A study with Mg28. *Proc Soc Exp Biol Med* 1960;103:363–366

92. Mandon B, Siga E, Chabardes D, Firsov D, Roinel N, De Rouffignac C. Insulin stimulates Na+, Cl-, Ca2+, and Mg2+ transports in TAL of mouse nephron: cross-potential with AVP. *Am J Physiol* 1993;265:F361–F369

93. Dai LJ, Ritchie G, Bapty BW, Kerstan D, Quamme GA. Insulin stimulates Mg2+ uptake in mouse distal convoluted tubule cells. *Am J Physiol* 1999;277:F907–F913

94. Nair AV, Hocheb B, Verkaart S, et al. Loss of insulin-induced activation of TRPM6 magnesium channels results in impaired glucose tolerance during pregnancy. *Proc Natl Acad Sci U S A* 2012;109:11324–11329

95. Hruby A, Ngwa JS, Renström F, et al. Higher magnesium intake is associated with lower fasting glucose and insulin, with no evidence of interaction with select genetic loci, in a meta-analysis of 15 CHARGE Consortium Studies. *J Nutr* 2013;143:345–353

96. Lee CT, Lien YH, Lai LW, Chen JB, Lin CR, Chen HC. Increased renal calcium and magnesium transporter abundance in streptozotocin-induced diabetes mellitus. *Kidney Int* 2006;69:1786–1791

97. Takayanagi K, Shimizu T, Tayama Y, et al. Down-regulation of transient receptor potential (TRP) M6 channel as a cause of hypermagnesiuric hypomagnesemia in obese type-2 diabetic rats. *Am J Physiol Renal Physiol* 2015;308:F1386–F1397

98. de Baaij JH, Groot Koerkamp MJ, Lavrijsen M, et al. Elucidation of the distal convoluted tubule transcriptome identifies new candidate genes involved in renal Mg(2+) handling. *Am J Physiol Renal Physiol* 2013;305:F1563–F1573

99. Chávez-Canales M, Arroyo JP, Ko B, et al. Insulin increases the functional activity of the renal NaCl cotransporter. *J Hypertens* 2013;31:303–311

100. Komers R, Rogers S, Oyama TT, et al. Enhanced phosphorylation of Na(+)-Cl(-) cotransporter in experimental metabolic syndrome: role of insulin. *Clin Sci (Lond)* 2012;123:635–647

101. Nishida H, Sohara E, Nomura N, et al. Phosphatidylinositol 3-kinase/Akt signaling pathway activates the WNK-OSR1/SPAK-NCC phosphorylation cascade in hyperinsulinemic db/db mice. *Hypertension* 2012;60:981–990

102. Sohara E, Rai T, Yang SS, et al. Acute insulin stimulation induces phosphorylation of the Na-Cl cotransporter in cultured distal mpkDCT cells and mouse kidney. *PLoS One* 2011;6:e24277

103. Takahashi D, Mori T, Nomura N, et al. WNK4 is the major WNK positively regulating NCC in the mouse kidney. *Biosci Rep* 2014;34:34

104. Schutta MH. Diabetes and hypertension: epidemiology of the relationship and pathophysiology of factors associated with these comorbid conditions. *J Cardiometab Syndr* 2007;2:124–130

105. Knoers NV, Levchenko EN. Gitelman syndrome. *Orphanet J Rare Dis* 2008;3:22

106. Davies DL, Fraser R. Do diuretics cause magnesium deficiency? *Br J Clin Pharmacol* 1993;36:1–10

107. Ishii M, Horio Y, Tada Y, et al. Expression and clustered distribution of an inwardly rectifying potassium channel, KAB-2/Kir4.1, on mammalian retinal Müller cell membrane: their regulation by insulin and laminin signals. *J Neurosci* 1997;17:7725–7735

108. Zhang Y, Xu G, Ling Q, Da C. Expression of aquaporin 4 and Kir4.1 in diabetic rat retina: treatment with minocycline. *J Int Med Res* 2011;39:464–479

109. Bockenbauer D, Feather S, Stanescu HC, et al. Epilepsy, ataxia, sensorineural deafness, tubulopathy, and KCNJ10 mutations. *N Engl J Med* 2009;360:1960–1970

110. Scholl UI, Choi M, Liu T, et al. Seizures, sensorineural deafness, ataxia, mental retardation, and electrolyte imbalance (SeSAME syndrome) caused by mutations in KCNJ10. *Proc Natl Acad Sci U S A* 2009;106:5842–5847

111. Galuska D, Kotova O, Barrès R, Chibalin D, Benziane B, Chibalin AV. Altered expression and insulin-induced trafficking of Na+-K+-ATPase in rat skeletal muscle: effects of high-fat diet and exercise. *Am J Physiol Endocrinol Metab* 2009;297:E38–E49

112. Rosta K, Tulassay E, Enzsoly A, et al. Insulin induced translocation of Na⁺/K⁺-ATPase is decreased in the heart of streptozotocin diabetic rats. *Acta Pharmacol Sin* 2009;30:1616–1624
113. Vague P, Coste TC, Jannot MF, Raccach D, Tsimaratos M. C-peptide, Na⁺,K⁺-ATPase, and diabetes. *Exp Diabetes Res* 2004;5:37–50
114. Sweeney G, Klip A. Regulation of the Na⁺/K⁺-ATPase by insulin: why and how? *Mol Cell Biochem* 1998;182:121–133
115. Alves DS, Thulin G, Loffing J, Kashgarian M, Caplan MJ. Akt substrate of 160 kD regulates Na⁺,K⁺-ATPase trafficking in response to energy depletion and renal ischemia. *J Am Soc Nephrol* 2015;26:2765–2776
116. Lennon EJ, Lemann J Jr, Piering WF, Larson LS. The effect of glucose on urinary cation excretion during chronic extracellular volume expansion in normal man. *J Clin Invest* 1974;53:1424–1433
117. Roy DR, Seely JF. Effect of glucose on renal excretion of electrolytes in the rat. *Am J Physiol* 1981;240:F17–F24
118. Ly JP, Onay T, Sison K, et al. The Sweet Pee model for Sglt2 mutation. *J Am Soc Nephrol* 2011;22:113–123
119. Cherney DZ, Perkins BA, Soleymanlou N, et al. Renal hemodynamic effect of sodium-glucose cotransporter 2 inhibition in patients with type 1 diabetes mellitus. *Circulation* 2014;129:587–597
120. Ferrannini E, Ramos SJ, Salsali A, Tang W, List JF. Dapagliflozin monotherapy in type 2 diabetic patients with inadequate glycemic control by diet and exercise: a randomized, double-blind, placebo-controlled, phase 3 trial. *Diabetes Care* 2010;33:2217–2224
121. Ahloulay M, Schmitt F, Déchaux M, Bankir L. Vasopressin and urinary concentrating activity in diabetes mellitus. *Diabetes Metab* 1999;25:213–222
122. Premaratne E, Verma S, Ekinci EI, Theverkalam G, Jerums G, MacIsaac RJ. The impact of hyperfiltration on the diabetic kidney. *Diabetes Metab* 2015;41:5–17
123. Le Grimellec C. Micropuncture study along the proximal convoluted tubule. Electrolyte reabsorption in first convolutions. *Pflugers Arch* 1975;354:133–150
124. Singh DK, Winocour P, Farrington K. Oxidative stress in early diabetic nephropathy: fueling the fire. *Nat Rev Endocrinol* 2011;7:176–184
125. Cao G, Lee KP, van der Wijst J, et al. Methionine sulfoxide reductase B1 (MsrB1) recovers TRPM6 channel activity during oxidative stress. *J Biol Chem* 2010;285:26081–26087
126. Li Y, Zhang W, Li P, Huang K. Effect of streptozocin-induced diabetes mellitus on expression of methionine sulfoxide reductases and accumulation of their substrates in mouse lenses. *Exp Eye Res* 2011;92:401–407
127. Dasgupta A, Sarma D, Saikia UK. Hypomagnesemia in type 2 diabetes mellitus. *Indian J Endocrinol Metab* 2012;16:1000–1003
128. Hess MW, Hoenderop JG, Bindels RJ, Drenth JP. Systematic review: hypomagnesaemia induced by proton pump inhibition. *Aliment Pharmacol Ther* 2012;36:405–413
129. Lameris AL, Monnens LA, Bindels RJ, Hoenderop JG. Drug-induced alterations in Mg²⁺ homeostasis. *Clin Sci (Lond)* 2012;123:1–14