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Glucagon and Amino Acids Are Linked in a Mutual Feedback Cycle: The Liver- α -Cell Axis

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Glucagon is usually viewed as an important counter-regulatory hormone in glucose metabolism, with actions opposing those of insulin. Evidence exists that shows glucagon is important for minute-to-minute regulation of postprandial hepatic glucose production, although conditions of glucagon excess or deficiency do not cause changes compatible with this view. In patients with glucagon-producing tumors (glucagonomas), the most conspicuous signs are skin lesions (necrolytic migratory erythema), while in subjects with inactivating mutations of the glucagon receptor, pancreatic swelling may be the first sign; neither condition is necessarily associated with disturbed glucose metabolism. In glucagonoma patients, amino acid turnover and ureagenesis are greatly accelerated, and low plasma amino acid levels are probably at least partly responsible for the necrolytic migratory erythema, which resolves after amino acid administration. In patients with receptor mutations (and in knockout mice), pancreatic swelling is due to α -cell hyperplasia with gross hypersecretion of glucagon, which according to recent groundbreaking research may result from elevated amino acid levels. Additionally, solid evidence indicates that ureagenesis, and thereby amino acid levels, is critically controlled by glucagon. Together, this constitutes a complete endocrine system; feedback regulation involving amino acids regulates α -cell function and secretion, while glucagon, in turn, regulates amino acid turnover.

Glucagon was discovered in 1923 as a hyperglycemic substance (1), and subsequent research documented its actions on glycogenolysis and gluconeogenesis. Indeed,

glucagon's actions on the cAMP system and its interactions with the glucagon receptor provided substrates for the Nobel prizes of Earl Sutherland, Christian de Duve, and Martin Rodbell (2). With the development of the radioimmunoassay for glucagon (3), delineation of its physiological role became possible, and Akira Ohneda working with Roger Unger, the nestor of modern glucagon research, demonstrated the reciprocal regulation of glucagon secretion by glucose (4). In elegant studies in both experimental animals and humans, typically involving blockade of glucagon (and insulin) secretion by the hormone somatostatin (5,6), it was demonstrated that hepatic glucose production, and thereby the regulation of blood glucose in the fasting state, is maintained in a balance between the inhibitory actions of insulin and the stimulatory actions of glucagon, with each hormone being approximately equally important for the regulation (7).

In further studies, inappropriate secretion of glucagon was observed in various forms of diabetes (8), and pioneering studies by Gerich et al. (9) pointed to an essential role for glucagon in the development of the hyperglycemia and ketoacidosis of type 1 diabetes, leading Unger and Orci (10) to suggest that inappropriate glucagon secretion underlies all forms of diabetic hyperglycemia. This hypothesis spurred attempts to develop glucagon receptor antagonists, which on one hand would allow conclusive studies about the essentiality of glucagon for diabetic hyperglycemia and, on the other hand, might be useful therapeutic agents for diabetes therapy (11). Eventually, the pharmaceutical industry managed to develop small molecule, orally available glucagon receptor antagonists (12,13), which have demonstrated excellent glucose-lowering efficacy

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in clinical studies in patients with type 2 diabetes, in fact providing convincing proof for the importance of (inappropriate?) secretion of glucagon for the hyperglycemia of type 2 diabetes (13).

In the delineation of the physiological role of a hormone, it may be useful to study conditions of hormone excess and hormone deficiency. Among endocrinologists with an interest in glucagon, such conditions were eagerly searched for, employing radioimmunoassays to reveal hyper- and hyposecretion. The latter turned out to be difficult because the glucagon concentrations in plasma are low and measurements, therefore, challenging in terms of assay sensitivity (14). In addition, there were specificity problems, caused by (as it turned out) gut-derived, alternative products of proglucagon, the glucagon precursor (15,16), so conditions with glucagon deficiency were not easily identified. Regarding hyperglucagonemia, patients with a glucagon-secreting tumor were eventually described (17), and with more cases being collected, it became possible to define a "glucagonoma syndrome" (18,19). Some glucagonoma patients did indeed show hyperglycemia, but not all, and overt diabetes is not the rule. It was assumed that this might reflect what is seen after prolonged infusion of glucagon, where the initial elevation of plasma glucose rapidly wanes (20,21). This evanescent glucose response to glucagon was much debated and may be due to the simultaneous stimulation of insulin secretion, whereby hepatic glucose production is reduced (21). The possible consequences of glucagon deficiency remained enigmatic, but because of the interest in the development of glucagon receptor antagonists for possible clinical use, it was important to identify the consequences of glucagon lack. Intensive research from several groups had identified glucagon secretion as the first-in-line defense against hypoglycemia in humans (22), and subcutaneous glucagon injections were used to treat insulin-induced hypoglycemia in patients with type 1 diabetes. Therefore, the expected outcome of glucagon antagonism was severe hypoglycemia. Surgical glucagon deficiency brought about by experimental pancreatectomy turned out not to solve the problem because of apparent production of glucagon from sites outside of the pancreas (as had been suspected since 1948 [23] and subsequently confirmed in humans [24,25]). The development of a high-affinity monoclonal neutralizing antibody against glucagon enabled Brand et al. (26) to bring about a state of apparently complete glucagon deficiency. This resulted in significant but very slight lowering of fed and postprandial glucose levels in rats, mice, and rabbits (26–29), whereas antibody treatment in combination with an extensive fast (up to 48 h) and a 10-h fast plus complete adrenergic blockade (sympathetic activation representing the second defense line against hypoglycemia [22]) did not cause hypoglycemia (27). The actions of glucagon appear to be mediated by a single glucagon receptor (30); therefore, it was also possible to create another condition with lack of glucagon signaling, namely glucagon receptor

knockout mice (31). These animals have similar plasma glucose levels as those treated with antibodies and also do not develop hypoglycemia, although they show pronounced changes in levels of the enzymes thought to be regulated by glucagon: pyruvate carboxylase, glycogen phosphorylases, phosphoenolpyruvate carboxykinase, fructose 1,6-bisphosphatase, and glucose 6-phosphatase. Obviously, life-long global deletion of the glucagon receptor may be associated with important adaptive changes, but the similarity with the immunoneutralization results and with results from further studies employing acute small interfering RNA-induced downregulation of the glucagon receptor (32) was reassuring. Two important lessons emerged from these studies: 1) glucagon is not essential for the maintenance of fasting glucose levels (for this, cortisol/corticosterone is more important because it provides gluconeogenic substrates for gluconeogenesis [33]) and, as a consequence, 2) glucagon antagonism does not cause hypoglycemia. The latter was of course of particular interest to those trying to develop glucagon receptor antagonists for diabetes therapy, and subsequent studies of the antagonists confirmed that they do not cause hypoglycemia (unless combined with inappropriate amounts of insulin) (11,13). Note that these conclusions are not incompatible with a role for glucagon in the regulation of normal and diabetic postprandial and fasting glucose levels, as indicated from studies with glucagon antagonism and deletions of the glucagon receptor (34).

Thus, neither glucagon deficiency nor glucagon excess had quite the expected consequences for plasma glucose regulation. In contrast, there were other, unexpected consequences of the hyperglucagonemia of the glucagon-producing tumors. The main features of the glucagonoma syndrome include a characteristic skin rash termed necrolytic migratory erythema, which is observed in most cases (82% of patients), often with painful glossitis and angular stomatitis (35). Other characteristics are normochromic normocytic anemia (61%), weight loss (90%), deep vein thrombosis (50%), and depression (50%). Mild hyperglycemia is seen in many (80%), but overt diabetes is only observed in about 30% and is related to pancreatic damage. The skin disease is characterized by superficial epidermal necrosis, fragile blister formation, crusting, and healing with hyperpigmentation (36). The histological appearance is quite unique and involves sudden death of the cells of the stratum basale. A biochemical feature of the syndrome is extreme hypoaaminoacidemia, which may involve all or only selected plasma amino acids (36,37). Since the skin is a tissue with an extremely high cell turnover, it was soon suspected that the low plasma amino acid levels might be involved, and, indeed, several subsequent studies have shown that infusions of mixed amino acids may lead to a complete resolution of the condition (37–39). These observations would point to an important role for glucagon in amino acid metabolism, confirmed in elegant

clamp studies demonstrating a critical role for glucagon in the disposal of amino acids (40). As already mentioned, one of glucagon's established effects is to promote gluconeogenesis (41). Among its actions on the liver, increased activity of key enzymes responsible for gluconeogenesis (phosphorylase kinase, phosphoenolpyruvate carboxykinase, fructose 1,6-bisphosphatase, glucose 6-phosphatase) is evident. During prolonged fasting, where glucagon secretion may be elevated (42), gluconeogenesis from glucogenic amino acids derived from the tissues may be particularly prominent. The glucose-alanine cycle, where alanine during prolonged fasting is released to the blood stream from the muscles, converted to pyruvate in the liver, and used as a substrate for gluconeogenesis, is an example (43). Both alanine transferase, responsible for conversion of alanine to pyruvate, as well as the key enzymes responsible for gluconeogenesis of this cycle are regulated by glucagon. Note, however, that the extent of this cycle is dependent on the supply of alanine rather than the actions of glucagon. Indeed, in mice with genetically induced deletion of glucagon gene products, mainly the enzymes related to hepatic amino acid metabolism were affected (as opposed to those regulating gluconeogenesis), and a major finding was hyperaminoacidemia (44). In addition, the isolated effect of glucagon on gluconeogenesis is short-lasting because it cannot provide substrates (45). Pancreatectomized humans exhibit pronounced hyperaminoacidemia, which may be rapidly relieved by glucagon administration (46). Patients with cirrhosis of the liver show hyperaminoacidemia and invariably have hyperglucagonemia (47,48) and impaired ureagenesis in response to glucagon (49).

Amino acid metabolism is complex and serves many purposes. Amino acids may be divided into three categories: glucogenic, ketogenic, or both glucogenic and ketogenic, depending on their ability to be transformed into pyruvate or related products of the Krebs cycle (the glucogenic amino acids) to contribute to the production of glucose (gluconeogenesis) or into acetyl-CoA (acetoacetyl-CoA), which cannot bring about a net increase in glucose production but supplies the Krebs cycle. Importantly, amino acids contain nitrogen and in the liver undergo transaminations and deaminations, eventually leading to ureagenesis. Glucagon strongly promotes ureagenesis (50), in this way removing from the body the ammonia that results from transamination, the accumulation of which potentially would be dangerous to the brain. The urea instead is eliminated with the urine. Indeed, it can be demonstrated, using glucagon immunoneutralization, that glucagon importantly regulates both the capacity for and the rate of urea formation in experimental animals (51). Particularly in diabetic animals with accelerated ureagenesis, glucagon immunoneutralization resulted in normalization of ureagenesis. In patients with glucagonoma, a similarly increased ureagenesis rate was found together with accelerated clearances of most amino acids; lean body mass was reduced in parallel

(52). Conversely, in patients with impaired glucagon secretion (chronic pancreatitis), the capacity for urea formation is reduced as a consequence of glucagon deficiency (53). Thus, it seems clear that excessive (long-term) production of glucagon has a major and potentially dangerous influence on plasma amino acid turnover, whereas glucose metabolism is less affected. However, amino acid metabolism may also be acutely affected, with significant reductions observed after minutes to hours after glucagon administration (45).

Now let us return to glucagon deficiency. As mentioned, the effects of glucagon receptor deletion in the knockout mice on plasma glucose and glucose tolerance were rather inconspicuous. Are there other prominent features of these animals? Indeed, the most dramatic consequence of the deletion is dramatic α -cell hyperplasia and hyperglucagonemia, the latter reaching a similar degree to that found in patients with glucagon-producing tumors (31), i.e., 100- to 1,000-fold elevations. And this is not solely a mouse phenomenon. A number of humans with inactivating mutations in the glucagon receptor gene have been described (11). These patients also develop massive α -cell hyperplasia (sometimes even appearing as an abdominal swelling), leading to dramatic hypersecretion of glucagon and other proglucagon-derived products from their hyperplastic α -cells (54–58). The massive hyperplasia suggests that an α -cell growth factor is produced in response to loss of glucagon action. In elegant experiments, Longuet and colleagues (59,60) showed that selective knockout of the glucagon receptor in the liver resulted in an identical phenotype to that of the global knockout, suggesting that the factor (or factors) is derived from the liver. A similar phenomenon results from blocking glucagon action with glucagon antagonists (11) (also in humans, where tumor-like levels of circulating glucagon may be reached [13]), and although there is data to indicate that the hypersecretion (and probably hyperplasia) is reversible (13), this phenomenon remains a challenge for the clinical development of glucagon receptor antagonists for diabetes therapy. But what might the nature of the hepatic growth factor be? In endocrinology, factors that stimulate the secretion of hormones often also exert trophic actions on the endocrine glands. Hypoglycemia is not known as a trophic factor for the α -cells, but the cells also respond to other metabolic substrates, namely amino acids. It has been known since the early days of the glucagon radioimmunoassay that amino acids are powerful stimulants of glucagon secretion (61,62), and, indeed, intravenous infusion of arginine is widely used as a test for the secretory capacity, not only of the insulin-producing but also the glucagon-producing cells. An intravenous arginine infusion (5 g) generally results in up to 10-fold increases in peripheral plasma levels (63). Recently, research from several laboratories has investigated the possibility that amino acids might provide the growth stimulus to the α -cells in the glucagon receptor knockout (*Gcgr*^{-/-})

mice. Thus, Solloway et al. (64) demonstrated that inhibition of the glucagon receptor leads to reduced catabolism of amino acids in the liver and increased plasma levels of amino acids, which then induce proliferation of pancreatic α -cells. Furthermore, inhibition of mTOR attenuated the proliferation of α -cells but without normalization of the plasma levels of amino acids (the liver was still without glucagon signal and thus unaffected). Similarly, antagonism of the glucagon receptor with glucagon antagonists or pancreatectomy in humans led to increasing amino acid levels (12,46).

As also suggested by Solloway et al. (64), these studies strongly support the idea that the growth factor responsible for the α -cell hyperplasia, and presumably also the hyperglucagonemia after *Gcgr* deletion, is one or more of the plasma amino acids. This supports the idea that a normal function of amino acids is to maintain and regulate the secretion of glucagon, partly by regulating the α -cell number. As already mentioned, the regulation by amino acids of glucagon secretion is very powerful, exceeding what is observed in response to falling blood glucose (with the exception of the branched amino acid, which may actually inhibit glucagon secretion) (62). In turn, the effect of glucagon released from the α -cells, when reaching the liver, would be to accelerate ureagenesis and increase amino acid clearance, thereby reducing plasma amino acid levels. A missing glucagon signal to the liver, in terms of regulation of glucose production, is apparently easily compensated for by decreasing insulin levels (because of falling glucose levels) and increasing cortisol levels to provide more amino acids to the liver. Such circuits might involve hypothalamic glucose-sensing with subsequent sympathetic activation of the adrenal cortex. Normally, glucagon secretion may be stimulated by this mechanism (65). However, without effective glucagon signaling (e.g., impaired liver function, diabetes), the liver is unable to accelerate ureagenesis and the rise in amino acid levels can no longer be controlled (53,66). Indeed, there is evidence that the fasting hyperglucagonemia of type 2 diabetes parallels development of signs of impaired liver function (nonalcoholic fatty liver disease) (67). It was recently proposed that angiopoietin-like 4, an inhibitor of lipoprotein lipase from the adipose tissue, might link glucagon antagonism and α -cell hyperplasia (68). However, the authors used an inefficient glucagon receptor antagonist, and the changes observed both with respect to glucagon secretion and α -cell hyperplasia were miniscule and clearly unrelated to the changes observed after effective antagonists and *Gcgr* deletion.

Taken together, it appears that a perfect feedback loop exists between the liver and the α -cells of the pancreas (Fig. 1), with amino acids as the link to the α -cells and with glucagon as the link to the liver. These data support a concept where a major role for glucagon in physiology and pathophysiology is to regulate amino acid metabolism and the idea that its role in glucose metabolism, although

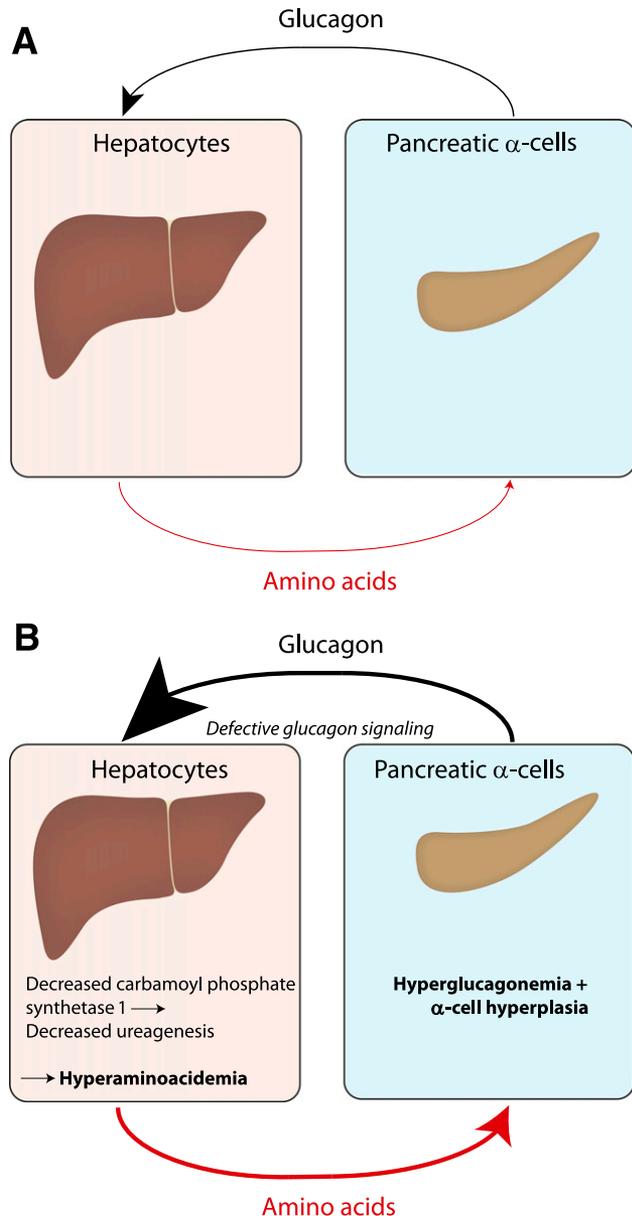


Figure 1—A physiological role of glucagon: regulation of amino acid metabolism. The proposed circuit includes hepatocytes in the liver and pancreatic α -cells. **A:** Under normal circumstances glucagon increases enzymes essential for the regulation of amino acid metabolism (ureagenesis) including in particular carbamoyl phosphate synthetase 1. Carbamoyl phosphate synthetase 1 is the first of two limiting steps in urea cycle, which takes place in the mitochondria and leads to the formation of carbamoyl phosphate from NH_3^- , HCO_3^- , and ATP. Amino acids then stimulate the secretion of glucagon. **B:** Upon disturbances of hepatic glucagon signaling, plasma levels of amino acids increase, causing hypersecretion of glucagon and eventually hyperplasia of pancreatic α -cells.

important, is discounted when disturbances in amino acid balance require restoration by glucagon.

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References

1. Kimball CP, Murlin JR. Aqueous extracts of pancreas. III. Some precipitation reactions of insulin. *J Biol Chem* 1923;58:337–348
2. Lefèbvre PJ. Early milestones in glucagon research. *Diabetes Obes Metab* 2011;13(Suppl. 1):1–4
3. Unger RH, Eisentraut AM, McCall MS, Madison LL. Glucagon antibodies and an immunoassay for glucagon. *J Clin Invest* 1961;40:1280–1289
4. Ohneda A, Aguilar-Parada E, Eisentraut AM, Unger RH. Control of pancreatic glucagon secretion by glucose. *Diabetes* 1969;18:1–10
5. Liljenquist JE, Chiasson JL, Cherrington AD, et al. An important role for glucagon in the regulation of glucose production in vivo. *Metabolism* 1976;25(Suppl. 1):1371–1373
6. Cherrington AD, Chiasson JL, Liljenquist JE, Jennings AS, Keller U, Lacy WW. The role of insulin and glucagon in the regulation of basal glucose production in the postabsorptive dog. *J Clin Invest* 1976;58:1407–1418
7. Alford FP, Bloom SR, Nabarro JD, et al. Glucagon control of fasting glucose in man. *Lancet* 1974;2:974–977
8. Müller WA, Faloona GR, Aguilar-Parada E, Unger RH. Abnormal alpha-cell function in diabetes. Response to carbohydrate and protein ingestion. *N Engl J Med* 1970;283:109–115
9. Gerich JE, Lorenzi M, Bier DM, et al. Prevention of human diabetic ketoacidosis by somatostatin. Evidence for an essential role of glucagon. *N Engl J Med* 1975;292:985–989
10. Unger RH, Orci L. The essential role of glucagon in the pathogenesis of diabetes mellitus. *Lancet* 1975;1:14–16
11. Bagger JI, Knop FK, Holst JJ, Vilsboll T. Glucagon receptor antagonism for the treatment of type 2 diabetes. *Diabetes Obes Metab* 2011;13:965–971
12. Mu J, Qureshi SA, Brady EJ, et al. Anti-diabetic efficacy and impact on amino acid metabolism of GRA1, a novel small-molecule glucagon receptor antagonist. *PLoS One* 2012;7:e49572
13. Kazda CM, Ding Y, Kelly RP, et al. Evaluation of efficacy and safety of the glucagon receptor antagonist LY2409021 in patients with type 2 diabetes: 12- and 24-week phase 2 studies. *Diabetes Care* 2016;39:1241–1249
14. Wewer Albrechtsen NJ, Veedfald S, Plamboeck A, et al. Inability of some commercial assays to measure suppression of glucagon secretion. *J Diabetes Res* 2016;2016:8352957
15. Bak MJ, Albrechtsen NW, Pedersen J, et al. Specificity and sensitivity of commercially available assays for glucagon and oxyntomodulin measurement in humans. *Eur J Endocrinol* 2014;170:529–538
16. Holst JJ. Glucagon and glucagon-like peptides 1 and 2. *Results Probl Cell Differ* 2010;50:121–135
17. McGavran MH, Unger RH, Recant L, Polk HC, Kilo C, Levin ME. A glucagon-secreting alpha-cell carcinoma of the pancreas. *N Engl J Med* 1966;274:1408–1413
18. Mallinson CN, Bloom SR, Warin AP, Salmon PR, Cox B. A glucagonoma syndrome. *Lancet* 1974;2:1–5
19. Holst JJ. Glucagonomas. In *Gut Hormones*. Bloom SR, Ed. Edinburgh, Churchill-Livingstone, 1978, p. 599–604
20. Sherwin RS, Fisher M, Hendler R, Felig P. Hyperglucagonemia and blood glucose regulation in normal, obese and diabetic subjects. *N Engl J Med* 1976;294:455–461
21. Holst JJ, Madsen OG, Knop J, Schmidt A. The effect of intraportal and peripheral infusions of glucagon on insulin and glucose concentrations and glucose tolerance in normal man. *Diabetologia* 1977;13:487–490
22. Cryer PE. Banting Lecture. Hypoglycemia: the limiting factor in the management of IDDM. *Diabetes* 1994;43:1378–1389
23. Sutherland EW, De Duve C. Origin and distribution of the hyperglycemic-glycogenolytic factor of the pancreas. *J Biol Chem* 1948;175:663–674
24. Holst JJ, Pedersen JH, Baldissera F, Stadil F. Circulating glucagon after total pancreatectomy in man. *Diabetologia* 1983;25:396–399
25. Lund A, Bagger JI, Wewer Albrechtsen NJ, et al. Evidence of extrapancreatic glucagon secretion in man. *Diabetes* 2016;65:585–597
26. Brand CL, Rolin B, Jørgensen PN, Svendsen I, Kristensen JS, Holst JJ. Immunoneutralization of endogenous glucagon with monoclonal glucagon antibody normalizes hyperglycaemia in moderately streptozotocin-diabetic rats. *Diabetologia* 1994;37:985–993
27. Brand CL, Jørgensen PN, Knigge U, et al. Role of glucagon in maintenance of euglycemia in fed and fasted rats. *Am J Physiol* 1995;269:E469–E477
28. Brand CL, Jørgensen PN, Svendsen I, Holst JJ. Evidence for a major role for glucagon in regulation of plasma glucose in conscious, nondiabetic, and alloxan-induced diabetic rabbits. *Diabetes* 1996;45:1076–1083
29. Brand CL, Rolin B, Jørgensen PN, Svendsen I, Kristensen JS, Holst JJ. Immunoneutralization of endogenous glucagon with monoclonal glucagon antibody normalizes hyperglycaemia in moderately streptozotocin-diabetic rats. *Diabetologia* 1994;37:985–993
30. Jelinek LJ, Lok S, Rosenberg GB, et al. Expression cloning and signaling properties of the rat glucagon receptor. *Science* 1993;259:1614–1616
31. Gelling RW, Du XQ, Dichmann DS, et al. Lower blood glucose, hyperglucagonemia, and pancreatic alpha cell hyperplasia in glucagon receptor knockout mice. *Proc Natl Acad Sci USA* 2003;100:1438–1443
32. Sloop KW, Cao JX, Siesky AM, et al. Hepatic and glucagon-like peptide-1-mediated reversal of diabetes by glucagon receptor antisense oligonucleotide inhibitors. *J Clin Invest* 2004;113:1571–1581
33. Magomedova L, Cummins CL. Glucocorticoids and metabolic control. *Handb Exp Pharmacol* 2016;233:73–93
34. Unger RH, Cherrington AD. Glucagonocentric restructuring of diabetes: a pathophysiologic and therapeutic makeover. *J Clin Invest* 2012;122:4–12
35. Vinik A, Feliberti E, Perry RR. Glucagonoma syndrome. In *Endotext*. De Groot LJ, Chrousos G, Dungan K, et al., Eds. South Dartmouth, MA, MDText.com Inc., 2000
36. Pedersen NB, Jonsson L, Holst JJ. Necrolytic migratory erythema and glucagon cell tumour of the pancreas: the glucagonoma syndrome. Report of two cases. *Acta Derm Venereol* 1976;56:391–395
37. Norton JA, Kahn CR, Schiebinger R, Gorschboth C, Brennan MF. Amino acid deficiency and the skin rash associated with glucagonoma. *Ann Intern Med* 1979;91:213–215
38. Alexander EK, Robinson M, Staniec M, Dluhy RG. Peripheral amino acid and fatty acid infusion for the treatment of necrolytic migratory erythema in the glucagonoma syndrome. *Clin Endocrinol (Oxf)* 2002;57:827–831
39. Thomaidou E, Nahmias A, Gilead L, Zlotogorski A, Ramot Y. Rapid clearance of necrolytic migratory erythema following intravenous administration of amino acids. *JAMA Dermatol* 2016;152:345–346
40. Charlton MR, Adey DB, Nair KS. Evidence for a catabolic role of glucagon during an amino acid load. *J Clin Invest* 1996;98:90–99
41. Ramnanan CJ, Edgerton DS, Kraft G, Cherrington AD. Physiologic action of glucagon on liver glucose metabolism. *Diabetes Obes Metab* 2011;13(Suppl. 1):118–125
42. Hojlund K, Wildner-Christensen M, Eshøj O, et al. Reference intervals for glucose, beta-cell polypeptides, and counterregulatory factors during prolonged fasting. *Am J Physiol Endocrinol Metab* 2001;280:E50–E58
43. Felig P. The glucose-alanine cycle. *Metabolism* 1973;22:179–207
44. Watanabe C, Seino Y, Miyahira H, et al. Remodeling of hepatic metabolism and hyperaminoacidemia in mice deficient in proglucagon-derived peptides. *Diabetes* 2012;61:74–84
45. Battezzati A, Simonson DC, Luzi L, Matthews DE. Glucagon increases glutamine uptake without affecting glutamine release in humans. *Metabolism* 1998;47:713–723
46. Boden G, Master RW, Rezvani I, Palmer JP, Lobe TE, Owen OE. Glucagon deficiency and hyperaminoacidemia after total pancreatectomy. *J Clin Invest* 1980;65:706–716
47. Marchesini G, Forlani G, Zoli M, et al. Insulin and glucagon levels in liver cirrhosis. Relationship with plasma amino acid imbalance of chronic hepatic encephalopathy. *Dig Dis Sci* 1979;24:594–601

48. Holst JJ, Burchard F, Kühl C. Pancreatic glucoregulatory hormones in cirrhosis of the liver: portal vein concentrations during intravenous glucose tolerance test and in response to a meal. *Diabetes Metab* 1980;6:117–127
49. Bugianesi E, Kalhan S, Burkett E, Marchesini G, McCullough A. Quantification of gluconeogenesis in cirrhosis: response to glucagon. *Gastroenterology* 1998;115:1530–1540
50. Hamberg O, Vilstrup H. Regulation of urea synthesis by glucose and glucagon in normal man. *Clin Nutr* 1994;13:183–191
51. Almdal TP, Holst JJ, Heindorff H, Vilstrup H. Glucagon immunoneutralization in diabetic rats normalizes urea synthesis and decreases nitrogen wasting. *Diabetes* 1992;41:12–16
52. Almdal TP, Heindorff H, Bardram L, Vilstrup H. Increased amino acid clearance and urea synthesis in a patient with glucagonoma. *Gut* 1990;31:946–948
53. Hamberg O, Andersen V, Sonne J, Larsen S, Vilstrup H. Urea synthesis in patients with chronic pancreatitis: relation to glucagon secretion and dietary protein intake. *Clin Nutr* 2001;20:493–501
54. Larger E, Wewer Albrechtsen NJ, Hansen LH, et al. Pancreatic α -cell hyperplasia and hyperglucagonemia due to a glucagon receptor splice mutation [article online], 2016. Available from <https://www.edmcasereports.com/articles/endocrinology-diabetes-and-metabolism-case-reports/10.1530/EDM-16-0081>. Accessed 27 August 2014
55. Balas D, Senegas-Balas F, Delvaux M, et al. Silent human pancreatic glucagonoma and “A” nesidioblastosis. *Pancreas* 1988;3:734–739
56. Martignoni ME, Kated H, Stiegler M, et al. Nesidioblastosis with glucagon-reactive islet cell hyperplasia: a case report. *Pancreas* 2003;26:402–405
57. Yu R, Nissen NN, Dhall D, Heaney AP. Nesidioblastosis and hyperplasia of alpha cells, microglucagonoma, and nonfunctioning islet cell tumor of the pancreas: review of the literature. *Pancreas* 2008;36:428–431
58. Zhou C, Dhall D, Nissen NN, Chen CR, Yu R. Homozygous P86S mutation of the human glucagon receptor is associated with hyperglucagonemia, alpha cell hyperplasia, and islet cell tumor. *Pancreas* 2009;38:941–946
59. Longuet C, Robledo AM, Dean ED, et al. Liver-specific disruption of the murine glucagon receptor produces α -cell hyperplasia: evidence for a circulating α -cell growth factor. *Diabetes* 2013;62:1196–1205
60. Longuet C, Sinclair EM, Maida A, et al. The glucagon receptor is required for the adaptive metabolic response to fasting. *Cell Metab* 2008;8:359–371
61. Unger RH, Ohneda A, Aguilar-Parada E, Eisentraut AM. The role of aminogenic glucagon secretion in blood glucose homeostasis. *J Clin Invest* 1969;48:810–822
62. Rocha DM, Faloona GR, Unger RH. Glucagon-stimulating activity of 20 amino acids in dogs. *J Clin Invest* 1972;51:2346–2351
63. Orskov C, Jeppesen J, Madsbad S, Holst JJ. Proglucagon products in plasma of noninsulin-dependent diabetics and nondiabetic controls in the fasting state and after oral glucose and intravenous arginine. *J Clin Invest* 1991;87:415–423
64. Solloway MJ, Madjidi A, Gu C, et al. Glucagon couples hepatic amino acid catabolism to mTOR-dependent regulation of α -cell mass. *Cell Reports* 2015;12:495–510
65. Thorens B. GLUT2, glucose sensing and glucose homeostasis. *Diabetologia* 2015;58:221–232
66. Heindorff H, Holst JJ, Almdal T, Vilstrup H. Effect of glucagon immunoneutralization on the increase in urea synthesis after hysterectomy in rats. *Eur J Clin Invest* 1993;23:166–170
67. Junker AE, Gluud L, Holst JJ, Knop FK, Vilsbøll T. Diabetic and nondiabetic patients with nonalcoholic fatty liver disease have an impaired incretin effect and fasting hyperglucagonaemia. *J Intern Med* 2016;279:485–493
68. Ben-Zvi D, Barrandon O, Hadley S, Blum B, Peterson QP, Melton DA. *Angptl4* links α -cell proliferation following glucagon receptor inhibition with adipose tissue triglyceride metabolism. *Proc Natl Acad Sci USA* 2015;112:15498–15503