



Hepatic Insulin Clearance in Regulation of Systemic Insulin Concentrations—Role of Carbohydrate and Energy Availability

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Hyperinsulinemia is the hallmark of insulin resistance in obesity, and the relative importance of insulin clearance, insulin resistance, and insulin hypersecretion has been widely debated. On the basis of recent experimental evidence, we summarize existing evidence to suggest hepatic insulin clearance as a major and immediate regulator of systemic insulin concentrations responding within days to altered dietary energy and, in particular, carbohydrate intake. Hepatic insulin clearance seems to be closely associated with opposite alterations in hepatic lipid content and glucose production, providing a potential mechanistic link to hepatic insulin sensitivity. The molecular regulation of insulin clearance in the liver is likely to involve changes in insulin binding and receptor internalization in response to the dietary alterations, the molecular mechanisms of which await further research.

Hyperinsulinemia is the hallmark of insulin resistance in obesity. Still, the pathogenesis of hyperinsulinemia has been debated for decades, in particular with respect to the interplay between insulin hypersecretion and insulin resistance. It is generally assumed that upregulation of insulin secretion compensates for insulin resistance (1), but it has also been suggested that insulin resistance develops secondary to insulin hypersecretion (2). The relative importance of peripheral versus hepatic insulin resistance and the role of insulin clearance have, however, received less attention (3). Insulin is cleared mainly by the liver, and hepatic insulin clearance therefore contributes to regulate

insulin action by controlling insulin availability to peripheral tissues. Recently, reduced hepatic insulin clearance has been suggested as the initial driver for systemic hyperinsulinemia in obesity, while insulin hypersecretion was observed in more advanced stages of insulin resistance (4,5). In accordance, we reported reduced insulin clearance in healthy lean subjects after 3 days on a diet with high carbohydrate (80E%) and low fat (9E%) content and a 75% increase in daily energy provision (6), whereas energy restriction induced by Roux-en-Y gastric bypass (RYGB) markedly increased insulin clearance within 1 week in obese subjects with normal glucose tolerance and obese patients with preoperative type 2 diabetes (T2D) (7) (Fig. 1A and B). Interestingly, in the dietary study, 3 days with high dietary fat (78E%) and reduced carbohydrate (10E%) intake under conditions of matched 75% caloric excess led to increased insulin clearance (6) (Fig. 1A), pointing toward carbohydrate availability rather than energy availability as the major regulator of insulin clearance. In both studies, the up- or downregulation of insulin clearance occurred in parallel with opposite changes in basal hepatic glucose production, as assessed by glucose tracer infusion (Fig. 1C and D), suggesting a mutual interaction between insulin clearance and insulin sensitivity of the liver. Notably, the changes in insulin clearance and hepatic glucoregulation occurred independently of changes in peripheral insulin sensitivity as measured by the hyperinsulinemic-euglycemic clamp (6).

These observations suggest that insulin clearance in the liver is rapidly modified under conditions of changed

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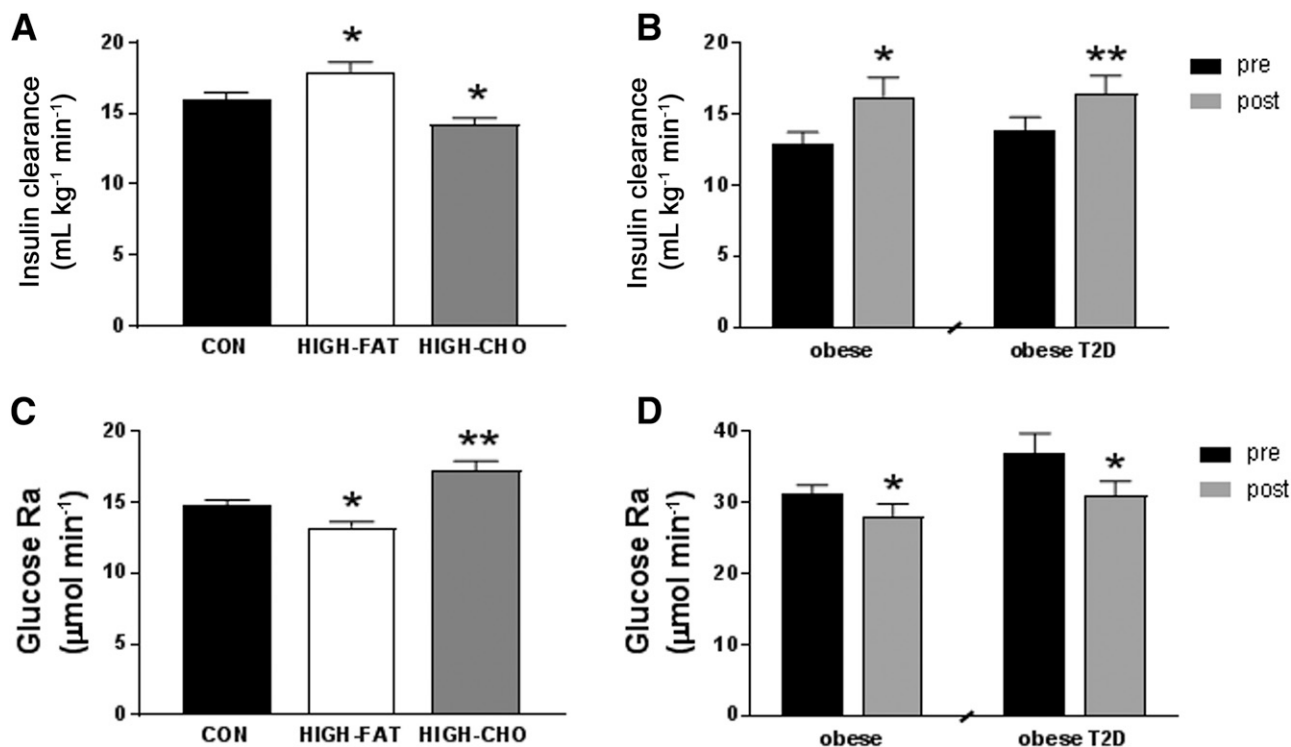


Figure 1—Insulin clearance and basal hepatic glucose production after 3 days manipulation with macronutrient and energy intake in healthy young subjects (A and C) and before (pre) and 1 week after (post) RYGB in obese subjects with preoperative normal glucose tolerance (obese) or T2D (obese T2D) (B and D). Data were previously published in Lundsgaard et al. (6) and Bojsen-Møller et al. (7). Insulin clearance was calculated from hyperinsulinemic-euglycemic clamps with insulin infusion rates of 56 and 40 mU/m²/min, respectively. Glucose rate of appearance (Ra) was calculated from the results of a 2-h primed continuous basal infusion of 6,6-2H₂ glucose tracer (0.055 and 0.036 mg/kg/min, respectively). CON: eucaloric diet (13.7 ± 0.2 MJ/day) with 65E% carbohydrate, 14E% fat, and 24E% protein. HIGH-FAT: 3-day hypercaloric diet (24.0 ± 0.4 MJ/day) with 10E% carbohydrate, 78E% fat, and 12E% protein. HIGH-CHO: 3-day hypercaloric diet (24.0 ± 0.4 MJ/day) with 80E% carbohydrate, 9E% fat, and 11E% protein. Data are means ± SE. For A and C, *n* = 9 except for *n* = 8 in HIGH-CHO. For B and D, *n* = 8 in each group. **P* < 0.05, ***P* < 0.01 compared with CON or pre.

dietary carbohydrate intake. In the following, we summarize the evidence supporting reduced hepatic insulin clearance induced by acute increase in energy and carbohydrate availability as an important mediator in the development of systemic hyperinsulinemia.

INSULIN CLEARANCE—SITE AND MEASUREMENT

The liver is the major site of insulin clearance. Splanchnic extraction of peripherally administered insulin is 50–70% when directly measured by the hepatic venous catheter technique, and this represents almost exclusively hepatic clearance (8–10). For endogenous insulin, hepatic extraction is of even greater importance due to the direct delivery to the portal vein (11,12). The kidneys are the main site of extra-splanchnic insulin clearance, with additional contributions resulting from uptake and degradation by peripheral insulin-sensitive tissues, i.e., skeletal muscle and adipose tissue (10). Renal clearance is, however, modest (~15–25%) under physiological conditions (10), and even in patients with end-stage renal failure, systemic insulin clearance is only slightly affected (13). Hence, the liver is the main organ responsible for clearance of exogenous and in particular endogenous insulin (10). Systemic insulin concentrations are thus excessively

high in liver cirrhosis explained by decreased hepatic insulin clearance (14), and liver transplantation normalizes the hyperinsulinemia (15). Hepatic insulin clearance involves insulin binding to the insulin receptor on hepatocytes and subsequent insulin receptor endocytosis (16). The insulin receptor has two isoforms derived from alternative splicing: type A and type B, with type B being primarily expressed in human liver (17). The molecular regulation of intrahepatic insulin degradation remains to be fully uncovered. It is believed that part of the insulin is degraded by lysosomal proteolysis in the hepatocytes. Here, an insulin-degrading enzyme (IDE) was proposed to be implicated, but molecular inhibition studies have recently questioned a major role of IDE in hepatic insulin clearance (18). Thus, insulin degradation was normal when measured *in vitro* in liver homogenates from cirrhotic compared with healthy subjects (19). This observation suggests that the main mechanisms behind insulin clearance involve insulin binding and receptor internalization. In agreement, inactivating insulin receptor mutations are associated with massive hyperinsulinemia resulting from severely impaired insulin clearance (20).

Whole-body insulin clearance can be estimated with reasonable accuracy during exogenous insulin infusions, as

during the hyperinsulinemic-euglycemic clamp (10) or the insulin suppression test (21). Coinfusion of somatostatin (or analogs thereof) allows for precise estimation of clearance of the exogenously administered insulin by eliminating endogenous insulin secretion (21). For endogenous insulin, the ratio of C-peptide to insulin concentrations may provide an estimate of hepatic insulin clearance. The latter approach builds on the assumption of absent hepatic extraction of C-peptide (8,11) and performs best in steady-state conditions (such as fasting) (22). The ratio can also be used during nonsteady state by use of areas-under-the-curves of C-peptide and insulin, provided that the concentration curves have returned to basal (22).

Absolute hepatic insulin clearance has been found to increase with increasing insulin concentrations, keeping the fractional insulin extraction stable (8,9). However, high portal insulin concentrations have been suggested to be associated with decreased insulin clearance due to receptor saturation (23), which is supported by findings of decreased fractional clearance during supraphysiological insulin infusions (10) and after excessive stimulation of endogenous insulin secretion with clamping at high glucose (300 mg/dL) concentrations for 2 h (24). Whether insulin clearance is saturable during normal physiological conditions (i.e., after meal intake) has been widely debated. Modeling of C-peptide and insulin concentrations showed reduced fractional hepatic insulin extraction with increasing oral loads of glucose in some (25,26) but not all (27) studies, whereas hepatic vein sampling demonstrated increasing absolute (and stable fractional) extraction of insulin with increasing glucose loads (28). Recently, a study with hepatic vein sampling suggested that hepatic insulin clearance increases in parallel with the arrival and passage of secretory insulin pulses, thus dampening systemic insulin oscillations (29), a finding which would be incompatible with acute saturation of the clearance mechanism during physiological conditions.

MODULATION BY DIETARY CARBOHYDRATE AND ENERGY EXCESS

In our dietary study, 3 days of 75% excess caloric intake with 80E% carbohydrate significantly decreased clearance of exogenous insulin (Fig. 1A) (6). Fasting plasma insulin concentrations were increased by 60%, whereas only minor changes in plasma C-peptide concentrations were observed, thus supporting decreased clearance of endogenous insulin as well. Observations in other studies support that short-term carbohydrate overfeeding decreases insulin clearance. Thus, in lean healthy men, 7 days of 50–70% energy excess with 60–86E% carbohydrate increased fasting plasma insulin concentrations by 51–188% (30,31) and augmented plasma insulin during an oral glucose tolerance test twofold (32). From these studies, the absolute carbohydrate intake seems to play an important role in modification of insulin clearance, though clearance was not measured directly. One study reported that progressive overfeeding with a 67E% carbohydrate-rich diet, reaching

+210% energy excess within 4 days, increased fasting plasma insulin concentration by 150%, while fasting C-peptide increased by only 50% (33), indicating that decreased insulin clearance was a major contributor to the hyperinsulinemia. As more direct evidence of reduced insulin clearance, 13 days of 62% caloric excess by supplementation of a carbohydrate-rich drink to the habitual diet of lean healthy men was associated with a 10% increase in plasma insulin concentrations during a hyperinsulinemic-euglycemic clamp (34). Finally, increasing energy intake in lean healthy men by adding a carbohydrate-rich liquid drink to their diet, resulting in 7% body weight gain within ~4 weeks, decreased insulin clearance under an oral glucose tolerance test, while insulin clearance was restored to baseline values when subjects shifted to a hypocaloric diet (35). Similarly, in the early phase of weight gain induced by prolonged overeating, insulin clearance rather than increased insulin secretion was the mediator of the hyperinsulinemia induced by the weight gain (36).

MODULATION BY DIETARY CARBOHYDRATE AND ENERGY RESTRICTION

After RYGB, clearance of exogenous insulin increased from 12.9 to 16.2 mL/min/kg within 1 week after surgery (7), while surgical reversal of the RYGB had the opposite effect (16.6 to 14.2 mL/min/kg) in our recent case report (37). The fasting C-peptide-to-insulin ratios were similarly increased and decreased in response to RYGB and the reversal surgery, respectively (7,37). Other studies showing lower plasma insulin concentrations during hyperinsulinemic clamps and increased fasting plasma C-peptide-to-insulin ratio (38–41) are consistent with these findings of increased clearance of exogenous and endogenous insulin as an early (days to weeks) consequence of RYGB. Interestingly, it has been shown that RYGB-induced weight loss changes the expression of the liver insulin receptor isoforms A and B (42), with potential consequences for insulin affinity/signaling and hence insulin clearance.

Studies of short-term very-low-calorie diets also showed increased insulin clearance (43) and declines in plasma insulin concentrations within 48 h (44), supporting that increased insulin clearance is related to energy restriction rather than RYGB itself (41). After RYGB, energy intake is indeed acutely reduced by at least 35–50%, as is the total carbohydrate intake, while acute changes in macronutrient composition have not been consistently reported (45). Interestingly, 47% energy restriction with only 10E% carbohydrate induced a greater lowering of plasma insulin concentration than similar energy restriction with 65% carbohydrate (44), again supporting a role for the absolute carbohydrate availability for insulin clearance.

HEPATIC FAT CONTENT AND INSULIN CLEARANCE

The clearance of insulin is an integral part of insulin's action on the liver, i.e., both rely on insulin binding to the insulin receptor. The short-term manipulation with energy

and carbohydrate availability discussed above was associated with concomitant opposite changes in insulin clearance and glucose production (Fig. 1), supporting a link between hepatic insulin clearance and hepatic insulin sensitivity. An inverse association between insulin clearance and basal as well as insulin-suppressed hepatic glucose production has also been demonstrated in cross-sectional studies (46).

Hepatic triacylglycerol (TG) accumulation is likely to be a common link, as hepatic TG content correlates negatively with both insulin clearance and hepatic insulin sensitivity (47). This association between hepatic fat and insulin clearance is further supported by in vitro observations in hepatocytes, in which TG accumulation acutely reduces insulin clearance (48). Accordingly, hepatic fat was shown to be the closest correlate to fasting plasma insulin concentration in 271 subjects without diabetes (49). Thus, when subjects matched on BMI were divided in accordance to hepatic TG, fasting plasma insulin concentration was twofold higher in the group with high hepatic TG content (50). Similarly, insulin clearance differs markedly between metabolically healthy and unhealthy obese subjects of comparable BMI (51). Also, obese women with polycystic ovarian syndrome, known to have a high prevalence of hepatic steatosis, have 43% reduced insulin clearance when compared with BMI-matched women (52), and insulin clearance increases with dietary energy restriction (53).

In summary, total adiposity is not the primary determinant of insulin clearance, which is rather related to hepatic TG content (54). Studying the pathogenesis of hyperinsulinemia in obesity thus requires estimation of hepatic versus subcutaneous fat, which could explain some of the divergent results regarding the relative importance of insulin clearance versus insulin hypersecretion in obesity-related hyperinsulinemia (51,55). Also, ethnicity (56) and genetics (57) may contribute to the relative contribution of insulin clearance in the pathogenesis of hyperinsulinemia.

ENERGY AND CARBOHYDRATE EXCESS— HEPATIC TG AS THE LINK TO INSULIN CLEARANCE

Excess energy and carbohydrate intake versus restriction would be expected to have opposing effects on hepatic TG content. Increased monosaccharide availability combined with carbohydrate-induced postprandial insulin hypersecretion promote hepatic de novo lipogenesis (58,59), even at eucaloric conditions (60), which thereby could lead to increased hepatic TG content. The fasting plasma TG concentration increased substantially ($1.7 \pm 0.2 \mu\text{mol/L}$) in healthy subjects after 3 days excess carbohydrate intake, while being reduced by 40% after the similarly hypercaloric high-fat diet with only 10E% carbohydrate (6). Likewise, fasting plasma TG was elevated 1.1- to 10-fold in other dietary studies with short-term overfeeding of carbohydrate-rich diets (30,32). A close and negative association was observed between the diet-induced changes in fasting plasma TG concentration and the changes in insulin clearance in the dietary study (Fig. 2). With fasting plasma VLDL-TG concentration reflecting hepatic TG content as

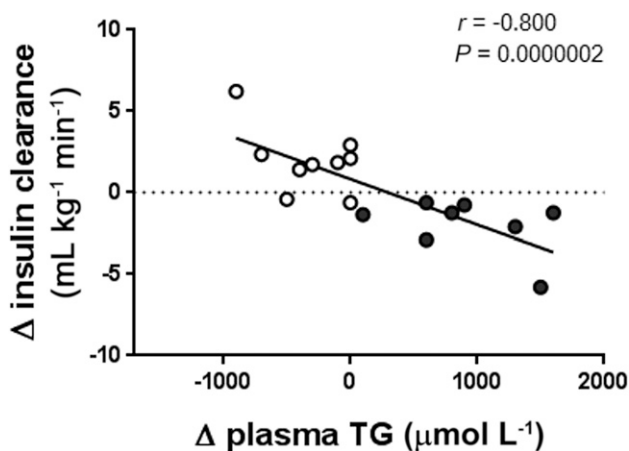


Figure 2—Scatter plot illustrating the association between the change in fasting plasma TG concentration and the change in insulin clearance after 3 days HIGH-CHO (black circles) and HIGH-FAT (white circles) dietary interventions compared with CON. CON: eucaloric diet with 65E% carbohydrate. HIGH-CHO: 3-day hypercaloric diet with 80E% carbohydrate and 11E% fat. HIGH-FAT: 3-day hypercaloric diet with 79E% fat and 10E% carbohydrate. Pearson correlation analysis was applied. Data from Lundsgaard et al. (6).

previously reported (61), these results suggest that accumulation of hepatic TG upon hypercaloric high carbohydrate intake is directly linked to decreased insulin clearance and vice versa during high fat/low carbohydrate intake.

The decreased insulin clearance after dietary carbohydrate excess may be mechanistically linked to the glycoprotein carcinoembryonic antigen-related cell adhesion molecule 1 (CEACAM1). CEACAM1 promotes internalization of the insulin-receptor complex (62) by insulin receptor tyrosine kinase-induced phosphorylation of CEACAM1 (63), which subsequently forms part of a protein complex mediating insulin receptor endocytosis (64). Hence, CEACAM1 knockout markedly reduces insulin clearance in mice (65), while CEACAM1 reconstitution restores insulin clearance and reverses the insulin-resistant phenotype (66). Interestingly, CEACAM1 can also be associated with the enzyme fatty acid synthase (FAS), which is central to de novo lipogenesis; thus, in hyperinsulinemic *ob/ob* mice that have elevated FAS activity, insulin failed to induce CEACAM1 phosphorylation (67). The resulting lower CEACAM1 activity might lead to reduced insulin receptor endocytosis in conditions with high de novo lipogenesis. Furthermore, fatty acid (FA) products of de novo lipogenesis are shown to be ligands for peroxisome proliferator-activated receptor- α (PPAR α) (68), with PPAR α activation reducing CEACAM1 expression (69). CEACAM1 could therefore be a candidate hepatic molecule linking insulin clearance and carbohydrate-induced de novo lipogenesis-derived TG accumulation. In support of this, hepatic CEACAM1 protein expression was recently found to be reduced in obese compared with lean subjects (70) and in subjects with fatty liver disease (71).

HEPATIC FAT AFTER ENERGY AND CARBOHYDRATE RESTRICTION

Energy restriction introduced by RYGB has been demonstrated to reduce hepatic fat within 1 week and before major body weight loss (72). Also, 1 week of energy restriction by diet alone resulted in 30% reduction in hepatic TG content and 50% reduction in fasting plasma TG, concomitantly with 38% reduction in fasting plasma insulin, with unchanged C-peptide concentrations (73). Of note, as little as 48 h of energy restriction with low (10E%) compared with high (65E%) carbohydrate diet induced a threefold greater decrease in hepatic TG content, concomitant with a greater decrease in plasma insulin concentration and fasting glucose production (44). Together these findings suggest that energy restriction, and in particular restriction of total carbohydrate availability, acutely decreases hepatic TG content independent of weight loss and that this appears to be associated with a simultaneous increase in insulin clearance. Accordingly, it was recently shown that carbohydrate restriction under eucaloric conditions reduced hepatic TG content over 2 weeks and that the change was evident within 3 days, concomitantly with lowering of fasting plasma TG and insulin concentrations in obese subjects with fatty liver disease (74). This implies that changes in absolute carbohydrate intake can impose on liver fat content and regulation of plasma insulin concentrations independent of changes in energy availability. To this end, pharmacological lowering of hepatic TG content by rosiglitazone, which acts as a ligand of PPAR γ receptors, is associated with a concomitant 20% increase in insulin clearance when administered to patients with T2D for 16 weeks, notably without weight loss (75).

A POTENTIAL ROLE OF HEPATIC GLYCOGEN OR GLUCOSE AVAILABILITY

Another potential link between carbohydrate intake and insulin clearance could be glycogen stores (or simply glucose availability) in the hepatocytes. The effect of hepatic glycogen on insulin clearance has not been directly investigated but may be of interest because acute prolonged exercise immediately increases insulin clearance in both lean and obese subjects (76), which is not related to altered exercise-induced peripheral insulin clearance (77). It could be speculated that this may relate to decreased hepatic glycogen content, as acute exercise of up to 2 h duration does not decrease hepatic TG content in healthy subjects (78).

THE ROLE OF THE FATTY ACIDS

In abdominal obesity, increased lipolytic activity of intra-abdominal adipose tissue could result in increased portal FA levels (79). It has therefore been speculated whether increased prehepatic FA availability plays a role in hepatic insulin metabolism. Associations between high FA availability and reduced insulin clearance have been suggested from lipid infusion studies in dogs (80) and *in vitro* studies in hepatocytes (81), although others have not been able to confirm this relationship in humans (47). An acute

regulation of insulin clearance by FA delivery to the liver seems unlikely, as indicated by several lines of evidence. First, fasting plasma FA concentrations are substantially decreased (–68%) during carbohydrate overfeeding (6), proving that increased FA provision to the liver is not involved in the onset of decreased insulin clearance. Similarly, short-term energy restriction by RYGB or diet increases fasting plasma FA concentrations while simultaneously increasing insulin clearance (7,38,39,41). To this end, clamp insulin clearance measured directly across the hepatic artery and vein was unaltered after 8 h infusion of a TG emulsion (82). When glucose was then coinfused with a TG emulsion for 3 h, raising plasma glucose to 11 mmol/L, insulin clearance was impaired (83).

CONCLUDING REMARKS

Insulin clearance in the liver is a dynamic process that can be modified within days under conditions of changing energy and particularly carbohydrate intake and notably before major changes in basal insulin secretion. Early increases in insulin clearance after reduced energy intake (and thus reduced carbohydrate availability) are likely to be associated with metabolic adaptations in the liver similar to those observed during fasting, *i.e.*, increased lipolysis and FA oxidation, resulting in lower TG accumulation. Interestingly, it has been shown that restriction of dietary carbohydrate rather than energy restriction *per se* initiates the metabolic response to fasting (84). Conversely, decreased insulin clearance is seen as an early response to carbohydrate overfeeding, which is potentially caused by concomitant increases in hepatic TG accumulation.

In the initial development of systemic hyperinsulinemia, reduced hepatic insulin clearance is therefore an important contributor, while insulin hypersecretion may contribute at later stages. Notably, hepatic insulin clearance is associated with hepatic rather than peripheral insulin sensitivity. In fact, it seems that the early impairments in insulin clearance precede the onset of peripheral changes in insulin action. Initially, increased systemic insulin concentrations induced by lower insulin clearance could serve to enhance peripheral glucose disposal, when hepatic substrate (carbohydrate) excess is present. This was clearly demonstrated in the dietary study where the carbohydrate-rich diet decreased hepatic insulin sensitivity and insulin clearance, concomitantly with 41% enhancement of peripheral glucose disposal (6). Also after energy restriction, a rapid improvement in hepatic insulin action and insulin clearance typically precedes the changes in peripheral insulin sensitivity, where improvements are not observed until substantial weight loss has been obtained (7,44,73). Thus, early changes in hepatic insulin clearance act to regulate systemic insulin availability and thereby affect the response of the peripheral tissues. In this context, it is of interest that 40 h of moderate hyperinsulinemia induces peripheral insulin resistance in healthy lean subjects (85) and that a low first-pass hepatic insulin extraction seems to determine peripheral insulin resistance in dogs (12).

In conclusion, hepatic insulin clearance is a major regulator of systemic insulin concentrations in the early response to altered energy and carbohydrate intake. While acknowledging the diversity of phenotypes in obesity and T2D including body composition, ethnicity, and genetics with additional contributions from a plethora of organ (e.g., β -cell) dysregulations in the development of overt T2D, our findings and the summarized evidence point to reduced hepatic insulin clearance as the initial culprit in the development of hyperinsulinemia. The causal link between hepatic carbohydrate availability, TG accumulation, and insulin clearance awaits further studies of potential mechanisms at the level of insulin receptor binding, internalization, and/or downstream intrahepatocellular signaling.

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References

- Kahn SE, Prigeon RL, McCulloch DK, et al. Quantification of the relationship between insulin sensitivity and beta-cell function in human subjects: evidence for a hyperbolic function. *Diabetes* 1993;42:1663–1672
- Le Stunff C, Bougnères P. Early changes in postprandial insulin secretion, not in insulin sensitivity, characterize juvenile obesity. *Diabetes* 1994;43:696–702
- Kim SH, Reaven GM. Insulin clearance: an underappreciated modulator of plasma insulin concentration. *J Investig Med* 2016;64:1162–1165
- Kim MK, Reaven GM, Kim SH. Dissecting the relationship between obesity and hyperinsulinemia: role of insulin secretion and insulin clearance. *Obesity (Silver Spring)* 2017;25:378–383
- Jung SH, Jung CH, Reaven GM, Kim SH. Adapting to insulin resistance in obesity: role of insulin secretion and clearance. *Diabetologia* 2018;61:681–687
- Lundsgaard AM, Sjøberg KA, Høeg LD, et al. Opposite regulation of insulin sensitivity by dietary lipid versus carbohydrate excess. *Diabetes* 2017;66:2583–2595
- Bojsen-Møller KN, Dirksen C, Jørgensen NB, et al. Early enhancements of hepatic and later of peripheral insulin sensitivity combined with increased postprandial insulin secretion contribute to improved glycemic control after Roux-en-Y gastric bypass. *Diabetes* 2014;63:1725–1737
- Bratusch-Marrain PR, Waldhäusl WK, Gasic S, Hofer A. Hepatic disposal of biosynthetic human insulin and porcine C-peptide in humans. *Metabolism* 1984;33:151–157
- Saccà L, Orofino G, Petrone A, Vigorito C. Direct assessment of splanchnic uptake and metabolic effects of human and porcine insulin. *J Clin Endocrinol Metab* 1984;59:191–196
- Ferrannini E, Wahren J, Faber OK, Felig P, Binder C, DeFronzo RA. Splanchnic and renal metabolism of insulin in human subjects: a dose-response study. *Am J Physiol* 1983;244:E517–E527
- Kryshak EJ, Butler PC, Marsh C, et al. Pattern of postprandial carbohydrate metabolism and effects of portal and peripheral insulin delivery. *Diabetes* 1990;39:142–148
- Asare-Bediako I, Paszkiewicz RL, Kim SP, et al. Variability of directly measured first-pass hepatic insulin extraction and its association with insulin sensitivity and plasma insulin. *Diabetes* 2018;67:1495–1503
- DeFronzo RA, Tobin JD, Rowe JW, Andres R. Glucose intolerance in uremia. Quantification of pancreatic beta cell sensitivity to glucose and tissue sensitivity to insulin. *J Clin Invest* 1978;62:425–435
- Kruszynska YT, Home PD, McIntyre N. Relationship between insulin sensitivity, insulin secretion and glucose tolerance in cirrhosis. *Hepatology* 1991;14:103–111
- Merli M, Leonetti F, Riggio O, et al. Glucose intolerance and insulin resistance in cirrhosis are normalized after liver transplantation [published correction appears in *Hepatology* 2000;32:446]. *Hepatology* 1999;30:649–654
- Sato H, Terasaki T, Mizuguchi H, Okumura K, Tsuji A. Receptor-recycling model of clearance and distribution of insulin in the perfused mouse liver. *Diabetologia* 1991;34:613–621
- Moller DE, Yokota A, Caro JF, Flier JS. Tissue-specific expression of two alternatively spliced insulin receptor mRNAs in man. *Mol Endocrinol* 1989;3:1263–1269
- Durham TB, Toth JL, Klimkowski VJ, et al. Dual exosite-binding inhibitors of insulin-degrading enzyme challenge its role as the primary mediator of insulin clearance in vivo. *J Biol Chem* 2015;290:20044–20059
- Antonello S, La Rocca S, Cavalcanti E, Auletta M, Salvatore F, Cacciatore L. Insulin and glucagon degradation in liver are not affected by hepatic cirrhosis. *Clin Chim Acta* 1989;183:343–350
- Højlund K, Hansen T, Lajer M, et al. A novel syndrome of autosomal-dominant hyperinsulinemic hypoglycemia linked to a mutation in the human insulin receptor gene. *Diabetes* 2004;53:1592–1598
- Pei D, Jones CN, Bhargava R, Chen YD, Reaven GM. Evaluation of octreotide to assess insulin-mediated glucose disposal by the insulin suppression test. *Diabetologia* 1994;37:843–845
- Polonsky KS, Rubenstein AH. C-peptide as a measure of the secretion and hepatic extraction of insulin. Pitfalls and limitations. *Diabetes* 1984;33:486–494
- Duckworth WC, Bennett RG, Hamel FG. Insulin degradation: progress and potential. *Endocr Rev* 1998;19:608–624
- Tillil H, Shapiro ET, Rubenstein AH, Galloway JA, Polonsky KS. Reduction of insulin clearance during hyperglycemic clamp: dose-response study in normal humans. *Diabetes* 1988;37:1351–1357
- Eaton RP, Allen RC, Schade DS. Hepatic removal of insulin in normal man: dose response to endogenous insulin secretion. *J Clin Endocrinol Metab* 1983;56:1294–1300
- Tillil H, Shapiro ET, Miller MA, et al. Dose-dependent effects of oral and intravenous glucose on insulin secretion and clearance in normal humans. *Am J Physiol* 1988;254:E349–E357
- Nauck MA, Homberger E, Siegel EG, et al. Incretin effects of increasing glucose loads in man calculated from venous insulin and C-peptide responses. *J Clin Endocrinol Metab* 1986;63:492–498
- Waldhäusl W, Bratusch-Marrain P, Gasic S, Korn A, Nowotny P. Insulin production rate following glucose ingestion estimated by splanchnic C-peptide output in normal man. *Diabetologia* 1979;17:221–227
- Meier JJ, Veldhuis JD, Butler PC. Pulsatile insulin secretion dictates systemic insulin delivery by regulating hepatic insulin extraction in humans. *Diabetes* 2005;54:1649–1656
- Acheson KJ, Schutz Y, Bessard T, Anantharaman K, Flatt JP, Jéquier E. Glycogen storage capacity and de novo lipogenesis during massive carbohydrate overfeeding in man. *Am J Clin Nutr* 1988;48:240–247
- Wadden D, Cahill F, Amini P, et al. Serum acylated ghrelin concentrations in response to short-term overfeeding in normal weight, overweight, and obese men. *PLoS One* 2012;7:e45748
- Walhin JP, Richardson JD, Betts JA, Thompson D. Exercise counteracts the effects of short-term overfeeding and reduced physical activity independent of energy imbalance in healthy young men. *J Physiol* 2013;591:6231–6243
- Clore JN, Helm ST, Blackard WG. Loss of hepatic autoregulation after carbohydrate overfeeding in normal man. *J Clin Invest* 1995;96:1967–1972
- Mott DM, Lillioja S, Bogardus C. Overnutrition induced decrease in insulin action for glucose storage: in vivo and in vitro in man. *Metabolism* 1986;35:160–165
- Brands M, Swat M, Lammers NM, et al. Effects of a hypercaloric diet on β -cell responsiveness in lean healthy men. *Clin Endocrinol (Oxf)* 2013;78:217–225

36. Erdmann J, Kallabis B, Ooppel U, Sypchenko O, Wagenpfeil S, Schusdziarra V. Development of hyperinsulinemia and insulin resistance during the early stage of weight gain. *Am J Physiol Endocrinol Metab* 2008;294:E568–E575
37. Svane MS, Toft-Nielsen MB, Kristiansen VB, et al. Nutrient re-routing and altered gut-islet cell crosstalk may explain early relief of severe postprandial hypoglycaemia after reversal of Roux-en-Y gastric bypass. *Diabet Med* 2017;34:1783–1787
38. Lima MM, Pareja JC, Alegre SM, et al. Acute effect of Roux-en-Y gastric bypass on whole-body insulin sensitivity: a study with the euglycemic-hyperinsulinemic clamp. *J Clin Endocrinol Metab* 2010;95:3871–3875
39. Campos GM, Rabl C, Havel PJ, et al. Changes in post-prandial glucose and pancreatic hormones, and steady-state insulin and free fatty acids after gastric bypass surgery. *Surg Obes Relat Dis* 2014;10:1–8
40. Bunt JC, Blackstone R, Thearle MS, Vinales KL, Votruba S, Krakoff J. Changes in glycemia, insulin and gut hormone responses to a slowly ingested solid low-carbohydrate mixed meal after laparoscopic gastric bypass or band surgery. *Int J Obes* 2017;41:706–713
41. Jackness C, Karmally W, Febres G, et al. Very low-calorie diet mimics the early beneficial effect of Roux-en-Y gastric bypass on insulin sensitivity and β -cell function in type 2 diabetic patients. *Diabetes* 2013;62:3027–3032
42. Besic V, Shi H, Stubbs RS, Hayes MT. Aberrant liver insulin receptor isoform a expression normalises with remission of type 2 diabetes after gastric bypass surgery. *PLoS One* 2015;10:e0119270
43. Bosello O, Zamboni M, Armellini F, et al. Modifications of abdominal fat and hepatic insulin clearance during severe caloric restriction. *Ann Nutr Metab* 1990;34:359–365
44. Kirk E, Reeds DN, Finck BN, Mayurranjan SM, Patterson BW, Klein S. Dietary fat and carbohydrates differentially alter insulin sensitivity during caloric restriction [published correction appears in *Gastroenterology* 2009;137:393]. *Gastroenterology* 2009;136:1552–1560
45. Nielsen MS, Christensen BJ, Ritz C, et al. Roux-en-Y Gastric bypass and sleeve gastrectomy does not affect food preferences when assessed by an ad libitum buffet meal. *Obes Surg* 2017;27:2599–2605
46. Kaga H, Tamura Y, Takeno K, et al. Correlates of insulin clearance in apparently healthy non-obese Japanese men. *Sci Rep* 2017;7:1462
47. Kotronen A, Vehkavaara S, Seppälä-Lindroos A, Bergholm R, Yki-Järvinen H. Effect of liver fat on insulin clearance. *Am J Physiol Endocrinol Metab* 2007;293:E1709–E1715
48. Strang BD, Bertics SJ, Grummer RR, Armentano LE. Relationship of triglyceride accumulation to insulin clearance and hormonal responsiveness in bovine hepatocytes. *J Dairy Sci* 1998;81:740–747
49. Kotronen A, Westerbacka J, Bergholm R, Pietiläinen KH, Yki-Järvinen H. Liver fat in the metabolic syndrome. *J Clin Endocrinol Metab* 2007;92:3490–3497
50. Fabbri E, Magkos F, Mohammed BS, et al. Intrahepatic fat, not visceral fat, is linked with metabolic complications of obesity. *Proc Natl Acad Sci U S A* 2009;106:15430–15435
51. Marini MA, Frontoni S, Succurro E, et al. Differences in insulin clearance between metabolically healthy and unhealthy obese subjects. *Acta Diabetol* 2014;51:257–261
52. Amato MC, Vesco R, Vigneri E, Ciresi A, Giordano C. Hyperinsulinism and polycystic ovary syndrome (PCOS): role of insulin clearance. *J Endocrinol Invest* 2015;38:1319–1326
53. Svendsen PF, Jensen FK, Holst JJ, Haugaard SB, Nilas L, Madsbad S. The effect of a very low calorie diet on insulin sensitivity, beta cell function, insulin clearance, incretin hormone secretion, androgen levels and body composition in obese young women. *Scand J Clin Lab Invest* 2012;72:410–419
54. Peiris AN, Mueller RA, Smith GA, Struve MF, Kissebah AH. Splanchnic insulin metabolism in obesity. Influence of body fat distribution. *J Clin Invest* 1986;78:1648–1657
55. Polonsky KS, Given BD, Hirsch L, et al. Quantitative study of insulin secretion and clearance in normal and obese subjects. *J Clin Invest* 1988;81:435–441
56. Piccinini F, Polidori DC, Gower BA, Bergman RN. Hepatic but not extrahepatic insulin clearance is lower in African American than in European American women. *Diabetes* 2017;66:2564–2570
57. Guo X, Cui J, Jones MR, et al. Insulin clearance: confirmation as a highly heritable trait, and genome-wide linkage analysis. *Diabetologia* 2012;55:2183–2192
58. Schwarz JM, Neese RA, Turner S, Dare D, Hellerstein MK. Short-term alterations in carbohydrate energy intake in humans. Striking effects on hepatic glucose production, de novo lipogenesis, lipolysis, and whole-body fuel selection. *J Clin Invest* 1995;96:2735–2743
59. Lundsgaard AM, Fritzen AM, Sjøberg KA, et al. Circulating FGF21 in humans is potently induced by short term overfeeding of carbohydrates. *Mol Metab* 2016;6:22–29
60. Hudgins LC, Hellerstein M, Seidman C, Neese R, Diakun J, Hirsch J. Human fatty acid synthesis is stimulated by a eucaloric low fat, high carbohydrate diet. *J Clin Invest* 1996;97:2081–2091
61. Mittendorfer B, Yoshino M, Patterson BW, Klein S. VLDL triglyceride kinetics in lean, overweight, and obese men and women. *J Clin Endocrinol Metab* 2016;101:4151–4160
62. Formisano P, Najjar SM, Gross CN, et al. Receptor-mediated internalization of insulin. Potential role of pp120/HA4, a substrate of the insulin receptor kinase. *J Biol Chem* 1995;270:24073–24077
63. Najjar SM, Philippe N, Suzuki Y, et al. Insulin-stimulated phosphorylation of recombinant pp120/HA4, an endogenous substrate of the insulin receptor tyrosine kinase. *Biochemistry* 1995;34:9341–9349
64. Choice CV, Howard MJ, Poy MN, Hankin MH, Najjar SM. Insulin stimulates pp120 endocytosis in cells co-expressing insulin receptors. *J Biol Chem* 1998;273:22194–22200
65. Poy MN, Yang Y, Rezaei K, et al. CEACAM1 regulates insulin clearance in liver. *Nat Genet* 2002;30:270–276
66. Russo L, Muturi HT, Ghadieh HE, et al. Liver-specific reconstitution of CEACAM1 reverses the metabolic abnormalities caused by its global deletion in male mice. *Diabetologia* 2017;60:2463–2474
67. Najjar SM, Yang Y, Fernström MA, et al. Insulin acutely decreases hepatic fatty acid synthase activity. *Cell Metab* 2005;2:43–53
68. Chakravarthy MV, Pan Z, Zhu Y, et al. “New” hepatic fat activates PPAR α to maintain glucose, lipid, and cholesterol homeostasis. *Cell Metab* 2005;1:309–322
69. Ramakrishnan SK, Khuder SS, Al-Share QY, et al. PPAR α (peroxisome proliferator-activated receptor α) activation reduces hepatic CEACAM1 protein expression to regulate fatty acid oxidation during fasting-refeeding transition. *J Biol Chem* 2016;291:8121–8129
70. Heinrich G, Muturi HT, Rezaei K, et al. Reduced hepatic carcinoembryonic antigen-related cell adhesion molecule 1 level in obesity. *Front Endocrinol (Lausanne)* 2017;8:54
71. Lee W. The CEACAM1 expression is decreased in the liver of severely obese patients with or without diabetes. *Diagn Pathol* 2011;6:40
72. Steven S, Hollingsworth KG, Small PK, et al. Calorie restriction and not glucagon-like peptide-1 explains the acute improvement in glucose control after gastric bypass in type 2 diabetes. *Diabet Med* 2016;33:1723–1731
73. Lim EL, Hollingsworth KG, Aribisala BS, Chen MJ, Mathers JC, Taylor R. Reversal of type 2 diabetes: normalisation of beta cell function in association with decreased pancreas and liver triacylglycerol. *Diabetologia* 2011;54:2506–2514
74. Mardinoglu A, Wu H, Bjornson E, et al. An integrated understanding of the rapid metabolic benefits of a carbohydrate-restricted diet on hepatic steatosis in humans. *Cell Metab* 2018;27:559–571.e5
75. Tiikkainen M, Häkkinen AM, Korshennikova E, Nyman T, Mäkimattila S, Yki-Järvinen H. Effects of rosiglitazone and metformin on liver fat content, hepatic insulin resistance, insulin clearance, and gene expression in adipose tissue in patients with type 2 diabetes. *Diabetes* 2004;53:2169–2176

76. Tuominen JA, Ebeling P, Koivisto VA. Exercise increases insulin clearance in healthy man and insulin-dependent diabetes mellitus patients. *Clin Physiol* 1997; 17:19–30
77. Richter EA, Mikines KJ, Galbo H, Kiens B. Effect of exercise on insulin action in human skeletal muscle. *J Appl Physiol* 1989;66:876–885
78. Bilet L, Brouwers B, van Ewijk PA, et al. Acute exercise does not decrease liver fat in men with overweight or NAFLD. *Sci Rep* 2015;5: 9709
79. Tchernof A, Després JP. Pathophysiology of human visceral obesity: an update. *Physiol Rev* 2013;93:359–404
80. Wiesenthal SR, Sandhu H, McCall RH, et al. Free fatty acids impair hepatic insulin extraction in vivo. *Diabetes* 1999;48:766–774
81. Svedberg J, Björntorp P, Smith U, Lönnroth P. Free-fatty acid inhibition of insulin binding, degradation, and action in isolated rat hepatocytes. *Diabetes* 1990; 39:570–574
82. Shah P, Vella A, Basu A, et al. Effects of free fatty acids and glycerol on splanchnic glucose metabolism and insulin extraction in nondiabetic humans. *Diabetes* 2002;51:301–310
83. Hennes MM, Dua A, Kissebah AH. Effects of free fatty acids and glucose on splanchnic insulin dynamics. *Diabetes* 1997;46:57–62
84. Klein S, Wolfe RR. Carbohydrate restriction regulates the adaptive response to fasting. *Am J Physiol* 1992;262:E631–E636
85. Rizza RA, Mandarino LJ, Genest J, Baker BA, Gerich JE. Production of insulin resistance by hyperinsulinaemia in man. *Diabetologia* 1985;28:70–75