



The rs7903146 Variant in the *TCF7L2* Gene Increases the Risk of Prediabetes/Type 2 Diabetes in Obese Adolescents by Impairing β -Cell Function and Hepatic Insulin Sensitivity

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OBJECTIVE

In this study, we aimed to explore the mechanism by which *TCF7L2* rs7903146 risk allele confers susceptibility to impaired glucose tolerance (IGT) or type 2 diabetes (T2D) in obese adolescents.

RESEARCH DESIGN AND METHODS

The rs7903146 variant in the *TCF7L2* gene was genotyped in a multiethnic cohort of 955 youths. All subjects underwent an oral glucose tolerance test with the use of the Oral Minimal Model to assess insulin secretion, and 33 subjects underwent a hyperinsulinemic-euglycemic clamp. In 307 subjects, a follow-up oral glucose tolerance test was repeated after 3.11 ± 2.36 years.

RESULTS

The *TCF7L2* rs7903146 risk allele was associated with higher 2-h glucose levels in Caucasians ($P = 0.006$) and African Americans ($P = 0.009$), and a trend was seen also in Hispanics ($P = 0.072$). Also, the T allele was associated with decreased β -cell responsiveness and IGT ($P < 0.05$). Suppression of endogenous hepatic glucose production was lower in subjects with the risk variant ($P = 0.006$). Finally, the odds of showing IGT/T2D at follow-up were higher in subjects carrying the minor allele (odds ratio 2.224; 95% CI 1.370–3.612; $P = 0.0012$).

CONCLUSIONS

The rs7903146 variant in the *TCF7L2* gene increases the risk of IGT/T2D in obese adolescents by impairing β -cell function, and hepatic insulin sensitivity predicts the development of IGT/T2D over time.

The landscape of childhood diabetes is changing, as an unprecedented phenomenon of type 2 diabetes (T2D) rarely seen in pediatrics before is unfolding. Looking ahead, projected incidence rates of T2D in youth are expected to triple over the next 40 years in the U.S. (1,2). Even more concerning is the ever-increasing number of those with impaired glucose tolerance (IGT) among obese adolescents (3,4). Youth-onset T2D heralds many years of disease with an increased risk of both micro- and macrovascular complications that will occur when affected individuals are still young (5–8). Therefore, it is imperative

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to identify adolescents at increased risk for T2D when they are in the prediabetic state, particularly because the tempo of progression from IGT to full-blown diabetes seems to be faster in youth than in adults (9,10).

β -Cell dysfunction and hepatic insulin resistance are key defects contributing to the development of T2D. Therefore, understanding the mechanisms associated with the decline of β -cell function and changes in hepatic insulin sensitivity (SI) is essential in combating the disease, particularly at a younger age. There is substantial evidence in adults for a relevant genetic determination of β -cell failure (11–17). Indeed, the discovery in 2006 by Grant et al. (18) that a common single nucleotide polymorphism (SNP) in the *TCF7L2* gene region was associated with T2D in an Icelandic case-control study has launched a new direction in diabetes research. Soon after, these results were replicated in other ethnic groups (19,20), with few cross-sectional studies in children (21,22). Notably, the T risk allele of the rs7903146 confers the strongest risk of T2D known to date in Caucasians (16,17,23). However, the molecular mechanisms by which variants in *TCF7L2* increase the risk of T2D remain unclear. *TCF7L2* encodes a transcription factor that is a member of the Wnt signaling pathway known to be active not only in the β -cells but also in other cell lineages and glucose-metabolizing tissues, including the liver (24–27). Studies in adults showed that the variant impairs β -cell function and had elevated rates of hepatic glucose production (HGP) (23,28,29). It should be noted, however, that most of the evidence linking *TCF7L2* variants to diabetes-related phenotypes in humans has been obtained in adults (23,29,30). There are no studies in adolescents that have yet combined detailed genetic/metabolic studies to help define the mechanisms by which genetic variations in *TCF7L2* may increase the risk of IGT/T2D. The pathogenesis of β -cell failure and its genetic underpinnings in youth remain elusive. Results from metabolic studies performed in adults cannot be extrapolated to pediatrics, as they are influenced greatly by the aging effects per se on both insulin secretion and sensitivity. Further, the advantage of studying pediatric subjects is that they lack the potential confounding effects of long-term obesity-associated chronic complications. There

is thus a compelling rationale to conduct studies focused on these unique age groups to understand disease etiology to better tailor prevention and treatment approaches to these groups. In this study, in a large multiethnic cohort of obese adolescents, we tested the hypothesis that the risk allele at rs7903146 locus favors the maintenance of IGT or its progression to T2D in youth because of 1) reduced β -cell function; 2) altered efficiency of proinsulin processing; and 3) increased hepatic insulin resistance during adolescence, a sensitive/critical time for youth-onset T2D.

RESEARCH DESIGN AND METHODS

Study Cohort

We recruited 955 obese children and adolescents (394 boys and 561 girls; mean \pm SD age 13.3 \pm 3.4 years; and mean \pm SD z score BMI 2.15 \pm 0.70) from a multiethnic cohort participating in the Yale Pathogenesis of Youth Onset Type 2 Diabetes (PYOD) study (NCT01967849), a long-term project aimed at studying early alterations in β -cell function and SI in obese youths. The subjects were not taking any medications that affect glucose metabolism when enrolled. All of them underwent a 3-h oral glucose tolerance test (OGTT), and 705 (74%) subjects showed normal glucose tolerance (NGT) and 250 (26%) IGT.

To evaluate the effect of the genotype on the risk of developing IGT, or progressing to T2D over time, 307 subjects underwent a second OGTT after a follow-up of 3.11 \pm 2.36 years. During the follow-up, all participants received standard nutritional guidance and recommendations for physical activity and were not on any medications affecting glucose metabolism. The longitudinal cohort was constituted of 114 Caucasians, 90 African Americans, and 103 Hispanics (113 boys and 194 girls; mean \pm SD age 12.4 \pm 3.0 years; and mean \pm SD z score BMI 2.28 \pm 0.55). Of them, at baseline, 202 (66%) were NGT and 105 (34%) IGT. The time interval for the follow-up was based on our previous study suggesting that changes in categories of glucose tolerance in obese adolescents are likely to occur over a relatively short period of time (10,31).

OGTT

A 3-h OGTT (1.75 g/kg body weight, up to 75 g) was performed in all 955 patients, as previously reported (4,10,32).

Two baseline samples were obtained at -15 and 0 min for measurements of plasma glucose, insulin, and C-peptide. Subjects were studied at the Hospital Research Unit of the Yale New Haven Hospital at 8 A.M. after a 12-h overnight fast. Thereafter, flavored glucose (Orangedex; Custom Laboratories, Baltimore, MD) was given orally, and blood samples were obtained at 10, 20, and 30 min and every 30 min thereafter for 180 min for the measurements of plasma glucose, insulin, and C-peptide. Proinsulin was available in 82 subjects with NGT and 38 with IGT. Glucose tolerance status was defined according to the American Diabetes Association guidelines.

Assessment of Insulin Secretion and Action: Oral Minimal Model Method

Out of the 955 patients, the data usable to perform the oral minimal model analysis were available for 664 subjects (290 Caucasians, 176 African Americans, and 198 Hispanics). The SI was estimated from plasma glucose and insulin concentrations measured during the 3-h OGTT using the Oral Glucose Minimal Model (31,33–36). β -Cell responsiveness was estimated from the C-peptide data measured during the 3-h OGTT using the Oral C-peptide Minimal Model (31,33–36). The model assumes that insulin secretion is made up of two components. The dynamic component is likely to represent secretion of promptly releasable insulin and is proportional to the rate of increase of glucose concentration through a parameter, Φ_d (10^{-9}), which defines the dynamic responsiveness index. The static component derives from provision of new insulin to the releasable pool and is characterized by a static index, Φ_s (10^{-9} min^{-1}), and by a delay time constant, T (min). From Φ_d and Φ_s , one can also calculate a single, overall β -cell responsiveness index, Φ_{total} (10^{-9} min^{-1}). Finally, a basal β -cell responsiveness index, Φ_b (10^{-9} min^{-1}) can also be calculated from basal C-peptide and glucose concentrations. To determine whether β -cell function is appropriate for the degree of insulin resistance, Φ_{total} can be expressed in relation to SI through the total disposition index (DI) = $\Phi_{\text{total}} \cdot S_I$.

Direct Measurement of Hepatic and Peripheral SI

The hyperinsulinemic-euglycemic clamp was performed in a subgroup of 33

subjects (20 male and 13 female; 9 CC, 19 CT, and 5 TT genotypes; mean \pm SD age 15.02 ± 2.77 years; and mean \pm SD BMI z score 2.19 ± 0.54). Twenty were NGT, and 13 were IGT. This subgroup did not differ from the main cohort for age, sex, race, BMI z score, glucose tolerance, hepatic fat fraction, and body fat.

Two intravenous catheters, one for blood sampling and one for infusion of glucose, insulin, and stable isotopes, were inserted in the antecubital vein of each arm after local lidocaine infiltration. The sampling arm was kept in a heated box for arterialization of blood. Hepatic and peripheral SI were measured by a hyperinsulinemic-euglycemic clamp by infusing insulin as a primed continuous infusion at a low insulin dose of $8 \text{ mU} \cdot \text{m}^{-2} \cdot \text{min}^{-1}$ and a high insulin dose of $80 \text{ mU} \cdot \text{m}^{-2} \cdot \text{min}^{-1}$, each step lasting 120 min. A primed continuous infusion of 6.6-deuterium-labeled glucose was used to quantify insulin's effects on glucose turnover (37). To maintain the plasma enrichment of ^2D -glucose constant at baseline value throughout the clamp, we used the hot glucose infusion method (37). Arterialized blood samples were collected every 10 min during the last 30 min of the baseline period and end of each step of the clamp for measurement of glucose enrichments, hormones, and substrates.

The glucose infusion rates were calculated during the last 30 min of each step of the clamp and expressed as milligrams of glucose per minute per meter squared. Endogenous HGP baseline and during the insulin clamp, along with the clamped glucose disposal rates, were calculated as previously reported (37).

Genotyping

Genomic DNA was extracted from peripheral blood leukocytes. Genotyping was performed with the use of a matrix-assisted-based laser desorption-ionization time-of-flight mass spectrometry on the MassARRAY platform (Sequenom) through the Yale Center for Genome Analysis (38).

Biochemical Analyses

Plasma glucose was determined using the YSI 2700 Analyzer (YSI, Yellow Springs, OH). Plasma insulin was measured by the Linco radioimmunoassay (Linco Research, Inc., St. Charles, MO), which has <1% cross-reactivity with C-peptide

and proinsulin. Plasma C-peptide was assayed with an assay made by Diagnostic Products (Los Angeles, CA). Plasma proinsulin was measured using a radioimmunoassay (Linco Research, Inc.).

Statistical Analyses

Distribution of continuous variables was examined for skewness and when appropriate data were log-transformed to approximate univariate normality before association analyses by inverse normal scores. A χ^2 test was used to assess the Hardy-Weinberg equilibrium for each of the studied SNPs and to compare proportions.

The primary outcome of the study was IGT. At baseline, the odds ratio (OR) of showing prediabetes according to the genotypes was evaluated by a logistic regression analysis, and age, sex, and z score BMI were used as covariates. The risk of subjects with NGT to progress to prediabetes or T2D at follow-up was calculated by running a logistic regression analysis and including as covariates age, sex, ethnicity, z score BMI at baseline, delta z score BMI, and follow-up time. The association between the genotypes and quantitative traits was evaluated by a regression model after coding the genotype with an additive model of inheritance (i.e., the genotype is coded with 0, 1, or 2 corresponding to the number of minor alleles carried by each individual), and age, sex, z score BMI, and glucose tolerance status were used as covariates when appropriate. Statistical analyses were performed with SPSS (19.0 for Windows; SPSS Inc., Chicago, IL).

RESULTS

Clinical Phenotypes of the Subjects According to *TCF7L2* rs7903146 Genotype and Ethnicity

The frequency of the *TCF7L2* rs7903146 minor allele (T) was 0.336 in Caucasians, 0.286 in African Americans, and 0.286 in Hispanics. The allele frequencies were comparable with those demonstrated in similar ethnic groups according to Ensembl (<http://useast.ensembl.org>). The genotype distribution within each ethnic group was in Hardy-Weinberg equilibrium ($P > 0.05$). The clinical and anthropometric characteristics of the study population according to the *TCF7L2* rs7903146 genotype are shown in Table 1. Age, sex distribution, and BMI z score were similar by *TCF7L2* rs7903146 genotype and by ethnicity. Fasting glucose

was significantly higher only in Caucasians carriers of the minor risk allele showed significantly higher 2-h glucose levels (Table 1). Indeed, the rs7903146 risk allele was associated with IGT in the three ethnic groups, in Caucasians (OR 1.573; 95% CI 1.100–2.250; $P = 0.0131$), African Americans (OR 2.011; 95% CI 1.265–3.196; $P = 0.003$), and Hispanics (OR 1.897; 95% CI 1.204–2.989; $P = 0.006$).

The rs7903146 Variant in the *TCF7L2* Gene Is Associated With Decreased β -Cell Responsivity (Φ_{total}) and a Low DI in Obese Adolescents

The evaluation of insulin secretion using the Oral Minimal Model (OMM) showed that subjects carrying the risk allele have a decreased β -cell responsivity (Φ_{total}) (in Caucasians, $P = 0.003$; in African Americans, $P = 0.002$; and in Hispanics, $P = 0.035$) (Table 1). Peripheral SI was, however, not affected significantly. To better express β -cell function in the context of ambient peripheral whole-body SI, we used the DI, which was significantly reduced in Caucasians ($P = 0.029$), African Americans ($P = 0.011$), and Hispanics ($P < 0.0001$), carriers of the risk genotype in *TCF7L2* (Fig. 1).

The rs7903146 Variant in the *TCF7L2* Gene Is Associated With Impaired Proinsulin Processing in Obese Adolescents

Out of the 955 study subjects, in a subgroup of 208 subjects (mean \pm SD age 13.2 ± 2.5 years; and mean \pm SD BMI 33.4 ± 2.8), the proinsulin levels were measured to determine whether the rs7903146 variant in the *TCF7L2* gene might affect the efficiency of proinsulin processing. Of them, 100 had NGT (45 CC, 45 CT, and 10 TT) and 108 IGT (40 CC, 40 CT, and 28 TT). The proinsulin to C-peptide ratio was calculated from the fasting proinsulin and C-peptide levels. The rs7903146 variant was associated with a higher proinsulin to C-peptide ratio than heterozygous in both the NGT ($P = 0.01$) and IGT groups ($P = 0.04$) (Fig. 2), even after adjusting for the effect of obesity and insulin resistance.

The rs7903146 Variant Is Associated With Hepatic but Not Peripheral Insulin Resistance

We explored the effect of the risk genotype of *TCF7L2* on hepatic SI during a hyperinsulinemic-euglycemic clamp in 33 obese adolescents with similar BMI,

Table 1—Clinical features of the study population according to the TCF7L2 rs7903146 genotype

	Caucasians				African Americans				Hispanics			
	CC	CT	TT	P value	CC	CT	TT	P value	CC	CT	TT	P value
N	174	187	42		140	107	24		141	119	21	
Age (years)	14.2 ± 3.9	13.6 ± 3.3	13.2 ± 2.9	0.118	13.3 ± 3.4	13.3 ± 3.2	14.3 ± 4.3	0.367	12.3 ± 3.1	13.1 ± 3.1	11.8 ± 3.1	0.186
Sex (male/female), %	32/68	44/55	43/57	0.041	42/58	38/62	38/62	0.800	48/52	39/61	62/38	0.085
BMI z score	2.0 ± 0.81	2.2 ± 0.58	2.1 ± 0.64	0.055	2.3 ± 0.51	2.2 ± 0.71	2.1 ± 0.55	0.297	2.1 ± 0.76	2.2 ± 0.65	1.9 ± 1.4	0.374
Tanner 1/2–3/4–5, %	7/25/68	5/37/58	12/26/62	0.061	13/36/51	7/43/50	4/33/63	0.308	11/45/44	8/38/54	14/48/38	0.429
Fasting glucose (mg/dL)	90.2 ± 8.3	91.2 ± 8.3	93.9 ± 10.3	0.036	91.5 ± 8.6	92.1 ± 8.6	93.3 ± 7.1	0.466	91.6 ± 7.7	92.7 ± 8.6	96.6 ± 12.0	0.082
2-h glucose (mg/dL)	118.4 ± 26.3	123.2 ± 27.4	137.1 ± 34.9	0.006	117.7 ± 28.5	129.5 ± 31.5	122.0 ± 26.4	0.009	117.8 ± 20.7	124.3 ± 24.4	129.0 ± 31.0	0.072
HbA _{1c} (%)	5.4 ± 0.30	5.4 ± 0.30	5.5 ± 0.34	0.057	5.5 ± 0.35	5.6 ± 0.37	5.7 ± 0.45	0.094	5.5 ± 0.27	5.5 ± 0.36	5.5 ± 0.37	0.990
HbA _{1c} (mmol/mol)	36 ± 2.3	36 ± 2.3	37 ± 2.39	0.057	37 ± 2.4	38 ± 2.4	39 ± 2.9	0.094	37 ± 1.98	37 ± 2.3	37 ± 2.4	0.990
Insulin (μU/mL)	30.3 ± 17.2	34.2 ± 19.7	33.2 ± 15.8	0.225	34.5 ± 18.2	37.4 ± 29.4	34.4 ± 13.7	0.320	30.3 ± 17.2	35.7 ± 18.0	42.7 ± 32.0	0.038
WBSI (l ² /mg/μU)	2.1 ± 1.3	1.9 ± 1.1	1.8 ± 1.2	0.267	1.8 ± 1.2	2.0 ± 1.6	1.6 ± 0.61	0.322	2.0 ± 1.1	1.6 ± 0.94	2.1 ± 1.7	0.062
IGI (μU/mg)	4.2 ± 4.0	4.2 ± 3.3	3.8 ± 3.4	0.769	6.2 ± 4.8	6.6 ± 9.7	6.4 ± 5.0	0.072	4.6 ± 3.8	4.6 ± 3.0	3.7 ± 3.5	0.266
DI (l ² /mg ²)	6.9 ± 6.8	6.0 ± 4.3	5.5 ± 3.0	0.090	9.2 ± 5.7	7.9 ± 7.8	8.6 ± 6.0	0.010	7.6 ± 5.9	6.6 ± 4.7	6.0 ± 5.9	0.051
OMM												
N	138	127	25		89	71	16		110	73	15	
Φ _b (10 ⁻⁹ min ⁻¹)	14.7 ± 6.3	14.9 ± 6.1	13.3 ± 7.2	0.490	13.8 ± 5.5	12.6 ± 6.7	12.2 ± 5.5	0.347	13.7 ± 4.5	15.7 ± 6.5	12.1 ± 6.0	0.839
Φ _d (10 ⁻⁹ min ⁻¹)	1,395.9 ± 765.6	1,431.5 ± 929.7	1,386.7 ± 996.2	0.880	1,963.7 ± 1,064.0	1,788.8 ± 1,270.1	1,431.6 ± 926.5	0.127	1,649.0 ± 1,007.9	1,679.4 ± 907.9	1,209.2 ± 525.6	0.626
Φ _s (10 ⁻⁹ min ⁻¹)	81.6 ± 49.8	76.4 ± 43.0	52.2 ± 23.0	0.010	87.4 ± 63.2	62.0 ± 36.3	75.1 ± 33.2	0.039	85.7 ± 62.7	78.2 ± 34.3	50.8 ± 19.8	0.858
Φ _{total} (10 ⁻⁹ min ⁻¹)	96.4 ± 55.0	97.1 ± 54.7	64.2 ± 27.8	0.003	111.347 ± 66.2	80.2 ± 40.8	88.1 ± 44.0	0.002	106.8 ± 70.9	92.8 ± 42.5	66.8 ± 26.2	0.035
SI (10 ⁻⁵ dl · kg ⁻¹ · min ⁻¹ /pmol · l ⁻¹)	26.1 ± 23.4	25.0 ± 20.2	30.6 ± 27.7	0.841	24.9 ± 18.4	26.9 ± 21.8	20.4 ± 15.1	0.742	27.1 ± 18.5	17.0 ± 12.1	24.3 ± 15.0	0.003

Data are mean ± SD unless otherwise indicated. IGI, insulinogenic index; WBSI, whole-body insulin sensitivity index.

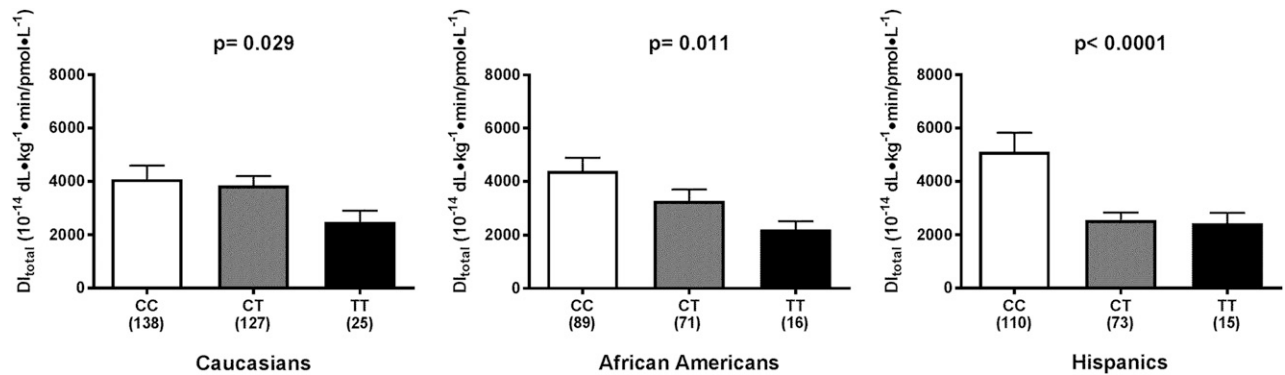


Figure 1—Association between the *TCF7L2* rs7903146 gene and the total DI as calculated by the OMM. The results showed that the variant is associated with a lower total DI in Caucasians, African Americans, and Hispanics.

fasting insulin, and glucagon (Supplementary Table 1). Of note, despite similar clamp insulin steady state in each genotype group and during each step of the clamp, homozygous carriers of the TT risk allele showed a significantly reduced suppression of HGP compared with those with the CC genotype, particularly during the high insulin dose of $80 \text{ mU/m}^2 \cdot \text{min}$, suggesting the presence of hepatic insulin resistance (Fig. 3).

The rs7903146 Variant in the *TCF7L2* Gene Predisposes to Transition From IGT to T2D in Obese Adolescents:

Lessons From the Longitudinal Cohort
To assess the effect of the rs7903146 variant on changes in glucose tolerance 307 subjects underwent a second OGTT after 3.11 ± 2.36 years. Two hundred two subjects (66%) were NGT at baseline (112 CC, 75 CT, and 15 TT), whereas 105 subjects (34%) were IGT at baseline (39 CC, 56 CT, and 10 TT). Compared with NGT, subjects with the diagnosis of IGT tended to show a shorter follow-up time (3.27 ± 2.43 vs. 2.28 ± 2.20 ; $P = 0.084$), but the years of follow-up did not differ

across genotypes neither in the baseline NGT nor in the baseline IGT group (all $P > 0.10$).

In the whole population, the odds of showing IGT/T2D at follow-up were higher in subjects carrying the minor allele (OR 2.224; 95% CI 1.370–3.612; $P = 0.0012$). Analysis using Kaplan-Meier curves in the whole population indicated that the carriers of rs7903146 CT/TT genotypes had greater risk of IGT/T2D than CC genotype carriers (Supplementary Fig. 1). The log-rank test comparing the plots of the complement of the Kaplan-Meier estimates between risk allele carriers (CT or TT) and noncarriers (CC) showed that this relationship was statistically significant ($P = 0.001$) (Supplementary Fig. 1).

To assess the odds of showing IGT/T2D at follow-up according to the glucose tolerance status at baseline, the study population was divided in two groups of glucose tolerance: NGT and IGT/T2D. In the group of subjects with NGT carrying the minor allele showed a higher risk to develop IGT/T2D as compared with the group of individuals carrying the major

allele (OR 2.648; 95% CI 1.326–5.291; $P = 0.005$) independent of age, sex, BMI, changes in BMI, follow-up time, 2-h glucose at baseline, and ethnicity. In subjects with IGT at baseline, the rs7903146 minor allele was associated with a higher risk of remaining IGT or transitioning to T2D at follow-up (OR 2.221; 95% CI 1.046–4.718; $P = 0.0379$) independent of age, sex, BMI, changes in BMI, follow-up time, 2-h glucose at baseline, and ethnicity.

CONCLUSIONS

In the current study, we combined detailed genetic/metabolic studies to help define how genetic variations in the *TCF7L2* gene may increase the risk of T2D, particularly during the critical developmental stage of adolescence. Remarkably, we found in this very young cohort that each copy of the T allele (rs7903146) is associated with almost twofold increased odds for IGT ($P = 0.0001$). Additionally, our longitudinal follow-up data from the cohort suggest that the *TCF7L2* risk genotype is associated with a high OR of maintaining IGT or progressing to T2D (OR 2.419; 95% CI 1.291–4.532; $P = 0.006$). To unravel the mechanisms that might explain the genotype/phenotype correlations, we used the OMM in a large multiethnic cohort of youths and assessed the proinsulin processing by measuring the circulating fasting proinsulin/C-peptide ratio and the clamp techniques coupled with tracer methodologies (in a subset of adolescents) to more accurately assess hepatic and peripheral SI. We found that the T allele of *TCF7L2* rs7903146 has profound effects on β -cell function as reflected by a reduced DI and an altered proinsulin secretory efficiency. Moreover, its effects seem

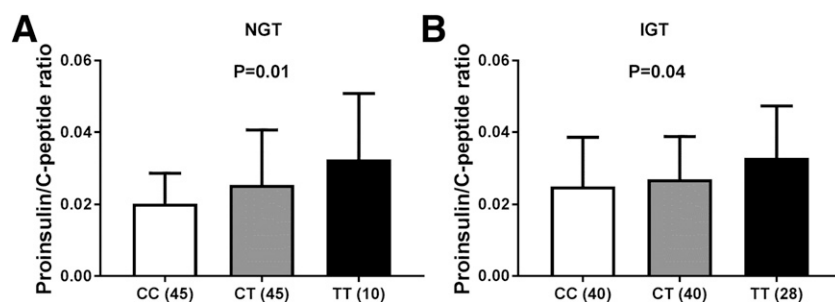


Figure 2—Association between the *TCF7L2* rs7903146 gene and proinsulin to C-peptide ratio. The variant is associated with a higher proinsulin to C-peptide ratio, which demonstrates that the variant impairs proinsulin processing.

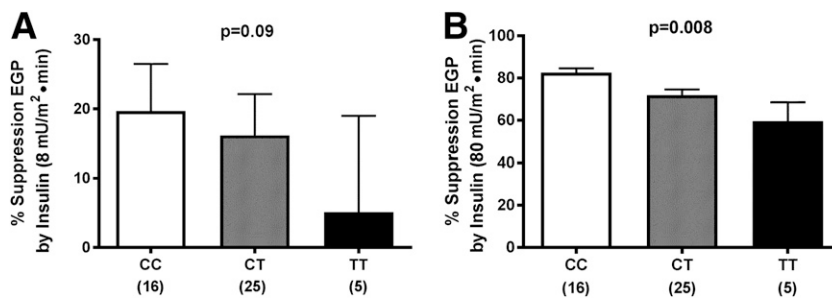


Figure 3—Association between the *TCF7L2* rs7903146 gene and percent suppression of endogenous glucose production (EGP) by low-dose (8 mU/m² · min) (A) and high-dose (80 mU/m² · min) (B) insulin infusion during a hyperinsulinemic-euglycemic clamp.

to extend to the liver by reducing insulin ability to suppress hepatic endogenous glucose production. These effects seem to occur in parallel, thus playing a critical role in two of the major defects that are key to the development of hyperglycemia.

The pathogenesis of the rapid β -cell failure and its genetic underpinnings in youth-onset IGT/T2D is poorly understood. Our study provides for the first time the evidence that the rs7903146 variant in the *TCF7L2* affects functional β -cell capacity early in the course of T2D development in obese adolescents. Longitudinal follow-up of these obese adolescents suggest the mechanisms by which homozygosity for the high-risk allele at rs7903146 favors the maintenance of IGT or progression from IGT to T2D in youth.

TCF7L2, the gene with the largest effect on disease susceptibility discovered to date, was identified pre-genome-wide association by Grant et al. (18) in 2006, with rapid replication of its consequence on diabetes susceptibility in multiple populations (19,20). In the Diabetes Prevention Program (DPP) study (39), participants who were homozygous for the T allele at rs7903146 were more likely to have progression to diabetes than were those who were homozygous for the C allele (hazard ratio 1.55; 95% CI 1.20–2.01; $P < 0.001$). The genotypic relative risk reported in this study in our obese adolescents might differ slightly from the OR documented by Grant et al. (18) and Florez et al. (39), owing to the younger age of our cohort. Our study is consistent with data reported in a population of young adults without diabetes participating in the Young Finns Study (40), indicating that the *TCF7L2* variant predicts the incidence of impaired fasting glucose.

Despite the evidence provided in this study in a very young cohort and by previous studies in adults of an effect of the rs7903146 variant on insulin secretion, resolving the functional mechanisms underlying these associations has been, so far, extremely challenging. The molecular mechanisms by which variants in *TCF7L2* increase the risk of T2D have not been fully elucidated. *TCF7L2* encodes a transcription factor that is a member of the Wnt signaling pathway known to be active not only in the β -cells (27) but also in other organs, including liver. Both increased and decreased *TCF7L2* expression in β -cells has been associated with altered insulin secretion and apoptosis in human islets (27) and altered proinsulin processing (27). Notably, rs7903146 in *TCF7L2*, which shows consistent T2D association in samples across diverse ethnic groups, is located in an islet-selective open chromatin site (41,42). Thus, in human islet cells, the chromatin state at rs7903146 is more open in chromosomes carrying the T allele, which is associated with increased T2D risk. Furthermore, there is also previous experimental support for the finding of elevated proinsulin/insulin ration in carriers of the T allele, showing that a number of genes involved in proinsulin processing is altered in animals with knockdown of *TCF7L2* or human carriers of the T allele (43).

Although the genetic association with T2D seems to arise from *TCF7L2*'s function in pancreatic β -cells, emerging studies provide evidence of the role of *TCF7L2* in the liver as well (30,44). Reports of functional studies in humans comparing hepatic SI between carriers of the risk SNP and control subjects are inconsistent. Human carriers of the *TCF7L2* rs7903146 risk T allele have been reported to have elevated HGP (29). Florez et al. (39)

reported impaired insulin secretion accompanied by higher hepatic SI in carriers of the T allele. In contrast, Schäfer et al. (29) could not confirm an association of the risk allele with clamp-measured SI. More recently, Varghese et al. (45) in a small group of overweight, adult subjects without diabetes did not find that genetic variation at *TCF7L2* alters either hepatic or extrahepatic insulin action. The contrasting differences on the effect of the *TCF7L2* variant between our study and that of Varghese et al. (45) are likely because of the younger age and greater severity of obesity of our subjects and the fact that we used a higher insulin to completely suppress HGP.

In the liver, the WNT- β -catenin pathway is involved in metabolic zonation (i.e., hepatocytes exhibit different metabolic profiles [glycolysis vs. gluconeogenesis] depending on their localization along the portocentral axis) (46). *TCF7L2* expression is highest in pericentral hepatocytes, in which gluconeogenesis is low. Recently, Neve et al. (46) reported a significant correlation between increased expression of alternative *TCF7L2* liver transcripts and the T2D-associated rs7903146 risk allele in obese normoglycemic individuals. These results underscore the importance of characterizing the role of *TCF7L2* in the pathogenic mechanisms leading to T2D. In light of the key role of the liver and the strong association found with the common non-coding *TCF7L2* SNP with prediabetes/T2D in adolescents, we explored if *TCF7L2* variant rs7903146 affects hepatic SI in obese adolescents and found that subjects with the TT risk genotype had a significantly impaired suppression in HGP during the hyperinsulinemic clamp. Although these results should be replicated in a larger number of subjects with the T risk allele, they do suggest that the risk variant is not only confined to the β -cell but also involves the liver. Additional studies aimed at deciphering the impact of this key gene variant on hepatic glucose fluxes and determining whether gluconeogenesis is increased according to the *TCF7L2* genotype are needed.

Despite this evidence, it cannot be excluded that the effect on the liver might be somehow mediated by the pancreas. In other words, the insufficient insulin levels, because of an impaired β -cell response in subjects carrying the *TCF7L2* risk allele, might be responsible for the

increase in glucose production in the liver.

Evidence today would indicate that the T2D seems to be more aggressive and deteriorating at a faster pace than what usually seen in adults (10,47). Of note, our data suggest that obese adolescents homozygous for the “risk” *TCF7L2* genotype followed over a 2-year window showed a greater risk of remaining IGT or developing T2D compared with those with the nonrisk *TCF7L2* genotype.

Our study has some limitations. We acknowledge the small sample size of the longitudinal cohort and of those undergoing the hyperinsulinemic-euglycemic clamp. The strengths of our study are the young age of our patients and the assessment of insulin secretion by the OMM and in a subgroup by the hyperinsulinemic-euglycemic clamp and the longitudinal component.

In conclusion, this is the first study showing in obese adolescents the association between the *TCF7L2* rs7903146 and β -cell dysfunction, with impaired proinsulin processing and decreased hepatic SI. The *TCF7L2* variant is implicated in the progression of prediabetes to T2D in the pediatric population.

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