Epigenetic overlap in autism-spectrum neurodevelopmental disorders: MECP2 deficiency causes reduced expression of UBE3A and GABRB3

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Autism is a common neurodevelopmental disorder of complex genetic etiology. Rett syndrome, an X-linked dominant disorder caused by MECP2 mutations, and Angelman syndrome, an imprinted disorder caused by maternal 15q11–q13 or UBE3A deficiency, have phenotypic and genetic overlap with autism. MECP2 encodes methyl-CpG-binding protein 2 that acts as a transcriptional repressor for methylated gene constructs but is surprisingly not required for maintaining imprinted gene expression. Here, we test the hypothesis that MECP2 deficiency may affect the level of expression of UBE3A and neighboring autism candidate gene GABRB3 without necessarily affecting imprinted expression. Multiple quantitative methods were used including automated quantitation of immunofluorescence and in situ hybridization by laser scanning cytometry on tissue microarrays, immunoblot and TaqMan PCR. The results demonstrated significant defects in UBE3A/E6AP expression in two different Mecp2 deficient mouse strains and human Rett, Angelman and autism brains compared with controls. Although no difference was observed in the allelic expression of several imprinted transcripts in Mecp2-null brain, Ube3a sense expression was significantly reduced, consistent with the decrease in protein. A non-imprinted gene from 15q11–q13, GABRB3, encoding the β3 subunit of the GABA_A receptor, also showed significantly reduced expression in multiple Rett, Angelman and autism brain samples, and Mecp2 deficient mice by quantitative immunoblot. These results suggest an overlapping pathway of gene dysregulation within 15q11–q13 in Rett, Angelman and autism and implicate MeCP2 in the regulation of UBE3A and GABRB3 expressions in the postnatal mammalian brain.

INTRODUCTION

Autism is a complex genetic neurodevelopmental disorder characterized by severe impairments in social interaction, communication and behavioral patterns that are restrictive and stereotypical (1). Rett syndrome (RTT; OMIM 312750) is an X-linked dominant neurodevelopmental disorder caused by mutations in MECP2, which encodes methyl-CpG-binding protein 2 (MeCP2) (2). Angelman syndrome (AS; OMIM 105830) is an imprinted disorder caused by maternal deficiency of chromosome 15q11–q13, methylation defects or maternal mutation of UBE3A encoding the ubiquitin ligase UBE3A/E6-AP (3). RTT and AS share overlapping clinical features with autism including developmental delay, language impairment, seizures and stereotypic behaviors (4). Furthermore, clinical assessments of social behavior have demonstrated a high frequency of autism in patients with RTT (5) and AS (6). A Mecp2 mutant mouse model of RTT also shows abnormalities in social interactions (7). In addition, MECP2 mutations have been found in a few patients diagnosed with AS (8) and autism (9,10) and 15q11–q13 duplications are present in ~1% of autism cases (11), suggesting overlap in the pathogenesis of these distinct genetic syndromes. GABRB3, encoding the GABA_A receptor β subunit, is located within the 15q11–q13 (12) and shows association with autism (13,14) but is biallelically expressed (15). We have previously demonstrated MeCP2 expression defects in autism and AS brain samples by a quantitative immunofluorescence approach (16). In contrast, expression level of proteins encoded by genes within 15q11–q13 in RTT, AS and autism have not been well characterized.

MeCP2 binds to methylated CpG sites (17), represses transcription of methylated constructs (18), and associates with chromatin modifying factors histone deacetylase (19,20) and...
DNA methyltransferase (21,22). MeCP2-null (23,24) or mutant (25) mice recapitulate the phenotype of RTT and demonstrate that MeCP2 is essential for postnatal mammalian brain development. Although MeCP2 was predicted to be a global transcriptional repressor, a paucity of MeCP2 target genes have been identified in MeCP2-null mouse brain (26), RTT brain (27) or cell lines (28) by gene expression profiling. The gene encoding brain derived neurotropic factor has been identified as a target of MeCP2 transcriptional repression where a low basal level of transcription prior to neuronal stimulation was increased in MeCP2-null neurons (29,30). The imprinted gene H19 also showed increased expression in MeCP2-null cells (21), but it was not previously determined whether the increase was due to de-repression of the methylated paternal allele. Because of the limited number of MeCP2 target genes, the role of MeCP2 in the pathogenesis of RTT and other autism-spectrum disorders remains elusive.

Although MECP2 mutation does not affect imprinted expression of several genes within 15q11–q13 (31), we hypothesized that MeCP2 may regulate the expression level of genes in this region without necessarily affecting allele-specific expression. Here, we demonstrate that MeCP2 deficiency results in significant reduction of UBE3A/Ube3a and Gabrb3/Gabrb3 expressions in mouse cerebrum without apparent alterations in allele-specific expression. Furthermore, significant reduction of UBE3A and Gabrb3 expressions was observed in AS, RTT and autism human cerebral samples compared with controls. These results demonstrate overlapping epigenetic defects in these phenotypically similar but genetically distinct neurodevelopmental disorders and implicate MeCP2 in the regulation of gene expression within 15q11–q13.

RESULTS
Reduced UBE3A expression in MeCP2 deficient mice
As a non-cell-autonomous dominant effect of MeCP2 mutation in the wild-type (wt)-expressing cells of mosaic MeCP2<sup>−/+</sup> females has been described (32), we performed single cell quantitative analyses of mutant (mt)-expressing (MeCP2-negative) and wt-expressing (MeCP2-positive) cells for UBE3A immunofluorescence on a tissue microarray by laser scanning cytometry (LSC). This approach demonstrated a reproducible 1.5–2-fold decrease in UBE3A immunoreactivity in 10 MeCP2<sup>−/−</sup> and two MeCP2<sup>−/−</sup> cerebral samples from two different MeCP2-null strains (23,24) compared with wt controls (Fig. 1A and B). Both mt-expressing cells from MeCP2<sup>−/+</sup> and MeCP2<sup>−/−</sup> (blue histograms) and wt-expressing cells from MeCP2<sup>−/+</sup> cerebrum (red histograms) showed significantly lower UBE3A expression compared with wt-expressing cells from MeCP2<sup>−/+</sup> cerebrum (orange histograms). To demonstrate that the defects observed in MeCP2<sup>−/+</sup> and MeCP2<sup>−/−</sup> brain samples were actually due to expression differences in UBE3A, not immunofluorescence detection, immunoblot analyses were performed (Fig. 1C). Reduced UBE3A/E6-AP expression was observed by immunoblot for both MeCP2<sup>−/+</sup> and MeCP2<sup>−/−</sup> brain samples from both MeCP2-null strains compared with controls but with lower significance than the LSC results. The precise sampling of single cells within the cerebral cortex in the LSC analysis compared with the whole brain analysis by immunoblot may explain the greater sensitivity of the LSC approach. These results demonstrate that UBE3A protein expression in the cerebral cortex is reduced by both cell-autonomous and non-cell-autonomous effects of MeCP2 mutation.

Ube3a expression level is lower in MeCP2 deficient mouse brain without alterations in imprinted gene expression
As the maternal expression of Ube3a in postnatal neurons is correlated with the paternal expression of an antisense Ube3a transcript from the imprinting control region (ICR) of the Snrpn/Snurf promoter (33), we examined the role of MeCP2 on the imprinting status of this locus. Chromatin immunoprecipitation (ChIP) demonstrated that MeCP2 was bound to the promoter of Snrpn and positive control U2af1-rs1 (34) in mouse cerebrum (Fig. 2A). In contrast, neither the Ube3a nor Gabrb3 promoter was found to be associated with MeCP2 at a detectable level, consistent with a lack of methylation. Allele-specific analyses of (MeCP2<sup>−/+</sup> × PWK)F1 progeny demonstrated the expected preferential maternal expression of Ube3a sense transcript, paternal expression of Ube3a antisense and Snrpn and biallelic expression of Gabrb3, regardless of MeCP2 genotype (Fig. 2B). These results demonstrated that reduced expression of UBE3A/Ube3a in MeCP2 deficient brain was not directly due to alterations in allele-specific expression and confirm similar results from human brain (31). Two additional imprinted genes from other loci (Rasgrf1 and H19) were also tested for allele-specific expression changes in MeCP2-null mice with similar results (Fig. 2B). These results further demonstrate that MeCP2 is not required for maintenance of imprinted gene expression.

To determine whether Ube3a transcript levels were also reduced, TaqMan PCR was performed on RNA samples obtained from MeCP2<sup>−/−</sup> and MeCP2<sup>−/+</sup> mice. The results showed a reduction in Ube3a (sense) and Gabrb3 transcripts, but not in Snrpn, in MeCP2 deficient brain (Fig. 2C). Fluorescence in situ hybridization with single stranded riboprobes was then performed on the tissue microarray in order to separately analyze the sense and antisense transcripts of Ube3a and confirm the reduction of the Ube3a sense transcript by another method. Figure 2D shows the results demonstrating a significantly lower expression of Ube3a sense but not Ube3a antisense transcripts. These combined results suggest that the increase in UBE3A protein expression in MeCP2 deficient brain is due to decreased expression of the Ube3a sense transcript without a significant change in Snrpn or the Ube3a antisense transcript or the maternal-biased expression of Ube3a.

UBE3A expression is deficient in RTT, AS and autism cerebral samples
To determine whether significant reductions in UBE3A expression were also observed in human brain samples, a postmortem brain tissue microarray described previously containing human cerebral samples (16) was analyzed by immunofluorescence and LSC. The histograms in Figure 3A demonstrate significant reductions in UBE3A expression of

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**Fig. 1A and B.** Both mt-expressing cells from MeCP2<sup>−/+</sup> and MeCP2<sup>−/−</sup> (blue histograms) and wt-expressing cells from MeCP2<sup>−/+</sup> cerebrum (red histograms) showed significantly lower UBE3A expression compared with wt-expressing cells from MeCP2<sup>−/+</sup> cerebrum (orange histograms). To demonstrate that the defects observed in MeCP2<sup>−/+</sup> and MeCP2<sup>−/−</sup> brain samples were actually due to expression differences in UBE3A, not immunofluorescence detection, immunoblot analyses were performed (Fig. 1C). Reduced UBE3A/E6-AP expression was observed by immunoblot for both MeCP2<sup>−/+</sup> and MeCP2<sup>−/−</sup> brain samples from both MeCP2-null strains compared with controls but with lower significance than the LSC results. The precise sampling of single cells within the cerebral cortex in the LSC analysis compared with the whole brain analysis by immunoblot may explain the greater sensitivity of the LSC approach. These results demonstrate that UBE3A protein expression in the cerebral cortex is reduced by both cell-autonomous and non-cell-autonomous effects of MeCP2 mutation.

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Figure 1. UBE3A expression in Mecp2-null mouse brain. (A) A multiple tissue microarray containing wt and knock-out mouse cerebral cortex samples (16) was stained for immunofluorescence using a C-terminal reactive anti-MeCP2 and an anti-UBE3A antibody. The MeCP2-negative cells (Mecp2 mt-expressing, blue histograms, percentage shown) were separately gated from MeCP2-positive cells (Mecp2 wt-expressing, red histograms) and the total population (black histograms) and compared with age-matched wt control female or male samples (orange histograms). (B) The graph shows combined LSC results (mean ± SEM) from four replicate slides using two different anti-UBE3A antibodies. (C) Protein extracts from whole adult mouse brain were probed on an immunoblot with anti-UBE3A or anti-GAPDH. A representative image shows lower expression in brain of UBE3A in both hemizygous male (−/+ ) and heterozygous female (−/+ ) brain compared with wild-type (wt, +/+ ) and littermate controls for both Mecp2tm1.1Bird and Mecp2tm1.1Jae strains. The graph demonstrates combined GAPDH-normalized results (mean ± SEM, quantitation as previously described (32), example ratios shown below each lane) from eight different −/+ and six different +/+ brain samples from Mecp2tm1.1Bird and Mecp2tm1.1Jae strains. * P < 0.05, **** P < 0.0001 by t-test.

RTT, AS and autism samples (black) compared with age-matched controls (gray). Ages of each sample are shown, but no significant differences were observed in the level of UBE3A expression with increasing age of the controls (data not shown), unlike the striking increase with age observed for MeCP2 (16,35). Mean UBE3A fluorescence was normalized to control histone H1 expression and graphed (Fig. 3B), demonstrating significant UBE3A reductions in RTT and autism samples. The reduced expression of UBE3A in the AS 15q11–q13 deletion sample was expected and consistent with a low level of paternal UBE3A expression in the cerebral cortex (36). The LSC results were confirmed by immunoblot (Fig. 3C shows a representative blot). Furthermore, reduced UBE3A expression in AUT1174 was also shown by immunoblot in a recent report (37).

GABRB3 expression defects are observed in Mecp2 deficient mouse and RTT, AS and autism human brain

GABRB3 has been strongly implicated in autism by multiple association studies (14,38,39), but alterations in expression level have not been previously demonstrated. Because Mecp2 deficiency affected not only Ube3a expression but also the biallelically expressed gene, Gabrb3 (Fig. 2C), we further investigated its protein product, the gamma aminobutyric acid (GABA) receptor β3 subunit, for defects in Mecp2/ MeCP2 deficient brain. Immunoblot analyses demonstrated reduced-GABRB3 expression in Mecp2−/− and Mecp2+/- mouse brain compared with controls (Fig. 4A), suggesting an additional target from the 15q11–q13 locus may also be dysregulated in RTT, AS and autism. As a transmembrane protein, GABRB3 was less amenable to LSC analysis than nuclear or cytoplasmic proteins because individual cells are identified by nuclear contouring. Immunoblot analyses were, therefore, performed on frozen frontal cerebral cortex, Brodmann area 9 (BA9), from a total of three RTT (with MECP2 mutations), two AS (with 15q11–q13 deletions), nine autism (no detectable MECP2 mutations or 15q11–q13 duplications) and 11 different age-matched normal controls. Two representative immunoblots are shown in Figure 4B. Table 1 shows the combined data of normalized GABRB3/GAPDH ratios for all individual samples. When analyzed as combined groups, RTT, AS and autism samples showed significantly lower GABRB3 expression compared with controls. The autism samples showed the highest significant decrease in GABRB3 as a group (P < 0.0001) despite higher variability between samples, with some showing extremely low expression (i.e., 797, 1174, 967 and 5173, with 5–10% of control expression) and others showing little change (i.e., 3924 and 5342, Fig. 4B and Table 1). The ages of samples are shown in Table 1, but reveal no significant change in GABRB3 expression with age of the control samples tested. The average GABRB3 expression for each patient category is, therefore, also represented as a group for statistical comparisons. Although GABRB3 expression deficiencies are not universal in all autism brain samples, the defects are more significant than in RTT and AS and occur at an unexpectedly high frequency (5/9 or 56%). The specificity of GABRB3 defects to AS, RTT and autism was tested by analyzing three
DISCUSSION

Uncovering the genetic basis of autism has been challenging because of its complex etiology. Although the heritability of autism in families is high, multiple gene loci and environmental influences are expected (1). Even for the autism-spectrum disorders with known genetic causes, the molecular pathogenesis is not well understood. Because of the overlap in phenotype among RTT, AS and autism (4), we tested the hypothesis that gene expression in the 15q11–q13 region may be altered in all three syndromes. We show for the first time that the AS gene, UBE3A/Ube3a, is significantly reduced in RTT and Mecp2-null mice, demonstrating that MECP2/Mecp2 deficiency causes reduced expression of Ube3a/UBE3A. In addition, we demonstrate UBE3A deficiency in autism brain samples, consistent with a recent report (37). Furthermore, we also show for the first time deficiencies in the expression of the GABA receptor β3 subunit gene (GABRB3) in RTT and autism brain and Mecp2-null mice.

Maternal duplications in 15q11–q13 have been observed in 1% of autism cases as the most common cytogenetic
abnormality in autism (11). Genome-wide linkage studies have identified the 15q11–q13 locus in some but not all scans (40–43). Furthermore, higher resolution scans of linkage within 15q11–q13 in autism families have identified significant linkage to $UBE3A$ and $GABRB3$ in some studies but not others (14,44–48). While the linkage of autism to 15q11–q13 may be somewhat disputed at the genetic level, here we show that expression level defects of two genes within 15q11–q13 are common to RTT, AS and autism. The reduction of $UBE3A$ and $GABRB3$ in AS cerebrum was expected because of the maternal 15q11–q13 deletion in these samples. Interestingly, neither $Ube3a$ deficient nor uniparental disomy mice showed evidence for decreased $Gabrb3$ expression (49,50), but the lack of AS brain samples with uniparental disomy or $UBE3A$ mutations prevented our confirmation of this finding in human samples.

Our results are consistent with a recent report showing decreased $UBE3A$ in autism brain samples (37), but extend the observation to demonstrate $GABRB3$ deficiency as a common defect in autism. The GABA$\ receptor genes are attractive candidate genes for autism as several studies have demonstrated linkage to one or more of these genes (13,14,48). Autoradiography has demonstrated reduced GABA$\$ binding in autism brain (51) and elevated circulating GABA levels have been found in autistic children (52–54). Although our sample size was small owing to the requirement for postmortem human brain tissue, the frequency of $UBE3A$ and $GABRB3$ expression defects within autism brain samples was much higher than for previously described genetic defects. The defects in $UBE3A$ and $GABRB3$ expressions were unlikely to be due to degradation in postmortem tissue as they were not observed in multiple normal and DS controls and because the results were all normalized to housekeeping gene controls. These results suggest that reduced expression of $UBE3A$ and $GABRB3$ are correlated and may be a downstream consequence of multiple different genetic or epigenetic etiologies. The phenotypic consequence of reduced expression of these genes is expected to be less severe than gene mutation ($UBE3A$ mutations in AS) or nullisomy. Both $Ube3a^{+/+}$ and $Gabrb3^{+/+}$ mice have learning and motor defects and inducible seizures, similar to AS and some cases of autism (55,56). While our autism brain sample population seems to be somewhat enriched for associated seizure disorders (Supplementary Material, Table S1), significantly reduced $GABRB3$ expression was observed in brain samples from individuals with and without seizures.

We also identify the X-linked gene, $MECP2$, as one genetic cause of $UBE3A$ and $GABRB3$ expression defects in the mammalian cerebrum. We have used two Mecp2 deficient RTT mouse strains (23,24) to demonstrate that $UBE3A$/ $Ube3a$ and $GABRB3$/Gabrb3 are significantly decreased in adult mouse brain compared with wt controls. Furthermore, we show significant defects in expression of $UBE3A$ and $GABRB3$ in human RTT brain samples with $MECP2$
mutations. Our results are consistent with GABRB3 showing reduced expression in RTT brain by genome-wide expression profiling in one report (27). The relatively subtle changes in expression of UBE3A and GABRB3 observed in this study may explain why these genes were not identified in other genome-wide profiling studies of MECP2/Mecp2 mutant tissues (26,28). The observation of down-regulation as well as up-regulation of MeCP2 target genes has also been observed in expression profiling studies, arguing against a single role for MeCP2 as a transcriptional repressor.

As we have previously demonstrated MECP2 expression defects in the same autism brain samples (16), perhaps MECP2 expression defects could be responsible for the lower UBE3A and GABRB3 expressions in these autistic individuals without characterized genetic defects. Alternatively, defects in expression of all three genes may be a downstream consequence of abnormal brain development. As the Mecp2308/+ RTT mouse model has been demonstrated to show some autistic behaviors, including forepaw stereopathies and social avoidance (7,25), the role of MeCP2 in the pathogenesis of at least some cases of autism appears likely (4).

In this report, we also begin to uncover a rather complex and unexpected mechanism by which MeCP2 regulates expression of UBE3A and GABRB3. Although MeCP2 binds to methylated DNA and is a predicted transcriptional

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*Replicate results from each individual sample were compared with the three closest age-matched controls by t-test. Total sample replicates were combined from each category and compared with total control sample replicates by t-test.

\( t < 0.01. \)

\( t < 0.05. \)

\( t < 0.001. \)

\( t < 0.0001. \)
repressor (17,18), it is not required for maintaining parental imprints (31). Here, we demonstrate that MeCP2 binds to the ICR in the Snrpn promoter, yet it is not required for silencing the maternal allele of Snrpn and Ube3a antisense or the paternal allele of the Ube3a sense transcript. Furthermore, expression level of Gabrb3 is affected even though this gene is biallelically expressed. We also demonstrate the maintenance of exclusive maternal expression of H19 in Mecp2-null mice despite its description as a ‘bona fide’ MeCP2 target gene (21). These results suggest that the effect of MeCP2 binding to the ICR is more complex than simply repressing imprinted genes in cis and may instead involve long-range chromatin interactions. Perhaps chromatin loop formation such as has been recently demonstrated in the imprinted Igf2/H19 region (57) is adversely affected by MeCP2 deficiency, as MeCP2 has been previously described to have matrix binding activity (58). The ~1 Mb region from UBE3A to GABRB3 also includes multiple small nuclear RNA genes (33,59,60) and binding activity (58). The H19 region (57) is adversely affected by paternal allele of the Ube3a (58).

The human tissue microarray contains triplicate frontal cerebral cortex (BA9, layers III–V) obtained frozen <30 h post-mortem. 5 μm section was processed for immunofluorescence as described previously for anti-MeCP2 (C-terminal, Affinity Bioreagents) and anti-histone H1 (Upstate Biotechnology) antibodies (66). For mouse tissues, combined staining was performed with anti-UBE3A antibody (BD Biosciences and monoclonal gift from Howley, Harvard Medical School) and a cascade-blue labeled goat-anti-mouse IgG (Molecular Probes). For human tissues, an anti-UBE3A antibody (Affinity Bioreagents) was used. LSC analysis of MeCP2 negative and positive populations was performed as described previously (32).

**In situ hybridization analysis of tissue microarrays**

Sense and antisense transcripts of Ube3a were detected by fluorescence in situ hybridization using single stranded riboprobes from a 796 bp cloned fragment spanning exons 6–10 (primers, 5′-ttatatgtagaactcagaa-3′ and 5′-tttctcttcatcctgg-3′) labeled with biotin (Lofstrand Laboratories). A control β-actin probe digoxigenin labeled (Roche) was simultaneously hybridized at 55°C. Hybridization conditions, fluorescent detection and quantitation by LSC were performed as described previously (16).

**Immunoblot analyses**

A total of 10 μg of protein extract from whole adult mouse brain was analyzed per lane by immunoblot as described previously (32) using anti-UBE3A (BD Biosciences), anti-GABRB3 (Affinity Bioreagents) or anti-GAPDH (Advanced Immunochemical) antibodies at recommended concentrations. Protein extracts were isolated from frozen human postmortem cerebral cortex (BA9) from samples previously described (16) and additional samples (Harvard Brain Bank and University of Maryland Brain and Tissue Bank). Similar to mouse samples, 10 μg of human protein was analyzed per lane with the same reagents except the substitution of a human-specific anti-Ube3a antibody (Affinity Bioreagents). Additional human cerebral (BA9) samples were obtained from autism, Down and age-matched control samples from the Autism Tissue Program for immunoblot analyses. Quantitation was performed using Nucleotech Gel Expert version 2.0.

**Allelic expression of imprinted genes in interspecies hybrids**

Mecp2<sup>tm1.1Bird/+</sup> (C57B6) female mice (23) were mated to wt PWK males and the resulting offsprings were genotyped for Mecp2 as described previously (32). RNA was isolated from adult (Mecp2<sup>tm1.1Bird/+ × PWK</sup>)F1 brain samples from all four genotypes as well as parental B6 and PWK mice. RT–PCR followed by restriction enzyme digestion was performed as described previously for Ube3a, Snrpn, Gabrb3 (67) and Rasgrf1 (68). H19 primers (5′-GAACCACTAACACAGTATTCCG-3′ and 5′-GGAACAGCTCCCAGACTAGG-3′, 58°C with 30 cycles) amplified a 585 bp fragment that was digested with BglII.

**Materials and Methods**

**Immunofluorescence analysis of tissue microarrays**

Construction of tissue microarrays used in this study have been previously described for Mecp2-null mouse (32) and human neurodevelopmental disorders (16). Briefly, for the mouse array, triplicate 600 μm cores of cerebral cortex were removed from five Mecp2<sup>tm1.1Bird/+</sup>, five Mecp2<sup>tm1.1Jae/+</sup> and three Mecp2<sup>+/+</sup> controls (P20–P27 w); one Mecp2<sup>tm1.1Bird/y</sup> and one Mecp2<sup>+/y</sup> littermates (P10 w); one Mecp2<sup>tm1.1Jae/y</sup> and one Mecp2<sup>+/y</sup> littermate controls (P10 w).
Chromatin immunoprecipitation

Chromatin from adult mouse cerebrum samples [C57B6, PWK or (B6 x PWK)F1] was isolated for ChIP as described previously (34,69). Anti-MeCP2 (C-terminal) was used to immunoprecipitate DNA fragments from ‘Input’ control as described previously for primers spanning 5' CpG islands for Snrpn and positive control U2af1-rs1(34), Ube3a (5'-CCCTTCTGGTCTTC'TCGAGGT-3'), 5'-CGAAGAGCACACACGATAAAA-3', 58°C anneal, 37 cycles) and Gabrb3 (5'-CCTCAGAGGACCCGTAC-3', 5'-GTCTAGGACCCGCGACA-3, 60°C anneal, 40 cycles).

TaqMan PCR

Each PCR contained 40 nM of each primer, 80 nM of the TaqMan probe and commercially available PCR mastermix (TaqMan Universal PCR Mastermix, Applied Biosystems) containing 10 mM Tris–HCl (pH 8.3), 50 mM KCl, 5 mM MgCl2, 2.5 mM deoxynucleotide triphosphates, 0.625 U AmpliTaq Gold DNA polymerase per reaction, 0.25 U AmpliErase UNG per reaction and 5 µl of the diluted cDNA sample in a final volume of 25 µl. Ube3a forward primer spanned exons 7–8 (5'-TCCAGATATGGATGTGTTACA TATG-3'), reverse primer (5'-GGGAAAATGGACATCCA GTATACA-3') and probe (5'-CTGATTGCGATAGTCTCT GGGTCTGGC-3') were from exon 8 (NM_173010). Snrpn forward primer spanned exons 3–4 (5'-TCAGGAAGAT CAAGCAGAGATG-3'), reverse primer (5'-AAGTCTT CCCCCAGTACAAAGAC-3') and probe (5'-ACATGGCAC AACGTGAAAGAAAAACGGGT-3') were from exon 4 (NM_013670). Gabrb3 forward primer spanned exons 6–7 (5'-CCGATTTTCTTCTACGAGAAA-3') and probe (5'- TCCGCCCAGGTTGCTATCTTCGCAG-3') were from exon 7. The samples were placed in 96 well plates and amplified in an automated fluorometer (ABI PRISM 7700 Sequence Detection System, Applied Biosystems). Amplification conditions were 2 min at 50°C, 10 min at 95°C, 40 cycles of 15 s at 95°C and 60 s at 60°C. Final quantitation was done using the comparative CT method (Applied Biosystems) and is reported as relative transcription or the n-fold difference relative to a calibrator cDNA (Mecep2+/- of each pair) following normalization to HPRT1 housekeeping control gene.

SUPPLEMENTARY MATERIAL

Supplementary Material is available at HMG Online.

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