A novel murine allele of Intraflagellar Transport Protein 172 causes a syndrome including VACTERL-like features with hydrocephalus

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The primary cilium is emerging as a crucial regulator of signaling pathways central to vertebrate development and human disease. We identified atrioventricular canal 1 (avc1), a mouse mutation that caused VACTERL association with hydrocephalus, or VACTERL-H. We showed that avc1 is a hypomorphic mutation of intraflagellar transport protein 172 (Ift172), required for ciliogenesis and Hedgehog (Hh) signaling. Phenotypically, avc1 caused VACTERL-H but not abnormalities in left–right (L–R) axis formation. Avc1 resulted in structural cilia defects, including truncated cilia in vivo and in vitro. We observed a dose-dependent requirement for Ift172 in ciliogenesis using an allelic series generated with Ift172avc1 and Ift172wim, an Ift172 null allele: cilia were present on 42% of avc1 mouse embryonic fibroblast (MEF) and 28% of avc1/wim MEFS, in contrast to >90% of wild-type MEFS. Furthermore, quantitative cilium length analysis identified two specific cilium populations in mutant MEFS: a normal population with normal IFT and a truncated population, 50% of normal length, with disrupted IFT. Cells from wild-type embryos had predominantly full-length cilia, avc1 embryos, with Hh signaling abnormalities but not L–R abnormalities, had cilia equally divided between full-length and truncated, and avc1/wim embryos, with both Hh signaling and L–R abnormalities, were primarily truncated. Truncated Ift172 mutant cilia showed defects of the distal ciliary axoneme, including disrupted IFT88 localization and Hh-dependent Gli2 localization. We propose a model in which mutation of Ift172 results in a specific class of abnormal cilia, causing disrupted Hh signaling while maintaining L–R axis determination, and resulting in the VACTERL-H phenotype.

INTRODUCTION

VACTERL association is a non-random association of congenital anomalies with no known etiology. First described as VATER association by Quan and Smith in 1973, the initial patients were noted to have vertebral anomalies (V), anal atresia (A), tracheoesophageal fistulas (TE) and renal (R) and limb (L) anomalies. The high incidence of congenital heart defects in patients with VATER association was subsequently described, resulting in the inclusive acronym VACTERL (1–3).

The etiology of VACTERL association has not been established. Currently, patients can be diagnosed with VACTERL association if they have two or more of the relevant anomalies (4). In the series reported by Khoury et al. (5), >90% of patients diagnosed with VACTERL association had three or fewer anomalies, and <1% of patients had all six anomalies. There have been few reported instances of familial recurrence of typical VACTERL, and therefore monogenic causation has been deemed unlikely. Interestingly, Mendelian inheritance of a distinct subgroup of VACTERL association, VACTERL
with hydrocephalus, or VACTERL-H, has been reported (6). Patients with VACTERL-H meet the diagnostic criteria for VACTERL, and in addition have hydrocephalus. Both autosomal recessive and X-linked recessive inheritance of VACTERL-H have been described ((7,8) reviewed in 9). These inheritance patterns have led some authors to argue that VACTERL-H should be classified as a syndrome (as opposed to an association) despite the unknown genetic etiology, given the evidence for monogenic inheritance (10).

Mice with disrupted Hedgehog (Hh) signaling display phenotypic overlap with VACTERL patients. In particular, Gli2 and Gli3 mutant mice have vertebral anomalies (11); Shh mutants develop truncated limbs (12); Gli3 mutants exhibit polydactyly (13); Shh, Gli2 and Gli3 mutants all show various foregut and hindgut anomalies (14–17); and Shh and Gli2;Gli3 double-mutants show renal anomalies (18–20). These findings have led to speculation that alterations of the Hh signaling pathway may contribute to the pathogenesis of human VACTERL (4). Recently, a human patient with typical VACTERL association was found to have a mutation of HOXD13 (21), a downstream target of Shh, providing possible indirect evidence linking human VACTERL with the Hh signaling pathway. However, no models of VACTERL-H have identified a single-gene etiology to date.

A growing body of work has demonstrated the crucial role played by primary cilia in vertebral developmental signaling pathways. Multiple studies have begun to elucidate the complex role played by cilia and intraflagellar transport in Hh signaling. Huangfu et al. (22) first made the connection between cilia and Hh signaling, demonstrating that wimple (wim), a null allele of intraflagellar transport protein IFT172 (Ift172), results in absent cilia and loss of Hh signaling. Subsequently, it has been established that multiple Hh signaling molecules, including Ptc1, Smo, Sufu and the Gli family of transcription factors, are all enriched in cilia (23–27). Intraflagellar transport appears to play an active role in Hh signaling, and IFT proteins are required for both Gli activator and Gli repressor function (25,28,29). Therefore, cilia mutations cause disruptions of the Hh signaling pathway, disrupting Hh signaling while maintaining the ciliary functions necessary for L–R determination, but defects, suggests that immotile cilia are unable to create the nodal flow required for L–R determination, mechanisms dis-}

RESULTS

**avcl:** an ENU-derived mouse mutant with phenotypic features of VACTERL-H

We performed a forward genetic screen for ENU-induced mutations that caused perinatal lethality and CHD in the mouse (31). avcl caused heritable syndromic atrioventricular septal defects (AVSDs) as part of VACTERL-H association (Fig. 1). avcl mutant newborns demonstrated hydrocephalus with highly penetrant vertebral, anal, cardiac, limb and palate abnormalities (Fig. 1U). Anomalies of the cervical vertebrae, including absent spinal processes or abnormally formed transverse processes, were present in 8/10 avcl mutant mice in contrast to 0/10 wild-type littermate controls (P = 0.02; Fig. 1A and B). Mutants had seven cervical vertebrae, as is standard, but each cervical vertebra lacked the vertebral body (Fig. 1B). Thoracic, lumbar and sacral vertebrae appeared normal. Anal atresia, evidenced by discontinuity between the anal squamous epithelium and rectal columnar epithelium, was present in three of five avcl embryos, whereas all the eight wild-type embryos showed a clear, intact transition between anal and rectal epithelium (P = 0.04; Fig. 1C and D). Cardiac anomalies, atrioventricular septal defects with common atrium, were present in 100% of avcl mutants analyzed (n > 100; P < 10^−6; Fig. 1E and F). Cardiac defects were not observed in any littermate control animals. Bone staining demonstrated anomalies of the long bones of the forelimbs and hindlimbs in mutant embryos (Fig. 1K–N). Mutant embryos demonstrated shortening of the humerus, radius, ulna, femur, tibia and fibula. In addition,
all mutant mice demonstrated preaxial polydactyly of both the forelimbs and hindlimbs ($n = 119$) (Fig. 1N). All newborn animals examined demonstrated abnormal facies, with shortening of the muzzle. Clefts of the secondary palate were similarly noted in 20/20 newborn mutants ($P < 0.01$) (Fig. 1P). Hydrocephalus was present in four of five mutant animals, but none of seven littermate wild-type controls ($P = 0.01$; Fig. 1Q and R).

The remaining features of the VACTERL-H phenotype, tracheo-esophageal and renal abnormalities, were present with variable frequency. Esophageal and tracheal anomalies were observed in 2/9 mutant embryos at E14.5 (esophageal stenosis with enlarged, cystic trachea in one case, and esophageal atresia in the other), whereas 0/10 wild-type embryos analyzed showed foregut anomalies ($P = 0.1$; Fig. 1G and H). Renal dysplasia with hypoplastic glomeruli was observed in four of nine avc1 mutant embryos compared with none of six wild-type ($P = 0.09$, Fig. 1I and J). No eye abnormalities were observed by gross or histologic analysis of avc1 mutant embryos ($n = 5$).
Interestingly, no cardiac looping anomalies were noted in any of the mutant embryos. All mutant embryos analyzed (n = 100) showed levocardia with normal, d-looping of the ventricles, as did all wild-type control embryos. Similarly, all mutant embryos demonstrated situs solitus of the abdominal organs. Thus, no situs anomalies were observed in avc1 mutant embryos.

avc1 is an allele of Ift172

A candidate gene approach was used to identify the avc1 mutation. Recombinant chromosomes were identified in affected animals from an intercross between known carriers that located the avc1 mutation between markers D5MIT334 and D5MIT419. The interval containing avc1 corresponded to 0.9 megabases (Mb) on the physical map containing 33 genes. Bioinformatic investigation of the 33 candidate genes revealed no known role for any in heart development. However, one gene in the interval, Ift172, encoded an intraflagellar transport protein required for cilia biogenesis and implicated in the Hh signaling pathway (22).

Sequencing of all 48 exons of the Ift172 gene, including adjacent intronic sequence, revealed an A-to-G transition in the splice donor site downstream of exon 24 (Fig. 2A). This polymorphism, located at position +3 of the intron of exon 24, altered a nucleotide conserved from humans to opossum. We hypothesized that the splice-site sequence alteration may affect the splicing of Ift172 transcripts. We evaluated products of Ift172 transcription by reverse transcriptase polymerase chain reaction (RT-PCR) in wild-type and

![Figure 2. Aberrant splicing of Ift172 in Avc1 mutant embryos.](image-url)
avc1 mutant embryos, using primers designed to amplify Ift172 transcriptional products generated between exons 22 and 26, spanning the exons adjacent to the sequence polymorphism. Semi-quantitative RT-PCR from wild-type whole-embryonic tissue at E10.5 revealed a single product, whose sequence corresponded to wild-type Ift172 cDNA. However, the sequence of the minority species, of higher molecular weight, corresponded to wild-type Ift172 cDNA. However, the sequence of the majorit species, of lower molecular weight, revealed exon 24 skipping (Fig. 2B and C). Theoretical translation of the mis-spliced message encountered a frame shift and premature termination in exon 25 (Fig. 2A).

The results of RT-PCR from avc1 mutant embryos suggested that the sequence alteration identified in Ift172 had functional consequences on Ift172 splicing. However, this perturbation of Ift172 had yet to be directly linked to the phenotype observed in avc1 mutant embryos. Because the avc1 allele was generated by ENU mutagenesis, it remained feasible that an undiscovered mutation affecting another gene in the physical interval containing avc1 contributed to the mutant phenotype. We sought to directly test whether avc1 and Ift172 were allelic. We performed a complementation test between avc1 and Ift172<sup>wim</sup>, an Ift172 null allele that causes mid-gestation embryonic lethality and loss of Hh signaling (22). The embryos from an intercross between Ift172<sup>wim</sup> and avc1/+ mice were evaluated. Mice carrying both the Ift172<sup>wim</sup> and avc1 alleles demonstrated cardiac, limb and craniofacial abnormalities, whereas none of the heterozygous carriers of either mutant allele demonstrated phenotypic abnormalities (Fig. 2D). These findings demonstrated that Ift172<sup>wim</sup> and avc1 failed to complement and provided further evidence that the identified mutation of the Ift172 gene is responsible for the phenotype observed in avc1 mutant mice. As Ift172<sup>avc1</sup> mice maintained some wild-type transcript and die at birth whereas Ift172<sup>wim</sup> mice die at E10.5, we conclude that avc1 is a hypomorphic allele. No homozygous avc1 mice were observed at weaning from 200 consecutive litters.

Whole-mount in situ hybridization against Ift172 in control embryos showed widespread expression of Ift172 in tissues affected by avc1 and known to be Hh-responsive (Fig. 3A–C, left column). Ift172<sup>avc1</sup> mutant embryos demonstrated greatly reduced but detectable expression (Fig. 3A–C, right column). These observations suggested that the majority product of mis-spliced mRNA in the mutant embryos underwent nonsense-mediated decay. To directly test this hypothesis, we performed western blotting with an antibody against the N-terminus of IFT172. Lanes represent whole-embryo extracts from wild-type (lane 1), avc1/+ heterozygote (lane 2) or avc1/avc1 mutant (lane 3) embryos at E10.5, with γ-tubulin used as a loading control. The filled arrowhead indicates IFT172 protein was quantitatively determined by western blot analysis using a rabbit antibody raised against the N-terminus of IFT172 from E10.5 embryonic extracts. avc1 mutant embryos expressed 18% of the amount of IFT172 compared with littermate wild-type controls after normalization (Fig. 3D). Heterozygous avc1 mutant embryos expressed 63% of the amount of IFT172 compared with littermate wild-type controls after normalization (Fig. 3D). These results provide additional evidence that the mis-spliced product of Ift172 RNA undergoes nonsense-mediated decay in avc1 mutant embryos, and that avc1 causes a dose decrease in IFT172 protein levels.

Ift172 mutant embryos have defects in Hh signaling

Given the known role of Ift172 in Hh signaling and the suspected link between defective Hh signaling and VACTERL syndrome, we sought to evaluate the impact of reduced Ift172 expression on Hh signaling in Ift172<sup>avc1</sup> mutant mice. We assessed the integrity of the Hh signaling pathway in Ift172<sup>avc1</sup> mutants in two ways, by whole-mount analysis of an Hh reporter and biochemically. First, we qualitatively evaluated the expression pattern of Patched1 (Ptch1) and Gli1.
The expression of *Ptch1*, the primary Hh membrane receptor, and *Gli1*, a member of the *Gli* transcription factor family responsible for transducing the Hh signal, are Hh dependent. We compared the expression of *Ptch1* using a previously reported *Ptch1-lacZ* Hh-reporter mouse line (32) and *Gli1* by *in situ* hybridization in *Ift172avc1* mutants and wild-type littermate controls at E10.5 (Fig. 4). In wild-type embryos, *Ptch1* and *Gli1* expression were confined to regions with known Hh signaling mirroring the expression of *Ift172* (Fig. 4A). In comparison, *Ift172avc1* mutant littermates demonstrated a global reduction in the expression of *Ptch1* and *Gli1* (Fig. 4B). Consistent with previous analysis describing an extra-cardiac population of Hh-receiving progenitor cells that subsequently migrate into the heart to form the atrial septum (33), there was minimal evidence of Hh signaling within the heart itself (Fig. 4A and B, lower panels).

We next quantitatively assessed Hh signaling in *Ift172avc1* embryos, examining the proteolytic processing of the Gli3 transcription factor. In mammals, Gli3 exists in two forms, a full-length protein that acts as a transcriptional activator (Gli3–190) and a processed form that acts as a transcriptional repressor (Gli3–83). Processing of Gli3 from the full-length activator form to the truncated repressor form is regulated by Hh signaling. Using western blotting and densitometry to quantify protein levels in tissue extracted from mutant and wild-type embryos, we found that there appeared to be a direct relationship between *Ift172* dosage and Gli3 processing (Fig. 4C). In wild-type embryos, the ratio of processed to unprocessed Gli3 was 1.33. The ratios of processed to unprocessed Gli3 in *avc1* mutants, *avc1/wim* compound heterozygote mutants and *wim* mutants were 0.39, 0.17 and 0.01, respectively (Fig. 4). This represented a 70.7% reduction of Gli3 processing in *Ift172avc1* mutant embryos, an 87.2% reduction in *Ift172avc1/wim* embryos and a 99.2% reduction in *Ift172wim* embryos. Thus, the *Ift172avc1* allele disrupted the Hh signaling, but to a lesser extent than the *Ift172wim* allele. These results supported the conclusion that *Ift172avc1* is a hypomorphic allele of *Ift172* and causes significant but incomplete loss of embryonic Hh signaling.

The intersection between the requirement for cilia and Hh signaling in heart development had not been demonstrated. We asked whether a decrement in Hh signaling could account for the atrioventricular septal defects observed in...
**If**t172 mutant embryos display truncated cilia

To evaluate the effects of reduced Ift172 expression on cilia structure *in vivo*, we used scanning electron microscopy to analyze the ventral node of Ift172<sup>avc1</sup> mutant, Ift172<sup>avc1/wim</sup> mutant and wild-type littermate embryos at E7.5 (Fig. 5). We observed a dose-dependent requirement for Ift172 in the production of full-length nodal cilia. The mean nodal cilia length was 3.56 ± 0.05 μm in wild-type E7.5 embryos (Fig. 5A, D, G and H), 2.26 ± 0.08 μm in Ift172<sup>avc1</sup> embryos (Fig. 5B, E, G and H) and 0.81 ± 0.02 μm in Ift172<sup>avc1/wim</sup> embryos (Fig. 5C and F–H). Reduced Ift172 dosage therefore impairs ciliogenesis *in vivo* at the ventral node.

To better characterize the mechanistic basis for the quantitative decrement in Hh signaling observed in Ift172<sup>avc1</sup> mutants, we evaluated the primary cilia of mutant mouse embryonic fibroblasts (MEFs). We predicted that fibroblast primary cilia would be abnormal in the *avc1* mutant setting and tested this hypothesis by isolating and comparing fibroblasts from wild-type, Ift172<sup>avc1</sup> and Ift172<sup>avc1/wim</sup> embryos. Analysis of the Ift172<sup>avc1</sup> allelic series revealed a dose-dependent requirement for Ift172 in the synthesis of primary cilia. Using acetylated tubulin immunohistochemistry, cilia were identified on 90.8% of wild-type cells (Fig. 6A, D and G), 41.5% of Ift172<sup>avc1</sup> mutant cells (Fig. 6B, E and G) and 28.4% of Ift172<sup>avc1/wim</sup> cells (Fig. 6C, F and G).

The documented deficiency in Hh signaling appeared greater in *avc1* mutants than could be explained by the decreased percentage of ciliated cells (*avc1* mutants showed an ~70% reduction in Gli3 processing, but only an ~54% reduction in the number of ciliated fibroblasts). We hypothesized that the cilia present on mutant cells may be abnormal and unable to support normal Hh signaling. We proceeded to structurally and functionally evaluate the cilia present on *avc1* and *avc1/wim* mutant MEFs. Quantitative analysis of primary cilia length revealed that there was a direct relationship between Ift172 dosage and cilia length. Cilia on fibroblasts derived from wild-type embryos were significantly longer than the cilia on cells derived from Ift172<sup>avc1</sup> embryos which were in turn significantly longer than cilia on cells from Ift172<sup>avc1/wim</sup> compound heterozygote embryos (Fig. 6H).

Closer analysis of cilia length suggested that there were two distinct populations of cilia present, full length and truncated to approximately half length, rather than a normal distribution of cilia length within each genotype. Histogram analysis of the length of cilia on Ift172<sup>avc1/avc1</sup> fibroblasts demonstrated two peaks, rather than a normal distribution of lengths. Ift172<sup>avc1</sup> cilia were approximately equally divided between two populations, a ‘full length’ population, with a peak centered at ~2.5 μm, and a ‘truncated’ population, with a peak centered at ~1.5 μm (Fig. 6I). Analysis of wild-type fibroblast cilia length revealed a greater population of ‘full length’ cilia, with a much smaller population of ‘truncated’ cilia, although analysis of Ift172<sup>avc1/wim</sup> fibroblasts revealed a large ‘truncated’ population, with a much smaller population of the ‘full length’ cilia (Fig. 6I). Cilia of both types were present in all genotypes, and the range of cilia length was nearly identical between the three genotypes (Fig. 6I). This observation suggested that the observed difference in mean cilia length...
Cilia were divided evenly between the two populations and 4.06 (28.4%) (C, F and G) mutant fibroblasts were ciliated (although significantly fewer (blue). Nearly all wild-type fibroblasts were ciliated (90.8%) (A, D and G), tubulin (red). Arrows identify primary cilia. Nuclei were stained with DAPI cilia growth, primary cilia were visualized in the cultured cells with acetylated mutant and wild-type littermate embryos. After serum-starving to promote fibroblasts. Figure 6. Abnormal number and morphology of primary cilia in Ift172 mutant fibroblasts. (A–G) Fibroblasts were isolated from Ift172avc1 and Ift172avc1/wim mutant and wild-type littermate embryos. After serum-starving to promote cilia growth, primary cilia were visualized in the cultured cells with acetylated tubulin (red). Arrows identify primary cilia. Nuclei were stained with DAPI (blue). Nearly all wild-type fibroblasts were ciliated (90.8%) (A, D and G), although significantly fewer Ift172avc1 (41.5%) (B, E and G) and Ift172avc1/wim (28.4%) (C, F and G) mutant fibroblasts were ciliated (P-values 8.12 × 10−13 and 4.06 × 10−23), respectively. (H) Ift172avc1 and Ift172avc1/wim mutant cilia were significantly shorter on average than wild-type cilia (2.18 and 1.62 versus 2.62 μm, P-values of 1.20 × 10−6 and 1.71 × 10−30, respectively). (I) Histogram analysis of primary cilia length demonstrates two distinct cilia populations, a full-length population and a truncated population that are approximately half normal length. Wild-type cilia were mostly full-length, Ift172avc1 cilia were divided evenly between the two populations and Ift172avc1/wim cilia appeared mostly truncated.

was due to varying the relative proportions between the normal and truncated cillum populations, as opposed to a generalized reduction in cillum length in mutant fibroblasts.

Ift172 mutant cilia demonstrate abnormal IFT88 and Gli2 localization

We predicted that Ift172 mutant cilia might have impaired intraflagellar transport, resulting in aberrant cillum structure and Hh signaling. To test this hypothesis, we performed immunocytochemistry with an antibody against aberrant cillum complex B, on wild-type, Ift172avc1 and Ift172avc1/wim cultured fibroblasts. It had been previously reported that IFT88 concentrates at two sites within the cillum, one at the cillum base and one at the distal tip (25). Interestingly, full-length cilia (>2 μm in length) demonstrated normal IFT88 distribution, with foci of staining at the distal and proximal ends of the axoneme, regardless of genotype (Fig. 7). We found that 92.4% of full-length wild-type cilia (Fig. 7A–D and M), 85.7% of full-length Ift172avc1 cilia (Fig. 7E–H and M) and 83.3% of full-length Ift172avc1/wim cilia (Fig. 7I–M) showed normal IFT88 distribution and there was no significant difference between the distribution of IFT88 in full-length cilia of all genotypes (wild-type versus Ift172avc1, P = 0.13; wild-type versus Ift172avc1/wim, P = 0.074). In contrast, the majority of truncated cilia showed only a single focus of IFT88 staining: only 42.1% of truncated Ift172avc1 cilia and 20.0% of truncated Ift172avc1/wim cilia showed the normal IFT88 distribution (wild-type versus avc1: P = 5.54 × 10−11 and wild-type versus avc1/wim: P = 1.53 × 10−25, respectively). Even truncated cilia on wild-type MEFs showed the normal distribution of IFT88 only 64.7% of the time, significantly less often than full-length wild-type cilia (P = 6.26 × 10−6). These results identified two distinct populations of primary cilia in cultured MEFs, full length and truncated, and documented abnormal IFT88 localization in the truncated population. A reduction in Ift172 dosage therefore established a truncated class of cilia with abnormal IFT.

We hypothesized that the observed IFT defect in Ift172 mutant cilia may impair the localization of Hh signaling components to the distal ciliary axoneme, providing a possible molecular explanation for the observed Hh signaling defects. We directly evaluated endogenous Gli2 localization by immunohistochemistry in wild-type and avc1 mutant MEFs to test this hypothesis. Gli2 undergoes localization to the distal tip of the ciliary axoneme by an Hh-dependent mechanism (34,35). We found that after the activation of Hh signaling with purmorphamine (PURM) (36), >60% of wild-type control MEFs contained concentrated Gli2 at the distal tip of the ciliary axoneme (Fig. 8A–D and I–L). There was no difference in the percentage of Gli2-positive wild-type full-length cilia (64 ± 3.5%) versus truncated cilia (65 ± 5.2%; P = 1). However, Gli2 localized to significantly fewer cilia in Ift172avc1 mutant MEFs after PURM treatment (Fig. 8Q). Compared with wild-type cilia, Gli2 localized to significantly
fewer full-length mutant cilia (41 ± 3.4%; P = 0.003) and truncated mutant cilia (28.5 ± 2.2%; P = 0.001) (Fig. 8E–H and M–Q). These results indicate that avcI impairs Hh-dependent Gli2 localization to the distal ciliary axoneme, a possible mechanism for the Hh signaling decrement observed in avcI mutant embryos.

**DISCUSSION**

We report that VACTERL-H may be added to the list of confirmed ciliopathies in mice, demonstrating that avcI, a novel hypomorphic allele of Ift72, results in VACTERL-H. We document abnormalities in cilia structure, intraflagellar transport and Hh signaling that provide mechanistic insight into how avcI causes the VACTERL-H phenotype. Our results provided empirical evidence for the previously suspected but unsubstantiated link between cilia, Hh signaling and VACTERL-H. We demonstrated for the first time the cellular mechanism causing a congenital heart defect in a cilia mutant: loss of an Hh-dependent lineage that generates the atrial septum, resulting in AVSDs with common atrium. Furthermore, by documenting the presence of a specific class of

Figure 7. Abnormal Ift88 localization in Ift172 mutant primary cilia. (A–M) Immunofluorescence was performed on serum-starved cultured fibroblasts from wild-type (A–D), Ift172avcI (E–H) and Ift172avcI/wim (I–L) embryos. Cilia were stained with acetylated tubulin (red) and IFT88 was stained with a specific anti-IFT88 antibody (green). Yellow arrows identify foci of IFT88 fluorescence. Size standard equals 3 μm. Cilia of each genotype were analyzed in two groups, full length (>2 μm) and truncated (<2 μm). The majority of full-length cilia of all genotypes displayed the normal distribution of IFT88, with one focus at the distal tip and one focus at the proximal tip (92.4% of wild-type cilia, 85.7% of Ift172avcI cilia and 83.3% of Ift172avcI/wim cilia) (M). There was not a significant difference in IFT88 distribution between any of these groups (wild-type versus Ift172avcI P = 0.13; wild-type versus Ift172avcI/wim P = 0.074; Ift172avcI versus Ift172avcI/wim P = 0.49) (M). Truncated mutant cilia revealed a different distribution (M); the majority of Ift172avcI truncated cilia (57.9%) and the majority of Ift172avcI/wim truncated cilia (80.0%) had a single focus of IFT88 staining. IFT88 distribution was significantly different between truncated cilia and full-length cilia of all genotypes. IFT88 distribution was also significantly different between truncated wild-type cilia and either truncated Ift172avcI cilia (P = 0.028) or truncated Ift172avcI/wim cilia (P = 1.65 × 10⁻⁶).
truncated cilia with disrupted IFT and Hh signaling in *avc1* mutants, we add a novel piece to the complex understanding of how cilia mutations impact the Hh signal transduction pathway and L–R determination.

The complete range of the VACTERL-H phenotypes is observed in *Ift172*mutant embryos with variable penetrance. Although several null alleles of IFT protein genes result in the absence of cilia, hypomorphic ciliary mutations allowing functional analysis of mutant cilia are less well described. One previously described hypomorphic allele of an IFT complex B protein, the ORPK allele of IFT88 (reviewed in 30), allows survival to adulthood and a milder subset of phenotypes than those caused by *avc1*. Like *avc1* mutants, ORPK mice have also been shown to have truncated cilia. The ‘gasping’ (*gsp*) phenotype reported by Ermakov et al. (37) demonstrates some similarities to VACTERL and to *avc1*, with abnormalities of foregut and hindgut, as well as polydactyly. The *gsp1*, *gsp3* and *gsp6* lines were all reported to have truncated nodal cilia, although the responsible gene has not been identified. Identification of *avc1* as a loss-of-function mutation in an IFT complex B gene provides the first identification of a monogenic cause for VACTERL-H association, which may now be considered a syndrome.

The *avc1* allele of *Ift172* decouples the role of the cilium in L–R patterning from its role in Hh signaling: L–R axis determination was preserved, whereas Hh signaling was significantly disrupted. The generation of L–R asymmetry is at least partially dependent on nodal flow established by rotating nodal monocilia (26), and L–R patterning abnormalities are common in cilia mutants (see reference 38 for partial list). For example,

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**Figure 8.** Abnormal Gli2 localization in truncated *avc1* mutant cilia. (A–P) Immunofluorescence was performed on serum-starved fibroblasts isolated from *Ift172*mutant embryos and their wild-type littermates. Fibroblasts were incubated with PURM to activate the Hh signaling pathway. Cilia were then stained with acetylated tubulin (green) and Gli2 with a specific anti-Gli2 antibody (red). Full-length and truncated cilia were analyzed separately. (Q) Approximately 64% of full-length or truncated wild-type cilia demonstrated Gli2 localization to the distal tip of the axoneme, whereas only 41% of full-length *avc1* mutant cilia (*P = 0.003) and 28% of truncated *avc1* mutant cilia (*P = 0.001) demonstrated similar localization.
Ift172<sup>wim</sup> mutants demonstrated absent nodal cilia and randomized L–R patterning [e.g. randomized heart looping (22)]. In contrast, Ift172<sup>avc1</sup> mutants demonstrated foreshortened nodal cilia but normal L–R patterning (e.g. normal heart looping in all cases). Our observations suggested that the proximal end of the cilium, maintained in avc1 mutants, may be sufficient to support normal rotation of nodal cilia and establishment of nodal flow, although insufficient to support normal Hh signal transduction. Although further studies are required to evaluate this hypothesis more definitively, this model could explain the decoupling of L–R axis formation from Hh signaling defects observed in avc1 mutant mice.

Our results suggest that decreased Ift172 dosage affects cilia synthesis in two distinct ways: (i) cells are less likely to produce cilia at all in the setting of decreased Ift172 dosage and (ii) the cilia that are formed are more likely to be truncated with abnormalities in IFT and Hh signaling. Fibroblasts with no Ift172 make no cilia, and carry out no Hh signaling transduction (22). Our analysis of the Ift172 allelic series utilizing our avc1 allele with the wim null allele of Ift172 suggests a dose-dependent requirement of Ift172 for the synthesis of functional primary cilia. Furthermore, we report that a hypomorphic mutation of Ift172, caused a ciliopathy with VACTERL-H-like features in mice. By providing a direct link between a single IFT gene mutation and VACTERL-H phenotypes, this work illustrates the diversity of phenotypes that can result from IFT gene mutations. IFT gene mutations have been significantly under-represented in the human ciliopathy literature. Establishing a Mendelian etiology for VACTERL-H in mice, we suggest that IFT genes, particularly IFT complex B genes including Ift172, be considered candidate genes for human VACTERL-H.

**MATERIALS AND METHODS**

**Mouse strains**

Avc1 was identified in a screen for recessive ENU-induced mutations that caused perinatal lethality and structural heart defects. Identification and characterization of wim has been previously described (22). The Gli1<sup>CreERT2</sup> line was obtained from the Joyner Laboratory (Sloan-Kettering Institute, NY, USA). Activation of CreER<sup>T2</sup> was accomplished by oral gavage with 2 µg of tamoxifen per dose in corn oil to pregnant dams, demonstrated to achieve optimal activation of CreER<sup>T2</sup> without toxicity (33).

**Patched1-LacZ** (B6;129-Petch<sup>tm1Mmps/J</sup>) and R26R (B6;129S4-Gt(ROSA)26Sor<sup>tm1Sor/J</sup>) mice were obtained from The Jackson Laboratory, and genotyping was performed as described (www.jax.org). All mouse experiments were performed in a mixed B6/129/SvEv background. All experiments involving mice were carried out according to a protocol reviewed and approved by the Institutional Animal Care and Use Committee of The University of Chicago, in compliance with the USA Public Health Service Policy on Humane Care and Use of Laboratory Animals.

**Genetic mapping and molecular identification of avc1**

SNPs that were previously identified as polymorphic between inbred strains of mouse, including C57BL/6J, FVB/NJ and C3H/FeJ (42) were genotyped on the Illumina platform (42) using DNA from affected mice. Additional polymorphic markers from publically available databases were utilized for high-resolution mapping. Avc1 was mapped to a 0.9 Mb interval on chromosome 5 between D5Mit229 and D5Mit420 in a mapping cross with more than 1000 opportunities for recombination. The only sequence alteration detected in the avc1-containing molecular interval was the A-to-G transition at position +3 in the Ift172 exon 24 splice donor site.
Embryo dissection and X-Gal staining

Embryos were dissected from maternal tissue samples and tail samples were taken for genotyping. Mice or dissected organs were then fixed for 1 h in 4% paraformaldehyde and stained with X-gal staining solution [5 mM K3Fe(CN)6, 5 mM K4Fe(CN)6, 2 mM MgCl2, 0.02% NP-40, 0.01% deoxycholate, 0.1% X-Gal in PBS] when appropriate or fixed in 10% neutral buffered formalin if no β-galactosidase-producing allele was present. To facilitate staining, embryos older than E10.5 were dissected open prior to staining. Mice were embedded in paraffin and sectioned at 5 μm. If X-gal stain was present, slides were counter-stained with 50% eosin for 1 s. If no X-gal stain was present, the slides were stained with hematoxylin and eosin.

Antibody production

Chemically synthesized peptides corresponding to KLRRDY YQWLMDTQQEER were sent to Pocono Rabbit Farm and Laboratory, Inc., for IFT172 antibody production in rabbits and antibody purification. The synthesized peptide corresponds to amino acids 738–755 of IFT172 encoded in exon 22 (NCBI m37 mouse assembly) and shared by both wild-type and Ift172avc1/wim mouse assembly) and shared by both wild-type and avc1/wim mouse lines.

Western blotting

Western blots were performed in triplicate. Mouse embryos were harvested in PBS at 4°C, an aliquot taken for genotyping and the remainder transferred into 0.5 ml of RIPA lysis buffer maintained at 4°C (Roche Complete protease inhibitor, 0.5 M Tris–HCl, pH 7.4, 1.5 mM NaCl, 2.5% deoxycholic acid, 10% NP-40, 10 mM EDTA). Tissues were lysed with rotor–stator-type homogenizer for 30 s. Lysates were rotated for 15 min and centrifuged for 15 min at 12,000g. The resulting supernatants were removed, assayed for protein concentration, normalized by the addition of RIPA buffer and diluted into Laemmli sample buffer (Bio-Rad, 161-0737EDU). The resulting samples were run into 8% Tris–glycine gels for SDS–PAGE and transferred to nitrocellulose blots for immunodetection. Following block (5% milk/TBST for 30 min) and wash (TBST 3×), primary antibodies (a gift of B. Wang) were added at 1:100 in 2.5% BSA/TBST and incubated overnight at 4°C. Alkaline phosphatase-conjugated secondary antibodies (Santa Cruz Biotechnology, Inc.) were each used at a dilution of 1:1000; rabbit anti-Ift88 (a gift from B. Yoder, University of Alabama, Birmingham, AL, USA) at a dilution of 1:500 and rabbit anti-Gli2 (a gift from Poatien Chuang) at a dilution of 1:400. Alexa488, Alexa568, goat anti-mouse AlexaFluor 488 and goat anti-rabbit AlexaFluor 594-conjugated secondary antibodies (Invitrogen) were each used at a dilution of 1:500.

Immunofluorescence

Cells were grown to confluence on eight well-chambered slides (Lab-Tek II). Fixation and immunofluorescence staining were performed as described by Ocbina and Anderson (28), with the following minor modifications: blocking solution was composed of PBS with 10% v/v normal donkey serum and 0.1% v/v Triton X-100. All antibodies were diluted in 1:1 blocking solution and PBS. The following primary antibodies were used: mouse anti-acetylated-alpha-tubulin (Sigma-Aldrich, T7451) at a dilution of 1:1000; rabbit anti-Ift88 (a gift from B. Yoder, University of Alabama, Birmingham, AL, USA) at a dilution of 1:500 and rabbit anti-Gli2 (a gift from Poatien Chuang) at a dilution of 1:400. Alexa488, Alexa568, goat anti-mouse AlexaFluor 488 and goat anti-rabbit AlexaFluor 594-conjugated secondary antibodies (Invitrogen) were each used at a dilution of 1:500.

Electron microscopy

E8.0 embryos were dissected in PBS at room temperature and immediately fixed in 2.5% glutaraldehyde and 2% PFA in 0.1 M sodium cacodylate buffer, pH 7.4 (Electron Microscopy Sciences), for at least 1 h at room temperature. Embryos were stored in a fixative for an additional 24 h at 4°C before processing as described in Huangfu and Anderson (44). Scanning electron micrograph images were taken on a Zeiss SUPRA 25 FESEM.

In situ hybridization

In situ hybridization was performed using digoxigenin-labeled probes. Protocol was as described in (45), with the following changes: embryos were washed six times in maelic acid buffer for 1 h (MAB; 0.1 M maelic acid, pH 7.5, 0.15 M NaCl, 0.1% Tween-20 and 0.002 M levamisole), followed by a 16 h overnight wash at room temperature. Color reactions were allowed to develop overnight, and images were taken prior to storage in 80% glycerol. The in situ probe for Ift172 was snap frozen in 80% glycerol for Gli1 as described (33).

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