Variable expressivity of ciliopathy neurological phenotypes that encompass Meckel–Gruber syndrome and Joubert syndrome is caused by complex de-regulated ciliogenesis, Shh and Wnt signalling defects

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The ciliopathies are a group of heterogeneous diseases with considerable variations in phenotype for allelic conditions such as Meckel–Gruber syndrome (MKS) and Joubert syndrome (JBTS) even at the inter-individual level within families. In humans, mutations in TMEM67 (also known as MKS3) cause both MKS and JBTS, with TMEM67 encoding the orphan receptor meckelin (TMEM67) that localizes to the ciliary transition zone. We now describe the Tmem67tm1(Dgen/H) knockout mouse model that recapitulates the brain phenotypic variability of these human ciliopathies, with categorization of Tmem67 mutant animals into two phenotypic groups. An MKS-like incipient congenic group (F6 to F10) manifested very variable neurological features (including exencephaly, and frontal/occipital encephalocele) that were associated with the loss of primary cilia, diminished Shh signalling and dorsalization of the caudal neural tube. The ‘MKS-like’ group also had high de-regulated canonical Wnt/β-catenin signalling associated with hyper-activated Dishevelled-1 (Dvl-1) localized to the basal body. Conversely, a second fully congenic group (F > 10) had less variable features pathognomonic for JBTS (including cerebellar hypoplasia), and retention of abnormal bulbous cilia associated with mild neural tube ventralization. The ‘JBTS-like’ group had de-regulated low levels of canonical Wnt signalling associated with the loss of Dvl-1 localization to the basal body. Our results suggest that modifier alleles partially determine the variation between MKS and JBTS, implicating the interaction between Dvl-1 and meckelin, or other components of the ciliary transition zone. The Tmem67tm1(Dgen/H) line is unique in modelling the variable expressivity of phenotypes in these two ciliopathies.

INTRODUCTION

Meckel–Gruber syndrome (MKS) is a lethal ciliopathy disorder that is usually diagnosed upon detection of a triad of manifestations that include renal cystic kidney dysplasia, polydactyly and neurodevelopment anomalies (1,2). The central nervous system (CNS) defects commonly observed in MKS include occipital encephalocele, rhombic roof dysgenesis and prosencephalon dysgenesis. The latter may include olfactory bulb dysgenesis, optic nerve hypoplasia, agenesis of the corpus callosum or total holoprosencephaly. These defects are thought to be caused by an underlying defect in ventral induction of the developing CNS by the prochordal mesoderm (3). Other occasional CNS features of MKS include...
microcephaly, cerebellar hypoplasia or total anencephaly (4). The human condition is both genetically and phenotypically heterogeneous, and displays marked phenotypic and genetic overlap with the allelic neurodevelopmental disorder Joubert syndrome (JBTS) (5). JBTS neurodevelopmental phenotypes involve cerebellar vermis hypoplasia or aplasia, a deep interpeduncular fossa and narrowing of the midbrain tegmentum. Collectively, these features are visualized on axial MRI with peduncular fossa and narrowing of the midbrain tegmentum.

To further elucidate the role of Shh and Wnt signalling in the severe human ciliopathies such as MKS with occipital encephalocele and JBTS, and the possible molecular basis for the variability of the neurodevelopmental anomalies, we studied embryonic neurodevelopmental, cilia and Wnt/β-catenin signalining phenotypes in the Tmem67−/− (Dgen/H) knockout mouse line (22). Tmem67 encodes Tmem67 (transmembrane protein 67 or meckelin), a 995 amino-acid transmembrane protein (Supplementary Material, Fig. S1A), with structural similarity to Frizzled receptors (23). Meckelin/TMEM67 contains an extracellular N-terminal domain with a highly conserved cysteine-rich repeat domain (CRD), a predicted β-pleated sheet region and seven predicted transmembrane regions (Supplementary Material, Fig. S1A). The knockout line recapitulated complex and variable features of both MKS-like and JBTS-like phenotypes: one group of incipient congenic animals developed frontal/occipital meningoencephalocele-like anomalies (Fig. 1), whereas a second group of fully congenic animals had a less variable range of features pathognomonic of JBTS (Fig. 2).

RESULTS

Tmem67−/− mutants display typical pathological features of Meckel–Gruber and Joubert syndromes

To study the mechanism of MKS and JBTS disease pathogenesis, we investigated the detailed neuroanatomical phenotypes of the Tmem67−/− (Dgen/H) targeted knockout mouse model. Reverse transcription (RT)–PCR analysis confirmed that targeted β-Gal-neo cassette insertion into Tmem67 abolished transcription of Tmem67 in Tmem67−/− embryos (Supplementary Material, Fig. S1B). Protein expression was assessed by immunoblotting cell lysates with a rabbit polyclonal antibody (‘Nt Ab’) against the Tmem67 CRD domain, which confirmed the absence of TMEM67 in Tmem67−/− embryos (Supplementary Material, Fig. S1C). Tmem67−/− mutants died by the first or second postnatal day (P0–P1) and recapitulated the histological features of MKS and JBTS (24), including multicystic renal dysplasia and embryonic developmental defects of the biliary tree (data not shown).

An MKS-like phenotype manifested in 35% of E11.5 embryos (n = 6/17; Table 1), of which n = 1/6 had an occipital meningocele that was detected externally as a bulge in the occipital area (Fig. 1A). Midbrain-hind brain exencephaly, with a failure of fusion of the neural folds, was noted in some E11.5 embryos (n = 4/17; Table 1, Fig. 1B), as was an anterior neuropore closure defect (n = 4/17; Table 1, Fig. 1C). At later developmental stages (from E13.5), frontal encephalocele had a variable degree of severity from a very mild defect (slight protrusion in the frontal area) to a gross encephalocele/meningocele-like anomaly seen in either a median or paramedian position (n = 5/19) (Fig. 1D and E). In the former type, the encephalocele protruded midline of the frontal area, while for the latter it protruded from one of the hemispheres (Fig. 1D). Prosencephalon dysgenesis was a consistent finding for all E11.5 embryos that could be diagnosed with MKS-like features (n = 7/7; Table 1). At later developmental stages, this manifested as semi-lobar holoprosencephaly in which the two lateral ventricles were fused with an absence of some midline structures such as the anterior commissar and variable degrees of corpus callosum dysgenesis (Fig. 1F). The ‘MKS-like’ group of animals also had enlargement of the hippocampus (25–27) and basal ganglia (28) (Fig. 1F), both suggestive of an up-regulation of canonical β-catenin–mediated Wnt signalling. IHC staining of encephalocele tissue demonstrated an increased level of activated β-catenin (Fig. 1G), and a mixed population of post-mitotic and progenitor neurons as indicated by the neuronal differentiation marker Tuj1 (Fig. 1G).

The second, JBTS-like phenotype manifested in 58% of later stage Tmem67−/− embryos and pups (n = 11/19; Table 2). The JBTS group of Tmem67−/− mutant embryos had no overt neural tube defect, but all animals had a reduced anteroposterior axis of the developing forebrain (Table 2), in addition to a small
hindbrain region, microcephaly or other overt facial dysmorphologies (Fig. 2A). Some embryos in the JBTS group of Tmem67²/² mutants had a deep interpenduncular fossa with a reduced anterioposterior axis of the midbrain tegmentum at the level of the isthmus (Fig. 2A), accompanied by cerebellar vermis hypoplasia or aplasia (Fig. 2B and C). These features comprise the MTS that is pathognomonic for JBTS in humans. Dilatation of the fourth ventricle was not noted in this group, but complex posterior fossa defects (Fig. 2D) and features compatible with the Dandy–Walker malformation (Supplementary Material, Fig. S2A) were seen, all of which are reported in human patients either in combination with the diagnostic MTS or as separate anomalies in the so-called ‘Joubert syndrome mimicry’ group (29).

In general, incipient congenic embryos (F6–F10 filial generations on the C57BL/6J background) manifested a very...
variable MKS-like phenotype, whereas fully congenic animals (F > 10) had the less variable JBTS-like phenotype (Table 2; P < 0.001, Chi-squared test). However, we also noted inter-individual variation in these phenotypes, even for Tmem67−/− mutant littermates (Supplementary Material, Fig. S2B).
The MKS-like phenotype is associated with cilia loss and caudal neural tube dorsalization, whereas the JBTS-like phenotype is associated with aberrant cilia structure and mild neural tube ventralization

To investigate ciliary phenotypes in ‘MKS-like’ Tmem67−/− mutants, we stained caudal neural tube sections from E11.5 and E14.5 embryos for primary cilia and basal bodies. This revealed near total loss of cilia at the developing neuroepithelial cell layer (Fig. 3A and B), confirmed by in vivo SEM of the ependymal cell layer of Tmem67−/− E18.5 lateral ventricle (Fig. 3C). In contrast, the JBTS group of mutant animals retained primary cilia at the E11.5 neuroepithelial cell layer of the caudal neural tube (Fig. 3D), and both IF microscopy and SEM revealed that ependymal cells had abnormal, bulbous cilia (Fig. 3E). Furthermore, in both the MKS-like and JBTS-like disease states, the planar organization of the ependymal cell layer was disrupted (Fig. 3C and E).

Loss of cilia causes suppression of the Shh pathway, since the activation of Shh signalling requires the translocation of Smo to the primary cilia following binding of Shh ligand to the Patched receptor (30). We therefore investigated the expression of the Shh protein and dorsoventral neural tube markers in the caudal neural tube of E11.5 Tmem67−/− embryos with an MKS-like phenotype. Embryos had a variable, thinning defect of the roof plate, Shh protein expression was reduced at the mutant floor plate and notochord, and a reduced number of cells expressing the floor plate marker Foxa2 (Fig. 4A). Cells labelled with the ventral markers Nkx2.1 and Nkx6.1 were also reduced in number and more ventrally located in the ‘MKS-like’ Tmem67−/− neural tube (Fig. 4A). There was a slight increase and ventral expansion of cells labelled with the dorsal marker Pax6, but localization of Pax7 expression was unaffected (Fig. 4A). Together, these data indicated a dorsalization of the neural tube in Tmem67−/− E11.5 embryos of the MKS-like group of mutants, mostly likely due to either the lack of cilia or lack of functioning cilia that culminated in reduced Shh signalling.

In contrast, dorsoventral patterning of the caudal neural tube of E11.5 Tmem67−/− embryos with the JBTS-like phenotype did not have a visible roof plate defect and had robust expression of the floor plate marker Foxa2 (Fig. 4B). There was slight dorsal expansion for the ventral markers Nkx2.1 and Nkx6.1, and the dorsal marker Pax6 (Fig. 4B), indicating a mild ventralization of the neural tube in this group of mutant embryos which suggested that the bulbous ‘JBTS-like’ Tmem67−/− cilia (Fig. 3E) failed to constrain Shh signalling.

The MKS-like phenotype is associated with high de-regulated Wnt/β-catenin signalling

To investigate in vivo canonical Wnt (β-catenin-associated) signalling in ‘MKS-like’ Tmem67−/− mutants, we stained a meningocele-like anomaly (Fig. 5A) or abnormal nodular masses in the lateral ventricle (Supplementary Material, Fig. S3A) for IF microscopy. This revealed extensive increases in the overall levels of β-catenin (Fig. 5A, Supplementary Material, Fig. S3A) and near total loss of cilia at the developing neuroepithelial cell layer of the developing fourth ventricle (Fig. 5A). Although some primary cilia were still seen in the surrounding normal head mesoderm, cilia were lost from the cells forming the meningocele like defect (Fig. 5A).

Mouse embryonic fibroblasts (MEFs) derived from MKS-like mutant animals lacked primary cilia, or developed very short (≏1 μm) cilia in 5% of cells, whereas 70% of Tmem67+/− MEFs had a single, uniform cilium of ~2 μm in length (Fig. 5B). Tmem67−/− MEFs also developed a range of centrosomal defects comprising widely separated mother/daughter centrosomes and multiple centrosomes (Fig. 5B), consistent with previously reported defects following TMEM67/meckelin loss (31).

Western blot analysis of whole cell extracts (WCE) from MEFs confirmed higher basal levels of active, non-phosphorylated β-catenin, cyclin D1 (a downstream target of the Wnt/β catenin pathway) and phosphorylated Dishevelled-1 (Dvl-1, a key signalling modulator of Wnt pathways) (32–34) in Tmem67−/− compared with either Tmem67+/− or Tmem67+/+ cells (Fig. 5C, Supplementary Material, Fig. S3B). Levels of phosphorylated Lrp6, a key co-receptor of the canonical Wnt/β catenin pathway, were markedly increased in Tmem67−/− compared with Tmem67+/+ MEFs.

Table 1. Summary of neurodevelopmental anomalies detected in Tmem67−/− mutants at the E11.5 stage of development

<table>
<thead>
<tr>
<th>Phenotype(s)</th>
<th>Tmem67−/− mutant animals</th>
<th>Total/17</th>
</tr>
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<tbody>
<tr>
<td></td>
<td>F6–F10</td>
<td></td>
</tr>
<tr>
<td></td>
<td>1            2  3  4  5  6  7  8  9  10  11  12  13  14  15  16  17</td>
<td></td>
</tr>
<tr>
<td>Meningocele-like</td>
<td>✓            ✓</td>
<td>1</td>
</tr>
<tr>
<td>MB exencephaly</td>
<td>✓            ✓            ✓   ✓     ✓</td>
<td>4</td>
</tr>
<tr>
<td>Rh. dysgenesis</td>
<td>✓            ✓            ✓   ✓     ✓</td>
<td>4</td>
</tr>
<tr>
<td>Expanded roof plate</td>
<td>✓            ✓            ✓   ✓     ✓</td>
<td>11</td>
</tr>
<tr>
<td>Forebrain</td>
<td>✓            ✓            ✓   ✓     ✓</td>
<td>11</td>
</tr>
<tr>
<td>Prog, dysgenesis</td>
<td>✓            ✓            ✓   ✓     ✓</td>
<td>3</td>
</tr>
<tr>
<td>Pros. normal but small</td>
<td>✓            ✓            ✓   ✓     ✓</td>
<td>4</td>
</tr>
<tr>
<td>sNTD</td>
<td>✓            ✓            ✓   ✓     ✓</td>
<td>4</td>
</tr>
<tr>
<td>Phenotype group</td>
<td>M  M  M  M  M – – M – – M – – – – – –</td>
<td></td>
</tr>
</tbody>
</table>

In total, 17 Tmem67−/− animals at E11.5 were examined morphologically or histologically, out of 49 animals of all genotypes from 5 timed matings. Individual animals (numbered 1–17) are separated on the basis of filial generation (F6–F10, or F—), as indicated) after back-crossing onto the C57BL/6J genetic background. The bottom row indicates if each individual animal had features compatible with MKS (M), or if a definitive diagnosis could not be made at this stage of development (—). Abbreviation: M, MKS-like phenotype group; MB, midbrain; Rh, thombencephalon; Pros, prosencephalon; sNTD, spinal neural tube defect.
following treatment with conditioned medium containing Wnt3a, a Wnt ligand known to activate canonical Wnt signalling (Fig. 5D). IF staining of β-catenin in MEFs detected higher levels of nuclear localization of active β-catenin in Tmem67−/− compared with Tmem67+/+ MEFs following treatment with Wnt3a (Supplementary Material, Fig. S3C). Jouberin, a positive regulator of the canonical pathway that binds to β-catenin (35), also translocated to nuclei in this group of Tmem67−/− mutant cells (Supplementary Material, Fig. S3D). Finally, levels of active RhoA (a downstream mediator of the non-canonical Wnt signalling pathway) were reduced in Tmem67−/− compared with Tmem67+/+ MEFs but could be increased by treatment with the non-canonical ligand Wnt5a (Fig. 5E).

We also used the TOPFlash assay to quantify the ability of Tmem67+/+ and Tmem67−/− MEFs to respond to Wnt ligands. After transfection of the reporter constructs, treatment with Wnt3a stimulated basal levels of Wnt/β-catenin signalling by 5.3-fold in Tmem67+/+ MEFS, but by 34.2-fold in Tmem67−/− MEFS following treatment with Wnt3a-conditioned medium (Fig. 6C). In contrast to ‘MKS-like’ MEFs (Fig. 5D), Wnt3a did not stimulate an increase in levels of β-catenin (Fig. 6D). Although Wnt3a stimulated increased levels of both non-phosphorylated and activated, phosphorylated Dvl-1 in ‘MKS-like’ MEFs (Supplementary Material, Fig. S3B), ‘JBTS-like’ Tmem67−/− MEFs down-regulated levels of Dvl-1 in response to this treatment (Fig. 6D). However, treatment of MEFs with Wnt5a induced the formation of prominent actin stress fibres in Tmem67−/− but not Tmem67+/+ cells (Fig. 6E). Active RhoA levels were slightly increased in Tmem67−/− compared with Tmem67+/+ cells, but were not affected by Wnt5a (Fig. 6E).

To further characterize the levels of Shh and Wnt/β-catenin signalling during neurodevelopment of ‘JBTS-like’ Tmem67−/− embryos, we used quantitative real-time PCR to measure the levels of Shh and Axin2 transcripts in neocortical tissue dissected from E12.5, E15.5 and P0 animals. At E12.5, there was a statistically significant increase in Shh levels in Tmem67−/− neocortex compared with Tmem67+/+ age-matched wild-type littermates (Fig. 6F), consistent with a loss of constraint for Shh signalling that could also cause the mild ventralization of the neural tube in ‘JBTS-like’ Tmem67−/− mutants (Fig. 4C). Conversely, canonical Wnt/β-catenin signalling (as measured by Axin2, a downstream target of the pathway) was significantly decreased in the P0 mutant cortex (Fig. 6F), consistent with the results in ‘JBTS-like’ MEFs (Fig. 6B–D).

### Table 2. Summary of neurodevelopmental anomalies detected in Tmem67−/− mutants at the E13.5 to P0 stages of development

<table>
<thead>
<tr>
<th>Phenotype(s)</th>
<th>F6–F10</th>
<th>F11</th>
<th>F12</th>
<th>F13</th>
<th>F14</th>
<th>F15</th>
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<th>F17</th>
<th>F18</th>
<th>F19</th>
<th>Total/19</th>
</tr>
</thead>
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<tr>
<td>Forebrain</td>
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<td>✓</td>
<td>✓</td>
<td>✓</td>
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<td>✓</td>
<td>✓</td>
<td>✓</td>
<td>✓</td>
<td>✓</td>
<td>5</td>
</tr>
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<td>Frontal meningocoele/encephalocele</td>
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<td>✓</td>
<td>✓</td>
<td>✓</td>
<td>✓</td>
<td>✓</td>
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<td>✓</td>
<td>✓</td>
<td>5</td>
</tr>
<tr>
<td>Occipital defect</td>
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<td>✓</td>
<td>✓</td>
<td>✓</td>
<td>✓</td>
<td>✓</td>
<td>5</td>
</tr>
<tr>
<td>Small forebrain</td>
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<td>✓</td>
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<td>✓</td>
<td>✓</td>
<td>✓</td>
<td>✓</td>
<td>✓</td>
<td>✓</td>
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</tr>
<tr>
<td>Enlarged forebrain</td>
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<td>✓</td>
<td>✓</td>
<td>✓</td>
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<td>✓</td>
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<td>Posterior fossa defects</td>
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<td>5</td>
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<tr>
<td>Cerebellar hypoplasia</td>
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<td>✓</td>
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<td>5</td>
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<tr>
<td>Deep interpeduncular fossa</td>
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<td>✓</td>
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<td>✓</td>
<td>✓</td>
<td>✓</td>
<td>✓</td>
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 Phenotype group: J, JBTS-like phenotype group; M, MKS-like phenotype group.

*Embryo sections shown in Supplementary Material, Fig. S2A.*
**DISCUSSION**

Ciliopathies are a group of developmental disorders with overlapping clinical phenotypes, but for allelic ciliopathies such as MKS and JBTS the mechanism of the variability of phenotype expressivity remains unclear. Allelism at a primary causative locus is not thought to solely explain this variability, implicating the effect of additional modifier alleles (38). We now describe the study of the Tmem67tm1(Dgen/H) mouse model for both MKS and JBTS, with targeted mutagenesis of the Tmem67 gene (also known as Mks3; Supplementary Material, Fig. S1A). Tmem67−/− mutants on an incipient congenic (F6–F10) C57BL/6J genetic background had typical ciliopathy features, comprising renal cystic dysplasia and hepatic developmental defects that are diagnostic for MKS in human patients. Tmem67−/− mutants also manifested a complex and variable suite of neurodevelopmental phenotypes (Table 1 and 2), in contrast to the mild hydrocephaly reported for both the Wpk rat (39) and the bpck mouse models (40). In fully congenic (F > 10) embryos and pups that survived to P0 or P1, neurodevelopmental phenotypes were less variable and pathognomonic for JBTS (Table 2). The Tmem67−/− embryos. Magnified insets (orange frames) show detail of cilia (arrowheads). Scale bar = 50 µm with 10 µm subdivisions.

**Figure 3.** Different cilia defects in Tmem67−/− embryos with MKS-like and JBTS-like phenotypes: (A, top panels) Horizontal section of the developing E11.5 caudal neural tube in litter-mate Tmem67+/+ and ‘MKS-like’ Tmem67−/− embryos at the level of the anterior limb buds stained for acetylated α-tubulin (red), centrosomes/basal bodies (γ-tubulin, green) and nuclei (DAPI, blue). (Bottom panels) Magnified insets (white frames) with primary cilia in the Tmem67+/+ and ‘MKS-like’ Tmem67−/− embryo stained as for (A). Scale bar = 10 µm. (B) E14.5 neuroepithelium of the lateral wall of the third ventricle (3V) in litter-mate Tmem67+/+ and ‘MKS-like’ Tmem67−/− embryo stained as for (A). Scale bar = 10 µm. (C) SEM images of E18.5 lateral ventricles showing loss of primary cilia and disruption in the planar polarization of the ependymal cell layer in an ‘MKS-like’ Tmem67−/− embryo. Magnified insets (orange frames) show detail of cilia (arrowheads) in a littermate Tmem67+/+ embryo. Scale bar = 50 µm with 10 µm subdivisions. (D, top panels) Horizontal sections as in (A) for littermate Tmem67+/+ and ‘JBTS-like’ Tmem67−/− embryo stained for acetylated α-tubulin (red), centrosomes/basal bodies (γ-tubulin, green) and nuclei (DAPI, blue). (Bottom panels) Magnified insets (white frames) with primary cilia indicated by arrowheads. Scale bar = 10 µm. (E) IF microscopy (top panels) and SEM images (bottom panels) of E18.5 lateral ventricles showing the presence of abnormal, bulbous cilia and defective planar organization of the ependymal layer in ‘JBTS-like’ Tmem67−/− embryos. Magnified insets (orange frames) show detail of cilia (arrowheads). Scale bar = 50 µm with 10 µm subdivisions.
mouse model has been described previously (22), and in the present study, Tmem67-/- mouse pups died at birth or during the early postnatal period, consistent with previous results (22). However, in contrast to our study, Garcia-Gonzalo et al. (22) described Tmem67-/- mutants with a normal external morphology and manifesting renal cystic dysplasia at E18.5, but did not perform extensive analyses of neurodevelopmental phenotypes and did not note the variability of the Tmem67-/- phenotype.

We separated the neurodevelopmental phenotypes in Tmem67-/- mutants on the basis of severity. The first group had an ‘MKS-like’ phenotype with the presence of cranial neural tube defects early in development (Fig. 1A–C), and the later development of a frontal encephalocele/meningocele (Fig. 1E) and prosencephalon dysgenesis (Fig. 1F), all of which are features of human MKS (3,41). The second ‘JBTS-like’ group had milder defects of the posterior fossa resembling clinical features of JBTS patients (Fig. 2B, Supplementary Material, Fig. S2A). Furthermore, there was inter-individual variation in these phenotypes, even for Tmem67-/- mutant littermates (Supplementary Material, Fig. S2B). The Tmem67 mouse is therefore a unique model for severe human ciliopathies and their phenotypic variability, since the Tmem67-/- mutants fully recapitulate the human MKS type 3 clinical presentation (42) and vermian hypoplasia (Fig. 2D), an essential diagnostic finding in JBTS patients (43).

In the present study, we now show that TMEM67/meckelin has complex roles in regulating canonical Wnt/β-catenin and Shh signalling in the developing brain. TMEM67/meckelin

Figure 4. Defective dorsoventral patterning and Shh signalling in Tmem67-/- mutants: (A) loss of Shh signalling and dorsalization of the caudal neural tube in ‘MKS-like’ embryos. (Top panel) Shh protein expression (red) at the notochord (nc, arrowheads) and induced expression at the floor plate (FP) in the Tmem67+/+ embryo. (Bottom panels) IHC staining for the indicated dorsoventral patterning markers and haemotoxylin counterstaining. Note the variable thinning of the roof plate (RP) in Tmem67-/- embryos. Scale bar = 100 μm. (B) Activation of the Shh signalling pathway and mild ventralization defects in ‘JBTS-like’ embryos using IHC staining for the indicated dorsoventral patterning markers. Note the absence of any roof plate defect in Tmem67-/- embryos.
is required for correct ciliogenesis in the CNS, and in MEFs derived from ‘MKS-like’ (Fig. 1) and ‘JBTS-like’ (Fig. 2) groups of animals: the MKS-like group failed to form cilia (Figs 3A–C, 5A and B), whereas a proportion of cilia in the JBTS-like group did form (Fig. 3D) but were structurally abnormal (Figs 3E and 6A). These ciliary phenotypes directly correlated with the type of de-regulation that we observed for both canonical Wnt/β-catenin and Shh signalling in these two groups. In the ‘MKS-like’ group, TMEM67/meckelin loss caused a very high de-regulated level of Wnt/β-catenin signalling both in vivo (Figs 1G and 5A; Supplementary Material, Fig. S3A) and in vitro (Fig. 5C, D, F). This is entirely consistent with previous reports that cilia act as a negative regulator of the canonical Wnt/β-catenin signalling (18,19,44). In contrast, cells that grew abnormally long cilia failed to activate the canonical pathway (Fig. 6B–D) and β-catenin translocation to the nucleus was disturbed (Fig. 6B), indicating that active β-catenin translocation into the nucleus is a process that requires a functioning cilium in the JBTS-like disease state. This finding is also consistent with a previous report describing a mouse model of JBTS, since the Ahi1 mouse model developed reduced canonical...
Wnt signalling accompanied with reduced proliferation in the developing cerebellum, although cilia morphology was not affected in this model (45). The aberrant down-regulation of canonical Wnt/β-catenin signalling in Tmem67−/− during later development (Fig. 6F), therefore, provides an explanation of the vermician hypoplasia in the ‘JBTS-like’ group.

Neuroepithelial cells of the ventricular zone in the developing ‘MKS-like’ Tmem67−/− E11.5 neural tube did not form cilia (Fig. 3A), and, as expected, we saw a reduction in the number of ventral neuronal and floor plate cell types in the cervical neural tube (Fig. 4A). The loss of cilia in the ‘MKS-like’ group therefore suppressed the Shh pathway (30), leading to typical ciliopathy mouse mutant phenotypes of the vermian hypoplasia in the ‘JBTS-like’ group. In the later development (Fig. 6F), therefore, provides an explanation for the variability of phenotype expressivity in Tmem67−/− embryos, and the differing Shh and Wnt signalling responses in the ‘MKS-like’ and ‘JBTS-like’ phenotypic groups. In support of this observation, recent studies have described both general and specific buffering or compensatory systems that combine to determine the outcome of an inherited mutation leading to inter-individual variation (53). The origin of inter-individual variation in levels of gene expression includes ancestral gene duplications and marked variation in the induction of the molecular chaperones such as Hsp90 that appear to act as random buffers of genetic variation (54). However, such mechanisms do not preclude the effect of environmental factors on inter-individual levels of gene expression. We also noted that more severely affected Tmem67−/− embryos were invariably at distal locations in the left and right uterine horns of pregnant dams, suggesting an effect due to variations in placental blood supply (data not shown). The molecular causes of the switch between aberrant up- and down-regulation of signalling in the Tmem67 knock-out line remain unknown, but are likely to include the effect of modifier alleles and stochastic effects that are likely to be context-dependent or highly sensitive to any environmental cues. We speculate that modifier alleles in the TMEM67/meckelin-Dvl-1 signalling axis, or other signalling modulators at the transition zone, may contribute to this phenotypic variation. The congenic Tmem67 line therefore provides a unique model system to elucidate the pathogenesis of MKS and JBTS phenotypic variability.

MATERIALS AND METHODS

Animals

The animal studies described in this paper were carried out under the guidance issued by the Medical Research Council in Responsibility in the Use of Animals for Medical Research (July 1993) in accordance with UK Home Office regulations under the Project Licence No. PPL/40/3349. C57BL/6J;129P2-Tmem67mel1(Dgen1b) heterozygous knock-out mice were derived from a line generated by Deltagen Inc. and made available from MRC Harwell through the European...
Mutant Mouse Archive, http://www.emmanet.org/ (strain number EM:02370). The targeted β-Gal-neo (‘geo’) construct inserts downstream of exon one of the Tmem67 gene (Supplementary Material, Fig. S1A). Genotyping was done by PCR on DNA extracted from tail tips or the yolk sac of the E11.5 embryos, or tail or ear biopsies of older embryos and adult mice, respectively. Animals were back-crossed onto the C57BL/6J genetic background for five filial generations (F5) before performing the anatomical and functional characterization.

Preparation of tissue sections, histology and immunohistochemistry

Mouse embryos or dissected tissues were fixed in 4% (w/v) para-formaldehyde and embedded in paraffin wax. Thin sections (4 μm) were cut onto ‘Superfrost Plus’ slides (VWR International Ltd.) and were deparaffinized and rehydrated by standard methods. Sections were stained with haematoxylin and eosin (BDH Chemicals Ltd.) for 2 min, then dehydrated in ethanol, cleared in xylene and mounted in DPX. For immunohistochemistry, tissue sections were deparaffinized and rehydrated through passing in xylene and descending grades of ethanol. Epitope recovery was obtained by boiling in 1 mM EDTA pH 8.0 for 2 min using pressure cooker, followed by 30 min cooling. Blocking and application of primary antibodies was as described (55). Appropriate HRP-conjugated secondary antibodies (Dako Inc.) were used (final dilutions of ×10 000–25 000). Sections were developed in ‘Sigma Fast’ 3,3′-diaminobenzidine (DAB) with CoCl₂ enhancer and counterstained with Mayer’s haematoxylin (Sigma-Aldrich Co. Ltd.).
Cells

Human embryonic kidney (HEK293) cells were grown in Dulbecco’s minimum essential medium (DMEM)/Ham’s F12 supplemented with 10% fetal calf serum at 37°C/5% CO₂. The derivation and culture of MEFs have been described previously (56). The MEFs were derived from E15.5 embryos in both phenotype groups, using identically similar experimental procedures, and assayed consistently at identical passage numbers of 2 or 3. Three separate MEF lines for both phenotype groups were derived and used in assays. MEFs were grown in DMEM/Ham’s F12 supplemented with 10% fetal calf serum and 1% penicillin streptomycin at 37°C/5% CO₂.

Cloning, plasmid constructs and transfections

Full-length human MKS3 was cloned into the pCMV-HA vector as described previously (37) and verified by bidirectional DNA sequencing. For transfection with plasmids, cells at 80% confluency were transfected using Lipofectamine 2000 (Invitrogen Inc.) according to the manufacturer’s instructions and as described previously (57).

Antibodies

The following primary antibodies were used: mouse anti-acetylated-a-tubulin clone 6-11B-1 (Sigma-Aldrich Co. Ltd.); rabbit-anti-a-tubulin and mouse anti-b actin clone AC-15 (Abcam Ltd.); mouse anti-Dvl-1 clone 3F12 and mouse anti-cyclin D1 clone A-12 (Santa Cruz Biotechnology Inc.); mouse anti-activated b-catenin (anti-ABC, clone 8E7) and mouse anti-Ki67 (Merck Millipore Inc.); and mouse anti-neuron-specific a3-tubulin (TuJ1) clone TU-20, rabbit anti-phospho-LRP6 (Ser1490), rabbit anti-b-catenin (Cell Signaling Technology Inc.). The monoclonal antibodies anti-Shh clone 5E1, anti Fox2A clone 4C7, anti-Pax6, anti-Pax7, anti-NKx6.1 clone F55A10 and anti-NKx2.2 clone 74.5A5 were obtained from the Developmental Studies Hybridoma Bank developed under the auspices of the NICHD and maintained by the University of Iowa, Department of Biology, Iowa City, USA. Rabbit anti-TMEM67/meckelin N-terminus or C-terminus (TMEM67 Nt or Ct, ‘Nt Ab’ or ‘Ct Ab’ in Supplementary Material, Fig. S1A) have been described previously (57). Secondary antibodies were Alexa-Fluor 350-, Alexa-Fluor 488-, Alexa-Fluor 568- and Alexa-Fluor 594-conjugated goat anti-mouse IgG and goat anti-rabbit IgG (Molecular Probes Inc.). Alexa-Fluor 488 phalloidin conjugate (Life Technologies Corp.) was used to visualize F-actin.

Immunofluorescence and confocal microscopy

Cells were seeded at 1.5 × 10⁵ cells/well on glass cover slips in six-well plates and fixed in ice-cold methanol (5 min at −20°C) or 2% paraformaldehyde (20 min at room temperature). Permeabilization, blocking methods and immunofluorescence staining were essentially as described previously (9). Primary antibodies were used at final dilutions of ×200–1000. Secondary antibodies were diluted ×500. Confocal images were obtained using a Nikon Eclipse TE2000-E system, controlled and processed by EZ-C1 3.50 (Nikon Inc.) software. Images were assembled using Adobe Illustrator CS2.

WCE preparation and immunoblotting

WCE from confluent MEFs were prepared by standard methods. 10 μg WCE total soluble protein was analysed by SDS–PAGE (using 4−12% acrylamide gradient gels) and western blotting according to standard protocols using either rabbit polyclonal antisera (final dilutions of ×200–1000) or mAbs (×1000–5000). Appropriate HRP-conjugated secondary antibodies (Dako Inc.) were used (final dilutions of ×10 000–25 000) for detection by the enhanced chemiluminescence ‘Femto West’ western blotting detection system (Pierce Inc.). Levels of b-actin were used as a loading control for all immunoblotting experiments.

Canonical Wnt activity (TOPFlash) luciferase assays

MEFs were co-transfected with 0.5 μg TOPFlash firefly luciferase construct (or FOPFlash, as a negative control); 0.5 μg of expression constructs (pCMV-HA-TMEM67, or empty pCMV-HA control); and 0.05 μg of pRL-TK internal control Renilla luciferase reporter construct (Promega Corp). Cells were treated with Wnt3a- or Wnt5a-conditioned media (58). Luciferase activities were assayed with the Dual-Luciferase Reporter Assay system (Promega Corp.) on a Mithras LB940 (Berthold Technologies Inc.) luminometer. Raw readings were normalized with Renilla luciferase values, and the results reported are from at least four independent biological replicates.

RhoA activation assays

The activated GTP-bound isoform of RhoA was specifically assayed in pull-down assays using a GST fusion protein of the Rho effector rhokin (Cytoskeleton, Inc.), using conditions recommended by the manufacturers, and as described previously (37).

Gene expression analyses by quantitative real-time PCR

For the relative quantification of gene expression, we used the standard curve method of quantitative real-time PCR. Total RNA (1 μg) was reverse transcribed using the Superscript III First-Strand cDNA System (Invitrogen Inc.). PCR analysis of cDNA was performed using dHPLC-purified primers (Sigma-Aldrich Ltd.) specific for mouse Shh and Axin2. Primer sequences are available upon request. Amplimer levels were quantified contiguously with the SYBR GreenER qPCR system (Invitrogen Inc.) using an ABI 7500 instrument, as described previously (8). Beta-actin was amplified for normalization.

Statistical analyses

Normal distribution of data (TOPFlash activities, cilia length measurements) was confirmed using the Kolmogorov–Smirnov test (GraphPad Prism). Pairwise comparisons were analysed with Student’s two-tailed t-test using InStat.
(GraphPad Prism). Results reported are from at least three independent biological replicates. Error bars on bar graphs indicate SEM. The statistical significance of pairwise comparisons shown on bar graphs is indicated by: n.s., not significant, *P < 0.05, **P < 0.01, ***P < 0.001 and ****P < 0.0001. For cell populations, a minimum of 150 cells were counted from 10 separate fields of view. Contingency tables were analysed with Pearson’s chi-squared test, using two degrees of freedom (GraphPad Prism).

SUPPLEMENTARY MATERIAL
Supplementary Material is available at HMG online.

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Conflict of Interest statement. None declared.

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