Genome-wide association and longitudinal analyses reveal genetic loci linking pubertal height growth, pubertal timing and childhood adiposity


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The pubertal height growth spurt is a distinctive feature of childhood growth reflecting both the central onset of puberty and local growth factors. Although little is known about the underlying genetics, growth variability during puberty correlates with adult risks for hormone-dependent cancer and adverse cardiometabolic health. The only gene so far associated with pubertal height growth, LIN28B, pleiotropically influences childhood growth, puberty and cancer progression, pointing to shared underlying mechanisms. To discover genetic loci influencing pubertal height and growth and to place them in context of overall growth and maturation, we performed genome-wide association meta-analyses in 18,737 European samples utilizing longitudinally collected height measurements. We found significant associations ($P < 1.67 \times 10^{-8}$) at 10 loci, including LIN28B. Five loci associated with pubertal timing, all impacting multiple aspects of growth. In particular, a novel variant correlated with expression of MAPK3, and associated both with increased prepubertal growth and earlier menarche. Another variant near ADCY3-POMC associated with increased body mass index, reduced pubertal growth and earlier puberty. Whereas epidemiological correlations suggest that early puberty marks a pathway from rapid prepubertal growth to reduced final height and adult obesity, our study shows that individual loci associating with pubertal growth have variable longitudinal growth patterns that may differ from epidemiological observations. Overall, this study uncovers part of the complex genetic architecture linking pubertal height growth, the timing of puberty and childhood obesity and provides new information to pinpoint processes linking these traits.
INTRODUCTION

Postnatal height growth is a heritable complex process characterized by rapid infant growth, slowly diminishing mid-childhood growth and a distinct pubertal height growth spurt. Whereas the genetics of adult stature has been evaluated in large-scale genome-wide association (GWA) analyses (1), few studies have addressed the molecular underpinnings of distinct growth phases. Moreover, specific growth patterns during childhood correlate with both altered pubertal timing and adult health risks. For example, increased height and body mass index (BMI) prior to puberty correlate with advanced pubertal onset (2–4), and early puberty associates with increased risk of adult obesity and related metabolic traits (5–7). Still, the specific mechanisms linking these traits remain elusive.

To elucidate part of the genetic architecture impacting adolescent growth, we focussed primarily on the dynamic and highly variable pubertal growth spurt that reflects both the activation of central puberty and local growth factors (8,9) while accounting for up to 15–20% of adult stature (10). Narrow-sense heritability estimates place the genetic contribution to variation in pubertal growth between 60 and 90% (11–13), and twin studies suggest a substantial proportion of shared genetic variance with other phases of childhood growth (13). Specifically, we aimed to (i) identify genetic variants associated with the onset, total magnitude and tail end of the pubertal growth spurt and (ii) investigate these variants’ longitudinal effects on overall childhood growth and the timing of puberty (study design outlined in Fig. 1).

Due to large variation in the timing and rate of the pubertal growth spurt (shown schematically in Fig. 2), an accurate model typically requires frequent height measurements spanning a large age range, often difficult to obtain. Furthermore, girls enter puberty and, thus, begin their growth spurt, an average of 2 years earlier than boys. Taking these challenges into consideration, we aimed to characterize loci influencing growth during puberty by leveraging heterogeneous height measurements taken at varied ages throughout childhood across participating cohorts to maximize statistical power. Therefore, we modelled the pubertal height growth spurt for GWA using three partially correlated simple measures (Supplementary Material, Table S1; Fig. 2) that also partly reflect the timing of puberty (14). In Analysis I, we targeted the take-off phase of the growth spurt (height standard deviation score (SDS) at 10 years in girls and 12 years in boys) by reasoning that increased height relative to the population mean in early puberty reflects either overall genetic height potential or entrance into the pubertal growth spurt. Because a large proportion of adult stature is achieved prior to the onset of puberty, we expected a significant part of the detected variants to associate with overall height growth potential, whereas a minority would have specific pubertal timing effects. In Analysis II, we assessed the overall contribution of growth across puberty to adult height (height change SDS between 8 years and adult) that reflects the total magnitude of growth during the pubertal growth spurt. Finally, in Analysis III, we approximated the timing of peak height velocity by looking at the height change SDS between age 14 years and adult because early maturing individuals grow less during late adolescence than late-maturing individuals, who still have much of their remaining growth to achieve after age 14. Similar simple height measurements across puberty have previously proven robust in the GWA setting for detecting common genetic variation influencing both height growth and pubertal timing (15).

RESULTS

Discovery and follow-up meta-analyses reveal 10 genome-wide significant loci associated with pubertal growth

Nine cohorts contributed partly overlapping population-based samples (Supplementary Material, Table S2) with childhood height measurements and approximately 2.5 million directly genotyped or imputed SNPs to three discovery GWA analyses (Supplementary Material, Table S3), in which we meta-analysed data from males and females both separately and combined for the three models. We observed significant deviation from the expected distribution of P-values for all three combined-gender analyses (I, II and III), males and females separately for Analysis I and females only for Analysis II (Supplementary Material, Fig. S1A).

All three models resulted in genome-wide significant loci, although we had most power (Supplementary Material, Table S4) to detect loci for Analysis I (height SDS at age 10 years in girls and 12 years in boys) (Table 1). In total, nine loci contained markers that reached P-values below the genome-wide significance threshold corrected for testing three primary phenotypes ($P < 1.67 \times 10^{-8}$, after genomic control). Of these, only rs7759938 nearby LIN28B was previously known to influence pubertal growth (15,16).

Due to the requirement of Analyses II and III to have both childhood and adult height measurements for the same individuals, there were no additional samples available for follow-up of suggestive signals for these analyses. Thus, we only performed follow-up for Analysis I. An additional 6 cohorts comprising up to 9710 samples were available for follow-up of the 22 suggestive signals ($P < 1 \times 10^{-5}$) for Analysis I (Supplementary Material, Table S5). Joint analysis of discovery and follow-up stages for Analysis I robustly confirmed a single novel variant, rs4788196 ($P = 9.49 \times 10^{-11}$, $n = 18,737$, Table 1), thus bringing the number of loci reaching the genome-wide significance threshold to 10, of which 7 were associated with Analysis I.

Expression quantitative trait loci (eQTL) analysis links rs4788196 (G) to decreased expression of nearby gene MAPK3 and pathway analyses highlight the TGF-beta signalling pathway and pathways in cancer

To link the identified association signals with putative biological processes, we tested all significantly associated gene regions for association with leukocyte gene expression levels and performed gene pathway analyses. Expression quantitative trait loci (eQTL) analysis in whole blood (17) linked rs960273 with the gene GNA12, as previously reported (1), as well as highlighting a previously unknown
role for the extracellular signal-regulated kinase 1 (MAPK3, also known as ERK1) in prepubertal height growth (Supplementary Material, Table S6). More specifically, the adolescent height-increasing allele (G) at rs4788196 on 16p11.2 (Analysis I) correlated with decreased expression of MAPK3, consistent with previous studies linking deactivation of the gene with increased bone growth in mice (18). We subsequently performed pathway analyses using the g:Profiler Gene Group Functional Profiler tool [g:GOST (19); Supplementary Material, Table S7A] and MAGENTA Gene Set Enrichment Analyses [GSEA (20); Supplementary Material, Table S7B] that commonly highlighted the TGF-beta signaling pathway and pathways in cancer for loci identified in Analysis I. Whereas g:Profiler identified the MAPK-pathway, the GSEA showed enrichment of lower than expected P-values for genes belonging to the TOB1 pathway, although the individual implicated gene regions were only suggestive-ly associated in Analysis I.

Figure 1. Study design. We performed a two-stage study to detect and characterize loci influencing the pubertal phase of childhood growth. Stage 1 consisted of locus mapping using a GWA approach on three related simple measurements of the pubertal growth spurt and joint analysis of discovery and follow-up studies of novel variants for height SDS at age 10 years in girls and 12 years in boys (Analysis I). Validated loci were then characterized using a variety of methods in Stage 2, including genetic characterization (conditioned analysis on previously reported nearby SNPs in low or partial pairwise linkage disequilibrium with pubertal growth signals), functional characterization (eQTL analysis of pubertal growth SNPs on the expression levels of nearby genes and pathway analysis on biological pathways using g:Profiler and MAGENTA), characterization of the growth and maturational effects of the identified loci (on height and BMI across puberty, on the timing of menarche for those signals not previously associated with AAM and on early height from 1 to 4 years for loci that influence the timing of menarche and pubertal growth) and the association between BMI-increasing alleles and total pubertal growth.

The novel locus near MAPK3 associates transiently with height growth in childhood and earlier menarche

Although there are no published studies implicating MAPK3 in human height growth, rare recurrent CNVs near MAPK3 on chromosome 16p11.2 have been shown to associate with early onset obesity (21,22). Nonetheless, the adolescent height effect that we observed did not appear to be CNV-mediated (Supplementary Material, Table S8). To characterize the MAPK3 variant in more depth, we evaluated the longitudinal height and BMI effects of rs4788196. We plotted the effect size (beta) against six age bins across puberty from 8 years to adult (Fig. 3A) and investigated early height yearly from ages 1 to 4 (Supplementary Material, Table S9). These analyses revealed a transient effect on height growth for the G allele from age 4 in both males and females that was diluted by adulthood, with no apparent effect on BMI (Supplementary Material, Fig. S2). Finally, because rapid
growth during childhood and early adolescence may correlate with early timing of sexual maturation (8), we also tested rs4788196 for association with age at menarche (AAM) (23) and found that the height-increasing allele associated with earlier AAM ($P = 1.42 \times 10^{-4}$, surpassing the significance threshold of 0.007, that corresponds to a Bonferroni-correction accounting for follow-up of seven loci not previously associated with AAM; Supplementary Material, Table S10).

Five of the discovered pubertal growth loci are also associated with pubertal timing, eight are adult stature loci and one is a BMI locus.

Even though the MAPK3-locus associated with both prepubertal height growth and the timing of puberty, it showed little evidence for association with adult anthropometric traits (Table 2). Nonetheless, epidemiological data support phenotypic correlations between earlier pubertal timing, increased adult obesity and decreased final height. To better understand how the loci we detected contribute to a genetic link underlying these traits, we performed a systematic analysis of all leading SNPs significantly associated with pubertal height and growth and noted substantial evidence for overlap (Fig. 4). Assessing the pubertal timing effect, both based on published GWA of AAM (23) and by in silico meta-analysis of leading signals not previously implicated in the timing of menarche, as described above, showed that all three discovery analysis approaches detected pubertal timing loci (near MAPK3, PXMP3, VGLL3, ADCY3-POMC and LIN28B). Moreover, eight signals overlapped with adult stature loci (1), of which ADCY3-POMC has further been implicated in childhood (24) and adult obesity (25). However, two of our signals within 1 Mb of reported height loci showed partial linkage disequilibrium ($r^2 < 0.6$) with previously published SNPs. Conditioning for the previously reported marker revealed evidence for a further independent association nearby CABLES1 (rs6507528; $P = 0.00011$) (Supplementary Material, Table S11). This variant may represent allelic heterogeneity or partially tag the same causative variant (26).

Longitudinal analyses across puberty show that the identified pubertal growth loci represent both overall growth potential and pubertal timing.

To further evaluate the leading signals, we compared their height effects longitudinally across puberty, revealing multiple distinct growth trajectories. This approach divided the loci associated with various measures of pubertal height and growth into two groups based on association with pubertal timing. One group of loci (near ZBTB38, EFEMP1, CABLES1, ADAMTS3 and GNA12), not associated with pubertal timing, all impacted height SDS across multiple growth phases, strongly and steadily from prepuberty to adulthood (Fig. 3C). Thus, these loci likely reflect overall growth potential, rather than puberty-specific effects. In contrast, the five pubertal timing-associated variants displayed distinct growth trajectories in both trajectories and timing of the pubertal growth spurt (Table 2; Fig. 3A and B).
Table 1. Pubertal growth loci reaching genome-wide significance ($P < 1.67 \times 10^{-8}$)

<table>
<thead>
<tr>
<th>SNP</th>
<th>Nearby gene(s)</th>
<th>Previously associated related trait$^a$</th>
<th>Chromosome</th>
<th>Position (bp)$^b$</th>
<th>Effect allele/other allele$^b$</th>
<th>Effect allele frequency</th>
<th>Discovery Relative height change $\beta$ (SE)$^c$</th>
<th>$P^d$</th>
<th>$P_{	ext{het}}$</th>
<th>$P_{	ext{sex-het}}$</th>
<th>Follow-up Relative height change $\beta$ (SE)$^c$</th>
<th>$P$</th>
<th>Joint Discovery + Follow-up Relative height change $\beta$ (SE)$^c$</th>
<th>$P$</th>
<th>$n$</th>
</tr>
</thead>
<tbody>
<tr>
<td>rs6764759 ZBTB38, RAS42</td>
<td></td>
<td></td>
<td>H</td>
<td>3</td>
<td>G/A</td>
<td>0.45</td>
<td>0.08 (0.012)</td>
<td>$4.6 \times 10^{-10}$</td>
<td>0.52</td>
<td>0.35</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>rs7846385 PXMP3, PKIA</td>
<td>H, AAM</td>
<td></td>
<td>H</td>
<td>8</td>
<td>C/T</td>
<td>0.26</td>
<td>0.09 (0.014)</td>
<td>$5.27 \times 10^{-10}$</td>
<td>0.13</td>
<td>0.13</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>rs1346789 EEF1AP1</td>
<td>H</td>
<td></td>
<td>H</td>
<td>2</td>
<td>C/T</td>
<td>0.22</td>
<td>−0.08 (0.015)</td>
<td>$1.15 \times 10^{-8}$</td>
<td>0.41</td>
<td>0.85</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>rs6507528 CABLESI</td>
<td>H</td>
<td></td>
<td>H</td>
<td>18</td>
<td>G/A</td>
<td>0.55</td>
<td>−0.09 (0.016)</td>
<td>$1.31 \times 10^{-8}$</td>
<td>0.44</td>
<td>0.44</td>
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<tr>
<td>rs1365198 ADAMTS13</td>
<td>H</td>
<td></td>
<td>H</td>
<td>15</td>
<td>G/T</td>
<td>0.76</td>
<td>0.08 (0.014)</td>
<td>$1.5 \times 10^{-8}$</td>
<td>0.66</td>
<td>0.41</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>rs4788136 MAPK2</td>
<td>Novel</td>
<td></td>
<td>H</td>
<td>16</td>
<td>G/A</td>
<td>0.44</td>
<td>0.06 (0.012)</td>
<td>$6.38 \times 10^{-7}$</td>
<td>0.51</td>
<td>0.43</td>
<td>0.08 (0.021)</td>
<td>$4.25 \times 10^{-5}$</td>
<td>0.96 (0.011)</td>
<td>13946</td>
<td>18737</td>
</tr>
<tr>
<td>rs960273 GNA42</td>
<td></td>
<td></td>
<td>H</td>
<td>7</td>
<td>C/T</td>
<td>0.32</td>
<td>−0.1 (0.018)</td>
<td>$2.51 \times 10^{-8}$</td>
<td>0.96</td>
<td>5.17</td>
<td>$10^{-4}$</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>rs1172294 ADCY3, DNAJC27, POMC</td>
<td>H, BMI</td>
<td></td>
<td>H</td>
<td>2</td>
<td>G/A</td>
<td>0.45</td>
<td>−0.08 (0.014)</td>
<td>$1.02 \times 10^{-8}$</td>
<td>0.87</td>
<td>0.92</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>rs7628864 VGLL3</td>
<td>AAM</td>
<td></td>
<td>H</td>
<td>3</td>
<td>G/A</td>
<td>0.38</td>
<td>−0.11 (0.019)</td>
<td>$3.17 \times 10^{-9}$</td>
<td>0.76</td>
<td>6.83</td>
<td>$10^{-6}$</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>rs7759938 LIN28B</td>
<td>H, AAM</td>
<td></td>
<td>H</td>
<td>6</td>
<td>G/A</td>
<td>0.32</td>
<td>0.11 (0.016)</td>
<td>$3.87 \times 10^{-9}$</td>
<td>0.23</td>
<td>0.23</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

$^a$Previously associated related traits are adult stature (H), AAM or BMI.
$^b$Marker position reported according to Build 36 and allele coding based on the positive strand.
$^c$Effect sizes are change in height or growth SDS score.
$^d$P-value adjusted for genomic control.
$^e$P-value assessed by t-test for sexual heterogeneity.
The locus associating with increased childhood BMI also associates with decreased pubertal growth but not with prepubertal height

Of the pinpointed loci associating with pubertal growth and timing, only one had previously been associated with BMI (Supplementary Material, Fig. S2). The strong correlation between childhood obesity and prepubertal height (2–4) and between prepubertal height and AAM (27) predicts that the BMI-associated marker, rs1172294 (ADCY3-POMC), would associate with increased prepubertal stature. However, the variant showed no association with stature before puberty (Fig. 3B). Nonetheless, the BMI-increasing allele (G) was associated with earlier menarche \( (P = 8.64 \times 10^{-6}) \) and a decline in pubertal growth in both males and females, as expected. Consistently, other variants previously associated with childhood obesity (24) showed a parallel between elevated BMI and diminished growth across puberty (Supplementary Material, Table S12). We also found that rs3817334 (MTCH2), previously associated with adult (25) but not childhood BMI, also associated with the same decrease in overall pubertal growth.

**DISCUSSION**

Taken together, the simple approach used in this study to model the pubertal growth spurt for GWAS in more than 18,000 study subjects of European descent identified 10 significantly associated pubertal growth loci. Utilizing unique longitudinal childhood measurements, we described the distinct height growth and BMI effects of these variants across puberty and noted significant longitudinal associations at each locus. More specifically, half of the identified loci also associated with pubertal timing and provided evidence linking a robust novel growth and menarche locus with MAPK3 expression levels. Despite prior association between several genes in the MAPK-pathway and skeletal growth
Table 2. Association of pubertal growth variants with height growth across puberty

<table>
<thead>
<tr>
<th>SNP (allele) nearby gene</th>
<th>Prepubertal height</th>
<th>Height at early puberty</th>
<th>Total pubertal growth</th>
<th>Late adolescent growth</th>
<th>Adult height&lt;sup&gt;b&lt;/sup&gt;</th>
<th>GIANT height&lt;sup&gt;c&lt;/sup&gt;</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Height SDS at 7–8 years</td>
<td>Analysis I height SDS at 10 years in females and 12 years in males</td>
<td>Analysis II height SDS change 8 years in adult</td>
<td>Analysis III height SDS change 14 years in adult</td>
<td>Adult stature SDS</td>
<td>Beta</td>
</tr>
<tr>
<td>rs7846385 (C) PXMP3</td>
<td>Females</td>
<td>0.090 0.016 3.73 × 10&lt;sup&gt;-8&lt;/sup&gt;</td>
<td>0.096 0.018 8.71 × 10&lt;sup&gt;-8&lt;/sup&gt;</td>
<td>−0.057 0.021 0.006</td>
<td>−0.043 0.023 0.068</td>
<td>0.03 0.019 0.114</td>
</tr>
<tr>
<td></td>
<td>Males</td>
<td>0.058 0.016 0.031</td>
<td>0.067 0.019 5.46 × 10&lt;sup&gt;-4&lt;/sup&gt;</td>
<td>−0.009 0.009 0.696</td>
<td>−0.068 0.026 0.008</td>
<td>0.038 0.02 0.055</td>
</tr>
<tr>
<td></td>
<td>Combined</td>
<td>0.074 0.012 1.48 × 10&lt;sup&gt;-10&lt;/sup&gt;</td>
<td>0.082 0.013 3.04 × 10&lt;sup&gt;-10&lt;/sup&gt;</td>
<td>−0.035 0.015 0.02915</td>
<td>−0.054 0.017 0.002</td>
<td>0.034 0.014 0.014</td>
</tr>
<tr>
<td>rs7628864 (G) VGLL3</td>
<td>Females</td>
<td>0.051 0.015 0.001</td>
<td>0.044 0.016 0.007</td>
<td>−0.112 0.019 3.17 × 10&lt;sup&gt;-9&lt;/sup&gt;</td>
<td>−0.059 0.021 0.006</td>
<td>−0.029 0.017 0.087</td>
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<tr>
<td></td>
<td>Males</td>
<td>0.009 0.015 0.549</td>
<td>0.023 0.018 0.201</td>
<td>0.011 0.020 0.578</td>
<td>−0.001 0.022 0.969</td>
<td>0.028 0.017 0.113</td>
</tr>
<tr>
<td></td>
<td>Combined</td>
<td>0.030 0.011 0.004</td>
<td>0.035 0.012 0.004</td>
<td>−0.054 0.014 9.41 × 10&lt;sup&gt;-8&lt;/sup&gt;</td>
<td>−0.031 0.015 0.043</td>
<td>0.00 0.012 0.967</td>
</tr>
<tr>
<td>rs7759938 (T) LIN28B</td>
<td>Females</td>
<td>−0.042 0.014 0.003</td>
<td>0.004 0.017 0.792</td>
<td>−0.027 0.020 0.169</td>
<td>−0.112 0.022 2.89 × 10&lt;sup&gt;-7&lt;/sup&gt;</td>
<td>−0.054 0.018 0.002</td>
</tr>
<tr>
<td></td>
<td>Males</td>
<td>−0.046 0.014 0.001</td>
<td>−0.011 0.018 0.547</td>
<td>−0.020 0.021 0.327</td>
<td>−0.074 0.023 0.002</td>
<td>−0.045 0.018 0.011</td>
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<tr>
<td></td>
<td>Combined</td>
<td>−0.044 0.010 8.80 × 10&lt;sup&gt;-6&lt;/sup&gt;</td>
<td>−0.003 0.012 0.804</td>
<td>−0.024 0.014 0.097</td>
<td>−0.094 0.016 3.87 × 10&lt;sup&gt;-9&lt;/sup&gt;</td>
<td>−0.05 0.013 1.0 × 10&lt;sup&gt;-4&lt;/sup&gt;</td>
</tr>
<tr>
<td>rs4788196 (G) MAPK3</td>
<td>Females</td>
<td>0.047 0.015 0.001</td>
<td>0.059 0.017 8.72 × 10&lt;sup&gt;-4&lt;/sup&gt;</td>
<td>−0.0003 0.018 0.958</td>
<td>−0.025 0.021 0.231</td>
<td>0.03 0.017 0.081</td>
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<td></td>
<td>Males</td>
<td>0.061 0.015 4.51 × 10&lt;sup&gt;-5&lt;/sup&gt;</td>
<td>0.066 0.017 1.49 × 10&lt;sup&gt;-5&lt;/sup&gt;</td>
<td>0.003 0.02 0.867</td>
<td>−0.046 0.023 0.04</td>
<td>0.039 0.017 0.023</td>
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<tr>
<td></td>
<td>Combined</td>
<td>0.054 0.011 3.56 × 10&lt;sup&gt;-7&lt;/sup&gt;</td>
<td>0.063 0.012 6.38 × 10&lt;sup&gt;-7&lt;/sup&gt;</td>
<td>0.001 0.014 0.921</td>
<td>−0.035 0.015 0.023</td>
<td>0.035 0.012 0.804</td>
</tr>
<tr>
<td>rs1172294 (G) ADCY3</td>
<td>Females</td>
<td>−0.001 0.013 0.945</td>
<td>−0.006 0.017 0.718</td>
<td>−0.078 0.019 3.16 × 10&lt;sup&gt;-5&lt;/sup&gt;</td>
<td>−0.046 0.021 0.028</td>
<td>−0.047 0.017 0.005</td>
</tr>
<tr>
<td></td>
<td>Males</td>
<td>−0.016 0.014 0.246</td>
<td>−0.045 0.017 0.01</td>
<td>−0.08 0.02 5.97 × 10&lt;sup&gt;-3&lt;/sup&gt;</td>
<td>−0.01 0.023 0.673</td>
<td>−0.045 0.017 0.008</td>
</tr>
<tr>
<td></td>
<td>Combined</td>
<td>−0.008 0.01 0.404</td>
<td>−0.026 0.012 0.04</td>
<td>−0.079 0.014 1.02 × 10&lt;sup&gt;-8&lt;/sup&gt;</td>
<td>−0.02 0.015 0.184</td>
<td>−0.046 0.012 1.3 × 10&lt;sup&gt;-4&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

<sup>a</sup>P-values taken from GWA discovery analyses, genomic-control corrected.

<sup>b</sup>Meta-analysis of association results for adult stature taken from the cohorts participating in the longitudinal analyses.

<sup>c</sup>Association to adult stature taken from publically available results of the GIANT Consortium adult height meta-analysis (http://www.broadinstitute.org/collaboration/giant/index.php/GIANT_consortium_data_files). Beta estimates were not available for public download.

All effect sizes are given for the menarche-advancing allele. In bold are associations reaching a P-value threshold of 0.002 (Bonferroni-corrected threshold accounting for follow-up analysis of five markers in five analyses).
syndromes (28–31), MAPK3 has not been associated with human height before. Interestingly, gonadotropin-releasing hormone, crucial for regulating the onset of puberty, activates MAPK3 (32), providing a putative biological link between hormone, crucial for regulating the onset of puberty, activates

Figure 4. Loci associated with pubertal height and their overlap with partially correlated phenotypes. The 10 genome-wide significant loci were assessed for their association with the correlated traits adult stature, pubertal timing (AAM) and adiposity (BMI) by examining previously published literature. Additionally, for loci not previously associated with pubertal timing, we queried the leading SNPs in GWA data of AAM by the ReproGen Consortium. All the pubertal growth loci showed pleiotropic associations with one or more related phenotype.

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In fact, our data affirm a complex genetic architecture underlying growth, pubertal timing and adiposity. In particular, specific genetic effects may contradict epidemiological correlations. Epidemiological studies have observed a developmental pattern linking taller prepubertal stature to earlier puberty, accelerated skeletal maturation and short adult stature due to early cessation of growth (2,3). Although the majority of loci we assessed showed the expected parallel association between early menarche and decreased overall pubertal height growth, their prepubertal height effects varied. Three variants (near MAPK3, PXMP3 and VGLL3) followed the expected pattern, linking taller prepubertal stature with earlier AAM, whereas the early puberty-associated allele (T) at rs7759938 (LIN28B) correlated with shorter prepubertal childhood height, as reported previously (15).

The relationship between puberty and adult stature is similarly complex, whereas epidemiological studies show a correlation between early puberty and reduced adult height (3), and a genetic association study found that early puberty alleles may associate with either increased or decreased adult stature (23). An example is rs7846385 (PXMP3), for which, contradictory to the predicted pattern, the early menarche allele associates with increased adult height. Our results show that tall adult height is achieved because the early-menarche allele (C) also influences tall childhood height and a limited reduction in total pubertal growth. These data, thus, agree with a recent candidate gene study suggesting that loci associated with adult height may have a stronger influence on prepubertal growth than during the pubertal growth spurt (16). However, utilizing a genome-wide approach with greater sample sizes, our study identifies loci previously missed that specifically target pubertal growth, and we find that they are associated with diverse and unique longitudinal growth patterns. We also find that not all loci influencing pubertal growth also impact adult stature.

Additionally, epidemiological studies link increased childhood adiposity with advanced puberty and increased prepubertal height. Although all childhood BMI-increasing alleles assessed in this study also showed an association with decreased overall pubertal growth, at the ADCY3-POMC locus, the same allele associated with both earlier puberty and increased childhood BMI, but not with prepubertal stature. The correlation between obesity and pubertal growth may be consequential of hormonal changes associated with childhood adiposity. However, because the same association pattern was also present at a locus uniquely associated with adult BMI (MTCH2), an underlying shared genetic effect remains likely.

Given the complexity of the relationships between these developmental traits, tracking unique gene effects across multiple growth periods may help to elucidate specific pathways linking childhood events to adult outcomes, as illustrated here with height growth, pubertal timing and adult stature. While epidemiological studies have described correlations between distinct childhood growth events and adult health, genetically defined association patterns may pinpoint molecular processes linking these traits. Characterization of these pathways may thus provide new insight towards a better understanding of the relationships between early growth patterns, pubertal timing and adult disease risk.

**MATERIALS AND METHODS**

**Phenotypes and study subjects**

Discovery study subjects were included from cohorts participating in the Early Growth Genetics Consortium (43), namely the Avon Longitudinal Study of Parents and Children (ALSPAC), 1958 British Birth Cohort (BC58-T1DGC and
BC58-WTCCC), Cardiovascular Risk in Young Finns Study (YFS), Helsinki Birth Cohort Study (HBCS), Lifestyle-Immune System-Allergy Plus Environment and Genetics Study (LISAplus), Northern Finland Birth Cohort 1966 (NFBC1966), Queensland Institute of Medical Research and Western Australia Pregnancy Study (RAINe). Cohort-specific details for all analyses can be found in Supplementary Material, Table S3. The data annotation, exchange and storage have been facilitated by the SIMBioMS platform (33).

Three primary phenotypes were analysed that were defined as follows.

Analysis I: single height: girls with height measurements available at age 10 (± 1 year) and boys with height measured at age 12 (± 1 year) were included. Sex-specific SDs for each individual were calculated within each study by dividing the difference between the individual’s measured height and the within-population height mean by the population standard deviation (SD). A total of 14 040 samples (7161 males and 6879 females) from 9 contributing cohorts were included.

Analysis II: total pubertal growth: individuals with a childhood height measurement at age 8 (± 1 year) and at adulthood (≥18 years of age) were included. Height difference was calculated between the two measurements, and sex-specific SDSs of this difference were calculated within each study as described above. Six cohorts with up to 10 799 samples (5043 males and 5756 females) contributed to the analysis.

Analysis III: late pubertal growth: subjects with a height measurement in adolescence at age 14 (± 1 year) and at adulthood (≥18 years of age) were included. Height difference was calculated between the two measurements, and logarithm-transformed sex-specific Z-scores were calculated within each study. Log transformation was performed prior to SDS calculation. Five cohorts with up to 9228 subjects (4282 males and 4946 females) were included in this analysis.

Genotyping and quality control

 Genome-wide genotypes were obtained using high-density SNP arrays on Illumina and Affymetrix platforms. Before imputation, SNPs with minor allele frequency of <1%, call rate <95% or Hardy–Weinberg equilibrium $P < 1 \times 10^{-6}$ were excluded. Samples were also excluded if they contained duplicates, excess heterozygosity, non-European ancestry or ambiguous gender. Imputation was performed using IMPUTE (34) or MACH (35) for roughly 2.5 million SNPs against HapMap Phase II (release 21 of 22). Imputed SNPs were filtered prior to meta-analysis to exclude poorly imputed SNPs (IMPUTE filter PROPER INFO <0.4, MACH filter $r^2 < 0.3$).

GWA analyses

Within each cohort, association analyses were performed by linear regression using an additive model across genotyped and imputed SNPs (dosages), for males and females separately. For all analyses, age at adolescent measurement was included as a covariate where available, and the first two principal components were adjusted within each study sample if necessary. For the association tests, PLINK (36), ProbABEL (38), SNPtest (34) or MACH2QTL (35) was used.

Meta-analyses

A fixed effects inverse-variance meta-analysis model was used to test the effect of each variant on height, total pubertal growth or late pubertal growth separately for males and females. Sex-specific results from each study were also meta-analysed for each phenotype in three combined-gender analyses. The $R$ package MetABEL (37) (v.2.11.1) was used to perform all meta-analyses. MetABEL corrects each individual result for its respective genomic inflation factor ($\lambda$) according to the genomic control method for population stratification. Subsequently, an additional genomic control correction was applied using the overall genomic inflation factor calculated for each of the nine meta-analysed results. The threshold for genome-wide significance was set at a conservative Bonferroni-corrected threshold of $P < 1.67 \times 10^{-8}$, accounting for testing three primary phenotypes. A further significance threshold of 0.002 (accounting for examining 10 loci in males and females) was applied to all follow-up analyses unless otherwise stated.

Conditional analyses

To determine whether our signals represent independent effects on growth during puberty from previously reported related phenotypes, we performed linear regression using an additive model on the primary pubertal growth phenotypes, adjusting each of the six markers (imputed genotype dose) for the previously reported marker nearby our signal. As in the primary analysis, age at adolescent measurement (where available) and optional adjustment for population substructure were included as covariates (Supplementary Material, Table S3).

Follow-up analyses of suggestive association signals

Genetic markers yielding association $P$-values of $1 \times 10^{-5}$ to $1.67 \times 10^{-8}$ in Analysis I and not previously associated with related traits adult stature, AAM or BMI were selected for follow-up genotyping ($n = 22$). Additional cohorts participated in the follow-up analyses, including in-silico analyses by ALSPAC (follow-up sample), Children’s Hospital of Philadelphia (CHOP), Finnish Twin Cohort Study, Genome-Wide Population-Based Association Study of Extremely Overweight Young Adults and Lifestyle-Immune System-Allergy study & German Infant Study on the influence of Nutrition Intervention plus environment and genetics (GINplus; follow-up sample). Association results for a marker showing borderline significance, rs281379, were also provided by Netherlands Twin Registry (NTR). De novo genotyping was done for selected markers (success rate >98%) from Northern Finland Birth Cohort 85–86 (NFBC8586) with TaqMan Pre-Designed SNP Genotyping Assays on LightCycler 480 Real-Time PCR System (Roche) according to the manufacturer’s instructions at the Finnish Genome Center (Helsinki, Finland).

Statistical analysis in replication samples was performed similarly as in the discovery analyses with PLINK (36), ProbABEL (38) or SNPtest (34), using linear regression models for each of the 22 markers under an additive model, with age at adolescent measurement and correction for population
substructure as optional covariates. Genomic control-corrected discovery results were meta-analysed together with the individual linear regression results from contributing cohorts for each SNP, using the MetABEL (37) package of R (v.2.11.1).

**CNV analysis of 16p11.2**

CNVs were genotyped using signal intensity distributions and B-allele frequency of the genotyping probes with PennCNV software (39) and adjusted for genomic waves according to genomic GC content, as previously described (40). The CNV scan was completed for 2310 individuals in YFS and 4931 in NFBC1966 (41).

**Expression quantitative trait loci**

We queried significant SNPs from the Dietary, Lifestyle, and Genetic determinants of Obesity and Metabolic syndrome study, an extension of FINRISK 2007. The eQTL methods are previously published (17). Briefly, whole blood was extracted from 518 unrelated individuals and genotyped on the Illumina 610-Quad SNP array. In parallel, mRNA expression was quantified with Illumina HumanHT-12 Expression BeadChips. Linear regression was used to test association between transcript expression levels and each SNP.

**Pathway analyses**

We entered the nearest gene to all signals at $P < 1 \times 10^{-4}$ (one per locus) into the g:Profiler Gene Group Functional Profiler tool (g:GOS2) (19), a webtool that, briefly, queries databases of biological pathways for enrichment of user-entered genes. For Analysis I, we also entered MAPK3 because the gene was implicated by eQTL evidence to be functionally relevant ($n = 93$, corresponding to 0.0003% of all discovery signals for Analysis I). We also ran GSEA using MAGENTA (20), a program that calculates a $P$-value for each gene in the genome based on GWA results and then searches biological databases for pathways showing an enrichment of genes with lower than expected $P$-values. Analysis II and III data are not reported here due to the lack of significant findings.

**Association analyses of age of menarche**

To assess the relevance of the pubertal growth-associated variants for pubertal timing, leading signals not previously implicated in the timing of menarche were queried from in silico meta-analysis data of 87,802 women published by the ReproGen Consortium (23).

**Cross-sectional height and BMI analyses**

Height or BMI measurements from childhood to adulthood were divided into six age bins: (i) prepuberty (6.5–8.5 years old), (ii) early puberty (8.6–10.5 years old), (iii) mid-puberty for females (10.6–12.5 years old), (iv) mid-puberty for males (12.6–14.5 years old), (v) late puberty (14.6–17.5 years old) and finally (vi) adult (>17.6 years old). In each cohort, each marker of interest (imputed genotype dosage) was tested for association with sex-specific height or BMI SDS for all age bins available, using linear regression assuming an additive model and adjusting for exact age at measurement (to the nearest month), along with optional correction for population stratification. A single measurement was included per study subject per bin, with the age closest to the mean used when more than one measurement was available. Altogether 23 SNPs were analysed for height and BMI across pubertal growth (only significantly associated markers are reported here). Summary statistics were meta-analysed like the primary analyses in each age bin, separately for males and females, for both height and BMI distinctly. Effect sizes were plotted versus age.

**Early growth analyses**

Cohorts with height measurements available at 1, 2, 3 or 4 years were included, namely the CHOP, Copenhagen Study on Asthma in Childhood, Generation R Study (Generation R), HBCS, INfancia y Medio Ambiente (Environment and Childhood) Project (INMA), LISAlplus&GINIplus, NTR, Northern Finland Birth Cohort 1966 (NFBC1966), Prevention and Incidence of Asthma and Mite Allergy birth cohort study and Western Australian Pregnancy study (RAINE). Length was measured at 12 months (range 6–18 months) and height at 24 (range 18–30), 36 (range 30–42) and 48 (range 42–54) months. If multiple measurements per individual were available, those closest to 12, 24, 36 or 48 months were used. Sex- and age-adjusted SDSs were calculated using Growth Analyser 3.0 (Dutch Growth Research Foundation, Rotterdam, The Netherlands) in each study separately (42). The sex-specific association between each marker genotype and height or BMI SDS was assessed using linear regression, assuming an additive model. Imputed genotypes were used, where directly assayed genotypes were unavailable. We meta-analysed the within-cohort sex-stratified linear regression results using the inverse-variance method. A fixed-effects model was assumed, using RMeta in R (v.2.7.0).

**URLS**


**SUPPLEMENTARY MATERIAL**

Supplementary Material is available at HMG online.
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Detailed acknowledgements by study can be found in the supplementary note.

Conflict of Interest statement. None declared.

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