Intrauterine instillation of diluted seminal plasma at oocyte pick-up does not increase the IVF pregnancy rate: a double-blind, placebo controlled, randomized study

M. von Wolff1,*, S. Rösner2, A. Germeyer2, J. Jauckus2, G. Griesinger3, and T. Strowitzki2

1Department of Gynaecological Endocrinology, University Women’s Hospital, and Reproductive Medicine, University of Berne, Effingerstrasse 102, Berne 3010, Switzerland 2Department of Gynaecological Endocrinology and Reproductive Medicine, University Women’s Hospital, University of Heidelberg, Vossstrasse 9, Heidelberg 69115, Germany 3Department of Gynecological Endocrinology and Reproductive Medicine, University Hospital of Schleswig-Holstein, Campus Luebeck, Ratzeburger Allee 160, Luebeck 23538, Germany

*Correspondence address. Tel: +41-31-632-1303; Fax: +41-31-632-1305; E-mail: michael.vonwolff@insel.ch

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STUDY QUESTION: Does intrauterine application of diluted seminal plasma (SP) at the time of ovum pick-up improve the pregnancy rate by ≥ 14% in IVF treatment?

SUMMARY ANSWER: Intrauterine instillation of diluted SP at the time of ovum pick-up is unlikely to increase the pregnancy rate by ≥ 14% in IVF.

WHAT IS KNOWN ALREADY: SP modulates endometrial function, and sexual intercourse around the time of embryo transfer has been suggested to increase the likelihood of pregnancy. A previous randomized double-blind pilot study demonstrated a strong trend towards increased pregnancy rates following the intracervical application of undiluted SP. As this study was not conclusive and as the finding could have been confounded by sexual intercourse, the intrauterine application of diluted SP was investigated in the present trial.

STUDY DESIGN, SIZE, DURATION: A single-centre, prospective, double-blind, placebo-controlled, randomized, superiority trial on women undergoing IVF was conducted from April 2007 until February 2012 at the University Department of Gynaecological Endocrinology and Reproductive Medicine, Heidelberg, Germany.

PARTICIPANTS/MATERIALS, SETTING, METHODS: The study was powered to detect an 14% increase in the clinical pregnancy rate and two sequential tests were planned using the Pocock spending function. At the first interim analysis, 279 women had been randomly assigned to intrauterine diluted SP (20% SP in saline from the patients’ partner) (n = 138) or placebo (n = 141) at the time of ovum pick-up.

MAIN RESULTS AND THE ROLE OF CHANCE: The clinical pregnancy rate per randomized patient was 37/138 (26.8%) in the SP group and 41/141 (29.1%) in the placebo group (difference: −2.3%, 95% confidence interval of the difference: −12.7 to +8.2%; P = 0.69). The live birth rate per randomized patient was 28/138 (20.3%) in the SP group and 33/141 (23.4%) in the placebo group (difference: −3.1%, 95% confidence interval of the difference: −12.7 to +6.6%; P = 0.56). It was decided to terminate the trial due to futility at the first interim analysis, at a conditional power of 62%.

LIMITATIONS, REASONS FOR CAUTION: The confidence interval of the difference remains wide, thus clinically relevant differences cannot reliably be excluded based on this single study.

WIDER IMPLICATIONS OF THE FINDINGS: The results of this study cast doubt on the validity of the concept that SP increases endometrial receptivity and thus implantation in humans.

STUDY FUNDING/COMPETING INTEREST(S): Funding was provided by the department’s own research facilities.

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Key words: endometrium / seminal plasma / in vitro fertilization / implantation
Introduction

Besides the quality of the embryo, endometrial receptivity plays an important role in the establishment of pregnancy. Endometrial functional disorders are likely of little relevance in a physiological context, e.g. spontaneous conception in a natural cycle. However, under non-physiological conditions such as high-dose gonadotrophin stimulation and absence of sexual intercourse, as it is typically the case in IVF treatment, endometrial dysfunction is likely to play a role in the non-achievement of pregnancy despite the transfer of vital embryos. Horcajades et al. (2005) were able to detect a dysregulation of endometrial gene expression under high-dose gonadotrophin stimulation. Furthermore, Gutscbe et al. (2003) and von Wolff et al. (2007) demonstrated that seminal plasma (SP) has a direct effect on endometrial cytokine expression.

In order to improve endometrial receptivity under IVF treatment, numerous treatments have been proposed, such as the administration of acetylsalicylic acid and/or heparin, as well as active and passive immunotherapy. The efficacy of these treatments, however, has never been unequivocally proved.

Increasing knowledge regarding the high immunomodulatory potential of SP on endometrial function has motivated various study groups to use SP as a physiological endometrial stimulant. Tremellen et al. (2000) showed that the proportion of transferred embryos that were viable at 6–8 weeks was significantly higher in women exposed to semen through intercourse compared with those who abstained. Von Wolff et al. (2009) carried out a prospective randomized double-blind study, where SP was applied intracervically at the time of follicle puncture. This pilot study showed that there was a higher clinical pregnancy rate in the patients treated with SP compared with those who received isotonic sodium chloride as placebo, although the difference was not statistically significant. The pregnancy rate per embryo transfer was ~12% higher in the SP group than in the placebo group and the pregnancy rate per randomized patient was ~8% higher than in the placebo group. Another recently published paper supported the result of this study, Chicea et al. (2013) performed an almost identical study but without the use of sodium chloride as placebo. They found a non-significantly higher pregnancy rate in 346 treated patients in the SP group than in the group without SP (55.5 versus 44.0%, P = 0.09) and a significant higher implantation rate (34.7 versus 27.5%, P = 0.026).

Despite inconclusive results, the study from von Wolff et al. gave a first estimate of potential treatment effect sizes on which subsequent study designs can be based. Accordingly, the present study was designed as a superiority study, employing a similar methodological rigour, e.g. a prospective, randomized, placebo-controlled, double-blind, single-centre study design. In contrast to the pilot study (von Wolff et al., 2009), SP was instilled into the lower part of the uterine cavity at the time of follicle aspiration. It was hypothesized that intratuterine instillation would potentially increase the effect when compared with intra-cervical instillation. Furthermore, the risk of protocol violation by patients ignoring the advice not to engage in intercourse required a protocol, which was less dependent on such effects, especially absence from intercourse can hardly be proved (Gallo et al., 2006).

As the intratuterine instillation of undiluted SP can cause uterine contractions, SP was used in a diluted form in the present trial. The in vitro studies mentioned above have not investigated whether the endometrial effects can also be achieved by using diluted SP.

Materials and Methods

This is a single-centre, two-armed, randomized, placebo-controlled, double-blind clinical study investigating superiority of SP administration at the time of oocyte pick-up in a German university affiliated IVF programme.

Subjects

All couples undergoing IVF or ICSI treatment at the Department of Gynaecology and Reproductive Medicine, Heidelberg, between April 2007 and February 2012 were considered for enrolment in the trial. Of these, 279 women (25.2–43.5 years of age) were eligible for enrolment and consented to participate in the study. Participation was limited to one treatment cycle per couple. The exclusion criteria included the presence of hepatitis B, C or human immunodeficiency virus, leukocytospermia or other signs of infection and men with <300 µl of SP. The study was approved by the local ethical committee and each patient’s approval was given by written consent.

Clinical IVF protocol

Either a long agonist (n = 267), short agonist (n = 2) or antagonist (n = 10) protocol was applied for ovarian stimulation.

For the long agonist protocol, pituitary desensitization was performed by intranasal application of the GnRH agonist 0.4 mg nafarelin acetate (Synarel®, Merck Serono, Darmstadt, Germany), starting in the luteal phase of the previous cycle. After down-regulation of the pituitary gland, patients received in addition either 150–300 IU of recombinant FSH (Gonal F®, Merck Serono) or highly purified HMG (Menogon HP®, Ferring, Hamburg, Germany) per day. Dosages were adjusted according to estradiol concentrations and vaginal ultrasound scan.

For the short agonist protocol, nasal application of 0.4 mg nafarelin acetate was started on Day 1 or 2 of the menstrual cycle and was continued until ovulation induction. FSH or HMG stimulation was started on Day 3 of the cycle using the dosages as described above.

For the antagonist protocol, FSH or HMG was initiated between Days 3 and 5 of the menstrual cycle using the dosages as described above. GnRH antagonists (Orgalutran®, MSD, Haar, Germany or Cetrodiode®, Merck Serono) were started between Days 6 and 7 of the menstrual cycle and continued until ovulation induction.

Once an adequate ovarian response had been confirmed, FSH and HMG were discontinued and 10 000 IU of urinary hCG (Predalon®, MSD, Haar, Germany) or 250 µg of recombinant hCG (Ovitrelle®, Merck Serono) was administered to induce ovulation. Transvaginal oocyte retrieval was scheduled 34–36 h after hCG administration and was performed under general anaesthesia. Fertilization was achieved by standard IVF or ICSI. One to three oocytes at the pronuclear stage were selected and cultured for another 1–2 days, when they were transferred back into the uterus. Selection of embryos for transfer was not performed due to the restrictions of the German embryo protection law. Intravaginal progesterone (600 mg/day) was used as luteal support.

Randomization

Physicians reported a scheduled oocyte pick-up with the information relevant for randomization to the IVF laboratory once the stimulation was started. The technician (J.J.) in the laboratory randomized the patient 1 day prior to the scheduled oocyte pick-up in a German university affiliated IVF programme. For randomization, two randomization lists (Stratums I and II) with block randomization for every 20 patients were prepared before the onset of the
study. Patients were consecutively added to the list on the day that follicle aspiration was scheduled. Accordingly patients were allocated to one of the treatment arms.

Stratums I and II were introduced as the pregnancy rate of IVF/ICSI treatments is highly dependent on the age of the patient and on the number of previous treatment cycles. Stratum I (expected low pregnancy rate) was defined as: ≥ 36 or < 36 years of age and transfer of ≥ 6 embryos in ≥ 3 embryo transfers without a pregnancy in previously performed IVF/ICSI cycles. All other patients were allocated to Stratum II (expected a high pregnancy rate).

Preparation and application of SP

Semen samples were obtained ≥ 1/2 weeks before follicle aspiration by masturbation from the patient’s partner and were collected in sterile flasks. SP was extracted by centrifugation of ejaculates at 600 g for 15 min. A second round of centrifugation was performed at 10,000 g for 15 min to extract all spermatozoa. SP was analysed several times to exclude remaining contamination with spermatozoa. None of the samples contained any spermatozoa after centrifugation. Then 400 µl of supernatant were diluted with 1600 µl of sterile sodium chloride and mixed. From this solution, 1500 µl was stored in 2 ml syringes at −20 °C (verum). For the placebo, 1500 µl of sterile sodium chloride was also stored in 2 ml syringes at −20 °C. Blinding of the samples was achieved by using a placebo (sodium chloride), which could not be differentiated from SP due to the same optical appearance. SP and placebo samples were stored in separate freezers in exactly the same syringes with the names of both partners attached for later allocation.

On the day of oocyte collection, blinded SP or placebo samples were taken out of the freezer by the technician around 30–60 min before the aspiration of the follicles and were transferred to the operating theatre. Follicles were aspirated and bleeding was stopped. An insemination catheter (Laboratoire C.C.D., Paris, France) was introduced just above the end of the cervix and 1500 µl SP or placebo was injected.

Outcome measures, statistics and sample size

The study was based on the hypothesis of superiority of SP administration in terms of clinical pregnancy likelihood. Based on an increase in pregnancy rates found in the pilot study (von Wolff et al., 2009), it was calculated that group sample sizes of 269 in the SP group and 269 in the placebo group could achieve 90% power to detect a difference in clinical pregnancy rate of 14%. This difference of 14% was considered as clinically meaningful by the authors. The clinical pregnancy rate in the SP group was assumed to be 24% under the null hypothesis and 38% under the alternative hypothesis. The clinical pregnancy rate in the placebo group was expected to be 22%. The test statistic is at an alpha level of 0.05 using a two-sided Z-test with continuity correction. These results assume that two sequential tests are made using the Pocock spending function to determine symmetrical test boundaries (Software program used for this calculation: PASS 2005, Kaysville, Utah, USA). Thus, an interim analysis was planned when 50% of the total sample had been randomized. If that interim analysis revealed a positive effect of SP administration at an alpha of 0.031, the study had to be prematurely stopped at a power of 60% and superiority had been proved. If this was not the case, the study had to be continued.

The primary outcome measure is the clinical pregnancy rate (fetal heart beat) per randomized patients 4–5 weeks after follicle aspiration. The SP and placebo groups are described in terms of age, etiology of infertility, stimulation protocol, proportion of patients undergoing IVF or ICSI, average number of transferred embryos and average number of blastomeres per transferred embryo. All data are presented descriptively as means (± standard deviation or ranges), or proportions with 95% confidence intervals, as appropriate. Fisher’s exact test was used to compare dichotomous characteristics of the SP and placebo groups, as well as pregnancy rates in both groups. The significance level alpha was set to 5%.

Results

A pre-planned interim analysis was performed at 50% recruitment, e.g. when 279 patients had been enrolled and randomized. The clinical pregnancy rate per randomized patient was 37/138 (26.8%) in the SP group and 41/141 (29.1%) in the placebo group (difference: −2.3, 95% confidence interval of the difference: −12.7 to +8.2%; P = 0.69). The live birth rate per randomized patient was 28/138 (20.3%) in the SP group and 33/141 (23.4%) in the placebo group (difference: −3.1, 95% confidence interval of the difference: −12.7 to +6.6%; P = 0.56).

Contrary to the hypothesis and the pilot study, no trend toward a positive treatment effect was seen with SP administration. The z-score value of −0.2858 was exceeding the lower boundary of −2.1570 as determined by the Pocock spending function for the interim analysis. At a conditional power of 62%, it thus was decided to terminate the trial due to its futility. If the comparator pregnancy rate had remained stable at 29% till the end of the trial (e.g. when the pre-planned sample size had been reached), a pregnancy rate of 59.7% would have been necessary to reach statistical significance in the remaining 130 patients to be allocated to the intervention.

Table I describes the demographics and outcomes of the two groups. Characteristics of the 279 trial participants were similar and not statistically different in the SP and placebo groups. There were 143 patients allocated to Stratum I and 136 patients allocated to Stratum II. In 239 patients, embryos were transferred. Of those, 122 patients received SP and 117 placebo. Even though diluted SP was injected into the uterine cavity, neither painful cramps nor infections were reported at oocyte pick-up or 2 weeks later (Fig. 1). An ~8% lower pregnancy rate in Stratum I (pregnancies: 34/143 = 23.8%), including patients ≥36 years or those with previous implantation failure, than in Stratum II (pregnancies: 44/136 = 32.4%), confirmed the adequacy of stratification. Adequacy of randomization was confirmed by the similar characteristics of trial participants in the two trial arms (Table I).

Discussion

The study revealed that intrauterine application of SP at the time of follicle aspiration in 279 IVF treatment cycles did not increase the pregnancy rate compared with placebo. It is important to emphasize that at the outset of this study some over-enthusiasm about a supposed effect of SP on implantation likelihood led to the decision to design a superiority trial, which could be prematurely terminated in favour of the alternative hypothesis using a common alpha spending function approach to group sequential testing. It is also important to emphasize that no distinct lower boundary of the alpha spending function to determine futility was prespecified. With the results at the first interim analysis in hand, it was then decided, on both clinical and statistical reasoning, to terminate the study early. There are, however, important drawbacks to this decision: first, there is still a chance, albeit very small, that SP instillation would be superior to placebo had the trial been continued. Secondly, the true effect of SP instillation into the uterine cavity could well be negative and with the limited sample size such a phenomenon cannot be ruled out with confidence.
The potential effect of ejaculate or SP on endometrial function has been the subject of numerous investigations for several years, mainly in animal models. However, when interpreting these studies, it should be remembered that in many species such as mouse, horse and pig, the semen reaches the uterine cavity physiologically, whereas probably very small amounts ascend through the cervical canal in the ejaculate. Furthermore, paternal antigens have been suggested to trigger a beneficial tolerant immune response towards paternal antigens that then favours the successful implantation of the resultant embryo sharing those same paternal antigens (Robertson et al., 2013). Even though SP has significant molecular and cellular effects on the endometrium in animals, SP does not appear to be essential for the establishment of a pregnancy in these animals. Tests have shown that intruterine insemination, even after extraction of SP, leads to pregnancy in horses and pigs (Katila, 2012).

In humans, investigations have been largely limited to in vitro experiments. Gutsche et al. demonstrated that SP stimulates the expression of leukaemia inhibitory factor (LIF) and interleukin 6 (IL-6) in vitro (Gutsche et al., 2003). Furthermore, a recent study demonstrated that SP increases the decidualization of stromal cells (Doyle et al., 2012). The concept of this study implies that active SP factors reach the endometrial stromal cells through the luminal epithelium. Based on investigations by Kucharski et al. (2008), who detected a transfer of cytokines administered into the uterine cavity into the bloodstream of pigs, such a mechanism is conceivable. The above study also implies that accelerated decidualization leads to an improvement in the endometrial implantation efficiency. However, recent studies have shown, by using antagonist protocols, that an early rise in progesterone and the probable resulting decidualization leads to a reduced pregnancy rate (Ochsenkühn et al., 2012).

Because studies on animal models and in vitro studies can only be applied to the clinical situation to a limited extent, only clinical studies can ultimately clarify whether SP has a relevant effect on pregnancy rates.

On the basis of this, the present study was carried out in succession of a study published in 2009 (von Wolff et al., 2009). The trend of the previous study which showed a positive effect of SP could not be confirmed in this study.

There are several possible reasons for the diverging results of these studies.

### Table I: Characteristics of trial participants and outcome of treatment assignment

<table>
<thead>
<tr>
<th></th>
<th>SP (n = 138)</th>
<th>Placebo (n = 141)</th>
<th>Difference (95% CI)</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean age (years)</td>
<td>34.6 (25.2–43.5)</td>
<td>34.9 (25.2–41.3)</td>
<td></td>
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<tr>
<td>Etiology of infertility (n/total)</td>
<td></td>
<td></td>
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<tr>
<td>Male factor</td>
<td>49/138</td>
<td>50/141</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tubal factor</td>
<td>19/138</td>
<td>15/141</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Idiopathic</td>
<td>5/138</td>
<td>10/141</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Others</td>
<td>8/138</td>
<td>7/141</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Unknown</td>
<td>57/138</td>
<td>59/141</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Long agonist protocol</td>
<td>134/138</td>
<td>133/141</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Short agonist protocol</td>
<td>0</td>
<td>2/141</td>
<td></td>
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<tr>
<td>Antagonist protocol</td>
<td>4/138</td>
<td>6/141</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Patients with previous implantation failure (n/total)</td>
<td>70/138 (Stratum I)</td>
<td>72/141 (Stratum I)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>IVF (n/total)</td>
<td>56/138</td>
<td>65/141</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ICSI (n/total)</td>
<td>82/138</td>
<td>76/141</td>
<td></td>
<td></td>
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<tr>
<td>Median no. of transferred embryos</td>
<td>1.83 (1–3)</td>
<td>1.86 (1–3)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>No. of blastocystes/ transferred embryo</td>
<td>3.8</td>
<td>3.8</td>
<td></td>
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<tr>
<td>Clinical pregnancy rate: Stratum I (n/total)</td>
<td>15/70 (21.4%)</td>
<td>19/73 (26.0%)</td>
<td>−4.6% (−18.3 to 9.4)</td>
<td>0.56</td>
</tr>
<tr>
<td>Clinical pregnancy rate: Stratum II (n/total)</td>
<td>22/68 (32.4%)</td>
<td>22/68 (32.4%)</td>
<td>0% (−15.4 to 15.4)</td>
<td>1.00</td>
</tr>
<tr>
<td>Total clinical pregnancies (n/total)</td>
<td>37/138 (26.8%)</td>
<td>41/141 (29.1%)</td>
<td>−2.3% (−12.7 to 8.2)</td>
<td>0.69</td>
</tr>
<tr>
<td>Abortions (n/total)</td>
<td>8/138 (5.8%)</td>
<td>6/141 (4.3%)</td>
<td>1.5% (−4.9 to 7.2)</td>
<td>0.59</td>
</tr>
<tr>
<td>Tubal pregnancies (n/total)</td>
<td>1/138 (0.7%)</td>
<td>1/141 (0.7%)</td>
<td>0% (−3.2 to 3.3)</td>
<td>1.00</td>
</tr>
<tr>
<td>Deliveries (n/total)</td>
<td>28/138 (20.3%)</td>
<td>33/141 (23.4%)</td>
<td>−3.1 (−12.7 to 6.6)</td>
<td>0.56</td>
</tr>
</tbody>
</table>

1Previous implantation failure: transfer of ≥6 embryos in ≥3 embryo transfers without a pregnancy.
2Stratum I: expected low pregnancy rate: ≥36 or <36 years of age and transfer of ≥6 embryos in ≥3 embryo transfers without a pregnancy in previously performed IVF/ICSI-cycles. Stratum II: expected high pregnancy rate: all other patients.
3Information about whether live birth occurred is missing for one patient with a clinical pregnancy in the placebo group. This pregnancy was not counted as a live birth. If this pregnancy resulted in live birth, the placebo group live birth rate is 34/141 (24.1%) and the difference between treatments is −3.8% (95% CI: −13.5 to 6.0; P = 0.47).
Firstly, the treatment concepts are different. In the study published in 2009, pure SP was instilled into the cervix, which is very close to the physiological situation during sexual intercourse. In our new study in which SP was injected into the uterine cavity, a treatment concept was chosen which is largely independent, but which represents a non-physiological form of treatment. Indeed, SP has never been proved to ascend in substantial volumes through the cervix into the uterine cavity. In humans, endometrial inflammatory responses induced by SP have only been shown in vitro (Gutsche et al., 2003), whereas cervical inflammatory responses following exposure to ejaculate have been demonstrated in vivo (Sharkey et al., 2012a). In addition, in vitro experiments have revealed that this effect is at least in part due to SP and not just due to semen (Sharkey et al., 2012b). However, even though the intracervical and intruterine treatment approach seems to be very different, in practice, they are possibly not. During the cervical instillation of SP, as performed in the first study, a substantial amount of SP probably also enters the uterine cavity. It can therefore be speculated that in both studies the effect of undiluted intracervical SP and diluted intruterine SP would exert the same effect on the uterine endometrium. As the effect on the cervix and the vagina was however much more pronounced in the first study and in the study by Chicea et al. (2013), this suggests, that in humans SP does act but more via the cervix and vagina than via the uterine cavity.

Furthermore, it could be speculated that the passage of the transfer catheter through the cervix already would have a positive effect, through which the positive effect of the SP was overshadowed. Meta-analyses have shown that a hysteroscopy, even without proof of any pathophysiological results, significantly increases the pregnancy rate of a subsequent IVF treatment (El-Touky et al., 2008). In addition, several studies have demonstrated an increased pregnancy rate after endometrial injury before IVF treatment (Nastri et al., 2012). This effect was not seen after endometrial injury at the time of follicle puncture. However, as the introduction of a transfer catheter possibly also causes some cervical trauma, it might be speculated that this would have a positive effect in our new study, although in reality such an effect was not observed.

Finally, there is the possibility that intracavitary treatment with SP does not have any clinically relevant immunomodulatory effects on the endometrium at all or that the dilution of SP reduced the effect to such an extent that it was not pronounced enough to have a substantial effect on endometrial function.

Indeed, the present study suggests that SP instillation into the uterine cavity is unlikely to increase the chance of clinical pregnancy, at least not by a rate ≥ 14% as initially hypothesized.

### Authors’ roles

M.V. was responsible for the study conception and design, acquisition, analysis and interpretation of data and drafting the article. S.R., A.G. and J.J. were responsible for acquisition and analysis of data and revising the article. G.G. was responsible for analysis and interpretation of data and revising the article. T.S. was responsible for the study conception and revising the article. All authors gave final approval of the manuscript.

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### Conflict of interest

There is no conflict of interest of any of the authors.

### References


