SU2C Phase Ib Study of Paclitaxel and MK-2206 in Advanced Solid Tumors and Metastatic Breast Cancer


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Abstract

Background: There is preclinical synergism between taxanes and MK-2206. We aim to determine the maximum tolerated dose, safety, and activity of combining MK-2206 and paclitaxel in metastatic cancer.

Methods: Patients received weekly doses of paclitaxel at 80 mg/m2 on day 1, followed by MK-2206 orally on day 2 escalated at 90 mg, 135 mg, and 200 mg. Treatment continued until progression, excessive toxicity, or patient request. Blood and tissue were collected for pharmacokinetic and pharmacodynamics markers. A cycle consisted of three weeks of therapy. Dose-limiting toxicity (DLT) was defined as unacceptable toxicity during the first cycle. All statistical tests were two-sided.

Results: Twenty-two patients were treated, nine in dose escalation and 13 in dose expansion. Median age was 55 years. Median number of cycles was four. Dose escalation was completed with no DLT. CTCAE Grade 3 or higher adverse events were fatigue (n = 2), rash (n = 2), hyperglycemia (n = 1), and neutropenia (n = 7). Four patients in the expansion phase required MK-2206 dose reduction. Phase II recommended dose was established as paclitaxel 80 mg/m2 weekly on day 1, and MK-2206 135 mg weekly on day 2. Paclitaxel systemic exposure was similar in the presence or absence of MK-2206. Plasma MK-2206 concentrations were similar to data from previous phase I monotherapy. There was a statistically significant decrease in expression of pAKT S473 (P = .01) and pAKT T308 (P = .002) after therapy. PI3K/AKT/mTOR downregulation in tumor tissues and circulating markers did not correlate with tumor response or clinical benefit. There were five objective responses, and nine patients had stable disease.

Conclusion: MK-2206 was well tolerated with paclitaxel. Preliminary antitumor activity was documented.

The PI3K/AKT/mTOR pathway is downstream of most growth factor tyrosine kinase receptors (TKRs) in cancer. It plays a key role in cell growth, protein translation, autophagy, metabolism, and cell survival (1,2). Pathway deregulation may occur through overexpression or activation of TKR, mutations and amplification of PIK3CA or AKT, and loss of negative regulators PTEN and INPP4B. Increased levels of phosphor-AKT and PTEN loss are poor outcome predictors (3). We showed that of 547 breast
cancers tested, 117 (21.4%) had mutations in PIK3CA (4). In breast cancer cells, PTEN levels inversely correlated with AKT phosphorylation (5). Thus, PTEN-low tumors and PIK3CA mutant tumors may rely on AKT for oncogenic signaling. Therefore, AKT inhibitors may have a broader utility than TKR inhibitors.

Preclinical work with MK-2206 shows that many PIK3CA mutant and PTEN loss lines are sensitive (6). Loss of PTEN and PI3K signaling activation are associated with resistance to endocrine therapy and trastuzumab (7–9). MK-2206 showed activity with improvement in breast cancer metastasis (10). In preclinical studies, MK-2206 demonstrated synergy with paclitaxel, and the combination had greater in vivo antitumor efficacy (6). Synergistic or additive inhibitory effects were also observed with docetaxel. Synergism was sequence-dependent and occurred when cells were treated with docetaxel followed by MK-2206 (11). Metabolism of MK-2206 in human liver is catalyzed by the cytochrome P450 3A4 isoenzyme (CYP3A4), as is docetaxel. In our previous phase I study using everolimus, there was a statistically significant pharmacokinetic (PK) interaction when combined with docetaxel, with severe adverse events (AEs) (12). Conversely, the same combination with paclitaxel had no PK interaction in our phase II neoadjuvant breast cancer trial (13).

The purpose of this study was to determine the MTD of the combination of weekly MK-2206 and paclitaxel (escalation) and to determine the safety and antitumor activity of the combination in metastatic breast cancer (expansion). Secondary objectives included PK of the combination, baseline molecular markers and pharmacodynamic markers in blood, and tumor tissue that may predict clinical activity.

Methods

The study was an open-label phase I study combining weekly paclitaxel with MK-2206 in advanced solid tumors with an expansion in advanced breast cancer (NCT01263145). Eligible patients had histologically confirmed metastatic tumors that had failed at least two therapy lines (escalation) and metastatic breast cancer that had progressed after maximum three therapy lines (expansion). Patients had measurable disease by Response Evaluation Criteria in Solid Tumors (RECIST) version 1.0 or evaluable disease (14), were age 18 years or older, had adequate organ function including HgbA1c under 8%, Eastern Cooperative Oncology Group (ECOG) performance status (PS) 0–2. Prior treatment with PI3K pathway inhibitors and paclitaxel for early disease was permitted. Patients were excluded if pregnant, breastfeeding, or taking CYP3A4 inducers or inhibitors. Washout period was 21 days. Radiographic evaluations were performed every nine weeks. The clinical trial was reviewed yearly and approved by institutional review boards. Patients provided written informed consent.

Study Therapy

MK-2206 was provided by Cancer Therapy Evaluation Program (CTEP), and paclitaxel was commercially available. Participants were considered for three dose-escalation levels and for a dose-expansion cohort once MTD was established. Paclitaxel was given at a fixed dose of 80 mg/m2 intravenously (IV) weekly on day 1, and MK-2206 was escalated at 90 mg, 135 mg, and 200 mg orally weekly on day 2. Premedication for paclitaxel consisted of dexamethasone 10 mg on week 1, 4 mg IV on week 2, and discontinued after if no infusion reaction occurred. Once MTD was reached, patients with metastatic breast cancer were treated. Cycle length was three weeks, and treatment was continued until disease progression, unacceptable toxicity, patient refusal, or physician’s decision.

Safety and Efficacy

Safety assessments were conducted at baseline, at weekly basis during the first cycle, then every cycle or earlier if toxicity occurred. Patients removed from study for AEs were followed until resolution or stabilization. Toxicity was graded according to National Cancer Institute Common Terminology Criteria for Adverse Events (CTCAE), version 4.0. A DLT was defined as any grade 3/4 nonhematologic toxicity, grade 3/4 rash or hyperglycemia lasting more than 72 hours, grade 4 febrile neutropenia that required hospitalization, and any grade 3 hematologic toxicity that required treatment delay beyond two weeks. If toxicity occurred, an appropriate treatment was used. If grade 3 or 4 toxicity occurred, drug was held until the toxicity was grade 1 or less. If grade 3 or 4 toxicity recurred, continuation of treatment was discussed with a decision made after consideration of relative risks and benefits. Therapy was restarted at a -1 dose level (MK-2206 of 135 mg PO weekly); if a second grade 3 or 4 toxicity occurred, therapy was stopped. Paclitaxel doses were reduced by 25% (60 mg/m2 IV weekly) after 12 weeks of therapy if patients develop grade 2 or greater neuropathy or nail changes.

All patients underwent anatomic imaging at presentation and every three cycles to establish response. A decrease in the size of the sum of the diameters of the index lesions of greater than or equal to 30% was considered a partial response (PR). Stable disease (SD) was considered if stability was documented at least once for four or more weeks. Clinical benefit was documented if PR or SD response lasted for 18 weeks (six cycles). Progression of disease (PD) was defined as 20% or greater increase of the index lesions or appearance of new lesions.

Pharmacokinetic Analysis

For paclitaxel, blood samples were drawn on days 1 and 15 of cycle 1 at predose, 10 minutes and 50 minutes following infusion, and then 10 minutes, 30 minutes, one hour, two hours, four hours, eight hours, and 24 hours after infusion completion. MK-2206 samples were collected on day 2 of cycle 1 at predose, 10 minutes and 50 minutes following infusion, and then 10 minutes, 30 minutes, one hour, two hours, four hours, six hours, eight hours, 24 hours, 72 hours, and 144 hours after MK-2206 dose. For paclitaxel, blood samples were drawn on days 1 and 15 of cycle 1 at predose, 10 minutes and 50 minutes following infusion, and then 10 minutes, 30 minutes, one hour, two hours, four hours, eight hours, and 24 hours after infusion completion. MK-2206 samples were collected on day 2 of cycle 1 at predose, one hour, two hours, four hours, six hours, eight hours, 24 hours, 72 hours, and 144 hours after MK-2206 dose; on day 16 of cycle 1 at predose, one hour, two hours, four hours, eight hours, and 24 hours after infusion completion. MK-2206 samples were collected on day 2 of cycle 1 at predose, 10 minutes and 50 minutes following infusion, and then 10 minutes, 30 minutes, one hour, two hours, four hours, six hours, eight hours, 24 hours, 72 hours, and 144 hours after MK-2206 dose; on day 16 of cycle 1 at predose, one hour, two hours, four hours, six hours, eight hours, 24 hours, 72 hours, and 144 hours after MK-2206 dose. Individual plasma concentration time data for paclitaxel and MK-2206 were used to generate PK parameter estimates using compartmental and noncompartmental methods utilizing WinNonLin Professional 5.3 (Pharsight Corp., St. Louis, MO). Peak plasma concentration (Cmax) and time to peak concentration (Tmax) were determined by data observation. Areas under the plasma concentration time curve (AUC) from zero to 24 hours postdose (AUC0-24), zero to 144 hours postdose (AUC0-144), and zero to infinity (AUC0-inf) were calculated by the linear trapezoidal method. Drug clearance (Cl) was determined by dose/AUC. Elimination half-life (t1/2) was calculated by 0.693/k, and apparent volume of distribution was calculated by Cl/k (15).
Biomarker Assessment

Fine-needle aspirations (FNAs) and core biopsies were obtained pretreatment and at two weeks. Samples were evaluated by reverse phase protein arrays (RPPA) to assess PI3K activation status. The antibodies used are listed in Supplementary Table 1 (available online). Core biopsies were assessed for PTEN and INPP4B expression by immunohistochemistry (IHC) and mutations at the PI3K pathway genes. Whole blood for pharmacodynamics assessments was collected at least at five time points: C1-D1 (pretreatment), C1-D2, C1-D3, C1-D5, C1-D8, C1-D15, C1-D16, C1-D17, C1-D19, and C2-D1 (prior to next dose). Peripheral blood mononuclear cells (PBMCs) and platelet-enriched plasma (PRP) were obtained and evaluated by RPPA. The full biomarker methodology including bioinformatics analysis is described in the Supplementary Methods (available online).

Statistical Analysis

For RPPA data analysis, paired t test was used to examine the difference between baseline and post-treatment samples by dose level (details are presented in the Supplementary Materials, available online). Two sample two-sided t tests were used to test the association between: 1) baseline protein expression, 2) pharmacodynamic changes (FDX), and 3) PI3K activity score with tumor response or clinical benefit. We defined FDX as the difference on protein expression after therapy exposure (FDX = post-treatment expression – pretreatment expression in tumor tissues and day 2-day 1, day 3-day1, and day 3-day 2 for circulating markers). PI3K activity score was defined as the sum of the phosphor-protein expression levels of AKT, mTOR, 4EBP1, S6K, and S6 (PI3K score = pS6-240/244 + pS6-235/245 [16] + pS6K-T389 + pmTOR-S2448 + p4EBP1-S65 + p4EBP1-T37/46 + pPRAS40-T246 + pAKT-S473 + pAKT-T308). For multiplicity adjustments, the Benjamini Hochberg (BH) (16) procedure was employed to control the False Discovery Rate (FDR). We used thresholds 0.3 or 0.1 for FDR to calibrate for the top proteins of interest. Fisher’s exact tests were performed to test the association of INPP4B, PTEN, or either protein presence or loss with tumor response or clinical benefit. A P value of less than .05 was considered statistically significant.

Results

Patients Characteristics and Disposition

Supplementary Figure 1 (available online) summarizes the patient disposition. Twenty-two patients were enrolled, nine patients in dose escalation, and 13 patients in dose expansion. Three patients in the dose expansion were replaced because of lack of compliance (did not show up or receive any treatment), early progression (new metastasis documented within the first week of therapy) and severe therapy-related toxicities at first dose. Table 1 summarizes the clinical characteristics of all patients. Median age was 55 years (range 34–79 years). There were five men and 17 women. All patients but one had an ECOG PS of 0 or 1. In addition to breast cancer (n = 14), other primary tumors included colorectal (n = 2), ovarian (n = 1), endometrial (n = 1), ocular melanoma (n = 2), and squamous cell carcinoma of the head and neck (n = 2). Median previous therapy lines were 3 (range 1–10) for dose escalation and 1 (range 0–3) for expansion.

Table 1. Patient and tumor characteristics*

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* ECOG = Eastern Cooperative Oncology Group.
Table 2. Treatment-related adverse events

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after MK-2206 first dose and required admission to the hospital for supportive management.

Pharmacokinetics

Plasma concentration time profiles for paclitaxel on C1D1 and C1D15 following 80 mg/m2 IV infusion over one hour on a weekly schedule in combination with 90 mg or 135 mg MK-2206 orally were similar with polyphasic elimination after achieving similar peak concentrations at the end of drug administration (Supplementary Figure 2, available online). Following a noncompartmental analysis to fit the concentration time data, all PK parameters estimated were not statistically significantly different between days or doses across both agents (Table 3). After two weekly doses of 200 mg MK-2206 on C1-D2 and C1-D9, mean paclitaxel exposure on C1-D15 measured by both AUC0–24 and AUC0–inf increased by 30% to 40% in addition to a more than two-fold increase in mean peak concentration when compared with 135 mg MK-2206. These PK changes in paclitaxel on C1-D15 are most likely caused by the increased dose of MK-2206 but the interpatient variability is high.

In published data, a mean paclitaxel Cmax of 2000–2500 ng/mL was observed and AUC did not exceed 5000 hr•ng/mL (17–19). There were no changes in paclitaxel terminal half-life. MK-2206 PK on a weekly administration schedule at all dose levels when combined with paclitaxel was characterized by a relatively slow absorption (mean T0.5 range = 4.8–6.8 hours) with a subtle biphasic elimination (Supplementary Figure 3, available online) leading to a mean terminal half-life of 50 to 60 hours, consistent with previous
Table 3. Mean pharmacokinetic parameters estimated for paclitaxel and MK-2206

<table>
<thead>
<tr>
<th>MK-2206 parameters</th>
<th>90 mg Qweekly</th>
<th>135 mg Qweekly</th>
<th>200 mg Qweekly</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>C1D2 (n = 3)</td>
<td>SD</td>
<td>C1D16 (n = 3)</td>
</tr>
<tr>
<td>AUC$_{0-24}$, nM.hr</td>
<td>2589.8*</td>
<td>N/A</td>
<td>3206.9</td>
</tr>
<tr>
<td>AUC$_{0-144}$, nM.hr</td>
<td>N/A</td>
<td>N/A</td>
<td>N/A</td>
</tr>
<tr>
<td>AUC$_{0-inf}$, nM.hr</td>
<td>1597.6*</td>
<td>N/A</td>
<td>22335.9</td>
</tr>
<tr>
<td>$C_{mean}$, nM</td>
<td>123.7</td>
<td>25.9</td>
<td>167.7</td>
</tr>
<tr>
<td>$t_{1/2}$, hr†</td>
<td>56.9*</td>
<td>N/A</td>
<td>49.9</td>
</tr>
<tr>
<td>$T_{max}$, hr†</td>
<td>190.0*</td>
<td>N/A</td>
<td>21.4</td>
</tr>
<tr>
<td>$V_{z}$, L</td>
<td>1563.0*</td>
<td>N/A</td>
<td>1399.9</td>
</tr>
<tr>
<td>Paclitaxel parameters</td>
<td>C1D1 (n = 3)</td>
<td>SD</td>
<td>C1D15 (n = 3)</td>
</tr>
<tr>
<td>AUC$_{0-24}$, hr.ng/mL</td>
<td>6780.1</td>
<td>2414.1</td>
<td>5036.2</td>
</tr>
<tr>
<td>AUC$_{0-inf}$, hr.ng/mL</td>
<td>7375.6</td>
<td>2881.2</td>
<td>5639.4</td>
</tr>
<tr>
<td>$C_{mean}$, ng/mL</td>
<td>4323.3</td>
<td>620.0</td>
<td>2408.9</td>
</tr>
<tr>
<td>$t_{1/2}$, hr</td>
<td>6.4</td>
<td>2.8</td>
<td>10.7</td>
</tr>
<tr>
<td>$Cl$, L/hr/m²</td>
<td>12.0</td>
<td>4.3</td>
<td>14.2</td>
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<tr>
<td>$V_{z}$, L/m²</td>
<td>99.7</td>
<td>25.6</td>
<td>222.2</td>
</tr>
</tbody>
</table>

* n = 1
† Harmonic mean ± pseudo standard deviation is reported. AUC = area under the curve.
‡ $t_{1/2}$ using four- to 24-hour postdose timepoints reported. Value in parentheses for C1D16 includes 72- and 144-hour time points.
single-agent data (10). Peak concentration and systemic exposure of MK-2206 were dose dependent. Mean peak concentrations and AUC\textsubscript{0-144hr} at 90 mg and 135 mg dose levels following the third dose at C1-D16 were similar to single agent data after steady state was reached (10). At a 200 mg dose, mean systemic exposure over 144 hours at C1-D2 (24,245 nM•hr) was similar to previous single agent data for 300 mg weekly (AUC\textsubscript{0-168hr}, 27,700 nM•hr) that led to three out of three patients with grade 3 skin rash. Less than a 2.5-fold increase in drug accumulation was observed based on AUC\textsubscript{0-24} and AUC\textsubscript{0-inf} systemic exposure comparison between C1-D2 and C1-D16, which is similar to published data (10,15,19).

**Antitumor Activity**

From the 21 patients who received therapy, there were five PR and nine SD. Three responses occurred in the expansion cohort and two in dose-escalation group, one in a patient with metastatic breast cancer and the other one in a patient with metastatic colorectal cancer. SD was documented in three patients in expansion cohort and in six patients in the dose-escalation cohort: two squamous cell carcinomas of the head and neck, two ocular melanomas, one ovarian, and one endometrial cancer. Maximum percentage of disease change by RECIST in patients with at least one follow-up scan is presented in Figure 1.

**Pharmacodynamics and Biomarker Analysis**

**Tumor Tissues**

Samples for RPPA were available from 21 patients at baseline and 16 patients post-treatment. Outcome information was available in 18 patients. There was a statistically significant difference between baseline and post-treatment expression of pAKT-S473 (P = .01) and pAKT-T308 (P = .002) (Figure 2A). However, at a false discovery rate (FDR) threshold of 0.1, there were no statistically significant differences in protein expression from baseline to post-treatment in any dose levels. At baseline level, no protein was found differentially expressed for tumors that had responses vs no responses. Two proteins were found differentially expressed among patients who had clinical benefit vs no benefit: pPKC\textsubscript{α}-S657 (P < .001) and VEGFR2 (P = .003). There was a statistically significant association of pharmacodynamic changes (PD\textsubscript{β}) of p27 with response (P < .001). There were no PD\textsubscript{β}-associated with clinical benefit. There were no statistically significant associations between baseline PI3K activity score with response

![Figure 1](https://academic.oup.com/jnci/article-abstract/107/3/dju493/925937)

Figure 1. Maximum percentage change of target tumor from baseline by Response Evaluation Criteria in Solid Tumors in patients with at least one follow-up scan.

![Figure 2](https://academic.oup.com/jnci/article-abstract/107/3/dju493/925937)

Figure 2. A) Differences between baseline and post-treatment expression of pAKT-S473 (P = .01), and pAKT-T308 (P = .003) with two-sided t test. B) Baseline PI3K activity scores showing no associations with tumor response (means -20.464 vs -21.801 for response and nonresponse, respectively, P = .7), or clinical benefit (Means -21.311 vs -21.846 for benefit vs no benefit, respectively, P = .07). Analysis was done with two-sided t test, P < .05 was considered statistically significant.
(means -19.0 vs -20.5 for response and nonresponse, respectively, \( P = .70 \)) or clinical benefit (Means -19.9 vs -20.7 for benefit vs no benefit, respectively, \( P = .63 \)) (Figure 2B). When assessing PI3K/ activation score from baseline to post-treatment, there was no association of pathway downregulation and response or clinical benefit (\( P = .28 \) and >.99, respectively) (Supplementary Tables 2 and 3, available online). Lastly, at an FDR threshold of 0.1, PDX changes were found not to be dose dependent.

Baseline tissues for IHC were available in 16 patients. PTEN loss was documented in three tumors and INPP4B loss in nine tumors. Two tumors had both PTEN and INPP4B loss. There was no association of INPP4B, PTEN or either protein loss with response (\( P > .99 \) for all cases), or clinical benefit (\( P > .99, .45, \) and >.99, respectively).

Adequate quality DNA for sequencing was available in 10 patients. A \( \text{PIK3CA} \) E545K mutation was detected in a patient with head and neck cancer who had SD for 10 weeks. No AKT or PTEN mutations were found.

**Circulating Markers**

Whole blood was collected on day 1 (pretreatment), day 2 (post-paclitaxel but pre-MK-2206) and day 3 (post-paclitaxel and MK-2206). Blood was available in 17 patients, and outcome information was available in 15. Comparisons were made from day 2 to day 1, day 3 to day 1, and day 3 to day 2.

In PRPs, at an FDR threshold of 0.1, there was a statistically significant decrease of pAKT-S473, pAKT-T308, and p70S6K-T389, (\( P < .001 \) in all) from day 2 to day 3 (Figure 3A). There was no association of PDX (day 2-day 1, day 3-day 1 or day 3-day 2) with tumor response. There were no statistically significant associations between baseline PI3K activity score with tumor response (Means -8.6 vs -9.3 for response and nonresponse, -0.5 vs -0.95 for tumor response vs nonresponse, \( P < .001 \) in all).

### Figure 3.

A) Decline of pAKT-T308 and pAKT-S473 in platelets on day 3, after treatment with paclitaxel + MK-2206 (MK-2206 administered D2). B) Dose dependence of pAKT levels at day 3 (D3) or by measuring pharmacodynamic (PD) changes from day 2 to day 3 (D3-D2) or from day 1 to day 3 (D3-D1).
Thus, extent of pathway inhibition was not a predictor of outcome, but rather tumors may differ in the extent they rely on PI3K signaling. Further studies with adequate power are needed to determine whether even greater pathway inhibition could further improve antitumor efficacy of AKT inhibitors. There was no association of INPP4B or PTEN loss with tumor response or stable disease. We found a single PIK3CA mutation. In the MK-2206/trastuzumab phase I study, all 37 patients had baseline analysis of circulating DNA for PIK3CA mutations. Only three PIK3CA mutations were found, but their analysis did not confirm the hypothesis that tumors with PIK3CA mutations are more sensitive to MK-2206 (20). At this point, our data does not support patient selection by the PI3K aberrations explored. The correlative work of I SPY2 results may show other predictors. Other molecular aberrations or a systems biology approach should be considered for optimal biomarker discovery.

Our study had some limitations. We studied weekly dosing of MK2206, as we expected that weekly dosing would be more tolerable than daily dosing, but we cannot exclude the possibility that other intermittent schedules such as every other day or every third day may not be tolerable and more efficacious. Further, we have not explored the safety, tolerability of different timing of delivery. In the dose expansion, we included tumors of different histologic types and genomic backgrounds; this and escalating doses limit our ability to assess efficacy. Although we were successful in getting pretreatment and on-treatment biopsies, the sample size and number of responders limit our ability to make significant correlations with outcome. Our biomarker analysis has shown inhibition of Akt phosphorylation, but whether this is sufficient target inhibition requires more study.

Our results show evidence of antitumor activity of the combination of paclitaxel and MK-2206 in solid tumors and in breast cancer, and the combination was well tolerated. Paclitaxel did not affect the PK profile of MK-2206 at 135 mg/week, suggesting that this AKT inhibitor can be safely combined with paclitaxel. Our data does not support patient selection by the PI3K aberrations explored, however this analysis was limited by sample size.

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Notes

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References