Analysis of tetracycline resistance \textit{tet}(W) genes and their flanking sequences in intestinal \textit{Bifidobacterium} species

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Objectives: The \textit{tet}(W) gene provides tetracycline resistance to a wide range of anaerobic intestinal and ruminal bacteria, but little is known about the molecular organization of the \textit{tet}(W) gene. The aim of this study was to gain new insights into the molecular organization of the \textit{tet}(W) gene in bifidobacteria strains from humans.

Methods: A segment of DNA encompassing the whole \textit{tet}(W) gene and its immediate upstream and downstream sequences was analysed in 10 representative strains of four \textit{Bifidobacterium} species, of which two have been shown to be tetracycline-susceptible. The non-conserved flanking regions of the \textit{tet}(W) gene were further analysed in six strains.

Results: All 10 strains share a core DNA domain of 2154 bp [starting 250 bp upstream of the \textit{tet}(W) gene start codon and ending 13 bp before the stop codon] with 98% to 100% DNA identity. Except for \textit{Bifidobacterium animalis} E43, all other strains further share 408 bp upstream and 70 bp downstream of the \textit{tet}(W) gene. An insertion-like element of 736 bp was found to interrupt the \textit{tet}(W) coding sequence in \textit{Bifidobacterium longum} M21, which may be the reason for its tetracycline susceptibility. However, genetic events explaining the susceptible phenotype of \textit{B. longum} LMG 13197 were not observed.

Conclusions: The \textit{tet}(W) genes from all 10 strains shared 98% to 100% DNA and amino acid identity, though large variation was found in their flanking regions.

Keywords: bifidobacteria, antibiotic resistance, probiotics

Introduction

The commensal intestinal microbiota of humans and animals may act as a reservoir of antibiotic resistance (AR) genes that could ultimately be transferred to pathogens. In fact, the transfer of AR genes has been shown to occur between bacterial species in the gastrointestinal tract (GIT) of mammals. Bifidobacteria are among the dominant bacterial populations of the human GIT, where they are thought to exert several health-promoting effects. Accordingly, species and strains of this genus are frequently used as probiotics in dairy products or dietary supplements. It is recommended that any strain used in food systems should be free of potentially transferable AR traits.

Of the 38 tetracycline resistance (\textit{Te}c) genes already described (http://faculty.washington.edu/marilynr/tetweb1.pdf), \textit{tet}(W), which encodes a ribosomal protection protein, is the most widespread \textit{Te}c gene in anaerobic gut and rumen bacteria. Indeed, it has recently been reported in \textit{Bifidobacterium} species. However, large variations in the MIC of tetracycline have repeatedly been reported in strains harbouring a \textit{tet}(W) gene. Also intriguing is the presence of susceptible strains in which the gene has been detected by either PCR or hybridization. For all these reasons, the molecular analysis of the \textit{tet}(W) genes in bifidobacteria has become the subject of much interest.

In a recent survey, we showed that chromosomally encoded \textit{tet}(W) genes are present in tetracycline-resistant and tetracycline-susceptible species of bifidobacteria isolated from the GIT of healthy humans not subjected to recent antibiotic treatment. In this paper, we report on the molecular characterization of the \textit{tet}(W) gene and its flanking regions in 10 representative strains belonging to four bifidobacterial species—\textit{Bifidobacterium}...
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animalis, Bifidobacterium bifidum, Bifidobacterium longum and Bifidobacterium thermophilum—of which 8 were tetracycline-resistant and 2 were tetracycline-susceptible. The aim was to investigate the molecular organization of the gene in both tetracycline-resistant and -susceptible strains, in order to supply the basis for a correlation between genotype and phenotype, and obtain information on the mechanism of this gene's spread. The analysis included non-shared regions upstream and downstream of tet(W) genes in six strains.

Materials and methods

Bacterial strains, media and culture conditions

Except for B. thermophilum LMG 21813T and B. longum LMG 13197T, which were kindly provided by the Belgian Coordinated Collections of Microorganisms (BCCM™, University of Ghent, Ghent, Belgium), all strains analysed in this work were isolated as part of a project devoted to the identification and typing of dominant intestinal microbial populations in healthy Spaniards. With the exception of the tetracycline-susceptible (Tc') strains B. longum M21 and B. longum LMG 13197T, all were considered to be Tc' strains. MICs of tetracycline for Tc' strains varied from 4 to 64 mg/L as determined by the Etest method (Table 1), when compared with an MIC ranging of tetracycline for Tcr strains varied from 2 to 64 mg/L as determined by the Etest method.

A positive tet(W)-specific PCR product and the sequencing of the resulting amplicons have suggested the presence of at least part of the tet(W) gene sequence in all Tc' and Tc' strains. All strains were maintained at ~80 °C until use, when they were recovered and cultured on MRS agar (Oxoid, Hampshire, UK) supplemented with 0.3 g/L cysteine–HCl (Sigma Chemical, St Louis, MO, USA) (MRS + C) [plus 4 mg/L tetracycline (Sigma) for the Tc' strains]. Incubations were performed at 37°C in an anaerobic chamber (Mac500, Don Whitley Scientific, West Yorkshire, UK) (atmosphere 10% H₂, 10% CO₂ and 80% N₂) for 48 h.

DNA techniques

Total genomic DNA was isolated from overnight cultures using the Wizard Genomic DNA Purification Kit (Promega Corporation, Southampton, UK). Plasmid DNA was isolated following the procedure of O’Sullivan and Klaenhammer. The general procedures for DNA manipulation, electrophoresis and visualization were essentially as described by Sambrook and Russell. Restriction endonucleases and Taq DNA polymerase from Takara (Otsu, Shiga, Japan) and T4 DNA ligase from Invitrogen (Carlsbad, CA, USA) were used according to the manufacturers’ instructions.

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Table 1. Strains used and their tetracycline resistance phenotypes

<table>
<thead>
<tr>
<th>Bifidobacterium strains</th>
<th>Source</th>
<th>Tetracycline resistance phenotype</th>
<th>MICa (mg/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td>B. animalis E43</td>
<td>human faeces</td>
<td>Tc'</td>
<td>4</td>
</tr>
<tr>
<td>B. bifidum L22</td>
<td>human faeces</td>
<td>Tc'</td>
<td>32</td>
</tr>
<tr>
<td>B. longum B93</td>
<td>human faeces</td>
<td>Tc'</td>
<td>32</td>
</tr>
<tr>
<td>B. longum B94</td>
<td>human faeces</td>
<td>Tc'</td>
<td>32</td>
</tr>
<tr>
<td>B. longum E111</td>
<td>human faeces</td>
<td>Tc'</td>
<td>16</td>
</tr>
<tr>
<td>B. longum H66</td>
<td>human faeces</td>
<td>Tc'</td>
<td>24</td>
</tr>
<tr>
<td>B. longum L42</td>
<td>human faeces</td>
<td>Tc'</td>
<td>12</td>
</tr>
<tr>
<td>B. longum M21</td>
<td>human faeces</td>
<td>Tc'</td>
<td>1</td>
</tr>
<tr>
<td>B. longum LMG 13197T</td>
<td>adult intestine</td>
<td>Tc'</td>
<td>0.75</td>
</tr>
<tr>
<td>B. thermophilum LMG 21813T</td>
<td>pig faeces</td>
<td>Tc'</td>
<td>64</td>
</tr>
</tbody>
</table>

Tc', tetracycline resistant; Tc', tetracycline susceptible (underlined). MIC, minimum inhibitory concentration as determined by the Etest method.

Sequencing of tet(W) genes and flanking sequences

Internal segments of the tet(W) genes from all strains had already been amplified as reported previously, using the universal primers based on Tc' genes encoding ribosomal protection proteins. The amplification and sequencing of the fragments were performed with the specific primers, tetW and tet2, for the tet(W) gene. Genome walking was performed from these internal regions of the gene to obtain the whole gene, and flanking upstream and downstream sequences were obtained by direct and inverse PCR strategies using a combination of primers (Table 2). For inverse PCR, total genomic DNA was digested with NcoI, ClaI or PsI and self-ligated overnight. The following morning, the ligated DNA was precipitated, centrifuged, and suspended in 100 μL of TE prior to use as a template in PCR. Appropriate pairs of primers (Table 2) were used to amplify the upstream and downstream region of the gene. Amplicons were purified using the GFX PCR DNA and Gel Band Purification Kit (GE Healthcare Biosciences, Buckinghamshire, UK) and sequenced by cycle extension in an ABI 370 DNA sequencer (Applied Biosystems, Foster City, CA, USA). Finally, sequences were assembled and aligned using the DNAMAN program (Lynnon Biosoft, Pointe-Claire, Quebec, Canada), and the nucleotide and deduced protein sequences were compared with those in the GenBank database using the BLAST program (National Center for Biotechnology Information of US National Library of Medicine; available at http://www.ncbi.nlm.nih.gov/blast/blast.cgi).

Copy number of tet(W) genes

The copy number of the tet(W) gene was determined using real-time PCR. Genomic DNAs were amplified with the tet(W) gene's internal primers tetW-1 and tetW-2 and with the single-copy housekeeping tuf gene's internal primers Bif-tuf-1 and Bif-tuf-2 (Table 2). PCR was performed in an ABI Prism 7500 Fast Real-Time PCR System (Applied Biosystems), SYBR Green I fluorophore was used to correlate the amount of PCR product with the fluorescent signal. Amplification was performed in a 25 μL final volume containing 50 ng of chromosomal DNA as a template, 10 nM of each primer and 12.5 μL of SYBR Green PCR Master Mix (Applied Biosystems). The amplification reaction was performed as follows: 2 min at 50°C, 10 min at 95°C for activating the AmpliTaq Gold DNA polymerase and forty cycles of: (i) denaturing at 95°C for 15 s; (ii) annealing/extension at 60°C for 1 min; and (iii) a dissociation step. The cycle number during which the fluorescence signal crossed the chosen cycle threshold (Ct; in this case, the default was set by the manufacturer) was recorded. All samples were processed in duplicate using independent DNA samples. Negative controls involving all the elements of the reaction mixture except the template DNA were also included.

Nucleotide accession numbers

The nucleotide sequences described in this paper were deposited in the GenBank database with the following accession numbers: B. animalis E43, EU434754; B. bifidum L22, EU434755; B. longum
Table 2. Primers used in this study

<table>
<thead>
<tr>
<th>Primer (forward/reverse)</th>
<th>Sequence (5’–3’)</th>
<th>Specificity</th>
</tr>
</thead>
<tbody>
<tr>
<td>DI(F)</td>
<td>GAYACICCGGGCAYRTIGAYTT</td>
<td>universal ribosomal protection-type Tc^R genes</td>
</tr>
<tr>
<td>DI(R)</td>
<td>CCRWAIGGGTRTGIGGIGIACYTG</td>
<td>universal ribosomal protection-type Tc^R genes</td>
</tr>
<tr>
<td>tet1(F)</td>
<td>GCTCAYGTGAYGCAAGAAGA</td>
<td>universal ribosomal protection-type Tc^R genes</td>
</tr>
<tr>
<td>tet2(R)</td>
<td>AGGTGAAAGCTGAGGTTGTTG</td>
<td>universal ribosomal protection-type Tc^R genes</td>
</tr>
<tr>
<td>tet(W)</td>
<td>AAGCGGCAGTCACTTCCTTCC</td>
<td>tet(W) gene</td>
</tr>
<tr>
<td>Utet(W)(F)</td>
<td>AATGCTCATGTACGGTAAG</td>
<td>shared sequence of the tet(W) loci</td>
</tr>
<tr>
<td>Detet(W)(R)</td>
<td>TTACATTACCTCTGAAAC</td>
<td>shared sequence of the tet(W) loci</td>
</tr>
<tr>
<td>tetW-1(F)</td>
<td>GCCTATCTTGGTCTTGG</td>
<td>upstream of tet(W)</td>
</tr>
<tr>
<td>tetW-2(R)</td>
<td>GACAGGCAGTCTGAGGTTG</td>
<td>tet(W) gene</td>
</tr>
<tr>
<td>tetW-2(F)</td>
<td>GACAGGCAGTCTGAGGTTG</td>
<td>tet(W)</td>
</tr>
<tr>
<td>Bif-tuf-1(F)^a</td>
<td>GACAACGCGTCTCTGATGC</td>
<td>B. longum tuf gene</td>
</tr>
<tr>
<td>Bif-tuf-2(R)^a</td>
<td>GACAACGCGTCTCTGATGC</td>
<td>B. longum tuf gene</td>
</tr>
</tbody>
</table>

^aBased on the complete genome sequence of B. longum NCC2705 (accession no. AE014295).

Results

Sequence conservation of the tet(W) gene, its upstream region and the resulting Tet(W) protein

The nucleotide sequence of the tet(W) gene and its immediate upstream region was determined in all 10 strains. All sequences shared a core DNA region of 2638 bp, except for B. animalis E43, which shared a sequence of only 2154 bp. This core region started 658 bp upstream of the tet(W) gene start codon and ended 70 bp after its stop codon (250 bp upstream of the start codon and 13 bp before the stop codon, respectively, in B. animalis E43). A genetic diagram of these regions is shown at the bottom of Figure 1. The common 2.1 kb segment showed a G + C content of 51.3% to 51.4%, which is lower than the G + C average for Bifidobacterium genomes (around 59%). The nucleotide identity of the sequences to one another ranged from 98.2% to 100%, the sequence of strain E43 being the least conserved. Putative conserved regulatory regions were observed in the first 250 bp upstream of the gene, including a short 14 amino

Figure 1. Genetic organization of the regions upstream and downstream of tet(W) in six Bifidobacterium strains. ORFs with the same shading (note transposase- and integrase-related ORFs in light grey) or grading represent similar or identical genes in the different species and strains. Arrows indicate complete ORFs, while broken ends indicate N- or C-terminal interrupted genes. The positions and orientations of the primers used in this work (Table 2) are indicated by the small black arrows below the core region.
acid (MLYMCMSATYNPQW) open reading frame (ORF) possibly involved in translational attenuation, a perfect inverted repeat (IR) of 17 bp (AATCCCGATGGTAAAGGATTCTGCTGGGATT) capable of forming a stable stem-loop with a ΔG of −18.8 kcal/mol (−17.7 kcal/mol for strain E43), followed by a stretch of thymine residues that may act as a σ-independent terminator, and a putative ribosome binding site (AAAGGGAGG) 9 bp upstream of the start codon of the tet(W) gene. Forty-three polymorphisms were scored in this core domain, of which 35 were within the tet(W) gene coding sequence. With respect to the nucleotide sequence of B. longum H66, the sequences analysed appeared as follows: (i) B. animalis E43: 39 bp changes, 32 in the coding tet(W) gene; (ii) B. longum LMG 131977 and B. thermophilum LMG 218133: the common region of these two strains was identical with 31 bp changes at positions identical to those in E43 except for a single nucleotide change outside the tet(W) gene in the coding region of the 14 amino acid peptide; (iii) B. longum B93 and B94: these two sequences were identical to each other and showed two nucleotide changes in the tet(W) coding region; (iv) B. longum E111 and B. bifidum L22: these showed a single bp difference in the tet(W) ORF but at different locations; and (v) B. longum L42 and M21: these showed sequences identical to those of H66.

The tet(W) gene was composed, in all cases, of 1912 bp, and the deduced Tet(W) protein of 639 amino acids (Figure 1). The nucleotide differences between the strains translated into 20 different amino acids for the protein of B. animalis E43 with respect to the Tet(W) sequence of B. longum H66 [17 differences due to nucleotide replacements in the common region, plus 3 amino acid differences as a consequence of the distinct 13 bp at the end of the tet(W) gene]. The protein sequences of B. longum LMG 131977 and B. thermophilum LMG 218133 showed a difference of 14 amino acids; all the amino acid replacements were as in B. animalis E43. The deduced proteins of strain E111 (G for A at position 248) and those of B93, B94 and L22 (P for H at position 430) showed a single amino acid difference with respect to the reference protein.

The B. longum M21 strain had an insertion of 736 bp at a position 1348 bp downstream of the tet(W) gene start codon (see below), explaining the non-functionality of the gene. However, nothing in the B. longum LMG 131977 sequence seemed to account for the organism’s susceptibility to tetracycline. The nucleotide and deduced amino acid sequences of this strain were identical to those of the highly resistant strain B. thermophilum LMG 218133 (Table 1).

Conserved flanking sequences

A combination of inverse and direct PCR and sequencing was used to obtain sequences from the upstream and downstream regions of the core conserved domain in six strains. Distinct sequence stretches were obtained for the different strains under study (Figure 1). These diverged abruptly 658 bp upstream of the tet(W) gene start codon: a segment of DNA considered within the core region in all strains except B. animalis E43. However, the sequences were more conserved for some strains in the downstream region of the gene. In addition to the 60 bp downstream of the gene’s stop codon (conserved in all strains except B. animalis E43), a further 106 bp segment was shared by B. longum H66, L42, M21 and B. thermophilum LMG 218133. Sequences from B. longum H66 and B. thermophilum were identical for a 142 nucleotide long segment. Finally, B. longum L42 and M21 showed complete identity for all the downstream sequences available for both strains (545 bp). These common regions are indicated by dotted vertical lines in Figure 1.

Analysis of ORFs in regions flanking the tet(W) genes

In B. animalis E43, the tet(W) gene was preceded by an ORF encoding a protein with a 55% amino acid identity (for a stretch of 198 bp) to an internal sequence (amino acids 127–318) of a transposase of the insertion sequence (IS) family of Pseudomonas aeruginosa 2192 (EAAZ58 611.1; COG3039). In contrast, no ORFs were identified in a downstream 730 bp sequence. However, a small segment of DNA of around 100 bp, which included the last nucleotides of the tet(W) gene (different from those of all other strains) and its stop codon, showed significant nucleotide identity to the equivalent part of the tet(W) gene from Lawsonia intracellularis PHE/M1-00 genome (NC_008011; locus tag LI0179).

At the 5’ end of the DNA segment sequenced from B. bifidum L22, an ORF was identified encoding a protein with 76% amino acid identity to the final sequence (135 amino acids) of the Cip protease from B. longum DIO10A (ZP_00121578.1; COG0542). This ORF ended 300 bp before the starting point of the common core sequence. No ORFs were identified in a small 180 bp stretch downstream of the tet(W) gene stop codon.

The DNA sequence upstream of the core region was not obtained for B. thermophilum LMG 218133, but a complete ORF was found downstream, which could encode a protein of 255 amino acids showing 100% identity to several SAM-dependent methyltransferases [e.g. that of orfY (FAEPRAM212_01960) from Faecalibacterium prausnitzii M21/2]. This same ORF was truncated at early but different positions in B. longum H66, M21 and L42 (Figure 1).

The N-terminal part of two truncated ORFs running in opposite directions was seen in the upstream region of B. longum H66. The first encoded a peptide of 124 amino acids with 47% amino acid identity to hypothetical proteins with an ARCH_ATPase domain (a conserved P-loop motif that is involved in ATP binding) from Clostridium leptum DSM 753 (EDSO9757.1; CLOLEP_03807). The second ORF encoded a 257 amino acid protein with 95% identity to the N-terminus sequence of a putative ABC-type multidrug transporter from B. longum DIO10A (ZP_00121338.1; COG1132). Downstream of the core region, an interrupted ORF was observed, whose initial 80 amino acids corresponded to an OrfY protein (methyltransferase) with a sequence identical to that of B. thermophilum LMG 218133. After this position, the final 240 bp of H66 showed 99% identity at the nucleotide level to a segment of the B. longum NCC2705 chromosome (AE014295.3; positions 1510367–1510602).

A long ORF (encoding 424 amino acids) starting before the sequence analysed was found in the remote 5’ end of the tet(W) gene in B. longum M21. The N-terminal part of the protein encoded by this ORF (289 amino acids) showed 91% identity to an internal segment of a hypothetical protein of the TraG/VirD4 family from Bacteroides capillosus (ZP_02039232.1; BACCAP_04884). Homologous proteins, which are thought to be involved in type IV secretion (a process necessary for bacterial conjugation), have also been found in many other anaerobic bacteria (such as Fusobacterium prausnitzii, Clostridium kluwyeri, Ruminococcus gravis and Eubacterium dolichum).
Downstream within the same coding region but in a different frame, an ORF encoding a 46 amino acid protein was identified. This small protein, designated MAFF for its first four amino acids, has been previously reported associated to tet(W) genes in *Butyrivibrio fibrisolvens* L230 and JK51 and *Roseburia* sp. A2-183. The MAFF protein of M21 showed significant identity (80% to 82%) to the MAFF proteins of other strains, but its closest relative was a hypothetical protein from *B. capillosus* ATCC 29799 (EDM97320.1; BAC01235_04885). Downstream of the tet(W) gene, after a truncated orfY (with an encoding capability of 30 amino acids), an ORF in the direction opposite to the Tc′ gene encoding the final 282 amino acids of a putative transposase (87% identity to a hypothetical protein from *Bifidobacterium adolescentis* L2-32; ZP_02029458.1; BIFADO_01916) of the mutator family (Pfam 00 872) was found to occupy all the sequence available.

In strain L42, a truncated ORF was found encoding an internal peptide 268 amino acids long, with high homology (89% identity) to a phage-related transposase of the IS family (Pfam 002022 and 00552) from *B. adolescentis* ATCC 15703 (BAF40060.1). As mentioned above, the entire downstream sequence of strain M21 was identical to that of strain L42, although in the latter the transposase ORF was complete and encoded a polypeptide of 393 amino acids with 84% homology to the hypothetical transposase sequence in *B. adolescentis* L2.32 (BIFADO_01916).

**An insertion sequence-like element inserted into the tet(W) ORF in B. longum M21**

As mentioned previously, the Tet(W)-encoding ORF in *B. longum* M21 was disrupted by a 736 bp sequence with a genetic organization resembling that of an IS (Figure 1). The G + C content of this IS was 62.4%, similar to that of the *B. longum* genome (60%).10 The DNA segment was flanked by two almost perfect IRs of 30 nt (CCGAGAATGCTTCCATTTAGAAGTGCACAA CCCC-674 nt-ACGGGTATTGACATTTACATTTGAGACCCG). A putative ORF was observed starting 78 bp from the 3′ end of the sequence and running in a relative orientation opposite to that of the tet(W) gene. Its deduced product showed a tra domain (motif Pfam00665, thought to be involved in integration of DNA into the chromosome) and homology to transposases of the IS30 family (54% identity over a 50 amino acid stretch to the product of locus AAK94953 from *Mycoplasma bovis*). A further downstream out-of-frame peptide showed higher homology (68% over an 82 amino acid stretch) to remnant inactivated IS30 derivatives present in the genome of *B. longum* DIO10A (ZP_00121392.1, COG2826). Apparently, the coding sequence has a point mutation around the middle of the coding region, and the transposase might be non-functional. The sequence of the IS-like element is flanked by two short direct repeats of 5 bp (GTTAC). Thus, it appears that the IS interrupted the tet(W) gene and, during the insertion event, duplication of the 5 bp sequence occurred. Indeed, the 5 bp sequence is present once at this same position in all tet(W) genes analysed so far.

**tet(W) gene copy number**

The single-copy housekeeping *tuf* gene was used as a reference. After normalization, the ΔC₇ values for all strains (which ranged between 0.01 and −0.56) indicated that the tet(W) gene was in single-copy state in the chromosome of all strains.

**Discussion**

Among bifidobacteria, the tet(W) gene has been reported in many species, including *B. animalis*, *B. bifidum*, *Bifidobacterium catenulatum*, *B. longum*, *Bifidobacterium pseudocatenulatum*, *Bifidobacterium pseudolongum* and *B. thermophilum*.5–7 tet(W) genes have been also reported in tetracycline-susceptible strains.6 In this paper, the genetic organization of tet(W) genes and their immediate upstream and downstream sequences was analysed in 10 strains. tet(W) had already been determined to be the only tetracycline-resistance-encoding gene in these strains.11

The nucleotide and deduced protein sequences of the tet(W) genes were all very similar (identity higher than 98%) and showed a close relationship to tet(W) genes from other intestinal bacteria.7,12–14 The differences, however, could be used to trace the phylogenetic relationships of tet(W) genes from GIT inhabitants.7,12–15 These relationships further support early suggestions of recent and independent transmission events.6,7

In the susceptible *B. longum* M21, an IS-like sequence of 736 bp was found to interrupt the structural tet(W) gene, giving rise to a non-functional protein. This IS element belongs to the IS30 family and has not been described before in bifidobacteria. Recently, an IS 1047 bp long was identified in the upstream region of the tetracycline-resistant *B. longum* F8.15 Five of the six bases duplicated in F8 (CAATGC) seem to mirror the 5 bp duplication in M21, which might suggest active insertion sites in *B. longum*. Further investigations will be required to unravel the non-functionality of the tet(W) gene in *B. longum* LMG 13197T.

The tet(W) genes from all 10 strains shared 98% to 100% DNA and amino acid identity, though variation was found in the upstream and downstream flanking regions.

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**Transparency declarations**

None to declare.

M. S. A. conducted research and wrote the article. A. B. F. and P. A.-M. characterized the strains and conducted research. A. M. supported the work and contributed to the discussion. B. M. directed the research and contributed to the writing and discussion of the results.
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