Prevalence and characteristics of β-lactamase and plasmid-mediated quinolone resistance genes in Escherichia coli isolated from farmed fish in China

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Objective: To determine the molecular epidemiology of extended-spectrum β-lactamases (ESBLs) and plasmid-mediated quinolone resistance (PMQR) in Escherichia coli isolated from farmed fish in China.

Methods: E. coli was isolated from fish gut samples from fish farmed throughout Guangdong province and tested for the presence of the β-lactamase genes and PMQR-encoding genes using PCR and DNA sequence analysis. Co-transfer of plasmids encoding for ESBLs as well as PMQR determinants was explored by conjugation into E. coli.

Results: A total of 218 non-duplicate E. coli were recovered from fish gut samples. β-Lactamase genes were identified in 19 (17%) of 112 strains with reduced susceptibility to ampicillin, and PMQR genes were identified in 59 (73.8%) of 80 strains with reduced susceptibility to ciprofloxacin. Only three ESBL genes were identified in three isolates: blaCTX-M-14, blaCTX-M-79 and blaSHV-27. PMQR gene screening identified qnr genes (n=59) as the most common, including qnrB (n=33), qnrS (n=21) and qnrD (n=5), with aac(6′)-Ib-cr (n=6) being rarely found. The co-carriage of two or three PMQR genes in one strain was found in 7 (11.9%) isolates. The ESBL gene blaCTX-M-79 was found to be co-carried with qnrS. Co-transfer of qnrS was observed with blaCTX-M-79.

Conclusions: Our study is the first to demonstrate the existence of high levels of mobile genes conferring reduced susceptibility to fluoroquinolones as well as the presence of ESBL genes in fish produced in China, and identifies a significant reservoir of antibiotic resistance genes relevant to human medicine.

Keywords: plasmid-mediated quinolone resistance, ESBL, aquaculture, molecular epidemiology

Introduction

Strains of Enterobacteriaceae that produce extended-spectrum β-lactamases (ESBLs), particularly Escherichia coli and Klebsiella spp., producing CTX-M, have emerged as significant antibiotic-resistant human pathogens across the world. A recent study of the acquisition of faecal carriage of CTX-M-type ESBL-producing E. coli by travellers to different parts of the world from Sweden showed an acquisition rate of 32% when travelling to Asia (excluding India). This high rate of acquisition possibly reflects exposure to food and water contaminated with ESBL E. coli, as the incidence of ESBL production in clinical isolates of E. coli in China is 65%. Antibiotic-resistant E. coli from animal sources in countries with high human carriage rates should not be ignored, as there is considerable evidence they contribute to the burden of human microbial resistance, particularly in parts of the world with variable regulation of antibiotic use. A study from China demonstrated high rates of plasmid-mediated quinolone resistance (PMQR) genes [aac(6′)-Ib-cr and qepA] predominately in companion animals. An alarmingly high rate of ESBL carriage has recently been reported in food animals from Hong Kong (63% in pigs; 58% in chickens; 33% in cattle); almost all being co-resistant to ciprofloxacin.

Fish farming (aquaculture) is growing rapidly, particularly in Asia. China is the world’s biggest producer of farmed fish, producing 32.7 million tons of the total world production of 52.5 million tons in 2008. The consequences of the extensive use of antibiotics in aquaculture are the selection in the fish gut...
flora of multiresistant bacteria that are passed to the human gut commensal flora when the fish are eaten. The heavy use of agents such as oxolinic acid, tetracycline, florfenicol and nitrofurantoin has selected resistance in fish pathogens, so quinolones are increasingly the preferred agents. Although China is the world’s largest producer of farmed fish, no studies of the prevalence of multiresistant Enterobacteriaceae with resistance to human medicine critical antibiotics (e.g. quinolones and extended-spectrum cephalosporins) in farmed fish exist to our knowledge in the published literature. We have undertaken a prevalence study in retail fish farmed across Guangdong province in southern China to identify the extent and molecular characteristics of the quinolone and ESBL resistance gene reservoir that has the potential to transfer to the human gut flora.

Materials and Methods

Bacterial strains
A total of 15 different fish markets, which were widely dispersed across the city of Guangzhou, were selected for the study in 2010. Each market received farmed freshwater fish from farms throughout Guangdong province, which is the largest centre for aquaculture in China. Twenty fish were sampled at each market, each from a different vendor, by opening the gut using a sterile scalpel following washing the gut surface with sterile saline and then swabbing the contents. The swabs were plated on MacConkey agar incubated for 18 h at 37°C and a single colony of E. coli was selected for further study. E. coli were identified using API20E (BioMerieux, Beijing, China).

Antimicrobial susceptibility testing
The MICs of ampicillin, ceftiofur, cefotaxime, chloramphenicol, florfenicol, spectinomycin, kanamycin, tetracycline, co-trimoxazole, nalidixic acid and ciprofloxacin were determined by agar dilution using CLSI methods. We tried to comply with recent recommendations and have used breakpoints applicable to human infections rather than animals, as we are interested in the effect of the movement of resistance genes from fish to humans rather than the prediction of therapeutic outcome in fish. All isolates demonstrating reduced susceptibility to ampicillin (MIC ≥ 8 mg/L) and reduced susceptibility to ciprofloxacin (MIC ≥ 0.06 mg/L) were retained for molecular studies. We chose a ciprofloxacin MIC ≥ 0.06 mg/L in order to stand the maximum probability of identifying PMQR-carrying isolates.

Results

Bacterial strains and antimicrobial susceptibility testing
A total of 218 E. coli isolates were recovered from the 300 fish gut samples. The MIC results are shown in Table 1. High rates of resistance to ampicillin, florfenicol, tetracycline and co-trimoxazole and reduced susceptibility to ciprofloxacin were detected. Reduced susceptibility to ampicillin was found in 112 isolates (51%) and reduced susceptibility to ciprofloxacin in 80 isolates (37%).

Characterization of ESBL and PMQR determinants
All ampicillin-resistant E. coli were screened using PCR as described previously for blaTEM, genotype groups 1, 2, 8, 9 and 25, blaSHV, blaCTX-M, blaGES, and blaOXA and blaKPC. All ciprofloxacin-resistant isolates were characterized by PCR for PMQR determinants aac(6’)-Ib-cr (identified by BtsCI digestion), qepA, qnr, B, S, and D as previously described. All PCR products from ESBL and PMQR genes including qnr variants were confirmed and analysed by DNA sequencing. The presence of E. coli sequence type (ST) 131 was determined in ESBL isolates by a PCR screening method.

Conjugative transfer of plasmids encoding ESBLs was studied by mating with azide-resistant E. coli J53, as described previously. Trans-conjugants were detected by plating mating mixtures on Luria–Bertani agar supplemented with 150 mg/L sodium azide and 2 mg/L cefotaxime. Co-transfer of resistance determinants was explored by amplifying the relevant genes in the transconjugants.

Table 1. Percentage resistant, MIC50 and MIC90 of 218 E. coli to 11 antimicrobial agents (breakpoints used are shown in the footnotes)

<table>
<thead>
<tr>
<th>Antimicrobial agents</th>
<th>AMP</th>
<th>CTF</th>
<th>CTX</th>
<th>CHL</th>
<th>FFC</th>
<th>SPC</th>
<th>KAN</th>
<th>TET</th>
<th>SMX/TMP</th>
<th>CIP</th>
<th>NAL</th>
</tr>
</thead>
<tbody>
<tr>
<td>Percentage resistant (n)</td>
<td>78.9 (172)</td>
<td>2.3 (5)</td>
<td>23.4 (51)</td>
<td>6.9 (15)</td>
<td>39 (85)</td>
<td>41.7 (90)</td>
<td>16.0 (35)</td>
<td>36.7 (80)</td>
<td>16.0 (35)</td>
<td>256</td>
<td>2</td>
</tr>
<tr>
<td>MIC50</td>
<td>32</td>
<td>0.25</td>
<td>0.25</td>
<td>8</td>
<td>16</td>
<td>32</td>
<td>4</td>
<td>&gt;512</td>
<td>2</td>
<td>256</td>
<td></td>
</tr>
<tr>
<td>MIC90</td>
<td>256</td>
<td>1</td>
<td>1</td>
<td>128</td>
<td>256</td>
<td>512</td>
<td>32</td>
<td>64</td>
<td>&gt;512</td>
<td>2</td>
<td>256</td>
</tr>
</tbody>
</table>

Antibiotic breakpoints used (in mg/L) are shown in parentheses. AMP, ampicillin (8); CTF, ceftiofur; CTX, cefotaxime (8); CHL, chloramphenicol (32); FFC, florfenicol; SPC, spectinomycin; KAN, kanamycin (64); TET, tetracycline (16); SMX/TMP, co-trimoxazole (4); CIP, ciprofloxacin (4); NAL, nalidixic acid (32). Reduced susceptibility MIC ≥ 8 mg/L.

All isolates with reduced susceptibility, MIC ≥ 0.06 mg/L.
genes, 33 were qnrB, 21 qnrS, and only 5 qnrD. No qnrA, qnrC or qepA genes were detected. Two or three PMQR genes co-existing in one strain were found in 7 (11.9%) of 59 PMQR gene-positive isolates, qnrB + qnrD (2), qnrB + aac(6’)-Ib-cr (2), qnrS + qnrD (1) and qnrS + aac(6’)-Ib-cr (1).

The ESBL gene blaCTX-M-79 was found in combination with qnrS, while the isolate with blaCTX-M-14 carried no PMQR genes. The co-transfer of qnrS was observed with blaCTX-M-79, and none of the other plasmids carrying other ESBL genes mobilized PMQR genes by conjugation. blaCTX-M-14 genes were also transferred alone by conjugation. None of the ESBL-producing strains belonged to the ST131 clone.

Discussion

As the world’s largest producer of farmed fish (32.7 million tons; 62% of world production in 2008), understanding the incidence and composition of the antibiotic resistance reservoir in farmed fish in China is important. We chose farmed fish, as there are no published data on the incidence of different antibiotic resistances in E. coli from this very important animal food source in China. The range of antibiotics that has been recorded as being used in aquaculture is wide, including aminopenicillins, aminoglycosides, nitrofurans, fluoroquinolones, sulphonamides and tetracyclines.9 Historically amphenicols (florfenicol and chloramphenicol), sulphonamides and tetracyclines have been used, usually by incorporation into fish food. A substantial body of data exist on the occurrence of tetracycline resistance in aquaculture, particularly but not exclusively in fish pathogens.9 Increasing rates of resistance to these older heavily used antibiotics in fish pathogens such as Aeromonas salmonicida has resulted in a shift to increased usage of quinolones. While high-level resistance to quinolones arises from mutations in the chromosomal gyrA and parC genes, it is the lower-level plasmid-mediated resistance conferred by the qnr and aac(6’)-Ib-cr genes that creates an environment in bacteria for the rapid selection of high-level resistance.12 Prior to our study there is only one study from Egypt looking at the prevalence of these genes together with ESBL genes in Gram-negative bacteria from water samples from fish farms.20 The quinolone and the third-generation cephalosporin (3GC) classes of antibiotics are important for the treatment of serious infections in humans caused by Enterobacteriaceae. The consequences for human health are more serious than from resistance to older antibiotics such as tetracycline and chloramphenicol. Because of the scale of fish farming in China, the impact on the prevalence of resistance in the gut flora of the general community is likely to be highly significant. The rates of resistance (using human clinical breakpoints) seen in the E. coli from fish guts in our study was generally high, particularly for tetracycline (39%) and co-trimoxazole (42%), with reduced susceptibility to ciprofloxacin (37%) and ampicillin (51%).

The ESBL rate was comparatively low (3/218, 1.5%) in the present study; however, 3GCs are unlikely to be widely used in aquaculture at the moment. The Egyptian study also found only 11/274 (4%) isolates carrying ESBL genes.20 In both China and Egypt, the rate of carriage of ESBLs in humans is high, so these isolates were most likely derived from contamination of the fish in the farm from human sewage, probably via river water. CTX-M-14 is the most common CTX-M genotype in China.1 CTX-M-79, which was also identified, is a double-nucleotide variant of CTX-M-15 that has been reported from human infections and faecal carriage in China.21

The ESBL frequency was interesting (Table 2), with qnrB and qnrS dominant, and just a small number of qnrD and aac(6’)-Ib-cr. The frequency was much higher than in Egypt (55/218 in China, 15/274 in Egypt), although the distribution was similar. The relative frequency of PMQR genotypes is similar to reports from food animals in China22 and human clinical isolates from China.23,24

The very high rates of resistance in the E. coli in our study to agents that have and most probably continue to be used (i.e. tetracyclines, florfenicol and co-trimoxazole) most probably reflects a strong selective pressure for continuing antibiotic resistance in E. coli on the fish farms. The recent change to increased usage of quinolones should result in increased resistance, which we have demonstrated with a very high rate (55/218 isolates, 25.2%) of PMQR in E. coli. This rate is higher than recent studies from China have reported for poultry and swine (14%),25 chickens (22.2%)22 and human clinical isolates (12.8% and 17.9%).18,25

The increasing industrialization of food production in Asia is resulting in greater usage of antimicrobials. A recent survey of faecal carriage of E. coli with resistance to ‘critically important’ antibiotics in food animals in Hong Kong showed a substantial increase in resistance in 2008 compared with an earlier study in 2002.2

Our study is the first to demonstrate the existence in fish produced in China (the world’s largest aquaculture environment) of high levels of PMQR genes as well as the presence of ESBL genes. Efforts should be made worldwide to more closely monitor and introduce control of antibiotic resistance in aquaculture, as this represents a major reservoir of resistance genes likely to threaten the human use of critical antibiotics in the future.

Table 2. Distribution of PMQR genes in 80 ciprofloxacin-resistant E. coli identified from 218 E. coli from farmed fish in Guangdong province collected at 15 markets

<table>
<thead>
<tr>
<th>PMQR genes</th>
<th>No. of isolates positive for gene(s)</th>
<th>No. of markets at which fish were positive</th>
</tr>
</thead>
<tbody>
<tr>
<td>qnrB</td>
<td>30</td>
<td>11</td>
</tr>
<tr>
<td>qnrS</td>
<td>16</td>
<td>8</td>
</tr>
<tr>
<td>qnrD</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>qnrB + qnrD</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>qnrS + qnrD</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>qnrB + aac(6’)-Ib-cr</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>qnrS + aac(6’)-Ib-cr</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>aac(6’)-Ib-cr</td>
<td>2</td>
<td>2</td>
</tr>
</tbody>
</table>

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Antibiotic resistance in farmed fish from China

Transparency declarations
None to declare.

References