Mitochondrial DNA transmission, replication and inheritance: a journey from the gamete through the embryo and into offspring and embryonic stem cells

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Introduction

The mitochondrion exists within the cytoplasm of nearly all eukaryotic cells. Along with mediating processes such as steroidogenesis, apoptosis, homeostasis and cell division, mitochondria are also essential generators of ATP through the process of oxidative phosphorylation (OXPHOS) which takes place within the electron transfer chain (ETC). OXPHOS, unlike any other cellular process, is highly dependent on the expression of proteins encoded by the mitochondrial genome [mitochondrial DNA (mtDNA)] and chromosomally encoded genes. Although the transmission of mtDNA is dependent on the mitochondrion as its vehicle, both the mitochondrion and mtDNA should be considered as independent entities that co-exist, just as the mitochondrion co-exists within the cytoplasm of a cell but its origins are bacterial (Cavalier-Smith, 2009). The role played by the mitochondrial genome in reproductive biology has been largely ignored. However, since the advent of more sophisticated reproductive technologies, such as nuclear transfer, its importance is becoming increasingly recognized, especially as mixed and diverse crosses. We then investigated the transmission of donor cell mtDNA following nuclear transfer and also sperm mtDNA in diverse crosses. After those events were regulated following natural fertilization which included the generation of offspring and stem cells through intra- and interspecific crosses. We then investigated the transmission of donor cell mtDNA following nuclear transfer and also sperm mtDNA in diverse crosses. After including interspecies nuclear transfer, we then probed the literature to understand how various other technologies could affect the normal transmission of mtDNA in order that we could make predictions about transmission from those technologies that have yet to be investigated. We restricted our search to mtDNA, but did discuss mitochondrial and ETC function when, specifically, the effects of heteroplasmy might affect cellular function and offspring survival. The literature was also interrogated for mtDNA-type disease and how this might be prevented through oocyte manipulation and stem cell therapy.

The mitochondrion

Mitochondria exist in various topographical forms, which reflect the type and functions of the cells they serve and thrive off. In maturing oocytes and early cleavage stage embryos, they tend to be oval and spherical while in the differentiating and mature cells, they adopt a more elongated configuration (Sathananthan et al., 2002). Furthermore, the density of mitochondria per cell is often indicative of the cell’s requirements for ATP production (Moyes et al., 1998). Consequently, cells such as neurons and muscles tend to have higher densities, though these are considerably fewer than the metaphase II stage oocyte, which possesses between 150 000 and 300 000 mitochondrial profiles (Jansen and De Boer, 1998).

Search methods

A computerized search was carried out using MedLine and Web of Knowledge for key areas associated with the mitochondrial genome, mtDNA replication, the factors controlling mtDNA replication, mtDNA transmission and mtDNA segregation. The search initially focused on somatic cell biology and then the relevance of those areas related to mtDNA disease. The search then concentrated on how those events were regulated following natural fertilization which included the generation of offspring and stem cells through intra- and interspecific crosses. We then investigated the transmission of donor cell mtDNA following nuclear transfer and also sperm mtDNA in diverse crosses. After including interspecies nuclear transfer, we then probed the literature to understand how various other technologies could affect the normal transmission of mtDNA in order that we could make predictions about transmission from those technologies that have yet to be.

What is mtDNA?

In humans, the circular, double-stranded mtDNA genome is ~16.6 kb in size (Fig. 1) and resides within the inner membrane of the mitochondrion. It consists of a heavy (H) and a light (L) strand, which encode 13 of the polypeptides contributing to Complexes I, III, IV and V of the ETC (Fig. 2). The remaining genes and those of Complex II are derived from chromosomal DNA (Anderson et al., 1981). Specifically, mtDNA codes seven subunits of NADH dehydrogenase (Complex I), one subunit of cytochrome c reductase (Complex III), three subunits of cytochrome c oxidase (Complex IV) and two subunits of the ATP synthase (Complex V; Fig. 1). mtDNA also encodes some of its own transcriptional and translational machinery, i.e. 22 tRNAs and 2 rRNAs, emphasizing the semi-autonomous nature of this genome as these processes are also dependent on chromosomally encoded genes. Furthermore, the chromosomally encoded transcription and replication factors interact with the only non-coding region of the
genome, the displacement loop (D-loop). Otherwise, the mtDNA genome contains no introns interspersed between the coding regions with some of the coding regions even overlapping each other, namely ATPase 6, ATPase 8, ND4 and ND4L (Anderson et al., 1981). Furthermore, some of the other genes do not possess termination codons, as these are generated through post-transcriptional polyadenylation (Ojala et al., 1981).

**mtDNA nucleoid**

mtDNA replication is mediated by several nuclear-encoded transcription and replication factors, which along with mtDNA are packed within a space of ~70 nm in diameter, forming the mitochondrial nucleoid (Chen and Butow, 2005). There are ~30 nucleoid proteins conserved between different species (Chen and Butow, 2005). Some of these are responsible for the maintenance and packaging of mtDNA, whereas the others have, in addition to their role in mtDNA maintenance, a metabolic function (Kucej and Butow, 2007). A structural model has been suggested where the nucleoid is composed of a central core and a peripheral region where the central core proteins are involved in nucleic acid synthesis (Fig. 3; Table I), whereas translation and protein assembly occur in the peripheral region (Bogenhagen et al., 2007). The number of mitochondrial nucleoids per mitochondrion ranges from 1 to 10 (Satoh and Kuroiwa, 2004). There is a dispute about mtDNA copy number as the estimated copies/mitochondrion ranged from 1 to 15 (Satoh and Kuroiwa, 2004), whereas others report that there are between 2 and 8 copies of mtDNA in each nucleoid (Legros et al., 2004).

Both mitochondrial transcription factor A (TFAM; Ekstrand et al., 2004) and ATAD3 (He et al., 2007) have been proposed to provide either the key-linking protein or the backbone to the nucleoid. ATAD3 (also known as TOB3) is an AAA domain protein, consisting of two subdomains, ATAD3f1 and ATAD3f2 (Wang and Bogenhagen, 2006; He et al., 2007). Although it is tightly associated with the inner mitochondrial membrane, its relationship with mtDNA is controversial. One study suggested that ATAD3 is attached to mtDNA at the
that mtDNA is extensively packaged with TFAM (Alam and mtDNA are Pohjoismaki et al., 2003; Kaufman co-localization to mtDNA through immuno-electron microscopy been further supported by more recent studies demonstrating its role in mtDNA transcription further supporting it functional similarity to other HMG proteins (Fisher et al., 1992; Takamatsu et al., 2002; Bianchi and Agresti, 2005). The latter two functions suggest an important role in mtDNA transcription further supporting it functional similarity to other HMG proteins (Fisher et al., 1992; Takamatsu et al., 2002; Bianchi and Agresti, 2005; Kaufman et al., 2007). Furthermore, up-regulation of TFAM expression results in an increase in expression of the mitochondrial-specific polymerase gamma (Polγ) (Ekstrand et al., 2004; Hance et al., 2005) implying a role in initiation of mtDNA replication.

POLG mediates the replication, recombination and repair of mtDNA (Grazieswicz et al., 2005), thus demonstrating its essential role in mtDNA maintenance (Spelbrink et al., 2000). Although it is capable of replicating mtDNA, it cannot replicate mammalian chromosomal DNA, perhaps resulting from its similarities to other members of the family A group of DNA polymerases. These include Escherichia coli DNA polymerase I and the T7 DNA polymerase (Fridlender and Weissbach 1971; Fridlender et al., 1972; Beese et al., 1993a, b). It consists of two subunits in animal cells and one in yeast (Grazieswicz et al., 2005): POLGA is the catalytic subunit that has 3' → 5' exonuclease activity and accounts for the high fidelity of this enzyme. POLGB is the accessory subunit that forms a heterotrimer in humans (in other mammalian species, it forms a heterodimer) at a ratio of 2:1 with POLGA (Carrodeguas et al., 1999, 2001), thus stabilizing the enzyme to increase efficiency. In addition, a recent study has suggested that POLGB is the key factor that determines mtDNA copy number within the nucleoid as it is required for D-loop synthesis (Di Re et al., 2009).

Two other factors play a key role in mtDNA replication. The helicase, Twinkle, was originally identified in association with autosomal dominant progressive ophthalmoplegia, a mitochondrial disorder characterized by mtDNA deletions (Spelbrink et al., 2001). It shares a structural similarity to the C-terminal helicase part of bacteriophage T7 gene 4 protein which exhibits both helicase and primase activities (Bernstein and Richardson, 1989; Spelbrink et al., 2001). Although mammalian Twinkle seems to lack primase activity (Spelbrink et al., 2001), Twinkle homologues in Plasmodium falciparum (Seow et al., 2005) and nearly all eukaryotes (Shutt and Gray, 2006) possess both helicase and primase activities. The mitochondrial single-stranded DNA-binding protein (mtSSB) mediates the unwinding of mtDNA through its physical interaction with Twinkle (Kirchonen et al., 2003), which co-localizes with both TFAM and mtSSB. These proteins act not only as structural components but also stabilize mtDNA during replication (Garrido et al., 2002).

**mtDNA replication**

Both the asymmetric and the coupled leading- and lagging-strand synthesis models have been described to explain mtDNA replication. The asymmetric model has been the traditional approach to understanding how mtDNA replication is mediated (reviewed in Shadel and Clayton, 1997). It is initiated from the origin of H-strand replication (OH), located within the D-Loop region, where TFAM binds to the enhancer of the light-strand promoter (LSP) and induces structural changes that expose the promoter region to the mitochondrial-specific RNA polymerase. This allows an RNA primer to be generated, which is used by POLGA to initiate mtDNA replication. mtDNA replication then continues two-thirds round the genome to the origin of L-strand replication (OL). L-strand synthesis then proceeds in the opposite direction.

The coupled leading- and lagging-strand synthesis model proposes that both H- and L-strands are replicated bidirectionally from the same initiation cluster site (Yasukawa et al., 2005). This mechanism is thought to occur in addition to the asymmetric model, but is also typical of cells repopulating mtDNA. The coupled leading- and lagging-strand synthesis model has now been refined to include the presence of replication intermediates which allow gaps within the replicating mtDNA on the lagging-strand to be filled to complete the replicon (Yasukawa et al., 2006). It has more recently been proposed that replication is driven from several origins of replication within the vicinity of the OH. However, to date, there has been little resolution as to which model is the most appropriate with entrenched views being expressed by the two opposing groups (Bogenhagen and Clayton, 2003; Holt and Jacobs, 2003).
### Table I: The key mtDNA nucleoid proteins associated with transcription and replication.

<table>
<thead>
<tr>
<th>Nucleoid protein</th>
<th>Molecular weight (kDa)</th>
<th>Chromosomal location (in human)</th>
<th>Function</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>TFAM</td>
<td>24.4</td>
<td>10q21</td>
<td>Transcription and packaging</td>
<td>Parisi and Clayton (1991)</td>
</tr>
<tr>
<td>POLGA</td>
<td>125–140</td>
<td>15q25</td>
<td>mtDNA maintenance</td>
<td>Ye et al. (1996); Lecrenier et al. (1997); Wernette and Kaguni (1986)</td>
</tr>
<tr>
<td>POLGB</td>
<td>35–55</td>
<td>17q</td>
<td>mtDNA maintenance, putative role as a regulator of mtDNA copy number</td>
<td>Carrodeguas et al. (2001); Di Re et al. (2009); Wernette and Kaguni (1986)</td>
</tr>
<tr>
<td>Twinkle</td>
<td>77</td>
<td>10q24</td>
<td>mtDNA unwinding</td>
<td>Li et al. (1999)</td>
</tr>
<tr>
<td>mtSSB</td>
<td>17.26</td>
<td>7q34</td>
<td>Stabilizes D-loop and stimulates Twinkle activity</td>
<td>Takamatsu et al. (2002); Tomaska et al. (2001)</td>
</tr>
<tr>
<td>ATAD3 f1</td>
<td>59</td>
<td>1p36.33</td>
<td>Chaperone-like activity; backbone to the nucleoid</td>
<td>Bogenhagen and Clayton (2003); He et al. 2007</td>
</tr>
<tr>
<td>ATAD3 f2</td>
<td>56.4</td>
<td>1p36.33</td>
<td>Chaperone-like activity; backbone to the nucleoid</td>
<td>Bogenhagen and Clayton (2003); He et al. 2007</td>
</tr>
<tr>
<td>TFB1M</td>
<td>39.5</td>
<td>16q25.1–q25.3</td>
<td>Accessory transcription factor and translation of mitochondrial proteins</td>
<td>Falkenberg et al. (2002)</td>
</tr>
<tr>
<td>TFB2M</td>
<td>45.3</td>
<td>1q44</td>
<td>Accessory transcription factor and translation of mitochondrial proteins</td>
<td>Falkenberg et al. (2002)</td>
</tr>
<tr>
<td>mTERF</td>
<td>45.7</td>
<td>7q21–q22</td>
<td>Control rRNA and mRNA synthesis</td>
<td>Roberti et al. (2006)</td>
</tr>
<tr>
<td>mtRNA pol</td>
<td>138.6</td>
<td>19p13.3</td>
<td>Initiation of transcription at a specific mtDNA site</td>
<td>Gaspari et al. (2004)</td>
</tr>
</tbody>
</table>

It has been generally accepted that the mtDNA genome is not under direct epigenetic regulation. For example, mammalian cells characterized for mtDNA methylation are highly hypomethylated (Pollack et al., 1984). However, an important question that remains to be determined is how the expression of the key genes associated with mtDNA replication is mediated. Recent analysis of the promoter region of Tfam produced inconclusive data as to whether its expression was regulated by DNA methylation (Choi et al., 2004; Gemma et al., 2009). Nevertheless, more recent analysis of PolgA has however suggested that specific CpG islands within exon 2 of this gene are targeted (Oakes et al., 2007) and could thus account for changes in PolgA expression during spermatogenesis. This approach would certainly compliment the observation of an 8- to 10-fold decrease in the number of mtDNA molecules as these cells differentiate (Hecht et al., 1984). At this stage, Tfam expression is also decreased in the human (Larsson et al., 1997) and, in the mouse, an isoform is expressed that does not possess a mitochondrial targeting sequence (Larsson et al., 1996). This reduction in Tfam expression and mtDNA copy number would match other reports that indicate an association between Tfam and mtDNA copy number (Ekstrand et al., 2004), most likely restricting it to a packaging role in this instance. This suggests that replication could then be regulated by either a conformational or a numerical change to TFAM that would result in exposure of Oxy or the broader initiation cluster coupled with a change in the DNA methylation status of PolgA. Indeed, DNA methylation of PolgA would present a truly interesting scenario, namely that the replication of a bacterial-originating genome was mediated by epigenetic regulation of a mammalian nuclear-encoded gene(s). It would also therefore be worth considering other members of the mtDNA nucleoid to determine whether their expression is also mediated in a similar manner, or by other processes such as microRNAs.

**mtDNA replication in the pluripotent preimplantation embryo**

Pluripotent blastomeres of mammalian preimplantation embryos and embryonic stem cells (ESCs) share a number of mitochondrial characteristics such as limited oxidative capacity and a greater dependence on anaerobic respiration (reviewed in Facucho-Oliveira and St John, 2009). This is further highlighted, following fertilization of the oocyte, by either whole embryo mtDNA copy number remaining constant during mouse preimplantation development up to the morula stage (Thundathil et al., 2005) or the reduction observed in porcine (Spikings et al., 2007) and bovine embryos (May-Panlopou et al., 2005). Nevertheless, during the early cleavage stages, mammalian embryos express either very low levels of mtDNA replication factors or there is no expression (May-Panlopou et al., 2005; Thundathil et al., 2005; Spikings et al., 2007). Consequently, mtDNA replication does not take place up to and including the morula stage and the number of mtDNA copies/blastoemere is progressively decreased after each embryonic cell division. This results in these blastomeres progressively losing their capacity to generate ATP through OXPHOS as they become more reliant on anaerobic respiration (Leese and Barton, 1984; Leese 1995; Van Blerkom et al., 2002). Indeed, mouse (Batten et al., 1987), cattle (Stojkovic et al., 2001), pig (Spikings et al., 2007), monkey (Squirrel et al., 2003) and human preimplantation embryos (Wilding et al., 2001) contain low numbers of spherical and immature mitochondria which are located in the perinuclear regions of the cytoplasm. The loss in mtDNA copy number is mirrored by mtDNA replication factor activity. For example, in pig and sheep, POLGA, POLGB and TFAM are only briefly detected between the 4- to 8-cell (Spikings et al., 2007) and 8-cell to 16-cell stages (Bowles et al., 2007b), respectively, whereas in cattle, Tfam is first up-regulated at the morula stage.
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(May-Panloup et al., 2005). These brief mtDNA replication events are most likely associated with the generic turnover of a multiple sets of genes, i.e. during embryonic genome activation (EGA; Ma et al., 2001), as demonstrated by the small mtDNA replication event, determined by 5-bromo-2′-deoxy-uridine staining, which takes place in pronuclear and 2-cell stage mouse embryos (McConnell and Petrie, 2004).

mtDNA replication and embryonic differentiation

At the blastocyst stage, there is an up-regulation in the expression of mtDNA replication factors which leads to reactivation of mtDNA replication in those mammalian species so far investigated (Piko and Taylor, 1987; May-Panloup et al., 2005; Thundathil et al., 2005; Spikings et al., 2007). At the same time, mitochondria start to differentiate into elongated organelles containing swollen cristae (Stern et al., 1971; Piko and Matsumoto, 1976; Wilding et al., 2001), acquire a higher membrane potential and levels of oxygen consumption, and increased OXPHOS activity leading to ATP production (Trimarchi et al., 2000; Wilding et al., 2001; Houghton, 2006). However, these replication events are most likely specific to the trophectodermal cells, the cells that give rise to the placenta and will mediate the process of implantation if the embryo is to continue to develop. These cells are no longer pluripotent, i.e. have the potential to differentiate in to any cell type of the body, and typically express markers such as Cdx2 (Niwa et al., 2005). The inner cell mass (ICM) cells, which are pluripotent and give rise to the fetus and ESCs, continue to express high levels of the pluripotency genes such as Oct4, Nanog and Sox-2 (Chen and Daley, 2008). They also continue to express low levels of POLG and TFAM (Spikings et al., 2007). As such, the high levels of expression of pluripotency genes appear to be associated with low levels of expression of mtDNA replication factors and anaerobic metabolism (reviewed in Facucho-Oliveira and St John, 2009).

mtDNA replication, pre-gastrulation and undifferentiated ESCs

The ICM cells continue to divide before they undergo gastrulation or give rise to ESCs and thus progressively gain an increased nucleocytoplasmic ratio. At the blastocyst stage, there are ~483 000 copies of mtDNA per single cell in the ICM (Cao et al., 2007). Consistent with the undeveloped mitochondrial network in the pluripotent ICM, undifferentiated pluripotent mouse and human ESCs contain ~20 mitochondria profiles with poorly developed cristae (Rivolta and Holley, 2002; Baharvand and Matthaei, 2003; St John et al., 2005a, b; Cho et al., 2006; Chung et al., 2007), and high levels of glycolytic enzymes (Kondoh et al., 2007). As the ICM cells progress to become ESCs, they also express low levels of mitochondrial replication factors, such as PolgA, PolgB and Tfam (St John et al., 2005a, b; Facucho-Oliveira et al., 2007) and progressively reduce their mtDNA copy number so that they possess 30–45 copies/cell (Facucho-Oliveira et al., 2007; Spikings 2007). Similarly, undifferentiated hESCs also contain very low levels of mRNA for Tfam, Tfb1m, Tfb2m and PolgA (St John et al., 2005a, b). Consequently, any mtDNA replication observed in the ongoing pluripotent cell will most likely be associated with replenishment rather than active increases in mtDNA copy number which will occur prior to division to ensure that each newly divided mitochondrion has sufficient copies of mtDNA for transcription and transmission.

mtDNA replication and cellular differentiation

The onset of mtDNA replication and cellular differentiation can be studied in ESCs and parallels studies in the mouse embryo. Indeed, the analyses of the expression of a panel of lineage-specific markers showed a strong correlation between days of spontaneous differentiation of mESCs and stages of mouse embryo development (Leahy et al., 1999). To this extent, it has been demonstrated that embryoid bodies (EBs) at Days 1 and 2 are equivalent to mouse embryos at E4.5 to E6.5; EBs at Days 3–5 are equivalent to E6.5 to E7.0 and that EBs after Day 6 are equivalent to E 7.5. Moreover, the embryos at E4.5 and E6.5 are at pre-gastrulation which correlates with the up-regulation of pluripotency genes in Days 1 and 2; R1 mESCs (Facucho-Oliveira et al., 2007). At E6.5 to E7.0, embryos are in gastrulation as determined by detection of markers associated with the endoderm, mesoderm and ectoderm (Leahy et al., 1999), and therefore, down-regulation of the pluripotency genes and up-regulation of lineage-specific markers would be expected (Days 3–4). At E7.5, mouse embryos are in the early stages of organogenesis (Leahy et al., 1999). During Days 1 and 2 of spontaneous differentiation of ESCs, the expression of the pluripotents markers Nanog, Dppa5, Prame7 and Ndp5211 were up-regulated (Facucho-Oliveira et al., 2007; Facucho-Oliveira and St John, 2009). These pluripotency genes were then down-regulated and the percentage of Vimentin+ and Nestin+ and β-tubulin III+ cells increased during Days 3 and 4. At this stage, the expression of PolgA, PolgB and Tfam and the number of mtDNA copies/cell were minimal. By Day 6, the expression of PolgA, PolgB (Facucho-Oliveira et al., 2007; Fig. 4), and the nuclear-encoded ATPase5b and the mtDNA-encoded COXI (Fig. 5) were up-regulated in line with an increase in the number of mitochondria and mtDNA copies/cell (Facucho-Oliveira et al., 2007). This indicates that the onset of organogenesis requires an increase in expression of the mtDNA replication factors and ETC subunits. This is further supported by homozygous PolgA knockout mice dying between days E7.5 and E8.5 (Hance et al., 2005) and homozygous Tfam knockout mice between E8.5 and E10.5 (Larsson et al., 1998).

Modification of mtDNA replication

Stem cells have the ability to respond to the environment in which they exist. For example, the stroma has been proposed as a mediator of stem cell maintenance and thus the regulator of the stem cell niche (Facucho-Oliveira and St John, 2009). These pluripotency genes were then down-regulated and the percentage of Vimentin+ and Nestin+ and β-tubulin III+ cells increased during Days 3 and 4. At this stage, the expression of PolgA, PolgB and Tfam and the number of mtDNA copies/cell were minimal. By Day 6, the expression of PolgA, PolgB (Facucho-Oliveira et al., 2007; Fig. 4), and the nuclear-encoded ATPase5b and the mtDNA-encoded COXI (Fig. 5) were up-regulated in line with an increase in the number of mitochondria and mtDNA copies/cell (Facucho-Oliveira et al., 2007). This indicates that the onset of organogenesis requires an increase in expression of the mtDNA replication factors and ETC subunits. This is further supported by homozygous PolgA knockout mice dying between days E7.5 and E8.5 (Hance et al., 2005) and homozygous Tfam knockout mice between E8.5 and E10.5 (Larsson et al., 1998).
BIII-tubulin, when compared with spontaneous differentiating cells, but it also produced significantly different patterns of mtDNA replication (Facucho-Oliveira et al., 2007) and was marked by a significant increase in expression of Tfam, PolgA and PolgB. During Days 1–5, the RA-induced D3 mESCs had higher levels of PolgA, PolgB and Tfam expression, a higher number of mtDNA copies/cell (Facucho-Oliveira et al., 2007; Fig. 4) and higher percentage of cells with increased ATPase5b and COXI expression (Fig. 5) than the D3 mESCs differentiated in the presence of 25 mM glucose (standard glucose concentration for ESC culture). RA exposure also enhanced neuronal differentiation in comparison to the D3 mESCs differentiated in 25 mM glucose. RA-induced D3 mESCs had higher percentages of Vimentin+ and β-tubulin III+ cells on Day 1, a lower percentage of Vimentin+ cells during Days 2–7 (Facucho-Oliveira et al., 2007; Fig. 4) and a higher percentage of β-tubulin III+ cells on Day 14 (Fig. 5).

The presence of glucose also influences metabolic activity. This is demonstrated by D3 mESCs differentiated in the presence of 1 and...
5 mM glucose having a higher percentage of cells with increased ATPase5b and COXI expression than the D3 mESCs differentiated in the presence of 25 mM glucose on Days 2 and 3 (Fig. 5). Furthermore, the D3 mESCs differentiated in 5 mM glucose showed a higher percentage of β-tubulin III+ cells than the D3 mESCs differentiated in 25 mM glucose on Day 14 (Fig. 5).

**Figure 5** Percentage of cells with increased ATPase5b and COXI expression during differentiation of D3 mESCs in the presence of 1 mM (A), 5 mM (B) and 25 mM (C) of glucose and RA-induced differentiation (D). D3 mESCs differentiated in the presence of 1 and 5 mM of glucose had a higher percentage of cells with increased ATPase5b and COXI expression than D3s differentiated in the presence of 25 mM of glucose on Days 2 and 3 (A–C). However, the D3s differentiated in the presence of 25 mM of glucose had a higher percentage of cells with elevated levels of ATPase5b and COXI than the D3 differentiated in 1 and 5 mM of glucose on Day 6 (A–C). The RA-induced D3 mESCs had the highest percentage of cells with increased ATPase5b and COXI expression on Days 1–5 and 7 (D). The percentage of β-tubulin III+ neurons was significantly higher in the RA-induced D3s and D3s differentiated in the presence of 5 mM of glucose than in the D3s differentiated in the presence of 1 and 25 mM of glucose on Day 14 (E). Bars represent mean ± s.e.m.; significant differences in the percentage of cells with increased ATPase5b and COXI expression are indicated (*P < 0.05).

**mtDNA transmission**

Mammals normally inherit their mtDNA from the population present in the oocyte just prior to fertilization (reviewed in Birky, 1995, 2001) when mtDNA replication has been completed (Spikings et al., 2007). These copies tend to be identical and thus the mtDNA transmitted to
the offspring is homoplasmic (Giles et al., 1980). Following natural fertilization, sperm mtDNA tends to be eliminated thus ensuring the maintenance of homoplasmyn, though this is primarily restricted to intra-specific crosses. This arises as, during early spermatogenesis, spermatagonia are labelled with ubiquitin, which appears to be ‘masked’ as spermatozoa transit through the epididymis and is then present following ejaculation (reviewed in Sutovsky et al., 2004). Having entered the oocyte, sperm mitochondria, and thus its mtDNA complement, are eliminated prior to embryonic gene activation (EGA) for that particular species. For example, in the mouse, sperm mitochondria and mtDNA persist to the 2-cell stage (Kaneda et al., 1995; Shitara et al., 2000), whereas in larger mammals, such as sheep (Zhao et al., 2004) and cattle (Sutovsky et al., 1999), it is eliminated by the 8-cell stage, and in the rhesus macaque (St John and Schatten, 2004; Sutovsky et al., 1999) and humans (St John et al., 2000), this tends to be at the 4–8-cell stage (St John et al., 2000). This suggests that the process of ubiquitination is unlikely to be associated with new genes transcribed during the initial round of transcription following EGA, but is dependent on mRNA carried over from the oocyte and subsequently translated.

Following interspecific crossing, e.g. the mating of two different strains of mouse (Gyllensten et al., 1991; Shitara et al., 2000) or different breeds of sheep (Zhao et al., 2004) or subspecies of rhesus macaques (St John and Schatten, 2004), sperm mtDNA can be transmitted through the offspring. Although only low levels of sperm mtDNA tend to be transmitted, there is no evidence to suggest that this is transmitted to subsequent generations (Gyllensten et al., 1991). Consequently, the molecules recognized by the ubiquitination process are likely to be species-specific. However, the mechanism that precludes sperm mtDNA from being transmitted within a species, such as in the human, is not absolute. Indeed, there is one reported case of sperm mtDNA recombining with oocyte mtDNA creating a new hybrid molecule that segregated to one but not other tissues giving rise to a muscle myopathy in a male patient (Schwartz and Vissing, 2002; Kraytsberg et al., 2004). Furthermore, abnormal human embryos can also fail to eliminate their sperm mtDNA (St John et al., 2000). These outcomes assert the argument that transmission is dependent on mtDNA replication, where sperm will contribute copies to the fertilization process, and if not eliminated, will be available for both replication and transmission. Interestingly, though poor-quality sperm have significantly higher numbers of mtDNA (Diez-Sanchez et al., 2003; May-Panloup et al., 2003; Amaral et al., 2007). However, although poorer quality sperm are then used in ICSI treatment, to date there have been no reported cases of transmission resulting from this treatment (Danan et al., 1999; Marchington et al., 2002).

Transmission of sperm mtDNA is evident in both invertebrates and vertebrates. For example, blue mussels transmit both maternal- and paternal-type genomes to male offspring and maternal only genomes to female offspring (Zouros et al., 1992) suggesting an important outcome for mtDNA transmission in sex selection and sex determination, although this is now contested (Kenchington et al., 2009). Both intra- and interspecific crosses of Drosophila can transmit both oocyte- and sperm-derived mtDNA to their offspring (Kondo et al., 1992). However, there is no reported evidence from such models to suggest that the transmission of potentially diverse mtDNA molecules would give rise to any type of mitochondrial-related disease or whether the resultant offspring exhibit specific advantageous or disadvantageous traits. Although increased genetic distance between the sperm and the oocyte is likely to influence fertilization outcome, the transmission of sperm mtDNA is normally at significantly low levels that it is unlikely to induce a phenotypic disadvantage, except in rare cases. Consequently, the potential for sperm mtDNA to be transmitted is most likely restricted to those crosses where the degree of genetic distance does not prevent mtDNA from restricting fertilization outcome. However, there is likely to be merit in conducting a wide-ranging inclusive study of sperm mtDNA transmission in humans. Although a few studies have indicated that sperm mtDNA is not transmitted (Taylor et al., 2003), they tend to have been centred on samples from patients with mtDNA disease. They would not necessarily provide an insight into the overall levels of sperm mtDNA leakage within a population; this necessitates a large, wide-ranging study.

There are a large number of mtDNA mutations and deletions, which have been associated with a large number of clinical phenotypes (see Wallace, 1999, for review). For example, Leber’s hereditary optic neuropathy (LHON; Wallace et al., 1988; Howell et al., 1991) and neurogenic weakness, ataxia and retinitis pigmentosa (Fryer et al., 1994) are associated with single point mutations in coding genes, whereas mitochondrial myopathy, encephalopathy, lactic acidosis and stroke-like episodes (MELAS; Goto et al., 1990) and deafness (Van den Ouweland et al., 1992) result from single point mutations to rRNAs and rRNAs. mtDNA disease is also associated with large-scale deletions, e.g. the 4977-bp deletion resulting in Kearns–Sayre syndrome (KSS; Schon et al., 1989). The onset of the phenotype for each of these diseases will be determined by the degree of mutant to wild-type molecule that persists within the affected tissue. In the case of LHON, it is predicted that over 60% mutant molecule needs to be present for the phenotypic onset of the disease to be clinically recognized (Chinnery et al., 2001), whereas for MERRF, over 85% mutant needs to be present (Boulet et al., 1992). However, this general rule has now been contradicted by recent findings associated with severe multisystem disorder and respiratory chain deficiency where the mutation was present at less than 25% (Sacconi et al., 2008).

Do the more sophisticated reproductive approaches affect mtDNA transmission?

The introduction of more sophisticated techniques for generating embryos, ESCs and offspring, such as cytoplasmic transfer and nuclear transfer (NT) have led to a re-fashioning of the concept of heteroplasmy. Cytoplasmic transfer, the transfer of cytoplasm from a donor oocyte, was introduced into human-assisted reproduction treatment to increase embryonic developmental outcome for those women who had suffered from repeated embryonic developmental failure, most often at the post-8-cell stage (Cohen et al., 1997). Here, donor oocyte cytoplasm was supplemented into such oocytes. As the oocyte cytoplasm contains not only mRNAs but also mitochondria, it was evident that two populations of mtDNA, one from the recipient oocyte and one from the donor cytoplasm, were transferred with the sperm to aid the fertilization process. Consequently, some of the resultant offspring were heteroplastic (Brenner et al., 2000). Similar outcomes have been observed following...
the transfer of either mouse mitochondria into mouse oocytes (Laips, 1996) or embryos (Nakada et al., 2001) or a germinal vesicle stage or pronuclear karyoplast (Jenuth et al., 1996; Meirelles and Smith, 1997, 1998; Palermo et al., 2002; He et al., 2003; Sato et al., 2005) into a mouse oocyte. As with human cytoplasmic transfer, it was evident that variable amounts of mtDNA from either the supplementing source or the karyoplast could be identified in the resultant offspring. In three independent experiments, this has been shown to range from 5% to 80% (Laips, 1996), 0% to 30% (Jenuth et al., 1996) and 16% to 69% (Meirelles and Smith, 1997). Interestingly though, karyoplast transfer is only successful when the karyoplast is introduced into a cytoplast from the same stage of development (Liu et al., 2003).

Although cytoplasmic transfer can result in live offspring, a longitudinal study of heteroplasmic mice has demonstrated a range of physiological abnormalities when compared with control animals (Acton et al., 2007). These included systemic and pulmonary hypertension, increased body mass and fat mass and abnormalities associated with electrolytes and haematological parameters. These abnormalities most likely arise from the genetic distance between the two populations of mtDNA where the genetic variability which will result in differing amino acids being encoded and could thus mimic mtDNA-type disease. This has perhaps been further exemplified through the generation of mice harbouring a large-scale deletion that induced multiple phenotypes when >85% of the deleted molecule was present (Nakada et al., 2001). Although there is no direct association with human mtDNA-type disease, cytoplasmic transfer has resulted in two cases of Turner’s syndrome leading to one selective and one spontaneous miscarriage, and one case of per-vasive development disorder (Barritt et al., 2001). Nevertheless, a recent study has demonstrated that isolated populations of mitochondria can be transferred from developmentally competent oocytes into their incompetent counterparts resulting in increased fertilization and developmental outcome. This suggests that (i) mitochondria do play a key role in early development (El Shourbagy et al., 2006) and (ii) improved fertilization outcome could be achieved with mtDNA derived from the same genetic source. This could have implications for human-assisted reproductive treatment, where, for a cohort of oocytes obtained from an individual following hormonal stimulation, mitochondria could be collected from several of her oocytes and pooled. These mitochondria could then be introduced into the remaining oocytes to increase fertilization and development outcomes.

Studies involving either embryonic cell NT (ECNT) and somatic cell NT (SCNT) have demonstrated that the mtDNA accompanying the donor cell can be transmitted to the offspring at varying levels along with recipient oocyte mtDNA. This occurs as the process of NT often uses a whole donor cell (Campbell et al., 1996), with donor cell mtDNA being either eliminated during preimplantation development, resulting in homoplasmic transmission of the recipient oocyte’s mtDNA only, or persists resulting in heteroplasmy (i.e. both donor and recipient mtDNA). Indeed, transmission of donor cell mtDNA ranges from 0% to 63% in preimplantation embryos (Meirelles et al., 2001) and 0% to 59% in live offspring (Takeda et al., 2003; see Table II). This tends to be independent of whether intra- or interspecific or interspecies ECNT or SCNT is performed (St John et al., 2004; Tables II and III), e.g. bovine embryos and offspring derived by both intra- and interspecific NT (Meirelles et al., 2001; Do et al., 2002; Takeda et al., 2003); and in caprine embryos (Jiang et al., 2004) and porcine (Takeda et al., 2006) and Macaca mulatta (St John and Schatten, 2004) offspring derived by interspecific ECNT and SCNT. It is thus evident that embryonic and somatic cell mtDNA can escape the selective mechanism that targets and eliminates intra-specific sperm mitochondria in the fertili-zed oocyte to maintain homoplasy (Shitara et al., 2000; Sutovsky and Schatten, 2000). Indeed, in the two non-human primates derived by interspecific ECNT, both oocyte and sperm mtDNA contributing to the donor blastomere were transmitted (St John and Schatten, 2004). However, the factors that allow for the large differences in heteroplasmy between individuals remain to be determined experimentally.

In order to determine why donor cell mtDNA might be transmitted, an early study indicated that the amount of mtDNA that was transmitted was proportional to the amount of mtDNA accom-panying the embryonic donor cell at the time ECNT was performed. The donor cells were from varying stages of preimplantation development thus containing decreasing levels of mtDNA following division during preimplantation development (Steinborn et al., 1998). However, closer examination of these data suggests that this is perhaps not as decisive an outcome as first thought. A further study set out to understand why donor cell mtDNA persists at levels inconsis-tent with its original contribution. Intra- and interspecific SCNT-ovine embryos, using donor cells depleted to 0.01% (residual – mtDNAα) and 0.1% of their original mtDNA content, were generated and compared with time-matched non-depleted (mtDNAβ) cells (Lloyd et al., 2006). Surprisingly, donor cell mtDNA persisted to the hatched blastocyst stage for all combinations. This outcome has been attributed to the persistent expression of mtDNA-specific replication factors, such as POLGA, POLGB and TFAM, during preimplantation development, which contrasts with their in vitro-derived counterparts (Bowles et al., 2007b).

In interspecies SCNT embryos (iSCNT; see Table III), donor cell mtDNA has been detected at the 16-cell stage in human-bovine crosses (Chang et al., 2003), the blastocyst stage in macaque–rabbit (Yang et al., 2003) and in a small minority of caprine-ovine embryos (4 of 40; range = 2–20+ cell embryos; Bowles et al., 2007b). In this small number of heteroplasmic embryos, significant increases in donor cell mtDNA were observed again suggesting that occasionally selective replication does take place. Although the generation of live offspring is highly unlikely through iSCNT, it remains to be determined whether residual levels of mtDNA would persist and proliferate or be eliminated in differentiating ESCs derived through iSCNT.

It is worth, however, stressing that considerable confusion has been created by the current zoological nomenclature that assigns species status to populations that are often only subspecies. This is very spring derived by both intra- or interspecific crosses (cf. Tables II and III). However, it should be stressed that per-forming mtDNA analysis with different detection methods will often result in different levels of mtDNA heteroplasmy being observed, thus we have listed the relative methods used. Several conclusions can be drawn: (i) different tissues will have different mtDNA
### Table II  mtDNA content in cloned embryos, fetuses and offspring generated by intra- and interspecific nuclear transfer.

<table>
<thead>
<tr>
<th>Species</th>
<th>Sample analysed</th>
<th>Degrees of heteroplasm</th>
<th>Detection methods</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bovine</td>
<td>Sampled blastomeres, blastocysts, tissues of a newborn clone</td>
<td>NT-blastomere zygotes and blastocysts had similar levels of heteroplasm (donor cell—11.0% and 14.0%, respectively), 5.8% at the 9- to 16-cell stage. Heteroplasmic levels in NT-fibroblast zygotes—4.7%, 9- to 16-cell—12.9%, blastocyst 2.2%. No donor cell mtDNA detected in gametes or tissues of newborn calf</td>
<td>Allele-specific PCR with fluorochrome- specific primers</td>
<td>Ferreira et al. (2007)</td>
</tr>
<tr>
<td>Sheep</td>
<td>Seven fetuses and five lambs cloned from fetal fibroblasts</td>
<td>Most of the heteroplasmic clones exhibited low levels of heteroplasm (0.1–0.9%, n = 6). High levels of heteroplasm (6.8–46.5%) in 1 clone</td>
<td>Quantitative amplification refractory mutation system (ARMS) PCR</td>
<td>Burgstaller et al. (2007)</td>
</tr>
<tr>
<td>Pig (S. vittatus and S. scrofa)</td>
<td>4 SCNT pigs derived from S. vittatus nuclear donor cells and S. scrofa recipient oocytes</td>
<td>0.1–1% heteroplasm in blood and hair roots. 0–44% in tissues of one animal</td>
<td>SSCP and agarose gel PCR-RFLP with image analysis and SNP-PCR</td>
<td>Takeda et al. (2006)</td>
</tr>
<tr>
<td>Sheep</td>
<td>2-cell to hatched blastocyst</td>
<td>Donor cell mtDNA persisted in some blastocysts (0.01–0.9%)</td>
<td>Allele-specific real-time PCR</td>
<td>Lloyd et al. (2006)</td>
</tr>
<tr>
<td>Pig (S. scrofa)</td>
<td>5 pigs derived from 2 nuclear donor cell types (double nuclear transfer)</td>
<td>Donor cell mtDNA eliminated but transmission of first recipient oocyte and zygote mtDNA—bimatal mtDNA transmission</td>
<td>Sequence analysis, RFLP, AS-PCR and primer extension polymorphism analysis</td>
<td>St John et al. (2000Sa, b)</td>
</tr>
<tr>
<td>Cattle (B. taurus)</td>
<td>3 calves</td>
<td>Heteroplasm detected in skin and blood (1–3.6%)</td>
<td>Agarose gel electrophoresis and image analysis</td>
<td>Theoret et al. (2006)</td>
</tr>
<tr>
<td>Cattle (B. taurus)</td>
<td>5 calves</td>
<td>Heteroplasm in skin samples</td>
<td>AS-PCR on agarose gel</td>
<td>Han et al. (2004)</td>
</tr>
<tr>
<td>Goat (C. bex and C. hircus)</td>
<td>1-, 2-, 4-cell, and morula stage</td>
<td>Heteroplasm only at 1- and 2-cell stage</td>
<td>DNA sequencing on agarose gel and DNA sequencing</td>
<td>Jiang et al. (2004)</td>
</tr>
<tr>
<td>Mouse (Mus musculus domesticus, Mus m. molossinus)</td>
<td>25 adult mice derived from 3 nuclear donor cell types</td>
<td>Heteroplasm in brain, kidney, liver and tail of all but one animal. Amount of donor cell mtDNA depended on donor cell type and tissue with significant differences between sertoli (0.78%) and fibroblast (2.4%) derived mice, and between brain (0.6%) and liver (1.7%) tissues, heteroplasm detected in all 20 placenta analysed</td>
<td>Allele specific real-time PCR</td>
<td>Inoue et al. (2004)</td>
</tr>
<tr>
<td>Cattle</td>
<td>12 day fetus derived from one donor cell type; 3 recipient oocyte cytoplasm: B. taurus mtDNA type B, type C, and B. indicus mtDNA</td>
<td>Mainly homoplasmic with occasional heteroplasmic at 0.5–0.7%</td>
<td>PCR-RFLP on agarose gel Cloned PCR fragment</td>
<td>Hiendleder et al. (2003)</td>
</tr>
<tr>
<td>Cattle (B. taurus)</td>
<td>16 embryos from 3 nuclear donor cell types Cumulus cells (holstein), ear skin cells (Japanese black) and fetal fibroblasts (Japanese black)</td>
<td>3 of 9 NT calves were heteroplasmic with donor cell mtDNA ranging from 0% to 59%</td>
<td>PCR-mediated single-strand conformation polymorphism analysis and southern blot hybridization</td>
<td>Takeda et al. (2003)</td>
</tr>
<tr>
<td>Cattle (B. taurus)</td>
<td>1-, 2-, 4-, 8- and 16-cell, morula and blastocysts</td>
<td>Heteroplasm at all stages</td>
<td>Allele specific PCR on agarose gel and direct sequencing of PCR product</td>
<td>Do et al. (2002)</td>
</tr>
<tr>
<td>Cattle (B. taurus, B. indicus)</td>
<td>8 B. taurus nucleus and B. indicus mtDNA (n = 11)</td>
<td>0.6–2.8% heteroplasm in 4 out of 11 I</td>
<td>Allele specific real-time PCR</td>
<td>Steinborn et al. (2002)</td>
</tr>
<tr>
<td>Cattle (B. indicus)</td>
<td>Reconstructed oocytes, blastomeres and whole morula, blastocysts and fetal and offspring tissues</td>
<td>0–63% in preimplantation embryos; reduction or complete elimination of donor cell mtDNA during development</td>
<td>PCR restriction fragment length polymorphism</td>
<td>Meirelles et al. (2001)</td>
</tr>
<tr>
<td>Cattle (B. taurus)</td>
<td>6 fetuses and 4 calves derived from 3 nuclear donor cell types</td>
<td>0.1–4% heteroplasm in all but one sample in blood, cerebellum, hear, kidney, liver, lung, muscle and skin</td>
<td>Allele-specific real-time PCR</td>
<td>Steinborn et al. (2000)</td>
</tr>
</tbody>
</table>
Mitochondrial DNA transmission, replication and inheritance

Table II Continued

<table>
<thead>
<tr>
<th>Species</th>
<th>Sample analysed</th>
<th>Degrees of heteroplasy</th>
<th>Detection methods</th>
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</thead>
<tbody>
<tr>
<td>Sheep (O. aries)</td>
<td>10 sheep derived from 3 nuclear donor cell types</td>
<td>No heteroplasy in blood, milk muscle and placenta</td>
<td>Southern blot hybridization with phosphorimager,</td>
<td>Evans et al. (1999)</td>
</tr>
<tr>
<td>Cattle blastomeres from Japanese back cow morula</td>
<td>21 cloned calves produced from recipient oocytes of undefined mtDNA genotypes, and 6 cloned calves from defined mtDNA genotype</td>
<td>20 of 21 calves homoplasmic; 1 calf heteroplasmic</td>
<td>PCR-mediated single-strand conformation polymorphism</td>
<td>Takeda et al. (1999)</td>
</tr>
</tbody>
</table>

content at specific developmental stages and in the adult; (ii) preimplantation embryos will carry different mtDNA patterns for cloned offspring when compared with natural fertilization and (iii) the mtDNA content in blastomeres either undergoes replication or escapes degradation during cleavage and can be degraded after the blastocyst stage or lost during somatic cell development, as revealed by the lack of donor cell mtDNA at birth in some offspring.

What are the consequences of introducing somatic mitochondria and donor cell mtDNA into oocytes?

Most studies of mitochondria in mammalian oocytes and preimplantation stage embryos have focused primarily on their metabolic capacity to generate ATP. However, mitochondria are also involved in other processes such as establishing developmental competence, calcium homeostasis and apoptosis (van Blerkom, 2004). The inner mitochondrial membrane potential, morphological changes and translocation of mitochondria also play key roles in embryo development. In pig, the translocation of mitochondria from the oocyte cortex to the perinuclear area indicates positive developmental potential that was reduced in porcine SCNT embryos compared with their in vitro counterparts (Zhong et al., 2008). Through transmission electron microscopy, these authors also demonstrated that unequal donor cell mitochondrial distribution and complex mitochondrial morphologies may influence mitochondrial translocation and affect development. Another report in goat–goat and goat–cattle embryos demonstrated that inter- and intra-species reconstructed embryos have a different pattern of developmental change to that of in vivo-produced embryos for mitochondria (Tao et al., 2008). These ultrastructural differences might contribute to the compromised developmental potential of reconstructed embryos. In cloned mice, again, there appears to be no difference observed in the number of mitochondria, but their mitochondria displayed ultrastructural alterations (Han et al., 2008). Interestingly, the authors revealed that the glucose present in the culture medium compensated for the potential effects of ATP-dependent processes being severely compromised during the first 2-cell cycles.

Somatic cell mitochondria do appear to adversely affect embryonic development. For example, it has been demonstrated that development rates to blastocyst of parthenogenetically activated oocytes supplemented with somatic mitochondria decreased in comparison to non-supplemented controls and those supplemented with oocyte cytoplasm (Takeda et al., 2005). Furthermore, SCNT blastocysts generated with mtDNA R donor cells (SFF1 line) had significantly more embryonic cells at blastocyst compared with those generated with non-mtDNA depleted cells (Lloyd et al., 2006). In addition, significantly more caprine-ovine iSCNT embryos generated with mtDNA R donor cells reached the 20-cell stage of preimplantation development than those generated with mtDNA R cells (Bowles et al., 2007a). To the contrary, transferring mitochondria from granulosa cells seems to boost preimplantation development, especially for those oocytes characterized by poor mtDNA content and cytoplasmic appearance (Hua et al., 2007). The difference between granulosa cells and other somatic cells may therefore be dependent on their differing maturation statuses and requirements for energy.

Although there appears to be a tolerance for interspecific crosses, the mtDNA genomes for different breeds of cattle and pigs have sequence variations in their coding genes. This is reflected in interspecific bovine (Steinborn et al., 2002) and porcine (St John et al., 2005a, b) offspring. It is thus likely that the subtle changes in the amino acid sequences for these two mtDNA genomes would generate an incompatibility that would affect the interactions with closely located nuclear-encoded polypeptides within the same complex of the ETC. Indeed, during the embryo-fetal period, expression of a mutated gene in specific tissues can attribute to the failure of the respiratory chain complex assembly and function (Minai et al., 2008). It has been suggested that mtDNA heteroplasm also may induce incompatibility between the nucleus and cytoplasm, which will again inhibit the development of cloned embryos (Chen et al., 2002; Bowles et al., 2007a, b, 2008), especially at the time of genomic activation (Sansinena et al., 2005). A recent report in cat-bovine cloned embryos demonstrated that the developmental block, which normally occurs just after EGA, was associated with the presence of foreign mtDNA (Thongphakdee et al., 2008). In a study of minke whale iSCNT, the granulosa–cumulus cells had the ability to dedifferentiate in bovine and porcine oocytes up to the 4-cell stage, which is again indicative of developmental block at EGA and mitochondrial incompatibility between the recipient oocytes and donor cells (Ikumi et al., 2004). When using iSCNT, it is thus critical to choose compatible donor and recipient cells. Furthermore, the regulation of mtDNA copy number in the preimplantation embryo may affect outcome. For example, in some species, such as mouse, the number of mtDNA copies/embryo is stable during cleavage, with subsequent blastomeres having fewer copies of mtDNA until replication begins post-implantation (Thundathil et al., 2005), whereas domestic species, such as cows and pigs, significantly reduce their mtDNA content during preimplantation.
### Table III mtDNA content in cloned embryos, fetuses and offspring generated by interspecies nuclear transfer.

<table>
<thead>
<tr>
<th>Donor cell</th>
<th>Recipient oocyte</th>
<th>Sample analysed</th>
<th>Degree of heteroplasmy</th>
<th>Detection methods</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mixed-breed domestic cat fibroblasts</td>
<td>Bovine</td>
<td>1-, 2-, 4- and 8-cell stage embryos</td>
<td>Cat mtDNA stable from 1- to 8-cell. Bovine mtDNA significantly lower at the 8-cell stage than 4-cell stage</td>
<td>Quantitative PCR</td>
<td>Thongphakdee et al. (2008)</td>
</tr>
<tr>
<td>Goat fetal fibroblasts</td>
<td>Sheep</td>
<td>1- and 4-cell embryos, morula and blastocyst</td>
<td>Pre-morula stage, goat mtDNA 2.0; 0.012 at blastocyst stage.</td>
<td>Real-time PCR</td>
<td>Ma et al. (2008a, b)</td>
</tr>
<tr>
<td>Ovine fetal fibroblast</td>
<td>Bovine</td>
<td>Individual blastocysts and blastomeres</td>
<td>Ovine mtDNA constant at 1% from 1- to 8-cell stage, 0.6% at 16-cell stage and 0.1% at blastocyst stage.</td>
<td>Real-time PCR</td>
<td>Hua et al. (2008)</td>
</tr>
<tr>
<td>Postmortem wolf somatic cells (Canis lupus)</td>
<td>Dog</td>
<td>Six cloned wolf pups</td>
<td>Homoplasmic for recipient oocyte mtDNA</td>
<td>Species specific real-time PCR</td>
<td>Oh et al. (2008)</td>
</tr>
<tr>
<td>Cattle (B. taurus)</td>
<td>Rabbit (Oryctolagus cunniculus)</td>
<td>1-, 2-, 4-/8-cell, morula and blastocyst embryos from: (i) heteroplasmic SCNT animal donor; and (ii) a homoplasmic nuclear donor</td>
<td>Heteroplasmy detected at all stages of preimplantation. Rabbit mtDNA remained constant to the morula stage and increased sharply in both types of blastocyst. Bovine mtDNA remained constant in SCNT donor cell group during preimplantation development but decreased sharply in the homoplasmic donor group post-morula</td>
<td>Species specific PCR on agarose gel electrophoresis and DNA sequencing</td>
<td>Jiang et al. (2006)</td>
</tr>
<tr>
<td>Ibex (C. ibex)</td>
<td>Rabbit (O. cunniculus)</td>
<td>2-, 4-, 8- and 16-cell morula and blastocyst embryos</td>
<td>Heteroplasmy detected at all stages; rabbit mtDNA increased from the 16-cell stage. Ibex mtDNA decreased after 16-cell stage</td>
<td>Species-specific real-time PCR</td>
<td>Jiang et al. (2005)</td>
</tr>
<tr>
<td>Human granulosa and fibroblast cells</td>
<td>Bovine</td>
<td>Preimplantation stages embryos</td>
<td>Most embryos heteroplasmic to blastocyst stage</td>
<td>Species specific PCR and DNA sequencing</td>
<td>Illmensee et al. (2006)</td>
</tr>
<tr>
<td>Takin (Budorcas taxicolor)</td>
<td>Yak (Bos grunniens)</td>
<td>2 blastocysts</td>
<td>Heteroplasmy in both blastocysts</td>
<td>Species specific PCR on agarose gel electrophoresis and DNA sequencing</td>
<td>Li et al. (2006)</td>
</tr>
<tr>
<td>Macaque (Macaca multatto)</td>
<td>Rabbit (O. cunniculus)</td>
<td>1-, 2-, 4-, 8- and 16-cell, morula, blastocyst</td>
<td>Macaque mtDNA: 2.0% (1-cell), 1.3% (2-cell), 1.7% (4-cell), 0.6% (8-cell), 0.8% (16-cell), 1.3% (morula) and 0.011% blastocyst</td>
<td>Species-specific real-time PCR</td>
<td>Yang et al. (2004)</td>
</tr>
<tr>
<td>Red common carp</td>
<td>Goldfish</td>
<td>Blastula, gastrula, somite, muscular reaction, blood circulation, larval and adult stages</td>
<td>Blastulae all heteroplasmic. In late stage embryos post-vascularization only recipient oocyte mtDNA detected</td>
<td>Species specific PCR on agarose gel electrophoresis with image analysis</td>
<td>Sun et al. (2005)</td>
</tr>
<tr>
<td>Human (Homo sapiens)</td>
<td>Bovine (B. taurus)</td>
<td>2-, 4-, 8- and 16-cell, morula and blastocysts</td>
<td>Heteroplasmic to 16-cell stage. Homoplasmic post-morula stage</td>
<td>Species specific PCR on agarose gel with image analysis, DNA sequencing</td>
<td>Chang et al. (2003)</td>
</tr>
<tr>
<td>Human (H. sapiens)</td>
<td>Rabbit (O. cunniculus)</td>
<td>Blastocysts</td>
<td>Heteroplasmic</td>
<td>In situ hybridization mtDNA interspecies PCR</td>
<td>Chen et al. (2003)</td>
</tr>
<tr>
<td>Macaque (M. multatto)</td>
<td>Rabbit (O. cunniculus)</td>
<td>1-, 2-, 4-, 8- and 16-cell, morula and blastocyst</td>
<td>Heteroplasmy detected at all stages.</td>
<td>Species specific PCR on agarose gel, DNA sequencing</td>
<td>Yang et al. (2003)</td>
</tr>
<tr>
<td>Giant panda (Ailuropoda melanoleuca)</td>
<td>Rabbit (O. cunniculus)</td>
<td>Blastocysts and 2 fetuses</td>
<td>Heteroplasmy detected in blastocysts. Both fetuses homoplasmic for donor cell</td>
<td>Species specific PCR on agarose gel, sequencing of PCR product</td>
<td>Chen et al. (2002)</td>
</tr>
<tr>
<td>Gaur (Bos gaurus)</td>
<td>Cattle (B. taurus)</td>
<td>3 fetuses</td>
<td>No heteroplasmic in brain eye, gonad, hear, intestine, kidney, liver, lung, muscle skin and tongue</td>
<td>PCR-PFLP and phosphorimager</td>
<td>Lanza et al. (2000)</td>
</tr>
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Mitochondrial DNA replication and transmission following induced pluripotency

Induced pluripotent cells (iPSCs) are somatic cells that have been induced to become ESC-like due to the introduction of defined factors (Takahashi and Yamanaka, 2006). They then express key pluripotent markers such as Oct4, Nanog and Sox-2 and have the ability to differentiate into multiple cell types. It however remains to be determined whether such cells have the ability to regulate their mtDNA copy number in their undifferentiated state as ESCs do. It further needs to be demonstrated whether these cells can effectively regulate their patterns of mtDNA replication as the process of differentiation takes place in ESCs (Facucho-Oliveira et al., 2007). It is unlikely that those cells generated through NT would be able to do so, due to the up-regulation of the mtDNA replication factors during early embryonic development (Bowles et al., 2007b). However, the iPSCs should, in line with pluripotent cells (Facucho-Oliveira et al., 2007; Facucho-Oliveira and St John, 2009), be able to demonstrate low levels of mtDNA replication to maintain existing populations. Attempts at reprogramming cells using cell fusion (Sumner et al., 2009), where an adult cell is fused to an ESC, may cause several problems in terms of the regulation of mtDNA. These may include heteroplasmy, because of two different sources of cells being used to generate the newly fused reprogrammed cell, and the failure to adopt appropriate numbers of mtDNA copies. As reprogrammed cells have been proposed as models that can be used for the study of specific diseases and to screen drugs for their efficacy (Bowles et al., 2007a), they need to be carefully investigated to determine their appropriateness for such analyses. This is because such experimental protocols may contain two experimental variables, namely increased numbers of mtDNA copy and/or heteroplasmy, which may result in irregular ATP production and the genetic rearrangement associated with the particular disease. Consequently, it would not be clear from the experimental outcome whether the cellular dysfunction arose through the genetic rearrangement or the cell’s inability to generate sufficient levels of ATP through OXPHOS. Equally so, the inability of the drug to induce, e.g. appropriate calcium oscillations in neuronal or cardiac cells, might also be due to impaired OXPHOS function. However, as such cells offer considerable hope as clinical entities, then it is worth investigating whether it is feasible to simply regulate the mtDNA content of these cells so that they are suitable for therapeutic and experimental use.

mtDNA segregation

The segregation of mtDNA molecules tends to follow a pattern of random genetic drift (reviewed in Shoubridge and Wai, 2007).
Consequently, it is not possible to predict which cell types or tissues mutant molecules will migrate to during early development. This is the case for both somatic and germ cells and specifically relevant to female primordial germ cells (PGCs), which, when harbouring mutant molecules, would thus ensure their continued transmission to subsequent generations. PGCs originate from the extra-embryonic endoderm and migrate to the germinal ridge (Surani et al., 2007). At this stage, they contain ~200 copies of mtDNA and maintain relatively fewer numbers during the early stages of oogenesis (Smith and Alcivar, 1993; Cao et al., 2007; Cree et al., 2008, Fig. 4). As PGCs differentiate and progress from meiosis I to II, mtDNA is clonally expanded (Smith and Alcivar, 1993). Consequently, those mature mouse oocytes that are competent for the process of fertilization or oocyte reconstruction will have between 180 000 and 200 000 mtDNA copies (Cao et al., 2007; Cree et al., 2008; Wai et al., 2008) with 1–2 copies/mitochondrion (Piko and Matsumoto, 1976; Piko and Taylor, 1987), whereas bovine (May-Panloup et al., 2005), porcine (Spikings et al., 2007) and human (Steuerwald et al., 2000; Reynier et al., 2001; Almeida-Santos et al., 2006) oocytes have between 100 000 and 700 000 copies (Fig. 4). Consequently, the combination of random genetic drift and clonal expansion of mtDNA would explain the large variation in mutant molecules found within the cohorts of oocytes from those women who are potential carriers of mtDNA mutations. For example, mtDNA mutation: wild type for a specific mitochondrial disorder, MELAS, ranged from 0% to 97% (Bliek et al., 1997).

Following fertilization of the oocyte, mtDNA copy number remains constant during mouse preimplantation development up to the morula stage but is reduced in porcine (Spikings et al., 2007) and bovine (May-Panloup et al., 2005) embryos. mtDNA replication is then initiated at the blastocyst stage in those mammalian species so far investigated (May-Panloup et al., 2005; Thundatih et al., 2005; Spikings et al., 2007). However, this replication is specific to the trophoectodermal cells, the cells that give rise to the placenta, whereas those cells giving rise to the embryo or ESCs, the ICM cells, have very few mitochondria present and thus copies of the mtDNA genome (Spikings et al., 2007). The ICM cells are likely to continue to reduce their mtDNA copy number as they continue to divide before they undergo gastrulation, as evidenced by human and mouse ESCs as they continue to divide before they undergo gastrulation, as evidenced by human and mouse ESCs at various stages of differentiation, the cell is capable of adaptation. The mitochondrial machinery is capable of sustaining phasic changes, thus ensuring that, at various stages of differentiation, the cell is capable of adaptation. Consequently, PGD may not reflect later stage outcomes. This is further likely as it appears that mtDNA rearrangements and wild-type molecules are able to intermix so that they do not remain as separate entities within distinct organelles or cells (Nakada et al., 2001).

The variable transmission of mtDNA has questioned the feasibility of using preimplantation genetic diagnosis (PGD) on embryos generated following either in vitro fertilization or intra-cytoplasmic sperm injection protocols to determine whether individual preimplantation blastomeres contain equal loading for mutant molecules. Interestingly, although mutant load can be different in various mature cell types (reviewed in McFarland et al., 2007), there appears to be uniformity in individual blastomeres (Dean et al., 2003) suggesting that segregation is a post-gastrulation event. Although this is a reassuring outcome, it suggests that PGD is only useful as a predictor that mature cells might have high mutant mtDNA loading when a critical threshold of mutant loading is reached in preimplantation embryos. For those preimplantation embryos with lower levels of mutation, it can inform patients that their offspring would most likely be affected (Poulten et al., 2009). However, the overriding question is: how sure can we be?

As mtDNA replication in fetal cells is initiated post-gastrulation and from a few molecules present in uncommitted cells, it allows individual cell types to be populated with the appropriate numbers of mtDNA copies to match their requirements for OXPHOS-derived ATP (Moyes et al., 1998). Such examples would be neuronal, muscle and cardiac-type cells where neuronal cells would have between 8 and 10 copies/mitochondrion (Satoh and Kuroiwa, 1991). It is likely that there is a refinement of mtDNA copy at various stages of development and these are likely to be linked to the expression of certain genes during the various different stages of development (Facucho-Oliveira et al., 2007; Facucho-Oliveira and St John, 2009). Early phases of replication are likely to be associated with replenishment of a genome and also to determine whether the replication machinery is capable of sustaining phasic changes, thus ensuring that, at various stages of differentiation, the cell is capable of adaptation. Consequently, PGD may not reflect later stage outcomes. This is further likely as it appears that mtDNA rearrangements and wild-type molecules are able to intermix so that they do not remain as separate entities within distinct organelles or cells (Nakada et al., 2001).

**Does heteroplasmy following reproductive technologies have implications for embryo, fetal and offspring survival and wellbeing?**

Currently, there are no effective intervention strategies available to prevent the transmission of mtDNA disease from mother to offspring. Experimental approaches concentrate on determining whether germinal vesicle or pro-NT might be appropriate approaches. In this case, the germinal vesicle or pronuclei from the affected oocyte would be transferred into an enucleated egg that was not carrying the mutant population (Meirelles and Smith, 1998). The degree of donor cell transmission that arises or transfer of donor mitochondria following cytoplasmic transfer when at its higher levels could mimic conditions associated with mitochondrial disease. Studies, for example, using mouse strains with variants in the coding sequences between the two strains would be similar to clinical-onset heteroplasmy and would produce similar outcomes to those observed following SCNT when different breeds of cattle and pig contributed by a donor cell and a recipient oocyte. As previously demonstrated, some replication of such mtDNA can take place during preimplantation development (Meirelles and Smith, 1998) and persist at high levels in a few offspring.
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(Sato et al., 2005). Consequently, it is worth questioning whether the degree of mtDNA heteroplasmy in cloned offspring would indeed account for some of the phenotypic diseases observed from NT. Although there is no direct evidence currently to support this, there is substantive evidence that is associated with those malformations in tissues from fetuses and offspring following SCNT (Cibelli et al., 2002). A recent study has also demonstrated that donor ooplasm transfer can affect growth in early life with interspecific germinal vesicle transfer causing growth abnormalities post-natally (Cheng et al., 2009). Such tissues that might be affected are lung, heart and liver (Bowles et al., 2008), which are high ATP users and would therefore be more likely to be indicative of failure to produce sufficient compatible genomes for the ETC. This would be further exaggerated following iSCNT where the nuclear and mtDNA-encoded subunits of the ETC would show varying degrees of incompatibility.

As the transmission of donor cell mtDNA in cloned offspring could also be explained by random genetic drift, as these molecules tend to segregate in a non-uniform manner, the up-regulation of mtDNA replication factors during preimplantation development (Bowles et al., 2007a, b) and their preferential selection for one molecule over another (Bowles et al., 2008) will influence the number of molecules present as segregation takes place. Consequently, any assisted reproductive technology that uses an oocyte cytoplasm to reprogramme the donor cell and allows mixing of mtDNA populations will affect any PGD outcomes as the population present in the preimplantation embryo will continue to be replicated unlike its fertilized counterpart and would be only predictive of that stage of development. This outcome may be equally valid for the transfer of a germinal vesicle or a pronuclei which will bring some accompanying mtDNA as part of the transfer. Consequently, entire elimination of donor cell mtDNA would be perhaps an appropriate safeguard for such approaches (Bowles et al., 2007a, b). A recent attempt in M. mulatta has suggested that this might be achieved (Tachibana et al., 2009) but these results must be viewed with considerable caution as it still needs to be verified whether the transferred spindle has accompanying mtDNA and whether this was transmitted to various tissues of the offspring.

Conclusions

mtDNA transmission and replication from the gamete through the preimplantation embryo into the fetus and into the offspring is a highly regulated process where mtDNA replication is initiated at precise stages throughout development. The task for scientists at the moment is to elicit when these key stages are and how many precise stages throughout development. The task for scientists at high and regulated process where mtDNA replication is initiated at

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