Clinical update

Diabetes and vascular disease: pathophysiology, clinical consequences, and medical therapy: part I

Francesco Paneni1,2, Joshua A. Beckman3, Mark A. Creager3, and Francesco Cosentino1,4*

1Cardiology and Cardiovascular Research, University of Zürich, Zürich, Switzerland; 2IRCCS Neuromed, Pozzilli, Italy; 3Cardiovascular Division, Brigham and Women’s Hospital and Harvard Medical School, Boston, MA 02115, USA; and 4Cardiology, Department of Clinical and Molecular Medicine, University of Rome ‘Sapienza’, Rome, Italy

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Hyperglycemia and insulin resistance are key players in the development of atherosclerosis and its complications. A large body of evidence suggest that metabolic abnormalities cause overproduction of reactive oxygen species (ROS). In turn, ROS, via endothelial dysfunction and inflammation, play a major role in precipitating diabetic vascular disease. A better understanding of ROS-generating pathways may provide the basis to develop novel therapeutic strategies against vascular complications in this setting. Part I of this review will focus on the most current advances in the pathophysiological mechanisms of vascular disease: (i) emerging role of endothelium in obesity-induced insulin resistance; (ii) hyperglycemia-dependent microRNAs deregulation and impairment of vascular repair capacities; (iii) alterations of coagulation, platelet reactivity, and microparticle release; (iv) epigenetic-driven transcription of ROS-generating and proinflammatory genes. Taken together these novel insights point to the development of mechanism-based therapeutic strategies as a promising option to prevent cardiovascular complications in diabetes.

Keywords
Diabetes • Vascular disease • Pathophysiology

Introduction

The number of people with diabetes mellitus is alarmingly increasing due to the growing prevalence of obesity, genetic susceptibility, urbanization, and ageing.1,2

Type 2 diabetes, the most common form of the disease, may remain undetected for many years and its diagnosis is often made incidentally through an abnormal blood or urine glucose test. Hence, physicians often face this disease at an advanced stage when vascular complications have already occurred in most of patients. Macrovascular complications are mainly represented by atherosclerotic disease and its sequelae. Diabetes-related microvascular disease such as retinopathy and nephropathy are major causes of blindness and renal insufficiency.1

Based on this scenario, a better understanding of the mechanisms underlying diabetic vascular disease is mandatory because it may provide novel approaches to prevent or delay the development of its complications. This review will focus on the most current advances in the pathophysiology of vascular disease (Part I) and will address clinical manifestations and management strategies of patients with diabetes (Part II).

Hyperglycemia, oxidative stress, and vascular disease

The alterations in vascular homeostasis due to endothelial and smooth muscle cell dysfunction are the main features of diabetic vasculopathy favouring a pro-inflammatory/thrombotic state which ultimately leads to atherothrombosis. Macro- and microvascular diabetic complications are mainly due to prolonged exposure to hyperglycemia clustering with other risk factors such as arterial hypertension, dyslipidemia as well as genetic susceptibility.3 Interestingly, nephropathy, retinopathy, and diabetic vascular disease are in line with the notion that endothelial, mesangial, and retinal cells are all equipped to handle high sugar levels when compared with other cell types.4 The detrimental effects of glucose already occur with glycemic levels below the threshold for the diagnosis of diabetes. This is explained by the concept of ‘glycemic continuum’ across the spectrum of prediabetes, diabetes, and cardiovascular risk.5–8 Early dysglycemia caused by obesity-related insulin resistance or impaired insulin secretion is responsible for functional and
structural alterations of the vessel wall culminating with diabetic vascular complications.

The initial trigger whereby high glucose concentrations alter vascular function is the imbalance between nitric oxide (NO) bioavailability and accumulation of reactive oxygen species (ROS), leading to endothelial dysfunction. Indeed, hyperglycemia-induced generation of superoxide anion (O$_2^-$) inactivates NO to form peroxynitrite (ONOO$^-$), a powerful oxidant which easily penetrates across phospholipid membranes and induces substrate nitration. Protein nitrosylation blunts activity of antioxidant enzymes and endothelial NO synthase (eNOS, Figure 1). Importantly, reduced NO bioavailability is a strong predictor of cardiovascular outcomes.

Overproduction of ROS by mitochondria is considered as a causal link between elevated glucose and the major biochemical pathways involved in the development of vascular complications of diabetes. Indeed, hyperglycemia-induced ROS production triggers several cellular mechanisms including polyol and hexosamine flux, advanced glycation end products (AGEs), protein kinase C (PKC) activation, and NF-$\kappa$B-mediated vascular inflammation. One of the main sources of ROS in the setting of hyperglycemia is represented by PKC and its downstream targets. The hyperglycemic environment induces a chronic elevation of diacylglycerol levels in endothelial cells with subsequent membrane translocation of conventional ($\alpha$, $\beta_1$, $\beta_2$) and non-conventional ($\delta$) PKC isoforms. Once activated, PKC is responsible for different structural and functional changes in the vasculature including alterations in cellular permeability, inflammation, angiogenesis, cell growth, extracellular matrix expansion, and apoptosis. An important consequence of PKC activation is ROS generation. In vascular endothelial cells, hyperglycemia-induced activation of PKC increases superoxide production via NADPH oxidase (Figure 1). Indeed, treatment with a PKC$\beta$ inhibitor suppresses NADPH-dependent ROS generation.

More recently, it has been reported that glucose-induced activation of PKC $\beta_2$ isoform phosphorylates p66$^{Shc}$ at serine 36 leading to its translocation to the mitochondria, cytochrome c oxidation and accumulation of ROS into the organelle. The p66$^{Shc}$ adaptor protein functions as a redox enzyme implicated in mitochondrial ROS generation and translation of oxidative signals into apoptosis. Interestingly, diabetic p66$^{Shc-/-}$ mice are protected against hyperglycemia-induced endothelial dysfunction and oxidative damage.

Figure 1  Mechanisms of hyperglycemia-induced vascular damage. High intracellular glucose concentrations lead to PKC activation and subsequent ROS production by NADPH oxidase and p66$^{Shc}$ adaptor protein. Increased oxidative stress rapidly inactivates NO leading to formation of the pro-oxidant ONOO$^-$ responsible for protein nitrosylation. Reduced NO availability is also due to PKC-dependent eNOS deregulation. Indeed, PKC triggers enzyme up-regulation thus enhancing eNOS uncoupling and leading to a further accumulation of free radicals. On the other hand, hyperglycemia reduces eNOS activity blunting activatory phosphorylation at Ser1177. Together with the lack of NO, glucose-induced PKC activation causes increased synthesis of ET-1 favouring vasoconstriction and platelet aggregation. Accumulation of superoxide anion also triggers up-regulation of pro-inflammatory genes MCP-1, VCAM-1, and ICAM-1 via activation of NF-$\kappa$B signalling. These events lead to monocyte adhesion, rolling, and diapedesis with formation of foam cells in the sub-endothelial layer. Foam cell-derived inflammatory cytokines maintain vascular inflammation as well as proliferation of smooth muscle cells, accelerating the atherosclerotic process. Endothelial dysfunction in diabetes also derives from increased synthesis of TXA$_2$ via up-regulation of COX-2 and inactivation of PGIS by increased nitrosylation. Furthermore, ROS increase the synthesis of glucose metabolite methylglyoxal leading to activation of AGE/RAGE signalling and the pro-oxidant hexosamine and polyol pathway flux. PKC, protein kinase C; eNOS, endothelial nitric oxide synthase; ET1, endothelin 1; ROS, reactive oxygen species; NO, nitric oxide; MCP-1, monocye chemoattractant protein-1; VCAM-1, vascular cell adhesion molecule-1; ICAM-1, intracellular cell adhesion molecule-1; AGE, advanced glycation end product.
Insulin resistance and atherothrombosis

Insulin resistance is a major feature of type 2 diabetes and develops in multiple organs, including skeletal muscle, liver, adipose tissue, and heart. The onset of hyperglycemia and diabetes is often preceded by many years of insulin resistance. Obesity plays a pivotal role in this phenomenon providing an important link between type 2 diabetes and fat accumulation. Indeed, a substantial proportion of diabetic patients are obese. Obesity is a complex disorder leading to alterations in lipid metabolism, deregulation of hormonal axes, oxidative stress, systemic inflammation, and ectopic fat distribution. Adipose tissue is an active source of inflammatory mediators and free fatty acids (FFAs). Accordingly, obese patients with type 2 diabetes display increased plasma levels of inflammatory markers. Free fatty acids bind Toll-like receptor (TLR) activating NF-kB through degradation of the inhibitory complex IκBα by IκKβ-kinase. As a result, NF-kB triggers tissue inflammation due to up-regulation of inflammatory genes IL-6 and TNF-α.

Toll-like receptor activation by FFA leads to phosphorylation of insulin receptor substrate-1 (IRS-1) by c-Jun amino-terminal kinase (JNK) and PKC, thereby altering its ability to activate downstream targets PI3-kinase and Akt. These molecular events result in the down-regulation of the glucose transporter GLUT-4 and, hence, insulin resistance. Insulin resistance is critically involved in vascular dysfunction in subjects with type 2 diabetes. Indeed, down-regulation of PI3-kinase/Akt pathway leads to eNOS inhibition and decreased NO production. Together with reduced NO synthesis, intracellular oxidation of stored FFA generates ROS leading to vascular inflammation, AGEs synthesis, reduced PGi2 synthase activity, and PKC activation.

Increased ROS levels associated with insulin resistance scavenge NO production and produce peroxynitrite, with a further reduction of NO bioavailability. Reduced cellular levels of NO facilitate pro-inflammatory pathways triggered by increased cytokine production. Indeed, TNF-α and IL-1 increase NF-kB activity and expression of adhesion molecules. TNF-α also stimulates the expression of C-reactive protein which down-regulates eNOS and increases the production of adhesion molecules and endothelin-1. A recent study clearly demonstrated that loss of insulin signalling in the vascular endothelium leads to endothelial dysfunction, expression of adhesion molecules, and atherosclerotic lesions in mice. Although insulin resistance development has been attributed to adipocyte-derived inflammation, recent evidence is overturning the adipocentric paradigm. Indeed, inflammation and macrophage activation seem to primarily occur in non-adipose tissue in obesity. This concept is supported by the notion that suppression of inflammation in the vasculature prevents insulin resistance in other organs and prolongs lifespan. Consistently, transgenic mice with endothelium-specific overexpression of the inhibitory NF-kB subunit IκBα were protected from the development of insulin resistance. In these mice, obesity-induced macrophage infiltration of adipose tissue and plasma oxidative stress markers were reduced whereas blood flow, muscle mitochondrial content, and locomotor activity were increased, confirming the pivotal role of the transcription factor NFKB in oxidative stress, vascular dysfunction, and inflam-
Insulin resistance as trigger of atherothrombosis. In subjects with obesity or type 2 diabetes the increase in FFA activates TLR leading NF-κB nuclear translocation and subsequent up-regulation of inflammatory genes IL-6 and TNF-α. On the other hand, JNK and protein kinase C phosphorylate insulin receptor substrate-1 (IRS-1), thus blunting its downstream targets PI3-kinase and Akt. This results in down-regulation of glucose transporter GLUT-4 and, hence, insulin resistance. Impaired insulin sensitivity in the vascular endothelium leads to increased FFA oxidation, ROS formation, and subsequent activation of detrimental biochemical pathways such as AGE synthesis, PKC activation, protein glycosylation as well as down-regulation of PGI2. These events blunt eNOS activity thereby leading to endothelial dysfunction. Lack of insulin signalling in platelets impairs the IRS1/PI3K pathway resulting in Ca²⁺ accumulation and increased platelet aggregation. FFA, free fatty acids; TLR, toll-like receptor; JNK, c-Jun amino-terminal kinase; IRS-1, Insulin receptor substrate-1; NO, nitric oxide; eNOS, endothelial nitric oxide synthase; IL-6, interleukin-6; TNF-α, tumor necrosis factor.
The atherogenic effects of insulin resistance are also due to changes in lipid profile such as high triglycerides, low HDL cholesterol, increased remnant lipoproteins, elevated apolipoprotein B (ApoB) as well as small and dense LDL. Once circulating FFA reach the liver, very low density lipoprotein (VLDL) are assembled and made soluble by increased synthesis of ApoB. VLDL are processed by cholesteryl ester transfer protein allowing transfer of triglycerides to LDL, which become small and dense and, hence, more atherogenic. Atherogenic dyslipidemia is a reliable predictor of cardiovascular risk and its pharmacological modulation reduces vascular events in subjects with type 2 diabetes and metabolic syndrome.

Coronary events in patients with insulin resistance are triggered by virtue of a prothrombotic state. Under physiological conditions, insulin inhibits platelet aggregation and thrombosis via tissue factor (TF) inhibition and enhanced fibrinolytic action due to modulation of plasminogen activator inhibitor-1 (PAI-1) levels. Indeed, patients with acute myocardial infarction receiving fibrinolitic therapy plus 48 h insulin infusion displayed a marked decrease in PAI-1 levels. In contrast, insulin resistance facilitates atherothrombosis through increased cellular synthesis of PAI-1 and fibrinogen and reduced production of tissue plasminogen activator. In platelets, lack of insulin leads to a down-regulation of the IRS-1/Akt pathway resulting in calcium accumulation upon basal conditions. This latter mechanism may explain why platelets from diabetic patients show faster response and increased aggregation compared with those from healthy subjects. Moreover, platelet reactivity and excretion of tromboxane metabolites are increased in obese patients with insulin resistance and this phenomenon is reversed by weight loss or 3-week treatment with pioglitazone. Body weight as well as impaired insulin sensitivity may also account for the faster recovery of cyclooxygenase activity despite aspirin treatment. Indeed, higher body mass index was an independent predictor of inadequate suppression of tromboxane biosynthesis in non-diabetics subjects treated with aspirin. In this study, the increase of aspirin dosage was sufficient to warrant platelet inhibition. This clinical observation may explain the residual cardiovascular risk in obese patients treated with anti-platelet medications.

Hyperglycemia and insulin resistance alone may not explain the persistent cardiovascular risk burden associated with type 2 diabetes. Indeed, normalization of glycemia does not reduce macrovascular events suggesting that mediators of vascular risk other than glucose significantly participate to increase the residual cardiovascular risk in diabetic patients. In this regard, adipose tissue dysfunction, inflammation, and aberrant adipokine release may be particularly relevant. In patients with abdominal obesity, an increased lipid storage leads to hypoxia, chronic inflammation together with changes in the cellular components of adipose tissue, leading to an altered secretory profile. Adipokines linked to vascular disease are leptin, adipocyte fatty acid-binding protein, interleukins, and novel ones like lipocalin-2 and pigment epithelium-derived factor. These molecules may drive vascular dysfunction via increased proliferation/migration of smooth muscle cells, eNOS inhibition, and activation of NFkB signaling with subsequent expression of adhesion molecules and atherosclerosis. Future work will need to address the potential role of these molecules as biomarkers and/or drug targets.

**MicroRNA and diabetic vascular disease**

MicroRNAs (miRs) are a newly identified class of small non-coding RNAs emerging as key players in the pathogenesis of hyperglycemia-induced vascular damage. These small non-coding RNAs orchestrate different aspects of diabetic vascular disease by regulating gene expression at the post-transcriptional level. Microarray studies have shown an altered profile of miRs expression in subjects with type 2 diabetes. Indeed, diabetic patients display a significant deregulation of miRs involved in angiogenesis, vascular repair, and endothelial homeostasis. Over the last few years, different studies have explored the mechanisms whereby deregulation of miRs expression may contribute to vascular disease in subjects with diabetes. In endothelial cells exposed to high glucose miR-320 is highly expressed and targets several angiogenic factors and their receptors, including vascular endothelial growth factor and insulin-like growth factor-1 (IGF-1). Elevated levels of this miR are associated with decreased cell proliferation and migration, while its down-regulation restores these properties and increases IGF-1 expression, promoting angiogenesis and vascular repair (Figure 3).

Hyperglycemia also increases the expression of miR-221, a regulator of angiogenesis targeting c-kit receptor which is responsible for migration and homing of endothelial progenitor cells (EPCs). miR-221 and 222 were also found to mediate AGE-induced vascular damage. Indeed, down-regulation of miR-222 both in human endothelial cells exposed to high glucose and in diabetic mice elicits AGE-related endothelial dysfunction via targeting, cyclin-dependent kinase proteins involved in cell cycle inhibition (P27KIP1 and P57KIP2). A recent study demonstrated that miR-503 is critically involved in hyperglycemia-induced endothelial dysfunction in diabetic mice and is up-regulated in ischaemic limb muscles of diabetic subjects. The detrimental effects of miR-503 in the setting of diabetes have been explained by its interaction with CCNE and cdc25A, critical regulators of cell cycle progression affecting endothelial cell migration and proliferation. Interestingly, miR-503 inhibition was able to normalize post-ischaemic neovascularization and blood flow recovery in diabetic mice. These findings provide the rationale to foresee a protective effect of the modulation of miR-503 expression against diabetic vascular complications.

Plasma miR profiling showed a profound down-regulation of miR-126 in a cohort of diabetic patients. Recent evidence suggest that reduced miR-126 expression levels are partially responsible for impaired vascular repair capacities in diabetes. miR-126 expression was reduced in EPCs isolated from diabetics and transfected with anti-miR-126 blunted EPCs proliferation and migration. In contrast, restored expression of this miR promoted...
EPCs-related repair capacities and inhibited apoptosis. miR-126 role in EPCs function is mediated by Spred-1, an inhibitor of Ras/ERK signalling pathway, a critical regulator of cell cycle.

Collectively, these studies support the notion that miRs drive complex signalling networks by targeting the expression of genes involved in cell differentiation, migration, and survival.

**Thrombosis and coagulation**

Individuals affected by diabetes display an increased risk of coronary events and cardiovascular mortality when compared with non-diabetic subjects. This phenomenon is largely explained by a deregulation of factors involved in coagulation and platelet activation. Both insulin resistance and hyperglycemia participate to the pathogenesis of this prothrombotic state. Insulin resistance increases PAI-1 and fibrinogen and reduces tissue plasminogen activator levels. The largest increase in PAI-1 has been reported in diabetic patients with poor glycemic control and treatment with glucose-lowering agents glipizide or metformin comparably decreased PAI-1. Hyperinsulinemia induces TF expression in monocytes of patients with type 2 diabetes leading to increased TF procoagulant activity and thrombin generation. These events are enhanced by hyperglycemia (Figure 4). Low-grade inflammation induces TF expression also in the vascular endothelium of diabetic subjects contributing to atherosclerosis. 

Microparticles (MPs), vesicles released in the circulation from various cell types following activation or apoptosis, are increased in diabetic patients and predict cardiovascular outcome. Microparticles from patients with type 2 diabetes have shown to increase coagulation activity in endothelial cells (Figure 4). Moreover, MPs carrying TF promote thrombus formation at sites of injury representing a novel and additional mechanisms of coronary thrombosis in diabetes. Among the factors contributing to the diabetic prothrombotic state, platelet hyperreactivity is of major relevance. A number of mechanisms contribute to platelet dysfunction affecting adhesion, activation as well as aggregation phases of platelet-mediated thrombosis (Figure 4). Hyperglycemia alters platelet Ca²⁺ homeostasis leading to cytoskeleton abnormalities and increased secretion of proaggregant factors. Moreover, up-regulation of glycoproteins Ib and IIb/IIIa in diabetic patients triggers thrombus via interacting with Von Willebrand factor (vWF) and fibrin molecules (Figure 4).
Vascular hyperglycemic memory

Recent prospective clinical trials have shown that normalization of glycemia failed to reduce cardiovascular burden in the diabetic population. In these trials, intensive glucose-lowering therapy was started after a median duration of diabetes ranging from 8 to 11 years. In contrast, early treatment of hyperglycemia was shown to be beneficial. These findings support the concept that hyperglycemic environment may be remembered in the vasculature. Reactive oxygen species are probably involved in this phenomenon.

The persistence of hyperglycemic stress despite blood glucose normalization has recently been defined ‘hyperglycemic memory’. A substantial understanding of its mechanisms has been achieved only in recent years. It has been recently demonstrated that transient hyperglycemia activates NF-kB, and this effect persists despite subsequent normalization of glucose levels. This finding is explained by epigenetic changes occurring at the level of DNA and histone-binding promoter of pro-oxidant and pro-inflammatory genes. Specifically, methylation and acetylation are critical epigenetic mark modulated by the hyperglycemic environment. Methylation of p65/NFkB promoter by the ROS-dependent methyltransferase Set7/9 is indeed the mechanisms whereby vascular inflammation is not reverted by restoration of normoglycemia. We have recently identified the source of ROS perpetuating vascular dysfunction despite normoglycemia restoration.

In diabetic mice and human endothelial cells, glucose normalization did not revert up-regulation of p66Shc protein, a mitochondrial adaptor critically involved in ROS generation. Persistent p66Shc expression is driven by epigenetic changes as reduced promoter methylation and acetylation of histone 3. Moreover, p66Shc-dependent ROS generation maintains up-regulation of PKCβII and inhibits eNOS activity, thus feeding a detrimental vicious cycle despite restoration of normoglycemia. Persistent oxidative stress is also responsible for sustained vascular apoptosis via caspase 3 activation. Gene silencing of p66Shc blunted persistent endothelial dysfunction and oxidative stress in the vasculature of diabetic mice, suggesting that this protein drives hyperglycemic memory. Interestingly enough, these findings are in line with the notion that both SIRT-1 and p53 control p66Shc transcription. Indeed, reduced SIRT-1 activity in diabetes favours acetylation of histone 3-binding p66Shc promoter. Moreover, increased p53 activity maintains p66Shc memory effect. All together, these pathways might be involved in self-perpetuating vascular damage of patients with diabetes despite optimal glycemic control.

*Figure 4* Coagulation and platelet reactivity in diabetes. In patients with diabetes chronic hyperglycemia and insulin resistance determine a significant alteration in the coagulation factors as well as increased platelet aggregation, leading to a prothrombotic state. Diabetes-induced increase of TF levels activates thrombin converting fibrinogen into fibrin. Fibrin organization is further enhanced due to high PAI-1 and reduced t-PA levels. Increased Ca²⁺ content, thrombin stimulation as well as interaction with vWF via gplb/IIIa receptor lead to platelet shape change, granule release, and aggregation. Release of MPs from injured endothelium and circulating platelets contribute to accelerate thrombus development. Endothelial dysfunction precipitates rupture of the endothelial layer leading to exposure of collagen and vWF thereby activating platelets and favouring vascular thrombosis. TF, tissue factor; t-PA, tissue plasminogen activator; PAI-1, plasminogen activator inhibitor-1; MPs, microparticles; vWF, von Willebrand factor; ECs, endothelial cells.
Oxidative stress plays a major role in the development of micro- and macrovascular complications. Accumulation of free radicals in the vasculature of diabetic patients is responsible for the activation of detrimental biochemical pathways, miRs deregulation, release of MPs, and epigenetic changes contributing to vascular inflammation and ROS generation. Since cardiovascular risk burden is not eradicated by intensive glycemic control associated with optimal multifactorial treatment, mechanism-based therapeutic strategies are in high demand. Specifically, inhibition of key enzymes involved in hyperglycemia-induced vascular damage or activation of pathways improving insulin sensitivity may represent promising approaches. Modulation of specific miRs might contribute to improve EPC-driven vascular repair. Moreover, the progressive identification of a complex scenario driven by epigenetic changes that modulate transcription of ROS-generating and pro-inflammatory genes may represent an attractive opportunity to dampen oxidative stress, vascular inflammation, and hence to prevent cardiovascular complications in patients with diabetes.

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**References**
studies of 95,783 individuals followed for 12.4 years. Diabetes Care 1999;22:
233–240.
low should we go? Re-discovering personalized care. Eur Heart J 2011;32:
2247–2255.
9. Creager MA, Luscher TF, Cosentino F. Beckman JA. Diabetes and vascular disease:
Pathophysiology, clinical consequences, and medical therapy. Part I. Circulation
11. Lerman A, Zeher AM. Endothelial function: cardiac events. Circulation 2005;111:
363–368.
MC, Beebe D, Oates PJ, Hammers HP, Giudino I, Brownlee M. Normalizing mitochondrial
superoxide production blocks three pathways of hyperglycemia damage. Nature
107:1058–1070.
14. Geraldes P, King GL. Activation of protein kinase C isoforms and its impact on dia-
glucose enhances apoptosis related to oxidative stress in human umbilical vein
endothelial cells: the role of protein kinase C and NAD(P)H-oxidation activation.
18. Paneni F, Mochola P, Akhmedov A, Costantino S, Osto E, Volpe M, Luscher TF, Cosentino F. Gene silencing of the mitochondrial adaptor p66shc suppresses vas-
gene expression in human peripheral blood mononuclear cells: relationship to oxy-
23. Shi Y, Cosentino F, Camici GG, Akhmedov A, Vannouette PM, Tanner FC, Luscher TF. Oxidized low-density lipoprotein activates p66shc via lectin-like oxi-
24. Shi Y, Cosentino F, Camici GG, Akhmedov A, Vannouette PM, Tanner FC, Luscher TF. Oxidized low-density lipoprotein activates p66shc via lectin-like oxi-
25. Shi Y, Cosentino F, Camici GG, Akhmedov A, Vannouette PM, Tanner FC, Luscher TF. Oxidized low-density lipoprotein activates p66shc via lectin-like oxi-
26. Shi Y, Cosentino F, Camici GG, Akhmedov A, Vannouette PM, Tanner FC, Luscher TF. Oxidized low-density lipoprotein activates p66shc via lectin-like oxi-
27. Shi Y, Cosentino F, Camici GG, Akhmedov A, Vannouette PM, Tanner FC, Luscher TF. Oxidized low-density lipoprotein activates p66shc via lectin-like oxi-
28. Shi Y, Cosentino F, Camici GG, Akhmedov A, Vannouette PM, Tanner FC, Luscher TF. Oxidized low-density lipoprotein activates p66shc via lectin-like oxi-


