RESEARCH PAPER

Cellulose and lignin biosynthesis is altered by ozone in wood of hybrid poplar (*Populus tremula* × *alba*)

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Abstract

Wood formation in trees is a dynamic process that is strongly affected by environmental factors. However, the impact of ozone on wood is poorly documented. The objective of this study was to assess the effects of ozone on wood formation by focusing on the two major wood components, cellulose and lignin, and analysing any anatomical modifications. Young hybrid poplars (*Populus tremula* × *alba*) were cultivated under different ozone concentrations (50, 100, 200, and 300 nl l⁻¹). As upright poplars usually develop tension wood in a non-set pattern, the trees were bent in order to induce tension wood formation on the upper side of the stem and normal or opposite wood on the lower side. Biosynthesis of cellulose and lignin (enzymes and RNA levels), together with cambial growth, decreased in response to ozone exposure. The cellulose to lignin ratio was reduced, suggesting that cellulose biosynthesis was more affected than that of lignin. Tension wood was generally more altered than opposite wood, especially at the anatomical level. Tension wood may be more susceptible to reduced carbon allocation to the stems under ozone exposure. These results suggested a coordinated regulation of cellulose and lignin deposition to sustain mechanical strength under ozone. The modifications of the cellulose to lignin ratio and wood anatomy could allow the tree to maintain radial growth while minimizing carbon cost.

Key words: Cellulose, lignin, ozone, poplar, tension wood.

Introduction

Wood is of primary importance for various industrial purposes such as paper manufacturing and construction (Plomion *et al.*, 2001), but also as a renewable source of energy for biofuels (Carroll and Somerville, 2009). The two main components of wood are cellulose and lignin. Their deposition in the cell wall occurs in a regulated manner during wood formation, which includes cambial cell division, cell expansion, secondary wall formation, and cell death (Hertzberg *et al.*, 2001; Mellerowicz *et al.*, 2001).

Cellulose comprises 40–50% of wood dry matter and, being the main component of the cell wall, constitutes a strong carbon sink within the plant (Delmer and Haigler, 2002). Cellulose is a linear polymer composed of 500–14 000 (1 → 4)-linked β-D-glucose residues (Somerville, 2006). In plant cell walls, the glucan chains are linked by hydrogen bonds to form insoluble cellulose microfibrils. Cellulose is synthesized at the plasma membrane by 36 cellulose synthase (CesA) subunits assembled in a rosette complex (Doblin *et al.*, 2002; Somerville, 2006; Joshi and Mansfield, 2007). The precursor of the β-1,4-glucan chain is uridine diphosphoglucose (UDPG) which results from the cleavage of sucrose by sucrose synthase (SuSy) or is derived from glucose-1-P via UDPG pyrophosphorylase (UGPase) (Delmer and Haigler, 2002). CesA proteins are encoded by
a gene superfamily. Characterization of cellulose-deficient mutants and genome sequencing revealed 10 CesA genes in *Populus trichocarpa* (Taylor, 2008). Eighteen CesA genes have been identified in *Populus trichocarpa*. Seven of these genes are specific to or are highly expressed in xylem tissue (Suzuki et al., 2006).

Lignin is the second most abundant component of wood after cellulose and accounts for 15–35% of wood dry matter. Lignin provides hydrophobicity and structural support that allows water transport in the vascular system. After the start of secondary wall formation, lignification begins in the middle lamella and primary wall and then continues in the secondary wall (Donaldson, 2001). Lignin is a phenylpropanoid derivative and heteropolymer of three monolignols: p-coumaryl, coniferyl, and sinapyl alcohols. Over the past two decades, the monolignol biosynthetic pathway has been redrawn several times (Humphreys and Chapple, 2002; Boudet et al., 2004; Davin et al., 2008). Significant information has been obtained by altering the expression of individual genes in the phenylpropanoid and monolignol biosynthetic pathway and studying the consequences on lignin content and composition (Vanholme et al., 2008). The shikimate pathway supplies phenylalanine that is converted to monolignols through a metabolic grid of 10 enzyme families (Humphreys and Chapple, 2002). In *A. thaliana*, 12 candidate genes for vascular lignification were identified (Raes et al., 2003). In poplar, 15–23 genes potentially involved in wood monolignol synthesis have been identified, based on transcript abundance in the xylem (Hamberger et al., 2007; Shi et al., 2010).

Although wood formation is commonly said to be highly influenced by the environment (Mellerowicz and Sundberg, 2008), few reports have dealt with the impact of abiotic stress on wood components and their biosynthesis. Most studies of abiotic stress have addressed variations in wood anatomy. Water-limited trees showed a decrease in vessel or anatomical properties in well-defined wood. Young hybrid poplars were bent and exposed to four different ozone concentrations (50, 100, 200, and 300 nl l⁻¹) in controlled chambers (Kostiainen et al., 2006, 2008). The distribution of TW follows no set pattern in such trees (Isebrands and Bensend, 1972). The production of TW could be related to internal axial stresses in fast-growing species (Isebrands and Bensend, 1972; Badia et al., 2006). The distribution of TW follows no set pattern in such trees (Isebrands and Bensend, 1972). The production of TW could be related to internal axial stresses in fast-growing species (Isebrands and Bensend, 1972; Badia et al., 2006) and high sensitivity to the gravity stimulus (Jourez et al., 2001; Jourez and Avella-Shaw, 2003). In fact, TW typically occurs in response to the gravity stimulus in bent trees and develops on the opposite side of stems in order to restore the verticality of their axis (Pilate et al., 2004; Mellerowicz and Sundberg, 2008). At an anatomical level, TW differs from normal wood or the opposite wood (OW) formed on the lower side of bent stems. In TW of many species, including poplar, the fibres develop a specialized cell wall layer known as the gelatinous layer (G-layer). Most of the G-layer (95%) consists of crystalline cellulose which therefore results in a higher cellulose content and lower proportion of lignin in TW compared with OW or normal wood.

The effect of ozone was therefore investigated in both kinds of wood (TW and OW) encountered in poplar. Moreover, the trees were bent so as to induce the formation of TW in the upper part of the stem and OW in the lower part, and thereby to analyse biochemical, chemical, and anatomical properties in well-defined wood. Young hybrid poplars were bent and exposed to four different ozone concentrations (50, 100, 200, and 300 nl l⁻¹) in controlled...
Materials and methods

Plant material and growth conditions

Micropropagated hybrid poplar plants (Populus tremula×alba, clone INRA 717-1-B4) were transplanted into 5 l pots containing compost fertilized with 20 g of slow-release 13:13:13 N:P:K (Nutricot T 100, Fertil, Boulogne-Billancourt, France). The plants were cultivated in controlled chambers at 75/85% relative humidity (day/night) with a 14-h light period (Sun T Agro, Philips, Eindhoven, The Netherlands; intensity: 250–300 μmol m⁻² s⁻¹) and 22/18 °C day/night temperatures. The young trees were artificially tilted at 42° from the vertical using a rigid stick in order to stimulate TW formation on the upper side of the stems. The development of TW was checked by cutting fresh transverse sections at different levels of the stem by hand and staining them with safranin O/Astra blue according to Vazquez-Cooz and Meyer (2002).

Ozone treatment

Plants developing six fully expanded leaves were subjected to ozone treatment in phytotrons used for plant acclimation. The young trees were exposed either to charcoal-filtered air (control) or ozone treatment. Plants developing six fully expanded leaves were subjected to ozone treatment.

Biomass and growth measurements

Leaves and stems were collected at the end of the experiment (46 d) and were dried at 60 °C for 3 weeks before biomass determination. Tree height and diameter were measured throughout the fumigation. Radial growth was measured 10 cm above the collar with a Vernier caliper. Analyses were done on six plants per treatment.

Preparation of enzyme extracts

Stems were harvested in the middle of the day. After removal of the bark, TW was sampled from the upper quarter of the section of the trees and OW from the lower quarter. Samples were then frozen in liquid nitrogen and extracts were obtained from the powders as described by Cabane et al. (2004). The resulting desalted extracts were used for enzyme assays.

Enzyme activities

The activities of enzymes involved in cellulose biosynthesis, namely SuSy (EC 2.4.1.13) and UGPase (EC 2.7.7.9), were determined with a Beckman DU 640 spectrophotometer (Beckman Coulter, Roissy, France). SuSy activity was based on the reduction of NAD⁺ at 340 nm with an enzyme coupling reaction (Hauch and Magel, 1998). UGPase activity was assayed at 340 nm by following the reduction of NAD⁺ with an enzyme coupling reaction (Ciereszko et al., 2001).

The activities of enzymes connected to lignin biosynthesis were measured as follows. Shikimate dehydrogenase (SHDH; EC 1.1.1.25) and cinnamyl alcohol dehydrogenase (CAD; EC 1.1.1.195) activities were determined at 30 °C with a microplate reader (Elx 808 iu BIO-TEK INSTRUMENTS). SHDH activity was determined by following the reduction of NAD⁺ at 340 nm (Fiedler and Schulz, 1985). CAD activity was monitored at 460 nm as described by O’Malley et al. (1992). Phenylalanine ammonia-lyase (PAL; EC 4.3.1.5) activity was assayed by measuring the release of cinnamate at 290 nm (Havir, 1987) with a Beckman DU 640 spectrophotometer (Beckman Coulter).

The protein content of enzyme extracts was determined with Bio-Rad Bradford protein reagent dye using bovine serum albumin as standard. The enzyme activities are reported as the mean value from three trees per treatment.

RNA extraction and cDNA synthesis

Tissues (~100 mg) were placed in teflon jars chilled with liquid nitrogen and ground to a fine powder for 2 min using a mixer mill MM301 (Retsch, France). Total RNA was isolated with TRIzol (Invitrogen), according to the manufacturer’s instructions. Any contaminating genomic DNA was removed by treating the RNA samples with DNase I, Amp Grade, (Invitrogen), then cleaning with RNeasy MinElute CleanUp columns (Qiagen). The absence of genomic DNA was checked by testing RNA samples (standard PCR) for amplification of sequences encoding ubiquitin-conjugated enzyme E2-17 kDa 10/12 (UBC10) (Sterky et al., 2004) with the following primers: UBC146A (5'-CCCCGCTTAAACATCTCA-3') and UBC146B (5'-GGGTCCAGCTTTTGCAGTC-3'). A 1 μg aliquot of total RNA was reverse transcribed using the iScript cDNA Synthesis Kit (Biorad) to generate cDNA.

Quantitative real-time PCR

Quantitative real-time PCRs were carried out using iQ SYBR Green Supermix (Biorad) in a MyQ Single-Color Real-Time PCR Detection System ICycler (Bio-Rad). The real-time PCR conditions were as follows: denaturation by hot start at 95 °C for 3 min, followed by 40 cycles of a two-step program consisting of denaturation at 95 °C for 15 s and annealing/extension at 60 °C for 45 s. The transcript abundance of genes involved in cellulose and lignin synthesis were quantified, namely CesA, PtCesA4, PtCesA5, PtCesA7, PtCesA8, PtCesA13, PtCesA17, and PtCesA18 [primers according to Suzuki et al. (2006)]; SuSy, PtSUS1 (JGI Protein ID:835735), PtSUS2 (5'-TTGTGAGGGAGTTGCGTGTGTTG-3'), PtSUS3 (5'-TTGGCGAGGGAAAGATGGC-3'), and PtSUS2 (JGI Protein ID:826368), PtSUS2F (5'-TTGATTCCTGTGTTGACAG-3'), PtSUS2R (5'-GGCTTGTGCGGCTTTTAGG-3'), UGPase, UGP1 and UGP2 [primers according to Meng et al. (2007)]; PAL; PAL1; cinnamoyl CoA reductase, CCR2; and cinnamyl alcohol dehydrogenase, CAD1 [primers according to Shi et al. (2010)]. The genes selected for transcript analysis were based on previously reported high levels or specific expression in poplar xylem (Li et al., 2005; Suzuki et al., 2006; Barakat et al., 2009).

Four genes were used as endogenous controls to normalize transcript quantity: 18S rRNA [primers according to Meng et al. (2007)]; polyaubiquitin (UBQ11), actin (ACT2) [primers according to Brunet et al. (2004)]; and UBC10 (primers described above in the cDNA synthesis section). Critical thresholds (Ct) for all genes were quantified in triplicate and normalized with GeNorm software (Vandesompele et al., 2002). Transcript relative abundance was calculated as the mean of three biological replicates (three trees per condition) and three analytical replicates.

Wood compounds

Cellulose content was determined on lyophilized wood samples. After grinding, the powders (10 mg) were subjected to acetic acid/nitric acid digestion as described by Updegraff (1969). The acid-insoluble material (cellulose) was recovered by centrifuging at 2200 g for 20 min.

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4000 g for 15 min. The dried pellets were resuspended in 67% sulphuric acid. Cellulose content was determined on a glucose basis by colorimetry using the phenol-sulphuric acid method (Dubois et al., 1956).

Lignin analyses were carried out on dry extract-free samples. These extract-free samples were prepared by subjecting the dried ground wood to exhaustive solvent extraction in a Soxhlet apparatus (toluene:ethanol, 1:1, then ethanol, and finally water). The lignin content was determined by the Klason method from 300 mg of sample according to the standard procedure (Dence, 1992) and was calculated as the weight percentage of the cell wall residue. All the Klason analyses were run in duplicate and with three trees per sample type. In addition, the acid-soluble lignin was determined according to the standard procedure and showed no differences between treatments. Thioacidolysis was performed as previously reported (Lapierre et al., 1999). The lignin-derived monomers were determined by gas chromatography–mass spectrometry (GC-MS) of their silylated derivatives (Lapierre et al., 1999).

Lignin and cellulose contents are reported as the mean values obtained for each of three individuals analysed trees.

Wood density
Wood samples (2 cm long) were saturated in distilled water. TW and OW were separated. The maximum volume of the wood fragment was determined from the apparent increase in mass using an electronic balance. Samples were then placed at 103 °C for 24 h to obtain the minimum dry mass. The density corresponded to the ratio of the minimum dry mass versus the maximum water-saturated volume. Analyses were performed on three different trees per condition. Two measurements were done on each tree.

Wood anatomy
Wood samples were dried in ambient air for 1 week. Transverse sections were prepared with a sliding microtome. Air-dried samples were analysed by electronic scanning electron microscopy (ESEM; FEI Quanta 200). Images were taken under a pressure of 1 Torr. Three poplars from each treatment were collected for wood anatomy analyses. For each tree, six random scanning zones were defined in the wood formed during the treatment: three in TW and three in OW. Vessel number, fibre number, and vessel lumen diameter were analysed on each scanning image using different types of software (Image J and Scion Image).

Cambium activity was analysed by saturating the stems with water under a vacuum, soaking progressively in polyethyleneglycol 1500, then obtaining transverse sections with a sliding microtome. The sections were stained with safranin O/Astra blue (Vazquez-Coaz and Meyer, 2002) and observed under a light microscope. The number of cambium cell layers was recorded on three trees per treatment.

Statistical analysis
The significance of ozone effects compared with controls was assessed in a two-way analysis of variance (ANOVA) followed by Tukey’s test.

Results
Young hybrid poplars (Populus tremuloides × alba, clone INRA 717-1-B4) growing upright (staked or not) in controlled chambers developed TW without any defined pattern (Supplementary Fig. S1 at JXB online). The young trees were therefore bent to limit the development of TW to the upper side of the stem (Supplementary Fig. S1) and were grown in charcoal-filtered air (control) or were fumigated with ozone (50, 100, 200, or 300 nl l⁻¹) in phytotronic chambers.

Biomass and plant development
Foliar and stem biomass were severely affected during the 46 d of ozone fumigation (Table 1). Biomass loss was significant at 100, 200, and 300 nl l⁻¹, with a reduction of up to 37% for leaves and 48% for stems. Leaf fall was enhanced at the same time as the reduction of foliar biomass. Leaf fall in hybrid poplars subjected to 300 nl l⁻¹ ozone was 2-fold higher than in control plants.

Tree height and radial growth were also reduced by ozone fumigation (Fig. 1). Height growth was decreased by 38% in plants subjected to 200 nl l⁻¹ ozone compared with the controls. The reduction of height growth was delayed at 300 nl l⁻¹ and corresponded to a growth break (visual observation of apical bud). A significant decrease in radial growth was also observed in plants exposed to ozone. This decline in radial growth attained 50% at 300 nl l⁻¹.

Cellulose biosynthesis
Two stem levels corresponding to two different developmental stages were sampled. The lower stem (LS) was located 10 cm above the collar and corresponded to wood that developed before and during ozone fumigation. The middle stem (MS) developed just after the beginning of ozone fumigation. TW from the upper side and OW from the lower side of the stem were collected from each level, and separated. Under control conditions, SuSy and UGPase activities were higher in TW than in OW at both LS and MS levels (Fig. 2). The transcript levels of SuSy genes (SUS1 and SUS2) were higher in TW than in OW (Table 2). The RNA levels for one UGPase gene, UGP1, were similar between TW and OW, whereas the transcript abundance of UGP2 was twice as high in TW than in OW. Among the 18 CesA genes identified in P. trichocarpa (Suzuki et al., 2006), seven genes specific to or highly expressed in xylem were selected for this study (PtCesA4, PtCesA5, PtCesA7, PtCesA8, PtCesA13, PtCesA17, and PtCesA18). The CesA5 and CesA13 genes are reported to be involved in cellulose

Table 1. Dry matter and percentage of fallen leaves of hybrid poplars cultivated for 46 d under different ozone treatments

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Dry weight (g)</th>
<th>Fallen leaves (% total leaves)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Leaves</td>
<td>Stem</td>
</tr>
<tr>
<td>Control</td>
<td>23.52±2.5 (100)</td>
<td>22.61±2.8 (100)</td>
</tr>
<tr>
<td>Ozone, 50 nl l⁻¹</td>
<td>19.50±4.1 (63)</td>
<td>15.60±6.1 (69)</td>
</tr>
<tr>
<td>Ozone, 100 nl l⁻¹</td>
<td>15.13±2.5° (64)</td>
<td>12.22±2.2° (64)</td>
</tr>
<tr>
<td>Ozone, 200 nl l⁻¹</td>
<td>15.10±0.9° (64)</td>
<td>11.67±1.6° (62)</td>
</tr>
<tr>
<td>Ozone, 300 nl l⁻¹</td>
<td>14.86±1.0° (63)</td>
<td>13.03±1.4° (68)</td>
</tr>
</tbody>
</table>

Data are mean values ± SD (n=6). The normalized values relative to the control are shown in parentheses.

* Significant differences (P < 0.05) between control and ozone treatments.
between control and ozone treatments. Synthesis in the primary cell wall, whereas the CesA4, CesA7, CesA8, CesA17, and CesA18 genes are believed to be involved in cellulose production in the secondary cell wall. The transcript abundance of CesA7 and CesA18 genes was higher in TW than in OW, but the RNA levels of the other CesA genes were unchanged between TW and OW.

Under ozone conditions, the SuSy and UGPase activities in TW were decreased in both LS and MS (Fig. 2). The transcript levels of the SUS1 and SUS2 genes were reduced in both TW and OW tissues (Table 2). The transcript levels of UGP1 and UGP2 were reduced in TW. However, the UGP1 transcript seemed to remain unchanged in OW, whereas the expression of UGP2 was stimulated. All seven CesA genes, with the exception of CesA13, were affected by ozone in both TW and OW. A substantial decrease in transcript level was observed for CesA7, CesA8, CesA17, and CesA18.

**Lignin biosynthesis**

The activity of CAD, which catalyses the reduction of p-hydroxycinnamaldehydes to p-hydroxycinnamyl alcohols, was measured. At the same time the activity of two enzymes involved in earlier steps of phenolic metabolism were monitored. These were SHDH, an enzyme of the shikimate pathway associated with phenylalanine synthesis, and PAL, the first enzyme of the phenylpropanoid pathway. Transcript levels were analysed by real-time quantitative PCR for PAL, CAD, and for cinnamoyl CoA reductase (CCR), the enzyme that provides substrates to CAD.

The enzymes involved in lignin metabolism showed similar activities in TW and OW under control conditions (Fig. 3) and similar CAD1 and CCR2 expression levels (Table 2). The PAL1 transcript level was higher in OW than in TW.

**Table 2.** Transcript abundance of cellulose and lignin biosynthetic genes in hybrid poplar wood (LS) cultivated for 46 d under control or ozone (300 nl l⁻¹) conditions

<table>
<thead>
<tr>
<th>Gene</th>
<th>Tension wood</th>
<th>Opposite wood</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Control</td>
<td>Ozone, 300 nl l⁻¹</td>
</tr>
<tr>
<td>SUS1</td>
<td>1.50±0.35</td>
<td>0.41±0.14</td>
</tr>
<tr>
<td>SUS2</td>
<td>1.64±0.22</td>
<td>0.47±0.16</td>
</tr>
<tr>
<td>UGP1</td>
<td>1.22±0.16</td>
<td>0.48±0.27</td>
</tr>
<tr>
<td>UGP2</td>
<td>2.28±0.71</td>
<td>1.59±0.42</td>
</tr>
<tr>
<td>CesA4</td>
<td>0.89±0.18</td>
<td>0.56±0.19</td>
</tr>
<tr>
<td>CesA5</td>
<td>0.81±0.03</td>
<td>0.60±0.16</td>
</tr>
<tr>
<td>CesA7</td>
<td>1.59±0.17</td>
<td>0.61±0.27</td>
</tr>
<tr>
<td>CesA8</td>
<td>0.95±0.26</td>
<td>0.31±0.13</td>
</tr>
<tr>
<td>CesA13</td>
<td>0.76±0.13</td>
<td>0.82±0.09</td>
</tr>
<tr>
<td>CesA17</td>
<td>0.85±0.15</td>
<td>0.46±0.16</td>
</tr>
<tr>
<td>CesA18</td>
<td>2.06±0.47</td>
<td>0.82±0.37</td>
</tr>
</tbody>
</table>

Transcript levels were normalized and expressed as fold changes compared with values of opposite wood in control conditions. Data are means ±SD of three biological replicates.
Under ozone, the LS was more affected than the MS. In the TW and OW of LS, the SHDH, PAL, and CAD activities were lower than in the controls. The PAL1, CAD1, and CCR2 transcript levels were also reduced by ozone, except PAL1 in TW. In MS, the PAL and SHDH activities were slightly enhanced at some ozone concentrations while CAD activity was only reduced at the highest ozone concentration.

Cellulose and lignin content

The cellulose content under control conditions and at both stem levels was higher in TW than in OW, whereas the TW Klason lignin content was lower (Fig. 4). Accordingly, the cellulose to lignin ratio was 2-fold higher in TW than in OW. OW composition remained unchanged under ozone, but TW was affected. Cellulose content was reduced by ozone, and Klason lignin was enhanced in LS TW (Fig. 4). Similar results were obtained when the acid-soluble lignin fraction was considered together with Klason lignin (data not shown). Ozone significantly reduced the cellulose to lignin ratio at every ozone concentration in LS and at the highest concentrations (200 nl l⁻¹ and 300 nl l⁻¹) in MS. In contrast, lignin composition was not affected by ozone, as revealed by the S/G thioacidolysis ratios (Supplementary Table S1 at JXB online).

Wood density and anatomy

Wood density was higher in TW than in OW in both MS and LS under each condition (Table 3). Ozone reduced the ratio between TW and OW densities at both levels of the stem, compared with control plants.

Images of wood anatomy were recorded by ESEM. Images were taken in the most external part of LS that was formed during fumigation. The TW images (Fig. 5) showed characteristic fibres with an additional G-layer. Under control conditions, TW contained fewer vessels with smaller lumens than OW. The cambial cell layers (Supplementary Fig. S2 at JXB online) and fibre frequencies were similar in TW and OW (Table 4).

Ozone reduced cambial growth in both TW and OW at 200 nl l⁻¹ (Table 4). Other parameters were not affected by ozone in OW. Wood anatomy was altered by ozone in TW. Vessel lumen diameter increased while vessel frequency decreased. Fibre frequency was enhanced in TW in hybrid poplars fumigated with ozone.
Discussion

Cellulose and lignin biosynthesis in tension wood formation

High proportions of TW develop in upright hybrid poplars cultivated in growth chambers, so the trees in the present experiment were bent so that TW formation would be limited to the upper side of the stem (Supplementary Fig. S1 at JXB online). In fact, many features of TW such as high wood density, low vessel frequency, fibres with a G-layer, and high cellulose content (Tables 2, 4, and Figs 4, 5) have been found in this part as compared with the OW (or normal wood) formed on the lower side of the hybrid poplar stem (Plomion et al., 2001; Pilate et al., 2004; Mellerowicz and Sundberg, 2008).

As described (Pilate et al., 2004), it was found that TW contained less lignin than OW and that their lignin S/G (syringyl/guaiacyl) ratios were slightly higher (Fig. 4, and Supplementary Table S1 at JXB online). However, the activities of different enzymes involved in lignin synthesis (CAD), and in supply pathways, the phenylpropanoid pathway (PAL), and the shikimate pathway (SHDH), were similar in TW and OW (Fig. 3). Additionally, CAD1 and CCR2 expression levels remained unchanged (Table 2). These findings are in accordance with different authors who concluded that TW cell walls contained the same quantity of lignin as OW and that the apparent decrease in lignin content resulted only from the increase of cellulose on a dry matter basis (Bentum et al., 1969; Joseleau et al., 2004; Pilate et al., 2004; Xu et al., 2006).

Table 3. Tension wood to opposite wood density ratio of hybrid poplar stem after 46 d of fumigation with different ozone concentrations

<table>
<thead>
<tr>
<th>Level</th>
<th>Treatment</th>
<th>Wood density (TW/OW)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lower stem</td>
<td>Control</td>
<td>1.209±0.007</td>
</tr>
<tr>
<td></td>
<td>Ozone, 50 nl l⁻¹</td>
<td>1.145±0.013</td>
</tr>
<tr>
<td></td>
<td>Ozone, 100 nl l⁻¹</td>
<td>1.111±0.032*</td>
</tr>
<tr>
<td></td>
<td>Ozone, 200 nl l⁻¹</td>
<td>1.075±0.010*</td>
</tr>
<tr>
<td></td>
<td>Ozone, 300 nl l⁻¹</td>
<td>1.089±0.018*</td>
</tr>
<tr>
<td>Middle stem</td>
<td>Control</td>
<td>1.088±0.032</td>
</tr>
<tr>
<td></td>
<td>Ozone, 50 nl l⁻¹</td>
<td>1.050±0.005</td>
</tr>
<tr>
<td></td>
<td>Ozone, 100 nl l⁻¹</td>
<td>1.033±0.037*</td>
</tr>
<tr>
<td></td>
<td>Ozone, 200 nl l⁻¹</td>
<td>1.039±0.017*</td>
</tr>
<tr>
<td></td>
<td>Ozone, 300 nl l⁻¹</td>
<td>1.037±0.007*</td>
</tr>
</tbody>
</table>

Data are mean values ± SD (n=3, individual replicates). *Significant differences (P<0.05) between control and ozone treatments. TW/OW, tension wood to opposite wood ratio.

Reduction of cellulose and lignin biosynthesis activities under ozone

The present growth data revealed a decrease in biomass production and growth rate of young hybrid poplars.

Fig. 5. Environmental scanning electron microphotographs of the transverse surface of tension wood (A, C) and opposite wood (B, D) in stems (LS) of hybrid poplars cultivated for 46 d under control conditions (A, B) or 200 nl l⁻¹ ozone (C, D). V, vessel; F, fibre; GF, G-layer fibre. Scale bars=200 µm.
exposed to ozone (Table 1 and Fig. 1). A marginal response, which would correspond to high sensitivity or high resistance, was avoided by using four different ozone levels (50, 100, 200, and 300 nl l⁻¹). Most of the responses showed the same trend, whatever the ozone level, and were more pronounced at the highest levels. The reduction of tree growth under ozone is a well-known effect (Wittig et al., 2009) and is generally attributed to a decrease in photosynthesis (Wittig et al., 2007). It was shown here that stems were more affected than leaves and that the reduction in stem diameter was stronger for the highest ozone concentration. Biomass allocation to stem was therefore reduced under ozone treatment since the leaf to stem biomass ratio decreased in poplar stems exposed to ozone (Table 1 and Fig. 1). A marginal response, which would correspond to high sensitivity or high resistance, was avoided by using four different ozone levels (50, 100, 200, and 300 nl l⁻¹). Most of the responses showed the same trend, whatever the ozone level, and were more pronounced at the highest levels. The reduction of tree growth under ozone is a well-known effect (Wittig et al., 2009) and is generally attributed to a decrease in photosynthesis (Wittig et al., 2007). It was shown here that stems were more affected than leaves and that the reduction in stem diameter was stronger for the highest ozone concentration. Biomass allocation to stem was therefore reduced under ozone treatment since the leaf to stem biomass ratio was significantly enhanced. Indeed poplars exposed for 35 d to 200 nl l⁻¹ ozone and labelled for 4 h with ¹³CΟ₂ showed a decrease in carbon allocation to stems. Newly incorporated carbon was reduced by nearly half in lower stems (data not shown). Carbon retention was probably increased in the leaves, resulting in a stronger reduction of carbon allocation to the stems. This greater retention in leaves may be explained by a higher carbon demand for repair and defence in damaged foliage (Sandermann, 2000).

Wood formation is initiated in the vascular cambium. In the present experiment, the decrease in radial growth under ozone probably resulted from reduced cambial activity (Table 4, and Supplementary Fig. S2 at JXB online), as previously suggested by Matyssek et al. (2002) and as observed in reaction to many other stresses (Savidge, 2001). Cambial derivatives develop into xylem cells through division, expansion, secondary wall formation, lignification, and finally cell death. This study is, to our knowledge, the first which combines analyses of the composition and metabolism (enzyme activities and RNA levels) of the two main cell wall components, cellulose and lignin, in relation to TW and OW formation under ozone fumigation. The results show that enzyme activities involved in cellulose and lignin metabolism decreased in poplar stems exposed to ozone (Figs 2, 3). Enzyme activities and transcript levels were in good agreement (Table 2) and support the hypothesis of a transcriptional control of cellulose and lignin metabolism in such stems. Moreover, these results suggest a coordinated regulation of metabolism for the biosynthesis of secondary walls which could be a direct or indirect consequence of carbon availability (Rogers et al., 2005). Hertzberg et al. (2001) showed that genes involved in lignin and cellulose synthesis were expressed in the secondary wall formation zone of poplar wood. In the present experiment, the decreased xylem differentiation zone associated with a reduction in cambial activity could explain the reduced cellulose and lignin biosynthesis activities.

**Modification of wood anatomy under ozone**

Ozone-induced variations in wood anatomy occurred mainly in TW (Table 4) which is a major consumer of carbon and may therefore be more susceptible to the reduced allocation of carbon to the stem under ozone. Vessel lumen diameter was enhanced by ozone, the vessel frequency was decreased, and the vessel lumen fraction was maintained. Other environmental conditions may also influence wood anatomy. The effects of drought and salt stress, unlike those of ozone, have been well characterized. Both stresses result in decreased vessel area and increased vessel frequency (Junghans et al., 2006; Arend and Fromm, 2007; Escalante-Perez et al., 2009). The formation of smaller vessels in response to drought appears to be an adaptive response to drought-induced xylem embolism. Conversely, the modifications in vessel anatomy found in this experiment might improve water transport efficiency and induce a greater risk of embolism (Tyree et al., 1994; Zanne et al., 2010).

**Decrease of the cellulose to lignin ratio in wood under ozone**

It was shown that cellulose and lignin biosynthesis activities (enzymes and transcripts) were reduced by ozone in hybrid poplar wood as a consequence of the decrease in cambial activity and cell wall production. Because the synthesis of both components was altered in the same manner, the content of cellulose or lignin could not be predicted from these results. Indeed, cellulose and lignin contents express the relative abundance of each polymer in the cell wall. Since cellulose and lignin represent the main components of the cell wall, the variation in abundance of one component impacts the content of the other. In the present case, the

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Tension wood</th>
<th>Opposite wood</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Cambium cell layers (number)</td>
<td>Fibre frequency (number mm⁻²)</td>
</tr>
<tr>
<td>Control</td>
<td>6.4±0.5</td>
<td>2890±200</td>
</tr>
<tr>
<td>Ozone, 50 nl l⁻¹</td>
<td>3700±190*</td>
<td>142±25*</td>
</tr>
<tr>
<td>Ozone, 100 nl l⁻¹</td>
<td>4500±730*</td>
<td>126±36*</td>
</tr>
<tr>
<td>Ozone, 200 nl l⁻¹</td>
<td>3700±170*</td>
<td>124±15*</td>
</tr>
<tr>
<td>Ozone, 300 nl l⁻¹</td>
<td>4050±240*</td>
<td>105±62*</td>
</tr>
</tbody>
</table>

Data are mean values ±SD (n=3, individual replicates).

*Significant differences (P < 0.05) between control and ozone treatments. ND, not determined.
decrease of cellulose and lignin biosynthesis activities suggested that the amounts of both lignin and cellulose decreased, but do not allow their relative abundance to be deduced. Interestingly, the present data indicate that ozone exposure led to a modification of cell wall composition (Fig. 4). The cellulose content was reduced in accordance with the decrease of biosynthesis activities (enzymes and transcripts). Conversely, lignin content increased, in opposition to enzyme activities and transcript levels. An apparent increase in lignin content can be explained by a strong decrease in cellulose content. All these results suggest that cellulose and lignin biosynthesis was reduced under ozone and that cellulose biosynthesis was more affected than lignin biosynthesis. Accordingly, the cellulose to lignin ratio of the cell wall was reduced by ozone.

Like the anatomical changes, the modifications of the cellulose to lignin ratio were mainly observed in TW, thereby reinforcing the hypothesis that this tissue shows higher sensitivity to ozone than OW. The LS exhibited greater alteration than the MS, although MS only developed during ozone fumigation. However, MS was closer to the carbon source organs (leaves) and might be less affected than LS by carbon deficiency under ozone fumigation.

The response of lignin in stems was completely different from that observed in poplar or beech leaves under ozone (Cabané et al., 2004; Betz et al., 2009). Lignin biosynthesis (enzymes and transcripts) was highly stimulated in leaves, resulting in an increase in lignin content. The newly synthesized lignin was structurally different from constitutive lignin and was thought to be involved in defence mechanisms.

The reduced cellulose to lignin ratio of ozone-treated hybrid poplar stem suggests that the tree would promote lignification rather than cellulose biosynthesis under ozone stress. Lignin and cellulose deposition might be regulated in a compensatory manner in order to sustain the mechanical strength of the cell wall, as already suggested for transgenic poplars (Hu et al., 1999; Li et al., 2003). Although the metabolic cost of lignin synthesis is high (Amthor, 2003), in the context of an absence of carbon skeleton, the carbon cost of lignin strengthening the cell wall would be moderate compared with that of cellulose reinforcement. Moreover, the present results showed that ozone reduced wood density in TW (Table 3), suggesting a decrease in cell wall production per unit volume. By reducing the cellulose to lignin ratio, the tree could continue some radial growth and save carbon. The maintenance of radial growth would allow an increase in height and hence a renewal of photosynthesis which would have been impaired in ozone-damaged leaves. Thus, reduction of the cellulose to lignin ratio in wood might serve as a specific adaptation to maintain photosynthesis in ozone-injured trees.

In conclusion, the first results concerning the effect of ozone on cellulose and lignin deposition during wood formation are reported here. Ozone altered cellulose and lignin biosynthesis in hybrid poplar wood, resulting in a lower cellulose to lignin ratio, together with modifications of anatomy and density. This response appeared to be coordinated and to maintain radial growth despite the lack of carbon, as an indirect consequence of ozone impact on leaves. The results suggest that the mechanical properties of the wood are affected by high concentrations of ozone.

Supplementary data

Supplementary data are available at JXB online.

Figure S1. Photomicrographs of a stem cross-section stained with safranin O/Astra blue showing tension wood crescents in hybrid poplar cultivated vertically or bent in phytotronic chambers.

Figure S2. Photomicrographs of a cross-section showing the cambium zone in tension wood and opposite wood in stems (LS) of hybrid poplars cultivated for 46 d under control conditions or 200 nl l⁻¹ ozone.

Table S1. Lignin-derived monomers (H, G, and S) and S/G ratio after thioacidolysis of tension wood or opposite wood at different stem levels from hybrid poplars cultivated for 46 d under control conditions or different ozone concentrations.

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