Analysis of the early-flowering mechanisms and generation of T-DNA tagging lines in Kitaake, a model rice cultivar

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Abstract

As an extremely early flowering cultivar, rice cultivar Kitaake is a suitable model system for molecular studies. Expression analyses revealed that transcript levels of the flowering repressor Ghd7 were decreased while those of its downstream genes, Ehd1, Hd3a, and RFT1, were increased. Sequencing the known flowering-regulator genes revealed mutations in Ghd7 and OsPRR37 that cause early translation termination and amino acid substitutions, respectively. Genetic analysis of F2 progeny from a cross between cv. Kitaake and cv. Dongjin indicated that those mutations additively contribute to the early-flowering phenotype in cv. Kitaake. Because the short life cycle facilitates genetics research, this study generated 10 000 T-DNA tagging lines and deduced 6758 flanking sequence tags (FSTs), in which 3122 were genic and 3636 were intergenic. Among the genic lines, 367 (11.8%) were inserted into new genes that were not previously tagged. Because the lines were generated by T-DNA that contained the promoterless GUS reporter gene, which had an intron with triple splicing donors/acceptors in the right border region, a high efficiency of GUS expression was shown in various organs. Sequencing of the GUS-positive lines demonstrated that the third splicing donor and the first splicing acceptor of the vector were extensively used. The FST data have now been released into the public domain for seed distribution and facilitation of rice research.

Key words: Flanking sequence database, flowering time genes, GUS, Kitaake, splicing donor and acceptor, T-DNA.

Introduction

Rice (Oryza sativa. L.) is a model plant for monocot species because its genome has been sequenced (International Rice Genome Sequencing Project, 2005) and it can easily be transformed (Toki et al., 2006; Hiei and Komari, 2008). In addition, insertion mutants have been generated by T-DNA, Tos17, dSpm, and Ds (Krishnan et al., 2009). Their insertion sites have now been determined and the flanking sequence tag (FST) information deposited into public domains such as the Rice Functional Genomic Express Database (RiceGE, http://signal.salk.edu/cgi-bin/RiceGE), Oryzabase (www.shigen.nig.ac.jp/rice/oryzabase/top/top.jsp), and orygeneDB (http://orygenesdb.cirad.fr). The current RiceGE release contains 319,969 insertion sites, including: 106,100 FSTs from POSTECH, Korea (Jeon et al., 2000; Jeong et al., 2002, 2006; An et al., 2003; Ryu et al., 2004); 77,701 from NIAS Tos17 insertion mutants, Japan (Miyao et al., 2003); 47,231 from Rice Mutant Data (RMD), Huazhong Agricultural University, China (Zhang et al., 2006); 28,540 from Eu-CIRAD/Genoplante Oryza Tag Lines (OTL), France (Sallaud et al., 2004); 17,684 sites from UC Davis, USA (Kumar et al., 2005); 11,794 from Taiwan Rice Insertion Mutant, Academia Sinica (Hsing et al., 2007); and 10,281 from SHIP T-DNA, China (Fu et al., 2009).
Insertion mutants are valuable resources, especially when their insertion positions are annotated into a chromosome. This allows one to identify the knockout mutants in a given gene. Systemic phenotyping of the insertion mutants can be used to determine the phenotypic alteration that co-segregates with the insertion element (Krishnan et al., 2009). This reverse-genetics approach has elucidated a number of gene functions (An et al., 2005). Mutants can also be used for verification of functional analysis for a map-based cloned gene (Liu et al., 2009; Piao et al., 2009). Without an insertion mutant allele, one must complement the natural mutant, which requires cloning a full-length gene and transforming the mutant plant. Because transformation efficiency is low in many rice cultivars, such complementation experiments often require great effort and much time.

Some of the T-DNAs used for generating insertion mutants contain a promoterless reporter gene, for example GUS or GFP (Jeong et al., 2002; Ryu et al., 2004). When T-DNA is inserted within a gene in the same orientation as the mutated gene, a translational fusion can be formed. This allows for easy analysis of temporal and spatial expression patterns associated with the tagged gene (Jefferson, 1989). T-DNA can carry enhancer elements, such as the 35S enhancer element of cauliflower mosaic virus (Jeong et al., 2002, 2006; Chern et al., 2007; Wan et al., 2009).

When such a T-DNA is inserted near a gene, its expression level can be elevated by the enhancer element. This activation tagging is useful, especially because it generates gain-of-function mutations that are not easily obtained through other mutagenesis procedures. Because gain-of-function phenotypes are dominant, they can be readily distinguished from the somaclonal mutations that may occur during the tissue culture period. Activation tagging saves time and effort when producing overexpression transgenic plants. Therefore, this resource can be used for elucidating gene functions, especially when knockout mutants do not show any phenotypic alteration. This tool can also be employed for identifying agro-nomically important genes that have been silenced during the domestication of modern cultivars.

Unfortunately, most mutant populations are generated in rice cultivars, such as Nipponbare and Dongjin, that take several months to complete a life cycle. They are difficult to grow to fertility in controlled environments because they require high light intensity and warm temperatures. To overcome this obstacle, it would be desirable for one to be able to create mutant populations in a cultivar with a short life cycle and which grows well in greenhouses and growth chambers year-round. Kitaake is an extremely early-heading rice cultivar originating from Hokkaido, Japan (42–45° N latitude). It flowers 7–8 weeks after sowing, and seeds are harvested in 11–12 weeks.

Early flowering is controlled by several quantitative trait loci (QTLs). These include Ghd71Hd4, a major QTL in early-heading cultivars (Nonoue et al., 2008). Ghd7 encodes a CCT domain-containing protein in the CO-like family (Xue et al., 2008). Ghd81Hd5 is another allele that exhibits early flowering in those cultivars (Fujino et al., 2013). Ghd8 encodes an OsHAP3 subunit of the CCAAT-box binding protein (Yan et al., 2010). OsPRR37 has been identified as the major locus that enhances the photoperiod sensitivity of flowering (Murakami et al., 2005). These traits have been used for generating extremely early flowering cultivars (Shibaya et al., 2011). The current work identified the genes responsible for this phenomenon in cv. Kitaake, generated T-DNA insertion mutants in the cultivar, and examined the efficiency of GUS trapping.

### Materials and methods

#### Plant materials and growth conditions

Along with the extremely early flowering rice cultivar Kitaake, rice, which originated from cv. Hokkaido (42–45° N latitude) and other cultivars (Ichitani et al., 1997; Supplementary Fig. S1, available at JXB online), this study also analysed the rice cultivar Dongjin, a mid- to late-flowering cultivar from the southern part of the Korean Peninsula (36–37° N latitude). Plants were grown in the paddy field or in a greenhouse under either short days (12/12 light/dark cycle, 28/25 °C) or long days (14/8 light/dark, 28/25 °C day/night). Light level of the greenhouse was approximately 1000 μmol m–2 s–1.

#### Single-nucleotide polymorphism analyses

A G to T point mutation occurred at 157 nucleotides from the Ghd7 ATG start codon in cv. Kitaake, generating the SpeI (ACTAGT) site. To distinguish between Kitaake and Dongjin, this region was amplified with primers Ghd7 SNP F and R, and digested it with SpeI (Supplementary Table S1 and Supplementary Fig. S2). Because the point mutations in OsPRR37 did not generate a restriction enzyme site, the OsPRR37 single-nucleotide polymorphism was detected by allele-specific PCR (Supplementary Table S1 and Supplementary Fig. S3).

#### Generation of T-DNA insertion lines

The binary pGA2715 vector used in this study has been described previously (Jeong et al., 2002). Rice transformation was performed via the Agrobacterium-mediated co-cultivation method (Lee et al., 1999). Briefly, Agrobacterium strain LBA4404 harbouring pGA2715 was cultured in an AAM medium to an OD_{600} of 0.1–0.2. Scutellum-derived calli were co-cultured with the Agrobacterium for 3 d under darkness at 22 °C in a 2N6-AS medium containing 0.2% phytagel. After extensive washing, the calli were cultured for 2 weeks under light at 28 °C on a 2N6-DCH30 medium containing 30 mg l–1 hygromycin and 250 mg l–1 ceftaxime. Afterward, they were transferred for an additional 2 weeks of culturing under light at 28 °C in a 2N6-BA medium containing 50 mg l–1 hygromycin and 250 mg l–1 ceftaxime. Actively growing calli were then transferred to an MSR16 medium and cultured under light at 28 °C. This resulted in a 70–85% regeneration frequency.

#### Isolation of the sequences flanking T-DNA

Genomic DNAs were obtained from leaves of primary transgenic plants as described previously (An et al., 2003). The DNA sequences flanking the inserted T-DNA were determined by the inverse PCR method (An et al., 2003). Briefly, genomic DNA was digested with various restriction enzymes and the cut DNAs were self-ligated with T4 DNA ligase. Nested PCR was conducted to amplify the flanking sequences. The PCR product was directly sequenced with an Applied Biosystems 3730 DNA sequencer. If sequences were not resolved due to multiple amplifications, the amplified bands were separated on an agarose gel and eluted for sequencing.
Histochemical GUS analyses
GUS staining was performed by a previously described method, with some modifications (Jeon et al., 2000). Plants were cut into approximately 1 cm pieces and submerged in staining solution containing 0.5 M Na₂HPO₄ (pH 7.0), 0.5 M NaH₂PO₄ (pH 7.0), 0.1% Triton X-100, 0.5 M EDTA (pH 8.0), 2% DMSO, 0.1% X-gluc (5-bromo-4-chloro-3-indolyl-[b-D-glucuronic acid/cyclohexylammonium salt), 1 mM K₃[Fe(CN)]₆, 1 mM K₄[Fe(CN)]₆·3H₂O, and 5% methanol. After incubation at 37 °C for 2–24 h, the samples were transferred into 70% (w/v) ethanol at 65 °C for more than 1 h to remove the chlorophyll. The GUS-stained samples were then soaked in a clearing solution (4 M urea, 10% glycerol, and 0.1% Triton X-100) for 30–60 min at room temperature (Hama et al., 2011). Afterward, the staining pattern was observed with a dissecting microscope (SZX16; Olympus, Tokyo, Japan). For sectioning, tissues were embedded in Technovit 8100 resin ( Heraeus Kulzer, Armonk, NY, USA), sectioned to 5-μm thickness, and observed with an optical microscope (BX61; Olympus) using bright- and DIC-field illumination (Jung et al., 2005).

RNA isolation and quantitative real-time PCR
Total RNA was isolated with Tri Reagent (TaKaRa, Shiga, Japan). First-strand cDNAs were synthesized and utilized for quantitative real-time RT-PCR in a Rotor-Gene 6000 (Corbett Research, Sydney, Australia). Osubi1 served to normalize the cDNA quantity. All experiments were performed at least three times, with three or more samples at each point. Primers sequences for quantitative real-time PCR are shown in Supplementary Table S1. Changes in expression were calculated via the ΔΔCt method. To ensure primer specificity, these experiments were performed when the melting curve showed a single peak. Some PCR products were sequenced to verify the specificity of the reaction.

Analysis of splicing patterns
For analyses of splicing patterns, genomic DNA was isolated from leaves of the GUS-positive lines, and cDNAs were synthesized from total RNAs of those GUS-positive tissues. The junction regions between the tagged genes and GUS were amplified using the forward primer located in the tagged gene and the reverse primer located in GUS (Supplementary Table S1).

PCR was performed with an initial 5 min of denaturation at 95 °C, then 35 cycles of 95 °C for 20 s, 50 or 55 °C for 30 s, and 72 °C for 1 min, followed by a final 10 min of extension at 72 °C. The PCR products were sequenced by Macrogen (Seoul, Korea).

Results
Characteristics of cv. Kitaake rice
This study compared growth characteristics of cv. Kitaake with those of cv. Dongjin, a cultivar that has previously been used to generate numerous T-DNA insertion lines. In the paddy field (latitude 37° N), Kitaake flowered approximately 65 days after germination (DAG), or 41 d earlier than Dongjin (Fig. 1A, C). To test whether days-to-flowering was affected by photoperiodic conditions, these cultivars were grown under both short and long days. Kitaake flowered earlier under either state: at 51 DAG under short days (7 d earlier than Dongjin) and at 60 DAG under long days (26 d earlier; Fig. 1B, C).

Growth characteristics varied under different environmental conditions. In the paddy, Kitaake plants were about 77 cm tall, or 27 cm shorter than Dongjin (Fig. 1D). At heading time in the paddy, Kitaake averaged seven tillers per plant versus nine for Dongjin (Supplementary Table S2). However, under short days, Kitaake was as tall as Dongjin (Fig. 1) and had more tillers (Supplementary Table S2). These findings indicated that Kitaake grew better than Dongjin under short day conditions.

Transcript levels of RFT1 and Hd3a, both florigens in rice, were induced earlier in Kitaake than in Dongjin (Fig. 2A, B). In the latter, these genes were not turned on until 50 DAG when plants were grown under long days. However, they were already expressed at 23 DAG in Kitaake and transcript levels were increased at 32 DAG. Similarly, expression of Ehd1, a positive regulator immediately upstream of those florigens, was low in Dongjin whereas expression was detectable at 23 DAG and increased at 32 DAG in Kitaake (Fig. 2C). Because the early flowering phenotype of Kitaake was much stronger under long days or in the paddy, it is likely that a long day-preferential repressor was being expressed in that cultivar.

Three major regulators—Hd1, OsMADS50, and Ghd7—repress flowering preferentially under long days. For example, Hd1 enhances flowering time under short days but delays flowering under long days (Yano et al., 2000). OsMADS50 is an long day-preferential repressor functioning upstream of Ehd1 (Lee et al., 2004). Ghd7 is another long day-preferential repressor that has been identified as a major QTL that determines yield (Nonoue et al., 2008; Xue et al., 2008). Whereas the levels of Hd1 and OsMADS50 transcripts were not significantly altered in Kitaake (Fig. 2D, E), those of Ghd7 were much lower in Kitaake than in Dongjin (Fig. 2F).

Transcript levels were measured at 3-day intervals, from 14 DAG until 47 DAG. This experiment showed that Ghd7 transcripts were lower in Kitaake from the early stage of plant development (Fig. 3A). In Kitaake, levels were highest at 2 h after the light was turned on and were reduced to low levels thereafter (Fig. 3B). A similar diurnal rhythm was observed in Dongjin, albeit at much higher levels.

Ehd3 is a plant homeodomain (PHD) finger protein that suppresses expression of Ghd7 (Matsubara et al., 2011). Because Ghd7 expression was lower in Kitaake, this study examined Ehd3 transcripts to see whether their levels were higher in Kitaake. However, those levels were similar between the two cultivars (Fig. 2G). The study measured OsGI, another repressor of Ghd7 (Hayama et al., 2003; Itoh et al., 2010), and found little difference in its transcript levels (Fig. 2H). Also, there were no alterations in levels of OsPRR37, OsPhyA, B, or C (Fig. 3C, D and Supplementary Fig. S4). Therefore, their expression levels were not the primary reason for this reduction in Ghd7 transcripts in Kitaake.

To determine whether alterations in protein sequences might cause a decline in Ghd7 expression and delay early flowering, genomic DNA of the flowering-time regulatory genes from Kitaake were sequenced and compared with those from cv. Nipponbare. No changes were detected in the genes or the 2- to 2.5-kb promoter area of OsPhyA, OsPhyC, Sc5, Ehd3, ELF3, OsGI, or Hdl5/ Ghd8. Along with synonymous missense mutations in the
Roles of OsPRR37 and Ghd7 mutations in controlling flowering time

The effects of the Kitaake OsPRR37 and Ghd7 alleles were investigated by crossing Kitaake and Dongjin. From the F1 hybrid, 124 F2 progeny were grown in the paddy field and genotyped according to those alleles. The progeny with Kitaake OsPRR37 and Ghd7 alleles flowered at 66 ± 2.9 DAG, similar to the Kitaake parent (Fig. 4). By comparison, the progeny with Dongjin OsPRR37 and Ghd7 alleles flowered at 104 ± 2.5 DAG, similar to the Dongjin parent. This indicated that the two alleles are major contributors to early flowering in Kitaake. When the progeny carried the defective ghd7 from Kitaake and the normal OsPRR37 from Dongjin, they flowered at 76 ± 1.1 DAG), or 28 d earlier than the progeny with the Dongjin alleles. This demonstrated that the mutation in Ghd7 causes early flowering in Kitaake. Finally, when the progeny carried the defective osprr37 from Kitaake and the normal Ghd7 allele from Dongjin, they flowered at 89 ± 2.0 DAG, or 15 d earlier that those with the Dongjin OsPRR37 allele, thereby supporting the conclusion that the mutations in OsPRR37 also contribute to early flowering in Kitaake.
This study group has previously generated T-DNA tagging lines in cultivars Dongjin and Hwayoung, both of which take approximately 100–110 d to flower in the paddy field (Jeong et al., 2002, 2006). Neither grows well in the greenhouse during the cold season. The current work produced 10 000 primary transformants in Kitaake and grew them to maturity in the greenhouse. Seeds were harvested from approximately 6000 plants. To find the insertion position, inverse PCR was used to determine the sequences flanking the T-DNA. This resulted in the identification of 6758 FSTs from 4614 lines. The FSTs were located on rice chromosomes via a blastn homology search. Among the 6758 inserts, 3122 (46.2%) were genic. The 300-bp UTR from ATG and the stop codons were considered to be genic regions. T-DNAs were located at the 5′-UTR (548 lines), exons (1138 lines), and 3′-UTR (374 lines). Among the 3122 genic inserts, T-DNA was inserted into the gene that was not previously tagged by T-DNA or transposons in 367 lines (11.8%). These FST data have now been released into the public domain RiceGE Database (http://signal.salk.edu/cgi-bin/RiceGE) for purposes of seed distribution (Table 1).

T-DNAs were evenly distributed among 12 chromosomes, proportional to the chromosome size (Table 2). The highest frequency was at chromosome 1, which is the largest in the genome. Distribution of those insertions over the 12 chromosomes was very similar in Kitaake to those for Dongjin and Hwayoung (Fig. 5). The frequency of insertion was lower in the heterochromatin regions near the centromere and higher in the distal euchromatin regions. Similar results were obtained when the current findings were compared with those from T-DNA lines recorded at RMD and OTL (Supplementary Fig. S5).

Gene trapping using the GUS reporter gene

The pGA2715 vector used here carries the promoterless GUS reporter gene, with an intron containing triple splicing donors/acceptors immediately next to the right border (RB; Supplementary Fig. S6). Therefore, some of the tagging lines were capable of generating a translational fusion between the tagged gene and GUS. Among the T-DNA tags found in the genic regions, the orientation of GUS was identical to that of the tagged gene in 1366 inserts: 267 in the 5′-UTR, 561 in the exons, and 538 in the introns. Inserts in the 3′-UTR were excluded because they did not form a translational fusion. This work assayed 1213 lines for GUS activity. The remaining 153 were not evaluated because they produced too few seeds. A total of 159 lines (13.4%) proved GUS-positive; 118 showed staining in the roots, 102 in the shoots, and 88 in germinating seeds. Some of these (63 lines) indicated preferential expression in only one of those plant parts, whereas 53 lines showed GUS staining in all three (Supplementary Table S3).
A majority of the positive lines (80) with staining in the roots showed a GUS expression pattern ubiquitous in all root organs while 61 also exhibited expression in the shoots or germinating seeds. Some of the remaining lines had staining in a specific region within the roots while others were GUS-positive in two or more organs (Supplementary Table S4). For example, GUS was preferentially expressed in the root primodium in line K-00491 (Fig. 6A, B). T-DNA was tagged in Os09g21000, encoding a putative potassium transporter. Preferential staining was observed in the elongation zone of line K-01188 (Fig. 6C, D). T-DNA was also found in Os05g49300, encoding an iron-sulphur cluster assembly enzyme, or ISCU.

Among the 102 positive lines in the shoots, 26 showed GUS expression in all organs, including the leaf blade, sheath, coleoptile, mesocotyl, and collar (laminal joint). Of these lines, 24 also had expression in the roots and germinating seeds. Some of the remaining lines displayed staining in only a specific region while others were GUS-positive in two or more shoot organs (Supplementary Table S4).

Mesophyll cell-specific GUS expression was observed in line K-04449, where T-DNA was inserted in Os02g13360 (Fig. 6E, F). In line K-00221, GUS was strongly expressed

Fig. 3. Expression profiles of Ghd7 and OsPRR37 in cv. Kitaake (open circles) and cv. Dongjin (closed circles) grown under long days. (A and C) Temporal expression patterns of Ghd7 and OsPRR37 in leaf blades from 14 to 47 DAG; samples were prepared at 2 h after turning on lights. (B and D) Diurnal rhythm in leaves at 23 DAG. Values are mean of two or more independent experiments and standard deviation. Y-axis, relative values between transcript levels and Ubi.

Fig. 4. Analysis of flowering time in F2 population for cross between cv. Kitaake and cv. Dongjin. Ghd7 and OsPRR37 are Dongjin alleles and ghd7 and ospr37 are Kitaake alleles. A total of 124 F2 progeny were analysed. Numerals above the bars indicate the number of double homozygotes in each genotype class.
in the stomata (Fig. 6G, H). T-DNA was also located in Os08g35190, which encodes a putative auxin-repressed protein. In line K-00157, expression was strong in the collar region. T-DNA was inserted in Os07g05190, which encodes a leucine-rich repeat (LRR) family protein (Fig. 6I). In line K-04010, expression was preferential in the mesocotyl region (Fig. 6J).

GUS expression was also observed in various tissues of germinating seeds (e.g. the aleurone layer, scutellum, and endosperm; Supplementary Table S3). Endosperm-preferential expression was found in five lines. For example, staining was noted in line K-04010, where T-DNA was inserted into Os02g13360 (Fig. 6K). This expression pattern coincided with that from public array data (www.ricearray.org; Jung et al., 2011). Finally, in line K-04980, GUS expression was observed in the scutellum (Fig. 6L), where T-DNA was inserted into Os12g38880, which encodes a pentatricopeptide-like helical protein.

This study also assayed 421 lines at various reproductive stages. Staining was found in the mature floral organs and developing seeds of 80 lines (19.0%; Supplementary Table S5). Preferential expression was observed in the anther (line K-00143), ovary (line K-00188), pollen (line K-00241), vascular bundle in the palea/lemma (line K-01531), and branch rachis (line K-00561; Fig. 7A–F). Details for these patterns of expression are presented in Supplementary Table S6. Seeds were examined during three periods of development: 3–5, 10–12, and 15–17 days after pollination. In some lines, such as line K-04584, GUS expression was detected preferentially in the aleurone layer (Fig. 7G). In K-03829, expression was preferential to the plumule in the embryo region (Fig. 7H). For K-00350, GUS was detected in the dorsal region of the endosperm (Fig. 7I), while expression was preferential to the scutellum in K-01208 (Fig. 7J).

The frequency of occurrence for GUS-positive lines was slightly higher in exons (21.4%, 108/505) than in either introns (15.7%, 75/478) or the 5′-UTR (13.0%, 30/230).

**Splicing patterns of GUS-positive lines**

Because the pGA2715 vector includes an intron with triple splicing donors/acceptors immediately next to the RB, this feature allows for a translational fusion to occur between the tagged gene and GUS at high frequency if all three splicing donors and acceptors are utilized. However, the frequency of GUS-positive lines was much lower here than expected. Therefore, this work investigated splicing patterns in the lines showing GUS expression. Genomic DNA and cDNA were prepared, and the junction regions between the tagged gene and GUS were determined.

When T-DNA was inserted into an intron, the splicing donor from the tagged gene and the acceptor from the vector were used to generate fusion transcripts. If all three acceptors were employed, three different transcripts would be created but only one would make a proper reading frame, thereby forming a translational fusion. GUS-positive lines where T-DNA was inserted into an intron were selected. Sequencing revealed that these plants produced only one type of fusion transcript. For example, in line K-01715, T-DNA was inserted into intron 1. Analysis of the fusion transcript between the tagged gene and GUS revealed only one type of transcript, which utilized the first splicing acceptor (Fig. 8A). Similar splicing patterns were observed from line K-04652, where T-DNA was inserted into intron 1 of Os05g49180 (Table 3). Therefore, this indicated that the first splicing acceptor of the intron located in front of GUS was dominantly utilized.

**Table 1. Description of T-DNA insertions in cv. Kitaake T-DNA tagging lines**

<table>
<thead>
<tr>
<th>Location of T-DNA insertion</th>
<th>No. of inserts</th>
<th>No. of genes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Genic</td>
<td>3122</td>
<td>2666</td>
</tr>
<tr>
<td>5'-UTR (300bp upstream)</td>
<td>548</td>
<td>475</td>
</tr>
<tr>
<td>Exon</td>
<td>1138</td>
<td>958</td>
</tr>
<tr>
<td>Intron</td>
<td>1062</td>
<td>910</td>
</tr>
<tr>
<td>3'-UTR (300bp downstream)</td>
<td>374</td>
<td>323</td>
</tr>
<tr>
<td>Intergenic</td>
<td>3636</td>
<td>3009</td>
</tr>
<tr>
<td>Total</td>
<td>6758</td>
<td>5675</td>
</tr>
</tbody>
</table>

**Table 2. Distribution of T-DNA insertions in cv. Kitaake T-DNA tagging lines**

<table>
<thead>
<tr>
<th>Chromosome</th>
<th>Size, Mb (%)</th>
<th>Pseudomolecule, n (%)</th>
<th>Full-length cDNA, n (%)</th>
<th>Genic insertion, n (%)</th>
<th>Intergenic insertion, n (%)</th>
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</thead>
<tbody>
<tr>
<td>1</td>
<td>43.3 (11.6)</td>
<td>6542 (11.7)</td>
<td>4026 (14.1)</td>
<td>421 (13.5)</td>
<td>450 (12.4)</td>
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<tr>
<td>2</td>
<td>35.9 (9.6)</td>
<td>5387 (9.6)</td>
<td>3196 (11.2)</td>
<td>336 (10.8)</td>
<td>452 (12.4)</td>
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<tr>
<td>3</td>
<td>36.4 (9.8)</td>
<td>5573 (9.9)</td>
<td>3569 (12.5)</td>
<td>405 (13.0)</td>
<td>520 (14.3)</td>
</tr>
<tr>
<td>4</td>
<td>35.5 (9.5)</td>
<td>5322 (9.5)</td>
<td>2531 (8.9)</td>
<td>305 (9.3)</td>
<td>356 (9.8)</td>
</tr>
<tr>
<td>5</td>
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<td>4579 (8.2)</td>
<td>2313 (8.1)</td>
<td>245 (7.8)</td>
<td>302 (8.3)</td>
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<td>31.2 (8.4)</td>
<td>4724 (8.4)</td>
<td>2292 (8.0)</td>
<td>218 (7.0)</td>
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<td>4462 (8.0)</td>
<td>2183 (7.6)</td>
<td>275 (8.8)</td>
<td>264 (7.3)</td>
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<td>8</td>
<td>28.4 (7.6)</td>
<td>4194 (7.5)</td>
<td>1933 (6.8)</td>
<td>198 (6.3)</td>
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<td>3408 (6.1)</td>
<td>1605 (5.6)</td>
<td>183 (5.9)</td>
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<tr>
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<td>1538 (5.4)</td>
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<td>4166 (7.4)</td>
<td>1685 (5.9)</td>
<td>171 (5.5)</td>
<td>204 (5.6)</td>
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<tr>
<td>12</td>
<td>27.5 (7.4)</td>
<td>4022 (7.2)</td>
<td>1693 (5.9)</td>
<td>206 (6.6)</td>
<td>202 (5.6)</td>
</tr>
<tr>
<td>Total</td>
<td>373.2 (100)</td>
<td>56,081 (100)</td>
<td>28,564 (100)</td>
<td>3122 (100)</td>
<td>3636 (100)</td>
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When T-DNA was inserted into an exon, the intron located in front of \textit{GUS} would have been spliced. Because this study had three splicing donors and three splicing acceptors, nine different, alternative splicing products could have been generated if all were utilized. However, when sequenced here, only one type of transcript was observed. For example, in line K-04431 where T-DNA was inserted into exon 8 of Os07g06300 (encoding an ethylene-insensitive protein), the third splicing donor and the first splicing acceptor were used to make the mature mRNA. This splicing allowed for a translational fusion between the ethylene-insensitive protein and \textit{GUS} (Fig. 8B). Like line K-04431, nine additional lines has T-DNA inserted into an exon (Table 3). This same splicing pattern was observed when T-DNA was inserted into the 5'-UTR (Table 3). All of these findings demonstrated that only one each of the splicing donors and acceptors was predominantly used, regardless of the insertion positions.

**Discussion**

The availability of in-depth genome sequences and abundant public genetics resources make rice a good model system for understanding plant gene functions. However, because most cultivars do not perform well in controlled environments, it would be desirable to use one that grows rapidly in greenhouses and growth chambers year-round. This study proposed Kitaake as its model. It flowers about 1 month earlier than most japonica cultivars. Because it requires less light for optimal growth and is more cold tolerant than other rice cultivars, it grows much better in greenhouses even during the cold season.

Sequencing the flowering-control genes in Kitaake revealed that the amino acid sequences of \textit{Ghd7} and \textit{OsPRR37} were altered when compared with Dongjin. A point mutation in the former caused early termination of \textit{Ghd7} translation, resulting in a complete loss of function. As expected, transcript levels for \textit{Ehd1}, which is immediately downstream of \textit{Ghd7}, were elevated in Kitaake. \textit{Ehd1} merges various environmental and developmental signals and controls the expression levels of \textit{Hd3a} and \textit{RFT1}, both florigens in rice (Komiya et al., 2008). This same point mutation has been found in several other early-heading cultivars, such as Hayamasari, Mudandian8, and Hejiang19 (Xue et al., 2008; Matsubara et al., 2011). Therefore, it appears that the mutant allele is common to early-flowering

**Fig. 5.** Distribution of T-DNA insertions and KOME full-length cDNA along rice chromosomes. Red = Kitaake T-DNA insertions; blue = Dongjin/Hwayoung T-DNA insertions; grey = KOME full-length cDNA. Grey circles indicate the centromeric regions.
In OsPRR37, three mutations cause amino acid substitutions. The cultivar Kasalath also has a severe lesion in that gene, leading to premature termination (Murakami et al. 2005).

To evaluate the roles of the Ghd7 and OsPRR37 alleles in Kitaake, this study generated F2 progeny from the cross between Kitaake and Dongjin. Analyses showed that the plants with Kitaake OsPRR37 flowered 15 d earlier than those with Dongjin OsPRR37, whereas Kitaake Ghd7 accelerated flowering by 28 d. This experiment indicated that both the Ghd7 and OsPRR37 alleles from Kitaake are linked to early flowering and that the effects are more severe with Ghd7 than with OsPRR37.

Asian rice can be classified into five types (Ghd7-0, -0a, -1, -2, and -3), according to their variations in Ghd7 transcripts (Xue et al., 2008). Early-flowering cultivars such as Kitaake belong to type Ghd7-0a, carrying a non-functional allele. Transforming members of this type with the functional Ghd7

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**Fig. 6.** Patterns of GUS expression in 10 DAG seedlings. (A, B) Preferential expression in root primordium (RP) from line K-00491. (C, D) Preferential expression in root elongation zone (RE) from line K-01188. (E, F) Mesophyll cell (MC)-specific expression from line K-04449. (G, H) Stomata cell (SC)-preferential expression from line K-00221. (I) GUS expression in collar region (CL) from line K-00157; LS = leaf sheath. (J) GUS expression in mesocotyl region (MS) from line K-04010. (K) GUS-staining in endosperm (ED) of germinating seeds from line K-04210. (L) Preferential expression in scutellum (S) of germinating seeds from line K-04980. Bars = 2 mm (A, C–E, G, I–L), 200 μm (B), and 100 μm (F, H).
Kim et al. allele delays flowering time significantly. In cultivars with the non-functional allele, expression of Ehd1 is greatly enhanced, suggesting that Ghd7 functions upstream of Ehd1 as a repressor.

The OsPRR37 protein is highly homologous to Arabidopsis PRR proteins, which operate within a transcriptional feedback loop of clock-associated genes. For example, the Arabidopsis PRR proteins act as transcriptional repressors of Circadian Clock-Associated 1 (CCA1) and Late Elongation Hypocotyl (LHY), which are the central oscillators of the circadian clock in plants (Nakamichi et al. 2010). In Arabidopsis, CCA1 and LHY control flowering time through a genetic pathway that appears to be independent of floral regulators GIGANTEA and CONSTANS (Fujiiwara et al. 2008). They repress expression of Short Vegetative Phase (SVP), encoding a MADS box transcription factor that inhibits FT expression (Fujiiwara et al., 2008). Therefore, it could be speculated that OsPRR37 also acts as a repressor of rice CCA1 and LHY, which control a flowering-time regulator. In Kitaake, expression of Ghd7 was reduced, possibly due to mutations in OsPRR37. However, analysis of T-DNA insertion mutants suggested that OsPRR37 controls Hd3a independently from Ehd1, Hdl, and Ghd7 (Koo et al., 2013). Reduced Ghd7 transcript levels in Kitaake might be caused by nonsense-mediated mRNA decay. Further analysis is needed to elucidate the OsPRR37-mediated genetic pathway.

Annually, this research group distributes more than 3000 T-DNA insertion lines generated in Dongjin and Hwayoung. However, many laboratories experience difficulty in growing those plants to maturity. Due to space limitations in the GMO-certified paddy field, this group is unable to satisfy the demand for amplying seeds for the scientific community. Therefore, T-DNA insertional mutant populations in Kitaake were generated instead. The transformation efficiency of cultivar Kitaake is comparable to that of Dongjin, a cultivar that historically transforms well. As a first step toward producing a larger population, approximately 10 000 insertion lines have now been generated and the positions of 6758 insertion sites have been determined. As expected, their patterns of T-DNA distribution on the chromosomes are similar to those previously reported (Sallaud et al., 2004; Jeong et al., 2006).

### Flower

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<tr>
<th>Seeds</th>
<th>Longitudinal</th>
<th>Cross</th>
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<td>3–5</td>
<td>10–12</td>
<td>15–17 (DAP)</td>
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**Fig. 7.** Patterns of GUS expression in reproductive organs. Mature florets (A–F) and developing seeds (G–J) at three developmental stages (3–5, 10–12, and 15–17 days after pollination) were examined. (A) Anther (An) from line K-00143. (B) Ovary (O) from line K-00188. (C, D) Pollen (P) from line K-00241. (E) Vascular bundle (V) of palea/lemma from line K-01531. (F) Branch rachis (BR) from line K-00561. (G) Aleurone layer (Al) from line K-04584. Ed, endosperm. (H) Plumule of embryo (Eb) from line K-03829. (I) Endosperm transfer cell (TC) at dorsal region of endosperm from line K-00350. (J) Scutellum (S) from line K-01208. Bars = 2 mm (A, C, E–J), 200 μm (B), and 100 μm (D).
The frequency of knocking out a gene is approximately 46%, which is also close to that achieved in earlier experiments (An et al., 2003; Jeong et al., 2006).

Because the tagging lines carry the promoterless GUS within the T-DNA, some are GUS-positive, including 9.7% of the lines in their roots, 9.0% in the shoots, and 7.3% in
germinating seeds. In reproductive organs, 23.5% of the lines are GUS-positive. Those showing organ-specific expression should be good resources for further study. For example, laminar joint-preferential GUS expression occurs in line K-00157, where T-DNA is inserted into Os07g05190, which encodes a putative LRR family protein. The laminar joint is a mechanical tissue that bends the leaf blade away from the vertical axis toward the abaxial side when the leaf blade and leaf sheath are fully elongated (Hoshikawa, 1989). This laminar joint angle normally increases in parallel with the concentration of brassinosteroid, but that angle is narrower in brassinosteroid insensitive 1 (OsBRI1) mutants than in the wild type (Yamamuro et al., 2000; Jeong et al., 2012). Because OsBRI1 (Os01g52050) also has an LRR domain, these two proteins may share a similar molecular mechanism. Therefore, this line would be a good resource for studying the regulation of laminar joint movement in relation to brassinosteroids.

As another example, line K-04980 shows scutellum-preferential GUS expression. T-DNA interrupts Os12g38880, which encodes a putative pentatricopeptide-like helical protein. Such proteins are involved in gibberellin, cytokinin, and auxin responses, as well as ethylene biosynthesis (Schapire et al., 2006). In Arabidopsis, mutations within pentatricopeptide repeat (PPR) genes, including EMB 175, CYK8, and DYH216, have a phenotype of embryo defects (Lurin et al., 2004; Cushing et al., 2005). Therefore, this particular rice line would be useful in studying embryo development.

The T-DNA vector used in this study contains an intron with multiple splicing donor and acceptor sequences, allowing for high-efficiency GUS-tagging. However, the efficiency of GUS trapping has been much lower than expected. This indicates that not all of the splicing sequences are utilized in the plants. Sequencing of the GUS-positive lines revealed that only the third donor and the first acceptor are efficiently used to process pre-mRNA. Therefore, two-thirds of the T-DNA inserts cannot make a translational fusion. However, because those T-DNA insertion sites are known and the splicing patterns can be predicted, it is possible to deduce the lines that are capable of generating a functional translation fusion between the tagged gene and GUS.

Although public databases contain more than 300,000 FSTs, approximately 40% of the rice genes have not yet been mutated. This work has described 367 T-DNA tags in new genes that were not previously reported and 2755 tags to genes tagged earlier, thereby providing additional alleles. Thus, generation of additional mutant lines in Kitaake will facilitate functional analyses of rice genes.

Supplementary material

Supplementary data are available at JXB online.

Supplementary Fig. S1. Pedigree of cv. Kitaake, with tree drawn based on information from Ichitani et al. (1997).

References


