Chloroplastic thioredoxin-f and thioredoxin-m1/4 play important roles in brassinosteroids-induced changes in CO2 assimilation and cellular redox homeostasis in tomato

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Received 21 January 2014; Revised 10 April 2014; Accepted 11 April 2014

Abstract

Chloroplast thioredoxins (TRXs) and glutathione function as redox messengers in the regulation of photosynthesis. In this work, the roles of chloroplast TRXs in brassinosteroids (BRs)-induced changes in cellular redox homeostasis and CO2 assimilation were studied in the leaves of tomato plants. BRs-deficient d³im plants showed decreased transcripts of TRX-f, TRX-m2, TRX-m1/4, and TRX-x, while exogenous BRs significantly induced CO2 assimilation and the expression of TRX-f, TRX-m2, TRX-m1/4, and TRX-x. Virus-induced gene silencing (VIGS) of the chloroplast TRX-f, TRX-m2, TRX-m1/4, and TRX-y genes individually increased membrane lipid peroxidation and accumulation of 2-Cys peroxiredoxin dimers, and decreased the activities of the ascorbate–glutathione cycle enzymes and the ratio of reduced glutathione to oxidized glutathione (GSH/GSSG) in the leaves. Furthermore, partial silencing of TRX-f, TRX-m2, TRX-m1/4, and TRX-y resulted in decreased expression of genes involved in the Benson–Calvin cycle and decreased activity of the associated enzymes. Importantly, the BRs-induced increase in CO2 assimilation and the increased expression and activities of antioxidant- and photosynthesis-related genes and enzymes were compromised in the partially TRX-f- and TRX-m1/4-silenced plants. All of these results suggest that TRX-f and TRX-m1/4 are involved in the BRs-induced changes in CO2 assimilation and cellular redox homeostasis in tomato.

Key words: Antioxidant, Benson–Calvin cycle, chloroplast, 2-Cys peroxiredoxin, glutathione, photosynthesis.

Introduction

Accumulating evidence supports the hypothesis that the cellular redox signalling and hormone signalling pathways form an integrated redox–hormone network that regulates many plant growth- and defence-related pathways (Bartoli et al., 2013). Redox regulation is often mediated by thioredoxins (TRXs), which are able to (de-)activate enzymes through the reversible reduction of disulphide bonds (Buchanan and Balmer, 2005; Montrichard et al., 2009).

TRXs are ubiquitous, low molecular weight (~12 kDa) proteins that catalyse thiol–disulphide exchange reactions in conjunction with a large number of enzymes and related proteins (Jacquot et al., 2002). The short peptide motif WC(G/P)PC, with two conserved cysteine residues, has been characterized as the conserved active redox site of TRXs (Jacquot et al., 1997; Buchanan and Balmer, 2005; Meyer et al., 2008). Plant cells contain numerous TRXs in the cytosol, nucleus, mitochondria, endoplasmic reticulum, and chloroplasts, which continue to be identified (Marcus et al., 1991; Baumann and Juttner, 2002; Meyer et al., 2005). Recent proteomic studies using TRX-trapping chromatography or labelled gel electrophoresis in combination with protein identification via mass spectrometry have identified >180 potential in vitro
TRX target proteins (Motohashi et al., 2001; Balmer et al., 2003, 2004, 2006; Wong et al., 2004; Marchand et al., 2006; Alkhalifioui et al., 2007). There are five types of typical TRXs present in the chloroplasts. The f- and m-type TRXs function as messengers in the Fdx (ferredoxin)/TRX system by transmitting the redox signal from Fdx:TRX reductase to the target enzymes. TRX-f has been shown to activate several chloroplast enzymes, including enzymes involved in the Benson–Calvin cycle, ATP synthesis, and fatty acid synthesis (Nishizawa and Buchanan, 1981; Wolosiuk et al., 1993; Sasaki et al., 1997; Schwarz et al., 1997; Zhang and Portis, 1999). TRX-m has been found to be involved in the inactivation of glucose-6-phosphate dehydrogenase, which is the first enzyme in the oxidative pentose phosphate cycle (Wenderoth et al., 1997). Both TRX-f and TRX-m can activate fructose-1,6-bisphosphatase (FBPase) and NADP-dependent malate dehydrogenase (NADP-MDH) in vitro, suggesting that there is some overlap in substrate specificity between the isoforms (Hodges et al., 1994; Geck et al., 1996). TRX-x, which is inactive toward FBPase and NADP-MDH, is the most efficient reductant of 2-Cys peroxiredoxin (2-CP). This property suggests that TRX-x functions specifically in resistance to oxidative stress (Collin et al., 2003). TRX-y exists in two isoforms and has been reported to act as an efficient electron donor for the chloroplast glutathione peroxidase (Navrot et al., 2006) and methionine sulfoxide reductase B2 (Vieira Dos Santos et al., 2007). Peroxiredoxin Q, which has been reported to function as an antioxidant (Rouhier et al., 2004; Lamkemeyer et al., 2006), was shown to be the best substrate of TRX-y (Collin et al., 2004). Lastly, TRX-z regulates plastid-encoded RNA polymerase-dependent transcription (Arsova et al., 2010). However, the biological functions of TRXs have been studied mostly in Arabidopsis through in vitro experiments, and detailed in vivo studies using different isoforms have not yet been conducted.

Brassinosteroids (BRs) are a group of plant steroid hormones that have been shown to play important roles in the growth, development, and stress response of plants (Clouse and Sasse, 1998; Müssig, 2005). For example, the tomato d"m mutant with the block in C-6 oxidation in BRs biosynthesis (Bishop et al., 1999). Previous studies showed that BRs can enhance stress tolerance against chilling, paraquat (PQ), and biotic stress, and this effect is largely dependent on BRs-induced hydrogen peroxide (H2O2) accumulation in the apoplast (Xia et al., 2009b). Furthermore, BRs are involved in the regulation of CO2 assimilation in several plants. In cucumber, BRs induce transient increases in RBOH (respiratory burst oxidase homolog) expression, NADPH oxidase activity, H2O2 levels in the apoplast, and nitric oxide (NO) production (Xia et al., 2009b; Cui et al., 2011). Additionally, BRs can induce the expression and activity of genes and enzymes involved in the Benson–Calvin cycle, and this effect is attributed to the apoplastic H2O2-induced activation of photosynthesis-related redox-sensitive enzymes (Jiang et al., 2012a). Apoplastic H2O2 can induce an increase in the ratio of reduced glutathione (GSH) to oxidized glutathione (GSSG), resulting in increased stability of redox-sensitive enzymes (Jiang et al., 2012b). Microarray analysis revealed that several TRX genes are targeted by BRs in Arabidopsis and rice (Müssig et al., 2002; Goda et al., 2004; Wu et al., 2008); however, their potential roles in BRs-induced cellular redox homeostasis and CO2 assimilation remain unknown.

Tomato is an important horticultural crop distributed world-wide and has been widely used to study the stress response. Virus-induced gene silencing (VIGS) has been well established and widely used for the analysis of gene functions in Solanaceous species (Liu et al., 2002a, b; Li et al., 2006). In this study, the roles of the different chloroplastic TRXs in the BRs-induced changes in cellular redox homeostasis and CO2 assimilation were examined. Accordingly, five chloroplastic TRXs were partially silenced individually using a VIGS approach, and the changes in CO2 assimilation as well as the activities of Calvin cycle and ascorbate (AsA)–GSH cycle enzymes and cellular glutathione redox homeostasis were subsequently determined in the presence or absence of exogenous BRs.

**Materials and methods**

**Plant materials and experimental design**

*Experiment I* Tomato seeds from wild-type (*Solanum lycopersicum* L. cv. Condine Red, CR) and its partially BRs-deficient d"m mutant were obtained from the Tomato Genetics Resource Center (University of California, Davis, CA, USA). These seeds were germinated and grown in a mixture of peat and vermiculite (1:1, v:v) under a 16 h light (200 μmol m−2 s−1; at 25 °C), 8 h dark (at 20 °C) cycle. To determine the role of BRs in the regulation of CO2 assimilation and the effects of BRs on the transcript levels of the chloroplastic TRX genes, plants at the four-leaf stage were sprayed with water or 0.2 μM EBR (24-epibrassinolide; Sigma, Santa Clara, CA, USA), one of the most active and stable forms of BRs. Twenty-four hours later, CO2 assimilation was measured in the third leaf from the bottom. Additionally, leaf samples were harvested from the water- or EBR-treated CR and d"m plants, frozen immediately in liquid nitrogen, and stored at −80 °C prior to gene expression analyses.

*Experiment II* Tomato (CR) seeds were germinated and grown in a mixture of peat and vermiculite (1:1, v:v) under a 16 h light (200 μmol m−2 s−1; at 25 °C), 8 h dark (at 20 °C) cycle. To examine the roles of the chloroplast TRXs in the regulation of CO2 assimilation and to determine whether they are involved in the BRs-induced increase in CO2 assimilation, VIGS was performed when the cotyledonary leaves were fully expanded but the true leaves had not yet appeared. Partially gene-silenced plants at the six-leaf stage were treated with 0.2 μM EBR via foliar spraying on all of the leaves; 10 ml of this solution was applied per plant, and distilled water was used as a control. After ~24 h, CO2 gas exchange was measured, and leaf samples were harvested for biochemical and gene expression analyses.

**VIGS constructs and Agrobacterium-mediated virus infection**

Five chloroplastic TRX isoforms were obtained, namely TRX-f, TRX-m2, TRX-m1/4, TRX-x, and TRX-y, from the tomato genome database using the BLAST tool (http://mpis.helmholtz-muenchen.de/plant/tomato/index.jsp) based on their similarity to the characterized chloroplastic TRX isoforms in Arabidopsis. To generate the Tobacco rattle virus (pTRV) VIGS constructs pTRV2-TRX-f, pTRV2-TRX-m2, pTRV2-TRX-m1/4, pTRV2-TRX-x, and pTRV2-TRX-y, which were used for partial silencing of the TRX-f, TRX-m2,
TRX-m1/4, TRX-x, and TRX-y genes, respectively, fragments of the corresponding genes with sizes of 210, 231, 312, 285, and 251 bp were PCR amplified from tomato cDNA with primers containing XhoI (CTCGAG) and SacI (GAGCTC) restriction sites. All of the primers used are listed in Supplementary Table S1 available at JXB online. The resulting plasmids were subsequently introduced into Agrobacterium tumefaciens strain GV3101.

For virus infiltration, a mixed culture of A. tumefaciens carrying the pTRV1:pTRV2-target gene in a 1:1 ratio was infiltrated into fully expanded cotyledonal leaves of tomato plants (Ekenrgen et al., 2003). An Agrobacterium culture carrying the empty pTRV2 vector was also infiltrated into a set of plants, which were used as a control. The inoculated plants were maintained at 20–22 °C in a growth chamber with a 16 h daylength. After ~4 weeks, quantitative real time–PCR (qRT–PCR) was performed to determine the gene silencing efficiency before the plants were used in assays (Supplementary Fig. S1 available at JXB online).

Leaf gas exchange measurements

Gas exchange analysis was conducted on the third leaf for Experiment I and on the fifth leaf for Experiment II using an open gas exchange system (LI-6400; LI-COR, Lincoln, NE, USA). The light-saturated rate of CO2 assimilation (Amax) was measured under an ambient CO2 concentration of 380 μmol mol−1 at a saturating photosynthetic photon flux density (1000 μmol m−2 s−1), at a leaf temperature of 25 ± 1.5 °C and a relative air humidity of 80–90%. An assimilation versus intercellular CO2 concentration (A/C) curve was determined according to von Caemmerer and Farquhar (1981). Assimilation was first measured at the ambient CO2 concentration under which the plants had grown. The atmospheric CO2 concentration was decreased to 50 μmol mol−1 in a stepwise manner and then returned to the growth concentration to check that the original rate could be regained; the rate was then finally increased stepwise to 2000 μmol mol−1 to complete the response curve. The maximum ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco) carboxylation rates (Vc,max) and the maximum ribulose-1,5-bisphosphate (RuBP) regeneration rates (Jmax) were estimated from the A/C curves using the method described by Ether and Livingston (2004).

Determination of lipid peroxidation

Lipid peroxidation was estimated by measuring malondialdehyde (MDA) equivalents, according to the method described by Hodges et al. (1999). Leaf samples (0.3 g) were ground in 3 ml of ice-cold 25 mM HEPES buffer (pH 7.8) containing 0.2 mM EDTA and 2% polyvinylpyrrolidone (PVP). The obtained homogenates were centrifuged at 4 °C for 20 min at 12 000 g, and the resulting supernatants were used for analysis of MDA equivalents. The samples were mixed with 10% trichloroacetic acid (TCA) containing 0.65% 2-thiobarbituric acid (TBA) and heated at 95 °C for 25 min. The content of MDA equivalents was corrected for non-MDA compounds by subtracting the absorbance at 532 nm of a TBA-less solution containing the plant extract.

Non-reducing SDS–PAGE and western blot analysis of 2-Cys peroxiredoxin

Total proteins were isolated from leaf tissues in a protein extraction buffer (100 mM HEPES, pH 7.5, 5 mM EDTA, 5 mM EGTA, 10 mM NaVO4, 10 mM NaF, 50 mM β-glycerophosphate, 1 mM phenylmethylsulphonyl fluoride, 10% glycerol, and 7.5% polyvinylpyrrolidone) supplemented with 10 mM N-ethylmaleimide (NEM) (thiol-blocking reagent). After centrifugation at 13 000 g for 20 min, the supernatants were transferred to clean tubes, quickly frozen in liquid nitrogen, and stored at −80 °C. The protein concentration in the extracts was determined using the Bio-Rad protein assay kit (Bio-Rad, Hercules, CA, USA), and bovine serum albumin (BSA) was used as a standard. Reducing agents and boiling were omitted in this protocol, as it is important to maintain the remaining disulphide bonds (Muthuramalingam et al., 2010). Total protein samples (15 μg) supplemented with 5× loading buffer [225 mM TRIS-HCl, pH 6.8, 5% (w/v) SDS, 50% glycerol, 0.05% bromophenol blue] were separated via 12% SDS–PAGE, and the redox state of 2-CP was detected through western blot analysis with a polyclonal antibody against 2-CP (Beijing Protein Innovation, Beijing, China). After incubation with a horseradish peroxidase (HRP)-linked secondary antibody (Cell Signaling Technology, Boston, MA, USA), the complexes on the blot were visualized using an enhanced chemiluminescence kit (Perkin Elmer, Wellesley, MA, USA), according to the manufacturer’s instructions. Band intensity was quantified using Quantity One software.

Determination of Rubisco, Rubisco activase (RCA), and FBPase activity

Rubisco activity was measured spectrophotometrically by coupling 3-phosphoglycerate acid formation with NADH oxidation at 25 °C according to the method described by Lilley and Walker (1974), with some modifications. Total activity was assayed after the crude extract had been activated in a 0.1 ml activation mixture containing 33 mM TRIS-HCl (pH 7.5), 0.67 mM EDTA, 33 mM MgCl2, and 10 mM NaHCO3 for 15 min. The initial measurements of Rubisco activity were carried out in 0.1 ml of reaction medium containing 5 mM HEPES-NaOH (pH 8.0), 1 mM NaHCO3, 2 mM MgCl2, 0.25 mM diithiothreitol (DTT), 0.1 mM EDTA, 1 U of glyceraldehyde 3-phosphate dehydrogenase (GAPDH), 0.5 mM ATP, 0.015 mM NADH2, 0.5 mM phosphocreatine, 0.06 mM RuBP, and 10 μl of extract. The change in absorbance at 340 nm was monitored for 90 s. RCA activity was determined using a Rubisco Activase Assay Kit (Genmed Scientifics, Washington, DC, USA). FBPass activity was determined by monitoring the increase in A540 using an extinction coefficient of 6.2 mM−1 cm−1 (Scheibe et al., 1986). Total activity was assayed after the crude extract had been activated in a 0.1 ml activation mixture containing 100 mM DTT, 2 mM FBP, 10 mM MgCl2, and 0.1 TRIS-HCl (pH 8.0). The initial activity was assayed immediately after homogenization. The assay mixture consisted of 0.1 mM HEPES-NaOH (pH 8.0), 0.5 mM Na2EDTA, 10 mM MgCl2, 0.3 mM NADPH+, 0.6 mM fructose-1,6-bisphosphate, 0.6 U of glucose-6-phosphate dehydrogenase from baker’s yeast (Sigma, Santa Clara, CA, USA), 1.2 U of glucose phosphate isomerase from baker’s yeast (Sigma, Santa Clara, CA, USA), and 100 μl of enzyme extract in a final volume of 1 ml.

Measurements of glutathione contents and the activity of enzymes involved in the AsA–GSH cycle

For the measurement of reduced glutathione (GSH) and oxidized glutathione (GSSG), plant leaf tissue (0.3 g) was homogenized in 2 ml of 6% metaphosphoric acid containing 2 mM EDTA and centrifuged at 4 °C for 10 min at 12 000 g. After neutralization with 0.5 M phosphate buffer (pH 7.5), 0.1 ml of the supernatant was added to a reaction mixture containing 0.2 mM NADPH, 100 mM phosphate buffer (pH 7.5), 5 mM EDTA, and 0.6 mM 5,5’-dithio-bis (2-nitrobenzoic acid). The reaction was initiated by adding 3 U of glutathione reductase (GR) and was monitored by measuring the changes in absorbance at 412 nm for 1 min. For the GSSG assay, GSH was masked by the addition of 40 μl of 2-vinylpyridine to the neutralized supernatant, whereas 40 μl of water was added for the ascorbate assay. The GSH concentration was obtained by subtracting the GSSG concentration from the total concentration (Rao and Örmsg, 1995).

To determine the enzymatic activities of proteins involved in the AsA–GSH cycle, leaf tissue (0.3 g) was ground in 3 ml of ice-cold buffer containing 25 mM HEPES (pH 7.8), 0.2 mM EDTA, 2 mM ascorbic acid, and 2% PVP. The homogenates were centrifuged at 4 °C for 20 min at 12 000 g, and the resulting supernatants were used to determine the enzymatic activity. The ascorbate peroxidase activity was assayed after the crude extract had been activated in a 0.1 ml activation mixture containing 33 mM TRIS-HCl (pH 7.5), 0.67 mM EDTA, 33 mM MgCl2, and 10 mM NaHCO3 for 15 min. The initial measurements of activities were carried out in 0.1 ml of reaction medium containing 5 mM HEPES-NaOH (pH 8.0), 1 mM NaHCO3, 2 mM MgCl2, 0.25 mM diithiothreitol (DTT), 0.1 mM EDTA, 1 U of glyceraldehyde 3-phosphate dehydrogenase (GAPDH), 0.5 mM ATP, 0.015 mM NADH2, 0.5 mM phosphocreatine, 0.06 mM RuBP, and 10 μl of extract. The change in absorbance at 340 nm was monitored for 90 s. RCA activity was determined using a Rubisco Activase Assay Kit (Genmed Scientifics, Washington, DC, USA). FBPass activity was determined by monitoring the increase in A540 using an extinction coefficient of 6.2 mM−1 cm−1 (Scheibe et al., 1986). Total activity was assayed after the crude extract had been activated in a 0.1 ml activation mixture containing 100 mM DTT, 2 mM FBP, 10 mM MgCl2, and 0.1 TRIS-HCl (pH 8.0). The initial activity was assayed immediately after homogenization. The assay mixture consisted of 0.1 mM HEPES-NaOH (pH 8.0), 0.5 mM Na2EDTA, 10 mM MgCl2, 0.3 mM NADPH+, 0.6 mM fructose-1,6-bisphosphate, 0.6 U of glucose-6-phosphate dehydrogenase from baker’s yeast (Sigma, Santa Clara, CA, USA), 1.2 U of glucose phosphate isomerase from baker’s yeast (Sigma, Santa Clara, CA, USA), and 100 μl of enzyme extract in a final volume of 1 ml.
and TRX-x transcript levels were upregulated ~1-fold following treatment with EBR in CR plants. Similarly, no significant change was observed in TRX-y transcript levels in the EBR-treated CR plants. Interestingly, the expression of those TRX genes was restored to a level similar to that in the water-treated CR plants when d\textsuperscript{gm} plants were treated with EBR (Fig. 2a).

The role of chloroplast TRXs in BRs-induced \(\text{CO}_2\) assimilation

BRs levels are closely related to \(\text{CO}_2\) assimilation in cucumber plants (Yu et al., 2004; Xia et al., 2009a). Here, it was found that the light-saturated rate of \(\text{CO}_2\) assimilation (\(A\text{\textsubscript{sat}}\)) was reduced by 24.7% in BRs-deficient d\textsuperscript{gm} mutant plants compared with wild-type CR plants. However, exogenous application of 0.2 \(\mu\text{M}\) EBR increased the \(A\text{\textsubscript{sat}}\) value in CR plants 24 h after EBR application. Furthermore, exogenous EBR increased the \(A\text{\textsubscript{sat}}\) of the d\textsuperscript{gm} plants to the level of untreated CR plants (Fig. 2b).

To determine the role of these TRXs in BRs-induced \(\text{CO}_2\) assimilation, the five TRX genes were partially silenced individually using a VIGS method. Transcript analysis of the leaflets in the middle of the fifth fully expanded leaves revealed that the transcripts for these genes were reduced by 65–85% in the respective silenced plants (Supplementary Fig. S1 available at JXB online). To investigate the underlying molecular mechanisms of BRs-induced \(\text{CO}_2\) assimilation, the effect of EBR on \(\text{CO}_2\) assimilation were analysed in partially silencing the TRX-y gene (pTR V-TRX-y), TRX-m2 silenced (pTR V-TRX-m2), TRX-m1/4 silenced (pTR V-TRX-m1/4), TRX-x silenced (pTR V-TRX-x), and TRX-y silenced (pTR V-TRX-y) plants. As shown in Fig. 3a, partially silencing the TRX-f, TRX-m2, TRX-m1/4, and TRX-y genes resulted in decreases in the \(A\text{\textsubscript{sat}}\) of 18.9, 26.0, 20.5, and 22.8%, respectively, compared with the pTRV control plants. However, partially silencing TRX-x did not result in significant changes in \(A\text{\textsubscript{sat}}\). The application of 0.2 \(\mu\text{M}\) EBR increased \(A\text{\textsubscript{sat}}\) in the pTR V-TRX-f, pTR V-TRX-m2, and pTR V-TRX-y plants by 48.0, 30.9, and 31.6%, respectively, but had little effect on the pTR V-TRX-f, pTR V-TRX-m2, and pTR V-TRX-y plants but were not changed in the pTR V-TRX-x plants (Fig. 3b, c). Accordingly, exogenous application of EBR increased \(V\text{\textsubscript{c,max}}\) and \(J\text{\textsubscript{max}}\) only in the pTR V-TRX-f, pTR V-TRX-m2, and pTR V-TRX-y plants and not in the pTR V-TRX-m1/4, pTR V-TRX-x plants. All of these results indicated that chloroplastic TRX-f, TRX-m2, TRX-m1/4, and TRX-y are involved in the regulation of \(\text{CO}_2\) assimilation, whereas only TRX-f and TRX-m1/4 play a role in the EBR-induced increase in \(\text{CO}_2\) assimilation.

TRX-induced changes in redox homeostasis influenced by BRs

MDA is a useful indicator of reactive oxygen species (ROS)-induced lipid peroxidation in plants. Increased MDA accumulation was detected in the partially TRX-f, TRX-m2, TRX-m1/4, and TRX-y silenced plants, but not in the
partially TRX-x-silenced plants. EBR treatment reduced the MDA content only in the pTR V, pTRV-TRX-m2, and pTRV-TRX-y plants and had no effect on the pTRV-TRX-f, pTRV-TRX-m1/4, and pTRV-TRX-x plants (Fig. 4a). Western blot analysis showed that 2-CP was mostly present in a reduced state (monomer), while ~22.7% was in an oxidized state (dimer) in the pTR V plants. In contrast, partial silencing of the TRXs resulted in an increase in the levels of oxidized 2-CP in the leaves, while the ratio of 2-CP monomers/2-CP dimers decreased by 20.5, 31.4, 37.5, 37.0, and 19.9% in the pTR V-TRX-f, pTR V-TRX-m2, pTR V-TRX-m1/4, pTR V-TRX-x, and pTR V-TRX-y plants, respectively, compared with the pTR V control plants. The application of 0.2 μM EBR increased the ratio of reduced 2-CP/oxidized 2-CP in the pTR V, pTR V-TRX-m2, pTR V-TRX-x, and pTR V-TRX-y plants by 55.1, 70.1, 129.3, and 74.0%, respectively, but had little effect on the pTR V-TRX-f and pTR V-TRX-m1/4 plants (Fig. 4b, c).

Involvement of glutathione redox homeostasis in BRs-induced CO₂ assimilation

TRX can regulate the activity of the AsA–GSH cycle, which plays a critical role in maintaining the cellular redox status. In this study, the changes in the activities of APX, MDAR, DHAR, and GR, which are four important enzymes involved in the AsA–GSH cycle, were analysed. As shown in Fig. 5a, partial silencing of TRX-f, TRX-m2, TRX-m1/4, and TRX-y decreased the activity of APX, MDAR, DHAR, and GR. However, no significant effects on the activities of these enzymes were observed in the pTR V-TRX-x plants. EBR treatment significantly increased the activity of the AsA–GSH cycle in the pTR V control, pTR V-TRX-m2, and pTR V-TRX-y plants, but had no effect in the pTR V-TRX-f, pTR V-TRX-m1/4, and pTR V-TRX-x plants. To analyse the EBR-induced changes in glutathione redox homeostasis further, the effects of EBR on the contents of GSH and GSSG and their ratios in the pTR V and various pTR V-TRX plants were compared. There was generally little change in GSH content observed in the pTR V and partially TRX-silenced plants (Fig. 5b). However, partial silencing of TRX-f, TRX-m2, TRX-m1/4, and TRX-y resulted in 31.5, 27.5, 30.2, and 30.1% increases in the GSH/GSSG content, respectively, leading to a decrease in the GSH/GSSG ratio. In contrast, EBR induced a slight increase in GSH content and decrease in GSSG, resulting in a significant increase in the GSH/GSSG ratio. Similar to the observed changes in the activities of the AsA–GSH cycle enzymes, the
EBR-induced increases in the glutathione pool and GSH/GSSG ratio were abolished by partially silencing TRX-f and TRX-m1/m4 but not by the silencing of other TRXs (Fig. 5a, b).

Gene expression and enzyme activities involved in the Benson–Calvin cycle influenced by the TRXs and BRs

Previously, it was reported that BRs-induced CO₂ assimilation is associated with increased expression of photosynthetic genes in cucumber plants (Yu et al., 2004; Xia et al., 2009a). In this study, the transcript levels of eight Benson-Calvin cycle-related genes in the leaves of plants lacking different chloroplastic TRX genes were analysed. These tested photosynthesis-related genes included the genes encoding Rubisco activase (RCA), Rubisco large subunit (rbcL), Rubisco small subunit (rbcS), glycerate-3-phosphate kinase (PGK), glyceraldehyde-3-phosphate dehydrogenase (GAPDH), fructose-1,6-bisphosphatase (FBPase), sedoheptulose-1,7-bisphosphatase (SBPase), and ribulose-5-phosphate kinase (PRK). As shown in Fig. 6, the transcripts of these genes were all up-regulated upon EBR treatment in the pTRV plants. In contrast, partially silencing the chloroplastic TRX genes resulted in down-regulation of these photosynthesis-related genes. Importantly, EBR-induced transcripts were detected only in the pTRV-TRX-m2 and pTRV-TRX-y plants and not in the other plants.

BRs levels are closely related to the activation of a subset of enzymes involved in the Benson–Calvin cycle, particularly the redox-sensitive enzymes (Yu et al., 2004; Xia et al., 2009a). Here, the changes in the activities of RCA, Rubisco, and FBPase were analysed in VIGS plants with and without exogenous application of EBR. As shown in Fig. 7, partial silencing of these genes did not result in significant changes...
Exogenous EBR application increased the activity or the respective enzymes, compared with the pTR V plants. Furthermore, the activity and the activation rates of the enzymes in the EBR-treated VIGS plants were much lower than in the pTRV control plants.

**Discussion**

TRX-f, TRX-m2, TRX-m1/4, and TRX-y are involved in the regulation of cellular redox homeostasis via the AsA–GSH cycle and 2-CP

TRXs are involved in the regulation of ROS metabolism and CO₂ assimilation. The results of the present study demonstrated that TRXs can differentially regulate oxidative stress in tomato plants. In rice, OsTrxRm RNAi plants exhibit increased H₂O₂ accumulation in the leaves (Chi et al., 2008). Histochemical staining demonstrated that more O₂⁻ and H₂O₂ accumulates in the leaves of the VIGS-TRX-f/TRX-m pea plants compared with control plants, confirming that TRX-f and TRX-m are important for the metabolic balance of ROS in plant cells (Luo et al., 2012). Here, it was found that partially silencing TRX-f, TRX-m2, TRX-m1/4, or TRX-y resulted in increased oxidative stress, as indicated by the increased MDA content in the leaves, while partially silencing TRX-x did not induce significant changes in the MDA content of the leaves (Fig. 4a). These findings revealed that chloroplastic TRX-f, TRX-m2, and TRX-m1/4 as well as TRX-y are involved in ROS metabolism. It seems likely that TRX-x plays an insignificant role in ROS metabolism, as partially silencing TRX-x did not induce oxidative stress in the leaves.

ROS accumulation is closely related to the ROS scavenging capacity of cells, which is largely dependent on antioxidant enzymes, such as APX, MDAR, DHAR, and GR, and non-enzymatic antioxidants, such as AsA and GSH. Proteomic studies in Arabidopsis have determined that several key enzymes involved in the AsA–GSH cycle, such as GR, are redox sensitive (Ströher and Dietz, 2008; Wang et al., 2012), and biochemical and genetic assays have identified the NADPH-dependent thioredoxin system as a backup system for GR1 (Marty et al., 2009). Until now, few studies have investigated the role of TRXs in the regulation of antioxidant activity in plants. Here, it is demonstrated that partially silencing the TRX-f, TRX-m2, TRX-m1/4, and TRX-y genes resulted in decreased activity of the enzymes APX, MDAR, DHAR, and GR, while partially silencing the TRX-x gene did not induce significant changes in the activity of these enzymes (Fig. 5a). Additionally, partially silencing TRX-f, TRX-m2, TRX-m1/4, and TRX-y did not alter the total glutathione (GSH+GSSG) or GSH content but did induce an increase in the GSSG content, leading to a decrease in the GSH/GSSG ratio (Fig. 5b). These results indicated that TRX-f, TRX-m2, TRX-m1/4, and TRX-y do not influence GSH biosynthesis but affect glutathione homeostasis by regulating GR activity, as observed in both the glutathione-deficient cad2 and GR-deficient gr1 Arabidopsis mutants (Meyer et al., 2007; Marty et al., 2009). However, such an effect was not observed in the pTRV-TRX-x plants. All of these results suggested that the enzymes involved in the AsA–GSH cycle are largely modified by TRX-f, TRX-m2, TRX-m1/4, and TRX-y.

in total Rubisco activity but significantly decreased the initial Rubisco, initial FBPase, and Rubisco activase activities as well as the Rubisco activation rate, except in the pTRV-TRX-x plants, where these parameters were not significantly altered. For example, initial Rubisco activities decreased by 31.9, 47.4, 35.6, and 39.6% while the Rubisco activation rate decreased by 32.1, 42.0, 29.8, and 41.3% in the pTRV-TRX-f, pTRV-TRX-m2, pTRV-TRX-m1/4, and pTRV-TRX-y plants, respectively compared with the pTRV plants. Furthermore, exogenous EBR application increased the activity or the
and that the oxidative stress induced in partially TRX-f, TRX-m2, TRX-m1/4, and TRX-γ-silenced plants is at least in part attributable to the reduced activity of the antioxidant enzymes. The finding that the cellular glutathione redox status could be regulated by chloroplastic TRXs further revealed the existence of cross-talk between the TRX system and the glutathione system.

In addition to the AsA–GSH cycle, peroxiredoxin, which is activated by TRX, acts as an alternative pathway to remove H₂O₂ from chloroplasts (Foyer and Shigeoka, 2011). The 2-CP enzyme exhibits broad substrate specificity, showing activity toward both hydrogen peroxides and complex alkyl hydroperoxides. During the peroxide reduction reaction, 2-CP is alternatively oxidized and reduced as it catalyses the electron flow from an electron donor to peroxide. In vitro tests have demonstrated that TRX-x is by far the most efficient in Arabidopsis (Collin et al., 2003). However, only the oxidized dimeric form of 2-CP was detected in the Ostrxm RNAi plants, suggesting that Ostrxm could also regulate the catalytic activity of 2-CP by reducing redox-active cysteine residues in plants (Chi et al., 2008). The present study revealed increased accumulation of oxidized 2-CP in all of the partially TRX-silenced plants, with the partial silencing of TRX-x, TRX-m1/4, and TRX-m2 being more significant, suggesting that these TRXs could differentially regulate 2-CP (Fig. 4b, c).

TRX-f and TRX-m1/4 play important roles in BRs-induced changes in cellular redox homeostasis

As observed in previous research, foliar application of EBR relieved oxidative stress (Xia et al., 2009b). In addition to increasing the activity of the AsA–GSH cycle, EBR also
induced an increase in the ratio of the reduced monomers to the oxidized dimers of 2-CP (Fig. 4c). Importantl, par-
tially silencing TRX-f and TRX-m1/4 compromised the EBR-
induced decrease in MDA content and increase in the activity
of antioxidant enzymes, as well as the associated changes in
 glutathione metabolism (Figs 4a, 5a, b), suggesting that EBR
partially alleviates oxidative stress by regulating TRX-
f and TRX-
m1/4.

It was found that the TRXs are actively involved in the
regulation of photosynthesis-related gene transcripts, enzyme
activity, and, ultimately, the CO2 assimilation capacity. Ostrxm
plants exhibit abnormal chloroplast development and growth
inhibition in rice plants, displaying decreased levels of several
chloroplast proteins that are critical for photosynthesis and
biogenesis (Chi et al., 2008). Silencing TRX-f and TRX-m
(VIGS-TRX-f/TRX-m) in pea plants results in a significant
reduction in Mg chelatase activity and the 5-aminolaevulinic
acid synthesizing capacity as well as decreased transcript
levels of RBCS and chlorophyll biosynthesis-related genes
and a decreased photosynthetic capacity (Luo et al., 2012).
In the present study, it was found that partially silencing the
chloroplastic TRXs, except for TRX-x, resulted in a significant
reduction in photosynthetic capacity in tomato leaves (Fig. 3), supporting
the hypothesis that TRX-f, TRX-m2, TRX-m1/4, and TRX-
y are actively involved in the regulation of CO2 assimilation.

In Arabidopsis, the maximal (Fv/Fm) and effective quantum
yields of photosystem II (PSII; ΦPSII) in the trx f1.1 and trx
f1.2 mutant plants are not different from those in wild-type
plants (Thormählen et al., 2013). In agreement with this find-
ing, it was observed that only the pTRV-TRX-m1/4 plants
showed a decrease in Fv/Fm, by 15.4% (Supplementary Fig.
S2 available at JXB online). All of these results suggested that
the TRXs do not affect CO2 assimilation by modulating PSII
electron transport.

Fig. 6. Changes in the expression of Benson- Calvin cycle-related genes for control and EBR-treated virus-induced gene silencing (VIGS) plants. Leaf
samples were taken at 24 h after EBR treatment. Data are the means of four replicates with SDs. Means followed by the same letter are not significantly
different according to Tukey's test (P<0.05).
TRX-f and TRX-m1/4 play important roles in BRs-induced changes in CO₂ assimilation

Previous studies have shown that chloroplastic TRXs participate in the regulation of the Calvin cycle and associated processes (Balmer et al., 2003). In addition to the down-regulation of genes involved in the Benson-Calvin cycle, partially silencing TRX-f, TRX-m2, TRX-m1/4, and TRX-y also resulted in a significant decrease in the initial Rubisco and FBPase activities as well as RCA activity (Fig. 7). An in vitro assay revealed that AtTRX-f is able to activate FBPase and NADP-MDH most efficiently, followed by AtTRX-m1, AtTRX-m2, and AtTRX-m4, whereas AtTRX-m3, AtTRX-x, and AtTRX-y cannot efficiently activate NADP-MDH from sorghum or from Chlamydomonas (Collin et al., 2003, 2004; Lemaire et al., 2003, 2005). The simultaneous decrease in ρVc,max and initial Rubisco activity, with little change in total Rubisco activity, indicates that chloroplastic TRX-f, TRX-m2, TRX-m1/4, and TRX-y mainly regulate the activation state of Rubisco via the action of Rubisco activase (Figs 3b, 7). The regeneration of RuBP is dependent on both the photosynthetic electron transport chain and the enzymes downstream of Rubisco in the Calvin cycle (Long et al., 2006). In addition to the decrease in RCA activity, initial FBPase activity was also decreased in the plants in which TRX-f, TRX-m2, TRX-m1/4, and TRX-y were partially silenced (Fig. 7). Accordingly, the lack of direct activation of RCA or FBPase was involved in the decrease in Jmax. Interestingly, the observed changes in the activities of these enzymes were in agreement with cellular glutathione homeostasis (Fig. 5b). In a previous study, it was found that several redox-sensitive Benson-Calvin cycle enzymes, such as RCA, can undergo disulfide bond interchange, leading to changes in the activities of these enzymes (Jiang et al., 2012b). It is likely that the oxidized cellular environment induced by the partial silencing of TRX-f, TRX-m2, TRX-m1/4, and TRX-y can directly modify the structure of these redox-sensitive enzymes.

Previously, it was found that inhibition of the biosynthesis of BRs using an inhibitor decreased CO₂ assimilation, while exogenously applied BRs increased CO₂ assimilation in cucumber plants (Yu et al., 2004; Xia et al., 2009a). Here, it was demonstrated that exogenous EBR application resulted in enhanced CO₂ assimilation in tomato plants, while BRs-deficient dmm mutant plants exhibited decreased CO₂ assimilation compared with wild-type CR plants (Fig. 2b), providing further evidence that BRs levels are closely related to the CO₂ assimilation capacity of plants.

BRs can differentially induce TRX transcripts in Arabidopsis and rice (Müssig et al., 2002; Goda et al., 2004; Wu et al., 2008). In the present study, it was found that BRs were able to up-regulate TRX-f, TRX-m2, TRX-m1/4, and TRX-x but
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Figure S1. Relative mRNA abundance of TRX-f, TRX-m2, TRX-m1/4, TRX-x, and TRX-y in respective gene silencing (VIGS) plants.

Figure S2. Changes in maximum quantum yield of PSII (Fv/Fm) in the leaves of pTRV and various partially TRX-silenced plants.

Table S1. PCR primers designed for vector construction.
Table S2. Gene-specific primers designed for qRT-PCR.

Acknowledgements

We are grateful to Professor C.H. Foyer for her critical suggestions regarding this study. This work was supported by the National Basic Research Program of China (2009CB119000).

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