Barley has two peroxisomal ABC transporters with multiple functions in β-oxidation

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Abstract

In oilseed plants, peroxisomal β-oxidation functions not only in lipid catabolism but also in jasmonate biosynthesis and metabolism of pro-auxins. Subfamily D ATP-binding cassette (ABC) transporters mediate import of β-oxidation substrates into the peroxisome, and the Arabidopsis ABCD protein, COMATOSE (CTS), is essential for this function. Here, the roles of peroxisomal ABCD transporters were investigated in barley, where the main storage compound is starch. Barley has two CTS homologues, designated HvABCD1 and HvABCD2, which are widely expressed and present in embryo and aleurone tissues during germination. Suppression of both genes in barley RNA interference (RNAi) lines indicated roles in metabolism of 2,4-dichlorophenoxybutyrate (2,4-DB) and indole butyric acid (IBA), jasmonate biosynthesis, and determination of grain size. Transformation of the Arabidopsis cts-1 null mutant with HvABCD1 and HvABCD2 confirmed these findings. HvABCD2 partially or completely complemented all tested phenotypes of cts-1. In contrast, HvABCD1 failed to complement the germination and establishment phenotypes of cts-1 but increased the sensitivity of hypocotyls to 100 μM IBA and partially complemented the seed size phenotype. HvABCD1 also partially complemented the yeast pxa1/pxa2Δ mutant for fatty acid β-oxidation. It is concluded that the core biochemical functions of peroxisomal ABC transporters are largely conserved between oilseeds and cereals but that their physiological roles and importance may differ.

Key words: Aleurone, gene duplication, germination, indole butyric acid, oil body, seed size.

Introduction

The peroxisome is the sole site of β-oxidation of fatty acids and related molecules in plants and fungi, and is required for very long chain fatty acid metabolism and signalling in mammals (Baker et al., 2006; Poirier et al., 2006; Wanders et al., 2006).
In plants, β-oxidation is important not only during germination, seedling establishment, fertilization, and dark-induced senescence, but also in a number of additional key roles (reviewed in Theodoulou and Eastmond, 2012; Linka and Theodoulou, 2013). These include biosynthesis of the hormones jasmonic acid (JA) and indole acetic acid (IAA), production of volatile benzenoids and benzoylloxyalkylglucosinolates, and also salicylic acid biosynthesis (Reumann et al., 2004; Kleibeinstein et al., 2007; Gfeller et al., 2010; Strader et al., 2011; Klemplien et al., 2012; Lee et al., 2012; Qually et al., 2012; Bussell et al., 2014).

Transport of β-oxidation substrates into the peroxisome is mediated by ATP-binding cassette (ABC) proteins belonging to subfamily D (Theodoulou et al., 2006). In mammals and fungi, peroxisomal ABCD transporters are expressed as 'half-size' proteins, each containing a transmembrane domain (TMD) and a nucleotide-binding domain (NBD), which homo- or heterodimerize to form a functional transporter (Theodoulou et al., 2006). Plants are unusual in having 'full-size' peroxisomal ABCD transporters, in which two dissimilar domains are expressed as a single polypeptide with 'full-size' peroxisomal ABCD transporters, in which two dissimilar domains are expressed as a single polypeptide with the structure TMD–NBD–TMD–NBD, which is considered to represent a fused heterodimer (Verrier et al., 2008). The prototypical plant member of the ABCD subfamily is the Arabidopsis thaliana protein, COMATOSE (CTS; also known as AtPXA1, PED3, ACN2, AtABCD1; Arabidopsis Genome Initiative, 2000). The prototypical plant member of the ABCD subfamily is the Arabidopsis thaliana protein, COMATOSE (CTS; also known as AtPXA1, PED3, ACN2, AtABCD1; Arabidopsis Genome Initiative, 2000).

CTS is thought to mediate OPDA transport into the peroxisome, since null mutants are JA deficient, but a second, minor import pathway also exists, as cts mutants are not male sterile (Theodoulou et al., 2005). It is possible that OPDA could enter the peroxisome by passive diffusion and be subject to vectorial acylation involving peroxisomal OPDA:CoA ligase (Kienow et al., 2008), though an as yet unidentified transporter cannot be ruled out.

Alleles of cts have been isolated in screens for resistance to the natural auxin, indole-3-butyril (IBA) and the artificial pro-auxin, 2,4-dichlorophenoxybutyric acid (2,4-DB) (pzx-1, Zolman et al., 2001; ped3 series, Hayashi et al., 1998, 2002). These compounds are metabolized by one round of β-oxidation to generate IAA and 2,4-dichlorophenoxyacetic acid (2,4-D), respectively, which inhibit root and hypocotyl growth (Hayashi et al., 1998; Zolman et al., 2001; Rashotte et al., 2003). This implies that CTS can transport these aromatic compounds or their CoA esters, although this has not been tested experimentally. Finally, a cts allele (acn2) was shown to be resistant to fluorooacetate, implying a role in acetate metabolism (Hooks et al., 2007). Thus, it appears that plants have apparently evolved a broad specificity peroxisomal ABC transporter to mediate the import of the wide range of substrates that they need to process by β-oxidation (Baker et al., 2006). This is in contrast to mammalian and yeast peroxisomal ABC transporters which exhibit restricted substrate selectivities as judged by cross-kingdom complementation experiments and characterization in yeast and in vivo (van Roermund et al., 2008, 2011, 2014; Zhang et al., 2011).

With the exception of a study implicating a CTS homologue in the control of seed size in tomato (Orsi and Tanksley, 2009), ABCD proteins have not been investigated in plant species other than Arabidopsis, and it remains to be determined to what extent the functions identified thus far are shared with other taxa and whether their relative importance differs. Arabidopsis and other oilseeds utilize oil stored primarily in cotyledons as the main source of energy during seedling establishment. In contrast, in cereals such as barley, starch stored in the endosperm fulfils this role, although the embryo, scutellum, and aleurone contain significant amounts of stored lipid (Jones and Jacobsen, 1991). Tissue-specific transcriptome analysis has shown that genes of the β-oxidation pathway are induced in both endosperm/aleurone and embryo from 24 h after imbibition of barley grains (Sreenivasulu et al., 2008), and enzymes of β-oxidation are present in both tissues. Gibberellin acid (GA) stimulates the breakdown of oil reserves and their conversion to sugar by gluconeogenesis in cereal aleurone (Doig et al., 1975; Newman and Briggs, 1976; Fernandez and Staehelin, 1987), although sensitivity to GA is cultivar dependent (Eastmond and Jones, 2005). It has been proposed that mobilization of stored triacylglycerol (TAG) from aleurone tissue provides energy and carbon skeletons for the synthesis of α-amylose that is needed to mobilize starch. An alternative hypothesis is that sucrose provided by gluconeogenesis in the aleurone could be transferred to the growing embryo via the scutellum, to support embryo growth (Eastmond and Jones, 2005). Although the embryo and scutellum are rich in oil, these tissues lack glyoxylate cycle enzyme activities (Holtman et al., 1994), and thus may respire fatty acids, rather than use them.
for gluconeogenesis (Eastmond and Jones, 2005). In conclusion, β-oxidation might play key roles in barley germination, an important trait in agriculture and malting.

In this study, the roles of peroxisomal ABC transporters in a model cereal, barley (*Hordeum vulgare* L.), were investigated and experiments were carried out to test whether they perform similar functions to those in oilseeds such as *Arabidopsis*. The isolation of two barley CTS homologues, *HvABCD1* and *HvABCD2*, is reported and their collective roles are analysed using RNA interference (RNAi). In parallel, the individual functions of *HvABCD1* and *HvABCD2* were investigated by testing their ability to complement the *Arabidopsis cts-1* mutant and the yeast *paxil/paxΔ* mutant which lacks the homologous transporter. Together, these approaches enabled the physiological roles of barley peroxisomal ABC transporters to be probed and the contributions of the two different genes to different known biochemical functions to be assessed.

It is concluded that the general capabilities of peroxisomal ABC transporters of *Arabidopsis* and barley are similar but that the two paralogues in barley may play distinct roles.

**Materials and methods**

**Plant material and growth conditions**

Barley, *Hordeum vulgare* L. var. Golden Promise, was grown under controlled conditions of 15 °C/12 °C (day/night) and a 16 h photoperiod [80% relative humidity, 500 μmol m−2 s−1 metal halide lamps (HQI) supplemented with tungsten bulbs]. Seeds were sown in 5 litre pots containing Levington’s C2 compost (http://www.everris.com/uk/Home/Ornamental-Horticulture/Products/Product.aspx/Professional-Growing-Media/Potting-and-Pot-Plant-Compost-/) (kind gift of Dr Ranjan Swarup, University of Nottingham) and leaf plants (var. Golden Promise) were generated by the BRACT transformation facility (John Innes Centre, Norwich, UK), using *Agrobacterium*-mediated transformation of immature embryos, according to the method of Harwood et al. (2009). Copy number and zygosity determination were carried out by DNA GENETICS (Norwich, UK).

**Identification and cloning of HvABCD1 and HvABCD2**

Analysis of the rice genome and barley expressed sequence tag (EST) sequences revealed two *CTS* homologues in grasses. The barley genes were designated *HvABCD1* and *HvABCD2*, according to nomenclature conventions prescribed in Verrier et al. (2008). *HvABCD1* was amplified from barley cDNA using primer pair CTS-a startF/Hv14973R2 (Supplementary Table S1 at JXB online) and blunt-end cloned into pBluescript, to give pCTS1/BS. Plasmids for the expression of *HvABCD1* and *HvABCD2* under the control of the native CTS promoter were constructed as follows: first, the promoter region of *AtCTS* was amplified from pG0229-T ProGFp (Footitt et al., 2007a) with primer pair AtCTS5’/NcoI/AtCTS3’/KpnI, excised with NcoI/KpnI, and cloned in the corresponding sites of pENTR11 (Invitrogen), to give pE-AtCTS5. The open reading frame (ORF) of *HvABCD1* was amplified from pCTS1/BS using primer pair HvCTS5’/HvCTS3’/KpnI and cloned into the corresponding sites of pENTR11 (Invitrogen), to give pE-HvCTS5. Amplification was conducted using the primer pairs HvABCD1 F/R and HvABCD2 F/R; cycle conditions: 96 °C 2 min; 30 cycles of (94 °C 30 s, 55 °C 30 s, 72 °C 30 s); 72 °C 7 min. Constitutively expressed *α-tubulin* was used as a loading control (Nicot et al., 2005; Jarošová and Kundu, 2010) and PM19 was used as a developmental control (Randolf et al., 2002). Amplifications were conducted using the primer pairs α TUB F/R [cycle conditions: 96 °C 2 min; 25 cycles of (94 °C 30 s, 58 °C 30 s, 72 °C 30 s); 72 °C 7 min] and PM19 F/R [cycle conditions: 96 °C 2 min; 30 cycles of (94 °C 30 s, 55 °C 30 s, 72 °C 30 s)] respectively.

**Generation of RNAi lines**

A 497bp fragment of *HvABCD2* with high sequence identity to *HvABCD1* (Supplementary Fig. S1 at JXB online) was amplified from cDNA using primers HvABCDNR1F and HvABCDNR1R, and recombinated sequentially into pGWC (kind gift of Dr Ranjan Swarup, University of Nottingham) and pBrac1207, via Gateway cloning (Invitrogen). Transgenic barley plants (var. Golden Promise) were generated by the BRACt transformation facility (John Innes Centre, Norwich, UK), using *Agrobacterium*-mediated transformation of immature embryos, according to the method of Harwood et al. (2009). Copy number and zygosity determination were carried out by DNA GENETICS (Norwich, UK).

**Analysis of RNAi lines**

Quantitative real-time PCR (Q-PCR). RNA was extracted from seedling leaf material (Zadoks code 15, which corresponds to five visible leaves), using a Nucleospin RNA plant mini-kit (Macherey-Nagel, Germany). Leaf material (70–100 mg) was ground to a powder in liquid N2 and resuspended in 400 μl of RA1 extraction buffer, containing 1% (w/v) PVP-40. Following clarification by centrifugation, the supernatant was processed according to the manufacturer’s instructions. Poly(dT) cDNA was prepared from 1 μg of total RNA with SuperScript III reverse transcriptase (Invitrogen), using primers as specified in Supplementary Table S1 at JXB online. Quantitative PCR was performed using SYBR-green Sensimix (Biorne), in 384-well optical reaction plates with a Roche LightCycler 480 apparatus (Roche/Applied Biosystems). PCR conditions and quantification were as specified by the manufacturer (Roche). The relative number of copies obtained for each transcript was normalized against *HvELF1* and *HvTubulin* (Nicot et al., 2005; Jarošová and Kundu, 2010) or *AtCTL3* and *AtCTL1* (De Rybel et al., 2010) transcript values for each sample as an internal reference.

**Genetic mapping**

The cDNA sequences of *HvABCD1* and *HvABCD2* were BLASTed against the recently released sequence assembly of the barley cv. Morex genome (IBSC, 2012) identifying corresponding genomic sequence contigs (*HvABCD1*=morex_contig_47855, *HvABCD2*=morex_contig_365253). Both contigs have been integrated into a genetically ordered sequence context using POPSEQ (Mascher et al., 2013) in two biparental segregating populations: the reference *Morex* × *Barke* recombinant inbred line (RIL) population and OWB doubled haploid population (IBSC 2012). Map positions are related to the positions of the barley *Select* markers as reported in Comadran et al (2012).

**Reverse transcription–PCR (RT–PCR)**

For the experiment presented in Fig. 1A, embryos and aleurone were isolated from barley grains and incubated in water for different periods (0, 4, 12, 24, 48, and 72 h). RNA was extracted from embryos using a modified TRIZOL extraction protocol, and aleurone tissue RNA was extracted according to Singh et al. (2003). Poly(dT) cDNA was prepared from 2 μg of total RNA with SuperScript III reverse transcriptase (Invitrogen). Amplification was conducted using the primer pairs HvABCD1 F/R and HvABCD2 F/R; cycle conditions: 96 °C 2 min; 30 cycles of (94 °C 30 s, 55 °C 30 s, 72 °C 30 s); 72 °C 7 min. Constitutively expressed *α-tubulin* was used as a loading control (Nicot et al., 2005; Jarošová and Kundu, 2010) and PM19 was used as a developmental control (Randolf et al., 2002). Amplifications were conducted using the primer pairs α TUB F/R [cycle conditions: 96 °C 2 min; 25 cycles of (94 °C 30 s, 58 °C 30 s, 72 °C 30 s); 72 °C 7 min] and PM19 F/R [cycle conditions: 96 °C 2 min; 30 cycles of (94 °C 30 s, 55 °C 30 s, 72 °C 30 s)] respectively.

**Figure 1A**

Expression of *HvABCD1* and *HvABCD2* in different embryonic stages of barley. A, Embryos (var. Golden Promise) were isolated from barley grains and incubated in water for different periods (0, 4, 12, 24, 48, and 72 h). Expression was detected using reverse transcription–PCR (RT-PCR) using specific primers for *HvABCD1* and *HvABCD2*.

**Figure 1B**

Expression of *HvABCD1* and *HvABCD2* in different aleurone and endosperm stages of barley. B, Aleurone and endosperm (var. Golden Promise) were isolated from barley grains and incubated in water for different periods (0, 4, 12, 24, 48, and 72 h). Expression was detected using reverse transcription–PCR (RT-PCR) using specific primers for *HvABCD1* and *HvABCD2*.
Seed size measurement Five samples of 50 grains from homozygous HvABCD1/2 RNAi lines and their respective null segregants were weighed. Seeds were photographed using a camera attached to a light microscope. Length and width were determined from micrographs, following correction for magnification.

Germination assays Fifty grains were placed in Petri dishes containing two layers of Whatman No. 2 filter paper and 4 ml of water. The dishes were sealed with Micropore tape and incubated at 22 °C under continuous white light (150 μmol m⁻² s⁻¹). Germinated caryopses, defined by the emergence of coleorhizae beyond the seed coats, were scored every 24 h over 7 d and removed from the dishes. Assays were performed in triplicate.

Root growth assays Grains from homozygous HvABCD1/2 RNAi lines and their respective null segregants were plated on 0.5× Murashige and Skoog (MS) medium with 0.5% (w/v) sucrose containing the indicated concentrations of 2,4-DB or IBA, or no supplements. Experiments were carried out in triplicate with four grains per replicate.

JA treatment Seedlings of HvABCD1/2 RNAi lines were grown in a growth chamber (20 °C day/15 °C night, 70% relative humidity, 16 h light). Ten-day-old plants were sprayed with 2 ml of methyl jasmonate (Sigma-Aldrich, Germany) at 2 mg ml⁻¹ in water. After 48 h, the treatment was repeated and leaf material was sampled 72 h after the first treatment. RNA extraction and Q-PCR were performed as described above.

Scanning electron microscopy Intact grains were imbibed in sterile distilled water (SDW) for 2–5 d, on moistened filter paper sealed in Petri dishes and placed in the dark at room temperature. Following imbibition, root and shoot were removed if present and the remaining tissue was mounted onto cryo-electron microscope stubs using OCT compound (Agar Scientific UK) and plunge-frozen in pre-slushed liquid N₂. They were then transferred under vacuum to the Alto 2500 (Gatan UK) cryo chamber stage which was pre-cooled to –180 °C. Here they were fractured using the cold blade mounted in the chamber and gentle etching was performed through sublimation by raising the temperature of the stage to –95 °C for 2 min. The stage temperature was returned to
-140 °C and the samples were sputter-coated with AuPd for 60 s to a thickness of ~10 nm. Samples were transferred to the JEOL JSM 6700 scanning electron microscope (SEM) chamber and mounted on the stage cooled to -150 °C, for examination. This temperature was maintained during examination of the fractured surface, and images were recorded using the on-board system and software.

Confocal microscopy
Grains were de-husked and de-embryonated using a sterile scalpel before sterilizing by immersion in 20% (v/v) sodium hypochlorite for 10 min. The samples were thoroughly washed for 1 min under a flowing stream of SDW. Each grain was placed in a 1.5 ml Eppendorf tube and covered with ~1 ml of 1 μM GA3, and left to imbibe at room temperature for 8-96 h. At the required time points, grains were washed in SDW and dried on filter paper to remove excess water. Grains were individually mounted onto cryostat holders using Tissue-Tek, OCT compound (Agar Scientific UK) and were quickly plunged into liquid N2. When bubbling ceased, the frozen samples were transferred to the chamber of the Leica 1850 Cryostat (Leica Microsystems UK) and left for 30 min to allow the temperature to equilibrate to the chamber temperature of -20 °C before sectioning. Sections were removed from the sample and discarded until the mid-region was reached ~1.6 mm from the tip. The following 10 sections, each 16 μm thick, were collected on glass slides for staining. Sections were stained using Nile Red at 1 μl ml-1 for 1 min, followed by washing with SDW and sealing with a cover slip. Sections were imaged using a Zeiss LSM 780 system, with laser 514 nm selected and emission collected between 539 nm and 753 nm.

Arabidopsis complementation
Wild-type (Ler) and cts-1 plants were transformed with pAtCTS::HvABCD1 and pAtCTS::HvABCD2 by floral dip. Lines expressing HvABCD2 in the cts-1 background were obtained by selecting cts-1 transformants which germinated on 0.5× MS medium and confirming genotype by selection and PCR analysis. As HvABCD1 did not complement the germination phenotype of cts-1 (see the Results), transgenic lines were obtained in the Ler background, crossed to cts-1 and a homozygous line identified by PCR genotyping. It was not possible to produce multiple independent homozygous lines for HvABCD1. Assays for germination, establishment, and hypocotyl growth were carried out as described in Dietrich et al. (2009). Seed size was determined as described above for barley (30 seeds per line). The effect of OPDA on root growth was determined as described in Zhang and Turner (2008).

Yeast complementation
HvABCD1 was amplified with primer pair FT206/FT207, restricted with KpnI/PstI, and cloned into the corresponding sites of pEL30 and pIJL30. The inserts were sequenced to confirm the absence of PCR-generated mutations. Yeast transformation and oleate growth tests were carried out as described in Nyathi et al. (2010). β-Oxidation measurements were carried out as described in van Roermund et al. (1995); in brief, 1-14C-labelled fatty acids were supplied to intact cells followed by quantification of 14CO2 and 14C-labelled β-oxidation products by liquid scintillation counting.

Results
Barley has two CTS homologues which are expressed in embryo and endosperm
Comparative analysis of sequenced plant genomes has revealed that cereals contain two CTS homologues, consistent with a gene duplication occurring after divergence of the Gramineae family (Nyathi et al., 2012). Full-length cDNAs corresponding to the barley CTS homologues were isolated by RT-PCR and designated HvABCD1 and HvABCD2, according to the naming convention outlined in Verrier et al. (2008). HvABCD1 and HvABCD2 were located on the barley genome by BLASTing the cDNA sequence against a sequence assembly of the Morex genome that had been genetically ordered using the POPSEQ approach (Mascher et al., 2013). HvABCD1 was located at 148.4 cM on chromosome 3H on the Morex×Barke map. HvABCD2 was located at 8.9 cM on chromosome 1H on the reference Morex×Barke RIL population map and 11.2 cM on 1H on the Oregon Wolfe barley doubled haploid map (International Barley Sequencing Consortium, 2012). The orthologous genes in rice mapped to syntonic positions on the corresponding chromosomes.

Examination of HvABCD1 and HvABCD2 expression profiles using the Bio-Array Resource barley eFP browser (Patel et al., 2012) revealed that both genes were widely expressed in different tissues, with HvABCD2 transcripts being generally more abundant (Supplementary Fig. S2 at JXB online). The expression of HvABCD1 and HvABCD2 was then investigated in more detail in germinating barley grains over 3 d imbibition, using RT-PCR. PM19, which encodes a putative plasma membrane transporter, was used as a developmental control. Consistent with previous reports (Ranford et al., 2002), transcripts were present in unimbibed seeds but declined upon germination (Fig. 1A). This confirmed the developmental status of the grains, since dormant embryos have been shown to retain high levels of PM19 message for up to 72 h of imbibition (Ranford et al., 2002). HvABCD1 transcripts were barely detectable in dry grains but were present in embryos at a similar level from 4 h to 72 h imbibition. HvABCD2 expression in embryos increased steadily over this period (Fig. 1A). Both genes were expressed transiently in aleurone tissue at 48 h imbibition, corresponding to a time point around germination. This suggests that β-oxidation could be important for embryo and aleurone function during germination and seedling establishment in barley.

Suppression of HvABCD1 and HvABCD2 affects grain size but not germination
In order to understand the function of CTS homologues in barley growth and development, expression of both HvABCD genes was reduced simultaneously using an RNAi approach. barley (cv. Golden Promise) was transformed with a construct designed to suppress both HvABCD transcripts (Supplementary Fig. S1 at JXB online), driven by the constitutive Ubi1 promoter (Harwood et al., 2009). Several HvABCD1/2 RNAi lines (hereafter referred to as HvABCD1/2i lines) were obtained and, for each one, a null segregant line was also selected for use as a control. Expression levels were analysed by Q-PCR analysis of leaf material. Abundance of HvABCD1 and HvABCD2 transcripts varied from 22% to 59% and 22% to 40% of levels measured in corresponding null segregant lines, respectively (Fig. 1B).

Since natural variation in seed size has been associated with ABCD transporter genes in tomato and Arabidopsis, and cts
alleles have small seeds (Russell, 1998; Alonso-Blanco et al., 1999; Orsi and Tanksley, 2009), it was investigated whether suppression of HvABCD1 and HvABCD2 affected grain size in barley. Grains of three different RNAi lines exhibited significantly reduced length, width, and weight, compared with null segregants (Fig. 1C–E). The germination of seeds from several HvABCD1/2i lines and their respective nulls was also measured over 7 d. Although the percentage germination of null lines at 7 d was somewhat variable, in none of the RNAi lines tested was germination markedly different from that of the respective null segregant (Fig. 1F; Supplementary Fig. S3 at JXB online). This is in agreement with results from tomato, where ABCD function does not appear to affect the rate of germination (Orsi and Tanksley, 2009), but in contrast to Arabidopsis, in which cts mutants are arrested late in phase II of germination (Russell et al., 2000; Footitt et al., 2002, 2006; Carrera et al., 2007).

**Suppression of HvABCD1 and HvABCD2 does not markedly affect seedling establishment and lipid mobilization**

In Arabidopsis, peroxisomal β-oxidation plays a critical role in mobilization of storage reserves during seedling establishment (Theodoulou and Eastmond, 2012). However, seedling establishment was not visibly affected in HvABCD1/2i lines, which is perhaps unsurprising, given that barley contains abundant reserves of starch in the endosperm. Nevertheless, since mobilization of TAG stored in the cereal aleurone has been suggested to facilitate the exploitation of endosperm starch and/or to support embryo growth (Eastmond and Jones, 2005), oil body morphology was examined over 5 d imbibition. After 2 d imbibition, scanning electron microscopy revealed that aleurone cells of wild-type cells contain numerous protein storage vacuoles (PSVs), surrounded by oil bodies (Fig. 2A–C). The oil body membrane is continuous with the PSV membrane, as previously reported (Fernandez and Staehelin, 1987); this is particularly evident where freeze-fracture has generated images showing extracellular (E)-faces with ‘scars’ where oil bodies have been removed along with PSVs (Fig. 2C). After 5 d, numerous oil bodies were still visible, though they appeared less spherical (Fig. 2D–F). It was also possible to visualize aleurone oil bodies by confocal microscopy using Nile Red staining of TAG in unfixed cryo-sections of de-embryonated grains treated with GA (Fig. 2G–N). After 5 d, the abundance of oil bodies had declined (Fig. 2K–N), in agreement with reports that GA stimulates lipid mobilization in this tissue (Fernandez and Staehelin, 1987; Eastmond and Jones, 2005). Since it was not possible to derive quantitative data from these images, fatty acids were quantified in de-embryonated, GA-treated barley grains, but were not found to be significantly different in RNAi and null segregant lines (data not shown). HvABCD1 and HvABCD2 differ in their ability to complement different phenotypes of Arabidopsis cts-1

While analysis of the barley HvABCD1/2i lines permitted assignment of several different physiological roles to peroxisomal β-oxidation (Figs 1–4), this approach does not provide information about the relative contributions of HvABCD1 and HvABCD2 to these processes. In order to probe their individual functions, HvABCD1 and HvABCD2 were expressed in the Arabidopsis cts-1 null mutant under the control of the native CTS promoter, as has been carried out previously for site-directed cts mutants (Dietrich et al., 2009). Two homozygous lines (D2.1 and D2.2) expressing HvABCD2 were obtained following transformation of the cts-1 mutant. However, it was not possible to recover lines for HvABCD1 by this method, as seeds were unable to germinate. Therefore, a line was established in the wild-type background (Ler) and crossed to cts-1. A homozygous cts-1 line expressing HvABCD1 (D1.1) was recovered by mechanically disrupting the seed coat and culturing on sucrose-containing medium.
All lines produced transcripts, as measured by Q-PCR analysis of seedlings; however, expression of HvABCD1 was low relative to the expression level of endogenous CTS (Supplementary Fig. S4 at JXB online).

Seed size was first measured in different genotypes. cts-1 seeds were smaller than those of Ler plants produced under identical conditions, being on average 88% of the length and 84% of the width of the wild type (Fig. 5A, B). This was accompanied by a reduction in average seed weight to 85% of the wild type, although this difference was not statistically significantly different (Fig. 5C). In cts-1 lines expressing HvABCD2, seed length and weight were restored to wild-type values, and seed width was significantly increased relative to cts-1 but still significantly different from Ler. Complementation was less complete in the line expressing HvABCD1 (Fig. 5A–C). Germination kinetics of lines D2.1 and D2.2 were indistinguishable from those of the wild type, whereas line D1.1 did not germinate, in agreement with the inability to recover transgenic lines via direct transformation of the cts-1 mutant (Fig. 5D). Thus HvABCD2 but not

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**Fig. 2.** Oil body morphology in barley grains. (A–F) SEM cryo-fractured images of wild-type (var. ‘Golden Promise’) barley aleurone cells. (A–C) Two day imbibed grains; (D, E), 5 d imbibed grains. (A, D) Aleurone cells, showing nucleus (n) and numerous protein storage vacuoles (psv). (B, E) PSV, surrounded by oil bodies (OBs), indicated by arrowheads. (C) PSV fractured across, surrounded by OBs; the arrowhead indicates the extracellular face (‘E-face’) where PSV and OBs have been removed during fracture. (F) Close-up of (E), showing connections between OBs and PSV. OBs are indicated by arrowheads. (G–N) Confocal images of unfixed barely grain sections showing Nile Red staining of OBs in aleurone cells. De-embryonated grains were imbibed for 24 h (G–J) or 96 h (K–N) in the presence of 1 μM GA. (G, H, K, L) HvABCD1/2 RNAi line 5; (I, J, M, N) null segregant. In G, I, K and M, the confocal image is overlaid on the bright field image. Scale bars: A=5 μm, B, C, E=1 μm; D=4 μm; F=400 nm; G–N=50 μm.
HvABCD1 is able to complement cts-1 germination. Since CTS performs distinct biochemical functions in germination and seedling establishment, the ability of transgenic lines to establish in the absence of sucrose was then tested. Sterilized seeds were plated on 0.5× MS medium, chilled for 2 d, and transferred to a growth chamber. Lines D2.1 and D2.2 produced green cotyledons after 5 d in constant light and were able to extend hypocotyls in the dark (Fig. 5E, F), although line D2.2 did not grow as well as the wild type (Fig. 5E). Seeds of line D1.1 were induced to germinate as described above and then transferred to medium lacking sucrose, but were unable to mobilize their storage lipid, as indicated by their inability to expand cotyledons and hypocotyls. However, development appeared normal in the presence of sucrose, indicating that there were no obvious deleterious effects of the transgenes (Fig. 5E, F).

Responses of the transgenic lines to hormones were also investigated. Seeds were induced to germinate and the effect of IBA on hypocotyl length was tested in sucrose-containing medium. As hypocotyls are less sensitive to IBA than roots (Rashotte et al., 2003), this assay is less influenced by transporter expression levels (discussed in Dietrich et al., 2009). Under these conditions, Ler exhibited a dose-dependent inhibition of hypocotyl elongation, whereas cts-1 was largely resistant up to 100 μM IBA (Fig. 5G). Partial sensitivity to 100 μM IBA was restored in cts-1 lines expressing either HvABCD1 or HvABCD2, indicating that both transporters can contribute to auxin metabolism, as suggested by the barley RNAi lines. The effect of the jasmonate precursor OPDA on root elongation was also tested (following rescue of germination for cts-1 and cts-1 pAtCTS:HvABCD1 seeds, as described above). OPDA inhibited elongation of wild-type (Ler) roots, but the opr3 mutant was resistant since it lacks 12-oxophytodienoic acid reductase and is unable to convert OPDA to JA (Stintzi and Browse, 2000; Zhang and Turner, 2008; Fig. 5H). cts-1 roots were shorter than those
of Ler under control conditions, but were also resistant to OPDA, consistent with the fact that this mutant has been shown to be impaired in JA production (Theodoulou et al., 2005). OPDA sensitivity was restored in roots of cts-1 lines expressing HvABCD2 but not in the line expressing HvABCD1.

HvABCD1 partially complements the yeast pxa1/pxa2A mutant for fatty acid β-oxidation

Since HvABCD1 did not complement the seedling establishment phenotype of cts-1, a role for this transporter in fatty acid metabolism was investigated further by expression in a Saccharomyces cerevisiae mutant which lacks Pxa1p/Pxa2p, the yeast heterodimeric transporter which is homologous to CTS. pxa1/pxa2Δ is defective in growth on oleate (18:1) as a sole carbon source, but expression of HvABCD1 under the control of the yeast catalase promoter restored growth to near wild-type levels (data not shown). pxa1/pxa2Δ cells transformed with HvABCD1 exhibited increased β-oxidation of several different long chain fatty acids compared with cells transformed with vector lacking an insert, indicating that the barley transporter can accept saturated and unsaturated long chain fatty acyl-CoAs in this heterologous system (Fig. 6). Expression of HvABCD2 was also attempted using a similar strategy, but no complementation of the pxa1/2Δ mutant was observed (data not shown).

Discussion

Using two complementary approaches, evidence has been provided that barley and Arabidopsis peroxisomal ABC transporters share several functions. When interpreting phenotypes of transgenic plants, it is important to consider the level of suppression in barley RNAi lines and the level of HvABCD expression in cts-1 complemented lines (Dietrich et al., 2009). Arabidopsis plants heterozygous for cts germinate and are able to establish in the absence of sucrose, but are resistant to 2,4-D and IBA (Hayashi et al., 1998; Zolman et al., 2001). Thus the auxin phenotypes are more sensitive to loss of function than germination and lipid mobilization in Arabidopsis. Therefore, a minor reduction in ABCD function in an RNAi line could lead to auxin resistance but might not impact markedly on other phenotypes. By this reasoning, a substantial knockdown of ABCD transporter expression is likely to be required to detect effects on lipid metabolism and perhaps also germination. However, the physiological thresholds which underpin the visible phenotypes may vary between barley and Arabidopsis, and the influence of the expression level on phenotypes of complemented plants is to some extent dependent on the substrate specificity of the heterologous transporters. Unfortunately, an antiserum raised to the relatively well-conserved C-terminus of Arabidopsis CTS, which recognizes both NBDs (Footitt et al., 2002; De Marcos Lousa et al., 2009), did not recognize barley CTS (data not shown); thus it was not possible to determine the level of HvABCD1 or HvABCD2 protein expression in transgenic lines. With these caveats in mind, several conclusions can be reached.

ABCD transporters play a role in seed size determination in diverse species

Both barley RNAi lines and the complemented Arabidopsis cts-1 lines support a role for ABCD transporters in control
Fig. 5. Complementation of cts-1 by HvABCD1 and HvABCD2. (A, B) Length and width of seeds of cts-1, complemented lines, and the wild type (Ler). Values are means ±SE (n=3; 30 seeds per replicate). t-test indicates significant difference of transgenic lines from cts-1 (***P<0.001). (C) Seed weight. Values are means ±SE (n=5; 80 seeds per replicate). (D) Germination over 7 d on water agarose. Values are means ±SE (n=3; 50 seeds per replicate). (E) Seedling establishment on 0.5× MS with or without 0.5% sucrose. In the upper panel, seeds of cts-1 and cts-1 AtCTS:HvABCD1.1 were induced to germinate by mechanically rupturing the seed coat and plating on sucrose medium for 2 d before transfer to medium lacking sucrose (–suc). Seedlings were rearranged on a fresh plate for photography. (F) Elongation of hypocotyls in the dark on 0.5× MS with or without 0.5% sucrose. (G) Elongation of hypocotyls in the dark on 0.5× MS containing 0.5% sucrose and different concentrations of IBA. Values are means ±SE (n=3; 20 hypocotyls per replicate) in F and G. (H) Effect of oxophytodienoic acid (OPDA) on root growth. Medium is 0.5× MS containing 0.5% sucrose; values are means ±SE (n=25). The opr3 mutant lacks oxophytodienoate reductase and is unable to convert OPDA to JA. (This figure is available in colour at JXB online.)
Barley peroxisomal ABC transporters | 4843

HvABCD2 contributes to OPDA metabolism

Transcript abundances of JA-inducible genes were reduced in leaves of barley RNAi lines, consistent with a role for HvABCD genes in conversion of OPDA to JA (Fig. 4). Whilst HvABCD1 did not complement cts-1 for germination, expression of HvABCD2 restored the germination rate to wild-type levels (Fig. 5D), which is supportive of the proposal that HvABCD2 mediates OPDA transport into the peroxisome. In agreement with this, roots of cts-1 lines expressing HvABCD2 but not HvABCD1 exhibited sensitivity to exogenously applied OPDA similar to that of wild-type controls (Fig. 5H), strongly supporting the notion that HvABCD2 is required for the conversion of OPDA to JA.

Although accumulation of OPDA is associated with inhibition of germination in Arabidopsis (Dave et al., 2011), the influence of different jasmonates on barley germination has not been studied in detail. The effects of jasmonates on seed germination appear to be species and even ecotype specific (Linkies and Leubner-Metzger, 2012); however, a recent transcriptome study demonstrated that jasmonate biosynthetic and putative receptor genes are up-regulated in coleorhiza tissue of imbibed after-ripened seeds, relative to dormant seeds, pointing to the importance of this class of compounds in barley germination (Barrero et al., 2009). Suppression of HvABCD1 and HvABCD2 had little or no effect on germination (Fig. 1F; Supplementary Fig. S3 at JXB online), perhaps suggesting that ABCD transport activity is not essential for this function in barley, as has also been proposed for tomato (Orsi and Tanksley, 2009). However, the level of HvABCD
knockdown may have been insufficient to impact on germination in the present experiments, and it is also possible that, as in Arabidopsis, an additional peroxisomal import pathway for OPDA operates in barley seeds (Theodoulou et al., 2005).

HvABCD1 and HvABCD2 are functional in fatty acid β-oxidation

Oil bodies have been shown to occupy >40% of the cell volume in barley aleurone, and embryo-derived GA stimulates oil breakdown during germination (Eastmond and Jones, 2005). The TAG composition is similar in Arabidopsis and barley, with the exception that Arabidopsis contains a high level (~25%) of the very long chain fatty acid, eicosanoic acid (20:1) (Holmer et al., 1973; De Man and Vervenne, 1988; Lemieux et al., 1990). In barley, genes involved in lipid catabolism and starch mobilization are expressed together with key transcripts of sucrose synthesis as early as 24 h after imbibition before radical protrusion, suggesting that lipid reserve mobilization could supply sucrose before the seedling becomes phoautotrophic (Sreenivasulu et al., 2008). Although oil bodies declined in abundance over 5 d of imbibition of intact grains and also in response to GA treatment of de-embryonated seeds (Fig. 2), increased oil body or fatty acid retention was not observed in HvABCD1/2i lines. It might have been predicted that suppressing HvABCD1 and HvABCD2 would result in retention of oil bodies, but it is possible that the RNAi lines are not sufficiently impaired in β-oxidation to give a detectable phenotype, consistent with the observation that Arabidopsis mutants in which lipid breakdown is reduced by 50% do not exhibit a marked growth phenotype (Kelly et al., 2011). However, complementation of the Arabidopsis cts-1 mutant for seedling establishment in the absence of sucrose indicates that HvABCD2 can transport a range of different fatty acid species (Fig. 5E, F). Although HvABCD1 did not complement the seedling establishment phenotype of cts-1, it did partially restore fatty acid β-oxidation to the yeast pxa1/pxa2Δ mutant (Fig. 6). The lack of complementation in planta may reflect the low expression level of HvABCD1 but could also result from differences between the two expression systems, such as the ability of the heterologous transporter to interact with the different endogenous peroxisomal acyl-CoA synthetases (van Roermund et al., 2012; De Marcos Lousa et al., 2013).

HvABCD1 and HvABCD2 contribute to IBA metabolism

Barley RNAi lines (Fig. 3) and the complementation of cts-1 (Fig. 5G) suggest a role for HvABCD proteins in IBA metabolism. IAA is synthesized by several different routes (Mano and Nemoto, 2012), including β-oxidation of IBA. Although IBA is a relatively abundant auxin, this was originally thought to be a minor biosynthetic route. However, the physiological importance of IBA-derived IAA has been demonstrated conclusively in Arabidopsis by elegant genetic analysis (Strader and Bartel, 2011; Strader et al., 2011). IBA has also been proposed to be a hormone in its own right (Tognetti et al., 2010). Very little is known regarding the functions of IBA in barley, but the data presented here demonstrate a role for IBA-derived IAA in root growth.

HvABCD1 and HvABCD2 may differ in substrate specificity

Since CTS is a broad specificity transporter which plays several distinct roles in growth and development of Arabidopsis, the presence of two transporters in cereals is intriguing. Classically, gene duplication is considered to be a mechanism which increases expression diversity and permits the evolution of tissue or developmental specialization. Duplicate genes also play a role in the establishment of new functions or in subfunctionalization (Li et al., 2005). The ABC transporter superfamily exhibits a particularly high level of gene birth and death, often associated with the acquisition of taxon-specific functions (Annilo et al., 2006; Verrier et al., 2008). HvABCD1 and HvABCD2 display similar expression patterns in embryo and aleurone, suggesting that they may play distinct roles in imbibed grains and indeed throughout the plant, since they are widely expressed in different tissues. This is consistent with their differential ability to complement cts-1. Whilst HvABCD2 complemented all phenotypes tested, HvABCD1 was apparently unable to mediate sufficient fatty acid transport to support seedling establishment when expressed in Arabidopsis and was unable to restore germination to cts-1 seeds (Fig. 5D), suggesting differences in substrate specificity. Although, as discussed above, complementation depends on expression in the correct subcellular location at an appropriate level, it is clear that HvABCD1 was expressed (and presumably targeted correctly) in the present experiments, since it conferred sensitivity of cts-1 hypocotyls to high concentrations of IBA and partially complemented the seed size phenotype (Fig. 5). Transport studies will be required to determine unequivocally the substrate specificities of HvABCD1 and HvABCD2, and further analysis of RNAi lines may reveal additional, perhaps cereal-specific functions for these proteins. In conclusion, monocot and dicot ABCD transporters share a core set of functions, but the retention of two ‘full-length’ HvABCD genes in the grass lineage points to an important functional or regulatory diversification which awaits further investigation.

Supplementary data

Supplementary data are available at JXB online.

Figure S1. Alignment of HvABCD1 and HvABCD2 cDNA sequences, showing the region used for the RNA interference construct.

Figure S2. Expression analysis of HvABCD1 and HvABCD2.

Figure S3. Germination of RNAi lines and corresponding nulls over 7 d.

Figure S4. Q-PCR analysis of gene expression in Arabidopsis transgenic lines and controls.

Table S1. Primers used in this study.
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References


Kelly AA, Quettier AL, Shaw E, Eastmond PJ. 2011. Seed storage oil mobilization is important but not essential for germination or seedling establishment in Arabidopsis. Plant Physiology 157, 866–876.


van Roermund CWT, Elgersma Y, Singh N, Wanders RJA, Tabak HF. 1995. The membrane of peroxisomes in Saccharomyces cerevisiae is impermeable to NAD(H) and acetyl-CoA under in vivo conditions. EMBO Journal 14, 3480–3486.

van Roermund CW, Ilijst L, Majczak W, Waterham HR, Folkerts H, Wanders RJ, Hellingwerf KJ. 2012. Peroxisomal fatty acid uptake
Barley peroxisomal ABC transporters


