Leukaemia-specific chromosome damage detected by comet with fluorescence in situ hybridization (comet-FISH)

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Acute myeloid leukaemia (AML) is associated with exposure to benzene and treatment with chemotherapeutic agents. It is thought to arise from damage to specific regions of DNA, resulting in chromosome rearrangements or loss. For instance, a deletion on the long arm of chromosome 5 [e.g. del(5q31)] is common in AML patients previously treated with alkylating agents, such as melphalan or exposed to benzene. Translocations of the MLL gene at 11q23 are frequently observed in AML arising from treatment with topoisomerase II inhibitors, such as etoposide. Our goal was to determine whether or not breakage at 5q31 and 11q23 is selectively induced by these chemical agents. To address this question, the comet assay combined with fluorescence in situ hybridization (comet–FISH) was used to detect DNA breakage in the specific chromosomal regions in an in vitro model. TK6 lymphoblastoid cells were exposed to melphalan, etoposide or the benzene metabolite, hydroquinone (HQ), at various concentrations. HQ, melphalan and etoposide induced DNA breaks at both 5q31 and 11q23 chromosome regions in a dose-dependent manner. However, HQ produced significantly more DNA damage at 5q31 than at 11q23. Etoposide produces slightly more DNA damage at 11q23 and melphalan had a somewhat greater effect at 5q31, but not significantly so. Thus, HQ and melphalan act similarly, perhaps explaining some similarities between benzene- and alkylating agent-induced AML. Comet–FISH also appears to be a useful approach for detecting and comparing damage to specific chromosome regions of significance in leukaemogenesis.

Introduction

Leukaemogenesis is a complex, multi-step process that involves transformation of a haematopoietic progenitor cell through the induction of several types of chromosomal aberrations and gene mutations followed by clonal expansion (1,2). The cause of most types of leukaemia is largely unknown, but several factors have been directly implicated, including inherited mutations, ionizing radiation and chemical exposure (3,4). Chemotherapeutic treatment with alkylating agents and topoisomerase II inhibitors can cause therapy-related acute myeloid leukaemia (AML) (5–7). However, depending on the agent used, patients display different cytogenetic characteristics (5,6).

AML induced by alkylating agents, such as melphalan, usually develops 7–10 years after exposure and commonly displays a deletion of all or part of chromosomes 5 and 7. Previous in vitro studies have shown that melphan preferentially induces breaks and structural aberrations in chromosomes 5, 7, 11 and 17 (8–10). Deletion of 5q31 was also detected in multiple myeloma patients treated with melphalan (11). Topoisomerase II inhibitors, such as etoposide, induce AML with a shorter latency period of 1–5 years after exposure and are usually associated with balanced translocations involving chromosome band 11q23, the location of the MLL gene (7,12). Topoisomerase II enzymes catalyse breakage of double-stranded DNA during replication, transcription and chromosomal segregation. Topoisomerase II inhibitors stabilize the complex of topoisomerase II, leaving a broken DNA 5’ strand end, slowing ligation, triggering double-strand break repair (illegitimate recombination) and, in some instances, apoptosis (12).

Occupational exposure to benzene also induces leukaemia including AML (13,14). Benzene is metabolized in the liver to its primary metabolite phenol, which is hydroxylated to hydroquinone (HQ) (15). HQ can be further oxidized in the bone marrow to 1,4-benzoquinone (16,17). This pathway is currently thought to play a major role in benzene-induced leukaemia. Leukaemias resulting from benzene exposure show many types of chromosomal aberrations including the loss of all or part of chromosomes 5 and 7, such as del(5q31) (18–20). 1,4-Benzoquinone has also been reported to be a topoisomerase II inhibitor and may affect the MLL gene at chromosome band 11q23 (21,22). In the present study, we have attempted to determine if leukaemogenic chemicals induce damage specifically at the 5q31 and 11q23 regions involved in leukaemogenesis, and critically to determine if HQ acts more like melphalan or etoposide.

The comet assay (single-cell gel electrophoresis) is a simple method for measuring DNA single- and double-strand breaks in eukaryotic cells. The more DNA breaks produced, the greater the fluorescent intensity of the comet tail relative to the head. These DNA breaks may be induced by oxidative stress (23–25). Recently, the comet assay has been combined with the specificity of fluorescence in situ hybridization (FISH) (26,27) to allow simultaneous detection of overall DNA damage and DNA breaks in specific regions of the genome. The use of a fluorescent-labelled DNA probe homologous to the region of interest in conjunction with comet (comet–FISH) permits the detection of DNA breaks within that region. Comet–FISH was introduced in 1997 (26) to study the spatial distribution of specific chromosome sequences and chromatin fibres in comet cells. Subsequently, it was used to examine initial DNA damage and subsequent repair in the TP53 gene region of bladder carcinoma cells after gamma irradiation exposure (28,29). It has recently been applied to study damage in a number of specific genes (28–32).

Here, we have used comet–FISH to compare the DNA damaging effects of melphalan, etoposide and HQ specifically at bands 5q31 and 11q23. Our a priori hypothesis was that the alkylating agent melphalan would produce more damage at
balanced saline solution (HBSS) without Ca$^{++}$ and Mg$^{++}$. An aliquot of the cell suspension was mixed with cold Hank’s solution and washed twice. Then, cold 70% ethanol was added to the cells and left at 4°C overnight. The cells were dehydrated with 100% ethanol, air-dried overnight and stored in a dry place before being processed for comet or comet–FISH analysis. All slides were blind-coded before they were stained with SYBR™ Green (Molecular Probes, Eugene, OR). The cell scoring was performed under a ×25 objective on a fluorescence microscope using an FITC filter. Twenty-five randomly chosen cells per slide and two slides per dose were analyzed with the CometScan (MetaSytens, Germany) imaging software.

### Chemical treatment and cytotoxicity

The TK6 cells were exposed to various concentrations of HQ (Aldrich, Milwaukee, WI), melphalan and etoposide (Sigma, St Louis, MO). Dimethyl sulfoxide was used as the vehicle for melphalan and etoposide and was present in cell cultures at a final concentration of 0.25%. HQ was dissolved in RPMI. When TK6 cells were in exponential growth phase, they were treated with these three chemicals for 1 h and then harvested for cell viability test, cell cycle analysis, comet assay and comet–FISH. All treatments were performed in duplicate for each dose and each experiment was repeated three times. Cytotoxicity was measured utilizing the Trypan Blue exclusion assay. The cells were mixed with Trypan blue 0.04% (Gibco, Carlsbad, CA) in a 1:1 ratio and the percentage of dead cells was determined for each chemical treatment at various concentrations. The percentage of dead cells was <10% at non-cytotoxic doses.

### Cell cycle analysis

An aliquot of the cell suspension was mixed with cold Hank’s balanced saline solution (HBSS) without Ca$^{++}$ and Mg$^{++}$ (Invitrogen) and washed twice. Then, cold 70% ethanol was added to the cells and left at 4°C overnight. The cells were washed with HBSS, and RNase A (1 mg/ml) and propidium iodide (PI) (34) were added. The solution was stored at 4°C until analysis. The cell cycle analysis was performed on a Beckman-Coulter EPICS XL-MCL. Using the intensity of PI fluorescence as a measure of the total DNA content of individual cells, cell cycle populations were quantified from a standard count of 10 000 cells. Cell cycle analysis was performed at 1 and 24 h after exposure.

### Comet assay

The alkaline comet assay was performed according to Singh et al. (23) with some modifications. Briefly, super-frosted microscope slides were pre-coated by dipping each slide into 1% normal melting agarose and air-dried overnight. Seventy-five microliters of low-melting agarose (Fisher Scientific, Pittsburgh, PA) mixed with $1 \times 10^7$ cells was placed on the pre-coated slide. Several slides were prepared at each dose to be used for both comet and comet–FISH assays. A glass coverslip was placed on top and slides were kept at 4°C for 10 min to allow the gel to solidify. Then a second layer of low-melting agarose was added and the slides were kept at 4°C for another 10 min to allow the gel to solidify. Slides were placed in cold lysis solution (2.5 M NaCl, 100 mM EDTA, 10 mM Trizma base and 1% Triton X-100, pH 10) and kept at 4°C until electrophoresis was performed. The Trizma base was from Sigma and other reagents in the lysis solution were purchased from EM Scientific (Darmstadt, Germany).

Unwinding was performed in an alkaline buffer (300 mM NaOH and 1 mM EDTA, pH 13) for 25 min and electrophoresis was run in the same alkaline buffer at 8°C at 0.73 V/cm, for 35 min. After electrophoresis, the slides were neutralized with 0.4 M Tris, pH 7.5. The gels were dehydrated with 100% ethanol, air-dried overnight and stored in a dry place before being processed for comet or comet–FISH analysis. All slides were blind-coded before they were stained with SYBR™ Green (Molecular Probes, Eugene, OR). The cell scoring was performed under a ×25 objective on a fluorescence microscope using an FITC filter. Twenty-five randomly chosen cells per slide and two slides per dose were analyzed with the CometScan (MetaSytens, Germany) imaging software.

### Comet–FISH

Comet slides from solvent controls and two concentrations of each of the three chemicals tested were subjected to FISH analysis. The slides were immersed for 20 min in absolute ethanol. All slides were hybridized with two specific chromosome probes, 5q31 and 11q23 (Vysis, Downers Grove, IL), each in separate spots outlined with a diamond pencil. Dual-colour human chromosome band probes 5q31 (200 kb) and 11q23 (350 kb) were used. To each spot, 10 µL of probe mixture was added (2 µL chromosome probe, 1 µL of salmon sperm DNA and 7 µL hybrid solution from Vysis). Coverslips (22 × 22 mm) were placed over each spot and sealed with rubber cement. An automatic denaturation and hybridization procedure was performed using the HyBrite™ denaturation/hybridization system (Vysis). Cells and probes were denatured at 68°C for 10 min and hybridized at 37°C for 72 h. After hybridization, the rubber cement was carefully removed and the slides were placed in water for a few seconds to allow the coverslips to slip off and prevent gel rupture. The slides were washed in 50% formamide/2× SSC at 45°C for 3 min and 0.1× SSC at room temperature for 2 min, three times. The cells were counter-stained with 1 µg/ml 4′,6-diamidino-2-phenylindole (DAPI). The hybridization signals were observed with a fluorescence microscope under triple band-pass filter for DAPI/FITC/Texas Red (excitation at 405, 490 and 570 nm, respectively; emission at 460, 525 and 635 nm, respectively). In each spot, only the spectrum orange signal was taken into account for the comet–FISH analysis because they represented the 5q31 and 11q23 chromosome bands.

### Comet–FISH scoring

We designed a 4×4 table to classify and evaluate the comet–FISH results (Figure 4). The figure describes four categories that quantify site-specific DNA breakage by the number of DNA probe signals (I–IV) and four categories that describe the localization of the probe signal in the comet head, head and tail or only tail) in relation to the presence of a comet tail which correlates with overall DNA damage (A–D). A total of 150–300 cells were randomly scored for each chemical (50–100 cells per dose). Each cell was categorized according to Figure 4 and the cumulative counts were used for the data analysis. The comet–FISH experiments were repeated three times.
**Statistical analysis**

The comet parameter used for data analysis was the percentage of DNA in the tail (% tail intensity) and linear regression analysis was performed to determine dose–response relationship shown as a $P_{\text{trend}}$.

The data generated from the comet–FISH assay, using Figure 4, were categorical and were analysed using an order logistic regression, which returns odds ratios (ORs) for an increase in the outcome by one category (35). The ordered logistic regressions analysis assessed the distribution of the chromosome probe signals among the different cell damage categories. Because there were multiple measurements per slide, our inference was based on robust standard errors (36), which adjusts for the residual correlation of measurements made on the same slide. The analyses were performed to determine which chromosomes were more affected by the chemicals of interest, and so we included interaction terms (dose × chromosome). For all analyses, we treated dose categorically by including the appropriate dummy variables in the model. For instance (using the data in Table III as an example), consider the following ordered logistic regression model [see Models for Ordinal Scales in McCullagh and Nelder (35) for equivalent notation]:

$$
\log \left( \frac{\gamma_j(Dose, Chrom)}{1 - \gamma_j(Dose, Chrom)} \right) = \beta_0 - \left[ \beta_1 I(Dose = 25) + \beta_2 I(Dose = 50) + \beta_3 I(Chrom = 11) \\
+ \beta_4 I(Dose = 25)I(Chrom = 11) \\
+ \beta_5 I(Chrom = 50)I(Chrom = 11) \right],
$$

where $\gamma_j(Dose, Chrom) = P(Y \leq j|Dose, Chrom)$, $Y$ is the ordered outcome (probe signals: 0 → 1–2, 2 → 3–4, 3 → 5–7, 4 → ≥8) and $I(\text{statement}) = 1$ if ‘statement’ is true, 0 otherwise. In this model, the test of the null $H_0$: $\beta_4 = 0$ tests whether the effect (OR for moving up one category) of HQ 25 versus 0 is the same for both chromosomes 11 and 5; likewise, the test of the null $H_0$: $\beta_5 = 0$ tests the equality of the OR’s for 50 versus 0. We used the Wald test of these coefficients (based on the robust standard errors) to derive the $P$-values reported below in the Results.

**Results**

**Overall DNA damage measured using the comet assay**

Exposure of human TK6 cells to three chemical leukaemogens, HQ, melphalan and etoposide, produced a significant increase in overall DNA damage, represented as an increase of the % tail intensity. HQ (up to 75 μM) caused increasing DNA damage ($P_{\text{trend}} = 0.04$) without significant cytotoxicity (Figure 1A). Melphalan ($P_{\text{trend}} = 0.006$) and etoposide ($P_{\text{trend}} = 0.005$) also caused DNA breakage with increasingly more DNA being present in the comet tails, without cytotoxicity at doses up to 25 μM (Figure 1B and C, respectively). Of the three compounds, etoposide was the most effective at producing DNA damage with % tail intensity content approaching 70% at doses above 5 μM.

**Cell cycle analysis**

Cell cycle analysis by flow cytometry revealed that HQ produced S-phase arrest at concentrations ≥25 μM (Figure 2A). Low doses of melphalan produced G2-phase arrest, whereas higher doses resulted in S-phase arrest (Figure 2B). At the highest (25 μM) dose, however, it produced G1/S phase arrest increasing the percentage of cells in G1 phase drastically (Figure 2B). Etoposide caused arrest at G2/M phase and increasing numbers of cells in G2 phase with increasing doses (Figure 2C).

**Site-specific DNA breaks detected by comet–FISH**

FISH probes were targeted to the p and q arms of chromosome 5 at the regions 5p15.5 and 5q31 (Figure 3A). The appearance of probe-specific fluorescent signals in the tail, after HQ exposure at 50 μM (Figure 3B), showed breakage at these locations on the chromosome. Using the scoring criteria described in Figure 4, we quantified the amount of site-specific probe breakage in TK6 cells induced by the three compounds. Figure 4 shows the distribution of the probe fluorescence between the head and tail of the comet, comparing categories A and B with C and D. If all or some of the probe fluorescence is located in the comet tail (categories C and D), this shows significant breakage of the DNA at the probe location. Additional probe signals appearing in the head of the comet could result from aneuploidy, gene amplification and duplication rather than breakage.
HQ at 25 and 50 μM caused a highly significant shift in location of the probe fluorescence from the head to the tail for FISH probes specific to both bands 5q31 and 11q23 (Table I). To test our a priori hypothesis, we directly compared the HQ-induced specific loci breakage on both bands and found that HQ treatment caused significantly more breakage at 5q31 (OR: 5.34) in the tail than at 11q23 (OR: 2.82) (*P* < 0.02, Table II). Melphalan, at doses that did not produce cytotoxicity or cell cycle changes, induced only modest breakage of probes to bands 5q31 and 11q23 and movement of the probes into the comet tail (Table I). This effect on bands, 5q31 (OR: 1.88) and 11q23 (OR: 1.32), was almost the same, but the OR at 5q31 was slightly more pronounced (Table II). Etoposide produced a large shift of probe fluorescence from the comet head to the tail especially at the higher dose of 2.5 μM. The effect of etoposide was slightly greater on band 11q23 (OR: 9.09) than on 5q31 (OR: 6.79), but this difference was not statistically significant (Table II).

Another method of scoring the amount of site-specific damage in each band is to count the number of probe signals detected by the fluorescent probes in each cell. We therefore counted fluorescent probe spots and categorized them into four categories, I–IV: I (1–2 spots), II (3–4 spots), III (5–7 spots) and IV (>8 spots) (Figure 4). Categorization in this fashion showed that HQ and melphalan caused site-specific damage and a shift from categories I and II to categories III and IV at both 25 and 50 μM (Table III). These two compounds induced a slightly higher shift at 5q31 than at 11q23. However, etoposide tended to induce more damage at 11q23 than 5q31, but these differential effects were not statistically significant (Table IV). Thus, in contrast to spot counting, the categorization of probe fluorescence into head and tail displayed a better assessment of the effects of the three leukaemogens to specific chromosomes.

The data were further classified into two more categories (Figure 4): number of probe signals in comets with no tail (A) and comets with tails (B, C and D), and into what we considered low (A, C and D), medium (AII, BII, BIII and CII) and high (CIII, CIV, DIV and DII) effects. The results from these classifications did not provide any additional information over that presented in Tables I and III.

**Discussion**

Measuring classical chromosomal aberrations in metaphase cells, either by Giemsa staining or FISH, is the most frequently used method to detect chromosomal damage. This method is time consuming and requires skilled personnel. We have explored the potential of comet–FISH as a simpler alternative method to assess damage to specific chromosomal regions.

Comet–FISH was introduced in 1997 and has been recently applied to study damage to the telomeres (37,38) and to specific genes such as *p53*, dihydrofolate reductase (*DHFR*) and the O\(^\text{6}\)-methylguanine DNA methyltransferase (*MGMT*)
To our knowledge, there are only two studies that used chromosome probes to study specific chromosomal damage (27,40) and these studies used whole-chromosome painting. In the present study, we quantified the amount of DNA damage induced by HQ, melphalan and etoposide in chromosome regions 5q31 and 11q23 using the comet–FISH technology. These chemicals and chromosome locations were chosen because of their relationship to chemically induced leukaemia (5,7).

Our *a priori* hypothesis was that the alkylating agent melphalan would produce more damage at band 5q31 than at 11q23 and, conversely, that the topoisomerase II inhibitor etoposide would cause more breakage at band 11q23 than at 5q31. For the benzene metabolite HQ, it was unclear which location, if any, would be preferred as it has been described as a topoisomerase II inhibitor (21), but is also known to cause deletion of 5q31 (20,33). Surprisingly, HQ exposure produced significantly more DNA breakage at 5q31 than at 11q23. Melphalan induced a modest overall effect, but tended to produce more breakage at 5q31 than 11q23 (Table I). Melphalan is a bifunctional alkylating agent that has the capability to induce DNA cross-linking. Cross-linking causes a retardation in DNA migration during the electrophoresis, which could account for the apparently low overall DNA breakage we observed with melphalan in the comet assay (Figure 1). Etoposide induced considerable DNA damage in a dose-dependant fashion, showing more damaging effects on chromosome 11q23 than at 5q31. Etoposide induced a modest overall effect, but tended to produce more breakage at 5q31 than 11q23 (Table I). Etoposide is a topoisomerase II inhibitor that induces a wide variety of DNA damage, but our findings are in agreement with those of Ng *et al.* (41), which showed an apparent preference for the *MLL* gene at chromosome 11q23. In the studies presented here, etoposide induced more DNA breaks than HQ, and HQ induced more than melphalan (Table III). Melphalan and etoposide showed preferential effects that were in line with our *a priori* hypotheses, but to our surprise there was evidence of

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Dose (μM)</th>
<th>Localization (%)a</th>
<th>5q31</th>
<th>11q23</th>
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<tr>
<td></td>
<td></td>
<td>Head</td>
<td>Tail</td>
<td>Head</td>
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<tr>
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<td>96.67</td>
<td>3.33</td>
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<td>27</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>0.5</td>
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<td>36</td>
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<td>6</td>
<td>69.33</td>
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*aLocalizations are classified from Figure 4 as head = A + B, head/tail = C and tail = D.
*bThe data show the cumulative counts of three independent experiments; in each experiment, approximately 50–100 cells were analysed. The percentage is calculated from 150 to 300 cells representing 100% total.

<table>
<thead>
<tr>
<th>Chemical treatment</th>
<th>ORa</th>
<th>P-valueb</th>
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<tr>
<td></td>
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<td>Melphalan</td>
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<tr>
<td>Etoposide</td>
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<td>9.09</td>
</tr>
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</table>

*aOR for trends, i.e. the OR for increasing one category of damage from jumping one dose category of the chemical. For example, the OR of 5.34 represents the ratio of odds of comparing one category of damage to the next one higher for changing dose of HQ from 0 to 25 (or 25 to 50) among chromosome 5.
*bComparison between chromosome bands 5q31 and 11q23.

![PROBE LOCATION & TAIL PRESENCE](https://academic.oup.com/mutage/article-abstract/22/5/321/1144447)

**Table I.** Distribution of 5q31 and 11q23 probe signals in the comet head, head and tail or tail, in relation to chemical dose

**Table II.** ORs and statistical significance after ordered logistic regression analysis comparing the distributions of 5q31 and 11q23 probe signals in the comet head, head and tail or tail

**Fig. 4.** Comet–FISH scoring criteria. (28–32,34,39). To our knowledge, there are only two studies that used chromosome probes to study specific chromosomal damage (27,40) and these studies used whole-chromosome painting. In the present study, we quantified the amount of DNA damage induced by HQ, melphalan and etoposide in chromosome regions 5q31 and 11q23 using the comet–FISH technology. These chemicals and chromosome locations were chosen because of their relationship to chemically induced leukaemia (5,7).

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The comet–FISH approach described here can measure whole-genome DNA damage plus specific chromosomal region damage in a manner that is faster and more streamlined than the classical chromosomal aberration assay. It may also be a useful method for biomarker research in human populations where one wishes to study effects on specific chromosome regions because it does not require metaphase cells for analysis.

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Conflict of interest statement

M.T.S. has received consulting and expert testimony fees from law firms representing both plaintiffs and defendants in cases involving exposure to benzene.

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