EBPG Guideline on Nutrition

Denis Fouque1, Marianne Vennegoor2, Piet Ter Wee3, Christoph Wanner4, Ali Basci5, Bernard Canaud6, Patrick Haage7, Klaus Konner8, Jeroen Kooman9, Alejandro Martin-Malo10, Lucianu Pedrini11, Francesco Pizzarelli12, James Tattersall13, Jan Tordoir14 and Raymond Vanholder15

1Département de Néphrologie, JE 2411- Université Claude Bernard Lyon1, Lyon, France, 2Department of Nephrology, Nutrition and Dietetics, Guy's and St Thomas' NHS Foundation Trust, London, UK (retired), 3Department of Nephrology, Institute for Cardiovascular Research, VU University Medical Center, Amsterdam, The Netherlands, 4Department of Medicine, Division of Nephrology, University Hospital, Würzburg, Germany, 5Department of Medicine, Division of Nephrology, Ege University Medical Faculty, Izmir, Turkey, 6Nephrology, Dialysis and Intensive Care Unit, Lapeyronie University Hospital, Montpellier, France, 7Department of Diagnostic and Interventional Radiology, Helios Klinikum Wuppertal, University Hospital Witten/Herdecke, Germany, 8Medical Faculty University of Cologne, Medicine Clinic I, Hospital Merheim, Germany (retired), 9Department of Internal Medicine, Division of Nephrology, University Hospital Maastricht, The Netherlands, 10Nephrology Department, Reina Sofia University Hospital, Cordoba, Spain, 11Division of Nephrology and Dialysis, Bolognini Hospital, Seriate, Italy, 12Nephrology Unit, SM Annunziata Hospital, Florence, Italy, 13Department of Renal Medicine, St. James’s University Hospital, Leeds, UK, 14Department of Surgery, University Hospital Maastricht, The Netherlands and 15Nephrology Section, Department of Internal Medicine, University Hospital, Ghent, Belgium

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Guideline 1. Prevalence of malnutrition and outcome

- Nutritional status should be assessed at the start of haemodialysis (Opinion).
- Protein–energy malnutrition should be avoided in maintenance haemodialysis because of poor patient outcome (Evidence III).
- In absence of malnutrition, nutritional status should be monitored every 6 months in patients <50 years of age (Opinion).
- In patients >50 years of age, and patients undergoing maintenance dialysis for more than 5 years, nutritional status should be monitored every 3 months (Opinion).

Rationale

Malnutrition is considered to be one of the late complications of chronic renal failure. A sub-analysis of the Modification of Diet in Renal Disease (MDRD) study, however, demonstrated that progressive renal insufficiency was associated with a spontaneous decline in protein intake. Predialysis patients appeared to have a spontaneous protein intake of <0.7 g/kg/day [1], which is below the minimal recommended daily intake.

Thus, malnutrition in haemodialysis patients may already originate during stage IV of chronic renal failure.

It has been demonstrated that serum albumin and creatinine increase during the first half year of haemodialysis [2,3], suggesting an improvement of
nutritional status after the initiation of dialysis. Nevertheless, many studies have reported on the presence of malnutrition in a large number of dialysis patients [4–7]. In the French national cooperative study [6], that included 7123 patients, nutritional status was determined by body mass index (BMI), normalized protein catabolic rate (nPCR) and several laboratory values. Life-threatening malnutrition was present in up to 36% of the patients. Low protein intake and low dialysis efficacy were associated with the presence of malnutrition. Several other studies demonstrated that haemodialysis patients eat less protein and fewer calories than prescribed, which is associated with a higher rate of malnutrition [4,5,7].

Several small and large scale cohort studies have revealed that protein–energy malnutrition is associated with increased morbidity, mortality and impaired quality of life [8–19]. Herselman et al. [10] demonstrated an association between a composite score for protein–energy malnutrition and infection-related morbidity in a group of haemodialysis patients. A recent paper demonstrated that in patients with an appropriate dialysis efficacy (Kt/V ≥ 1.2) low serum albumin and low protein intake, measured as low nPCR, were associated with a higher risk of hospitalization and mortality [17]. Data from the United States Renal Data System (USRDS) database [13] as well as data from the large Dialysis Outcomes and Practice Patterns Study (DOPPS) cohort [14] confirm that malnourished dialysis patients have an increased risk of mortality. In the USRDS DMMS-1 cohort analysis protein–energy malnutrition was established through serum albumin levels, BMI and notification by the treating physicians in the patient medical files of the existence of malnutrition [13]. From this data set of 5058 patients it was concluded that patients who were considered malnourished by their physicians, had a 27% greater risk of cardiovascular death. In addition it was shown that for each one-unit decrease in BMI the risk for cardiovascular death rose by 6% and each 1 g/dl fall in serum albumin level was associated with a 39% increase in risk of cardiovascular death. A recent study reported that both malnutrition, established by measurement of total body nitrogen by in vivo neutron activation analysis, and serum albumin were independent predictors of mortality in incident haemodialysis patients [19]. Hypoaalbuminaemia appeared also to be a predictor of vascular morbidity. In DOPPS, a prospective observational study, nutritional status is investigated by means of a modified subjective global assessment (mSGA), BMI, serum albumin and some other laboratory parameters at baseline (n = 7719) and after 6 months (n = 3739; [14]). Patients with severe malnutrition according to mSGA had a 33% higher mortality risk and patients with moderate malnutrition a 5% increased risk. In patients with the lowest BMI quartile the mortality risk was 60% higher than that of patients in the highest quartile. In addition it was demonstrated that patients who had a loss in BMI of ≥ 3.5% had an increase in mortality risk. Likewise, both a low serum albumin level as well as a fall in serum albumin was strongly associated with an elevated mortality risk.

Apart from the elevated risk of mortality, results from the HEMO study have revealed that malnutrition, established with anthropometric measurements, serum albumin and PCR, was associated with impaired physical functioning [20] and impaired quality of life [12]. Likewise, Koo et al. [18] reported an association between depression and malnutrition in a group of chronic haemodialysis patients.

It is widely appreciated that age negatively affects outcome of dialysis patients. It has been demonstrated only in a few studies that malnutrition contributes to the increased mortality risk of older dialysis patients [21,22]. In the HEMO study, it was demonstrated that middle age (≥50 years) and older (≥65 years) dialysis patients had lower dietary energy and protein intake, serum albumin levels and nPCR compared with young dialysis patients (<50 years) despite similar dialysis efficacy measured as equilibrated Kt/V (eKT/V) [21]. Such indications for malnutrition in the older patients were associated with higher morbidity. In a French cohort study, it was shown that in dialysis patients over 75 years, malnutrition negatively affected overall survival despite adequate dialysis treatment [22].

In several small and larger studies dialysis vintage appeared to have a clear negative effect on nutritional status of dialysis patients [16,23–29]. In a cohort study of 3009 patients, Chertow et al. [27] were able to demonstrate that dialysis vintage was associated with a decline in nutritional parameters and that every year on dialysis was associated with a 6% increase in the risk of dying. Likewise, in the HEMO study it was shown that patients over 5 years on dialysis had significantly lower anthropometric parameters suggesting an impaired nutritional status compared with patients shorter on dialysis [16]. This was also shown in studies with more sophisticated tools for determination of nutritional status [24,29].

Thus an adequate monitoring of nutritional status is an important step of haemodialysis patients care and allows for the identification of body composition alterations associated with increased morbimortality.

References

EBPG guideline on nutrition


**Guideline 2. Diagnosis and monitoring of malnutrition**

Protein–energy malnutrition and wasting are strong predictors of death among haemodialysis patients. There is not a single measurement that provides complete and unambiguous assessment of the nutritional status of haemodialysis patients (see below Guideline 2.1). Ideally, a nutritional marker should not only predict outcome, but it should also be an inexpensive, reproducible and easily tested test that is not affected by such factors as inflammation, gender, age and systemic diseases. No such ideal nutritional marker is available at present. Thus the use of a panel of anthropometric and biochemical measurements that correlate with nutritional status is required to assess protein–energy malnutrition in a given individual.

**Guideline 2.1. Diagnosis of malnutrition**

- Malnutrition should be diagnosed by a number of assessment tools including (Opinion):
  - (A) Dietary assessment
  - (B) Body mass index
  - (C) Subjective global assessment (SGA)
  - (D) Anthropometry
  - (E) nPNA
  - (F) Serum albumin and serum prealbumin
  - (G) Serum cholesterol
  - (H) Technical investigations (bioimpedance, dual X-ray absorptiometry, near-infrared reactance)
(A) Dietary assessment

- Every haemodialysis patient should have access to a qualified dietitian (Opinion).
- All haemodialysis patients should receive a care plan and individualized dietary information in writing. Both the care plan and dietary information should be reviewed frequently depending on individual medical conditions and personal circumstances (Opinion).
- All haemodialysis patients should be reassessed and counselled within 1 month after haemodialysis has started (Opinion).
- Malnourished haemodialysis patients should be reassessed and counselled more frequently (Opinion).

Rationale

Dietitians are qualified professionals and experts in the application of science in nutrition and metabolism. Training includes interview and counselling techniques. They enable patients to adapt their regular diet to a diet that includes individual requirements for maintenance haemodialysis (MHD), based on personal circumstances while also recommending nutritional support as and when needed. Most but not all patients will have received nutritional assessments and counselling prior to starting MHD. It is most important to adjust their diet as soon as possible, preferably within 1 month. All dietary information provided should be in writing and details should be recorded in the patient’s care plan. It is essential to evaluate and modify individual dietary regimens after a further month or sooner as needed. Stable MHD patients should be interviewed every 3 or 6 months according to age (<50 years, every 6 months, >50 years every 3 months, see Guideline 1), and dialysis vintage (<5 years, every 6 months, >5 years every 3 months, see Guideline 1) as indicated to improve dietary compliance [1,2]. Hospitalized patients and patients requiring naso-gastric tube feeding or intra-dialytic parenteral nutrition (IDPN) should be assessed within 2–3 days and require follow-up at least once weekly for 2 weeks or until stable. Thereafter follow-up and monitoring can be extended to once per month or as required [1].

Assessment of dietary intake can be obtained by dietary records and/or food questionnaires:

Dietary records. Existing methods to record food intake of individual patients range from 24 h-recall to 3 and 7 days diet diaries, the expertise of a qualified dietitian is essential to complete and calculate these accurately. Dietary assessments are essential as there are no alternatives to calculate nutrient intake, now using special computerized food composition programmes and they are part of a set of methods assessing the overall nutritional status of patients on MHD. Data obtained from unsupervised food recordings and covering a short period of time should be interpreted with caution as results can be subjective and incomplete. Patients may overestimate when their intake is poor or underestimate when their intake is good. Also perceptions of portion sizes differ resulting in inaccurate food assessments [3]. The latter can be overcome by using commercial replica food models or a photographic food atlas [4].

Twenty-four hour dietary recall: Recalling what a patient consumed as food and drink during the previous 24 h is a simple method that requires a minimum of professional input and may be used in the routine follow-up in nutritionally stable patients when there are constraints on dietetic input [2]. It can reveal major imbalances or obvious dietary inadequacies or highlight areas of concern which need further investigation. It is a good starting point for more detailed discussions and counselling. The 24-h recall interview technique depends on memory and patients may underestimate actual intake. Recalling food intake even during the previous 24 h may be difficult for the elderly suffering from memory impairment. Intake is confined to a short period and may not represent a typical food intake reflecting daily variations. Longer recall periods may provide inaccurate information as patients become less motivated and several shorter periods of 2–3 days may provide more accurate information to assess protein and energy intake.

Three days food records: In patients with a stable food intake a period of <7 days may be adequate to assess protein and energy intake. The Kidney Disease Outcomes Quality Initiative (K/DOQI) Recommendations for Nutritional Management [2] suggest a 3-day diary which includes a dialysis day, a weekend day and a non-dialysis day. This provides a closer insight into dietary habits. A 3-day diary is preferred as patients do not always comply with accurate recordings for a longer period due to lack of motivation. Patients should be taught how to complete diaries using household measures and food models if available. A dietary record must include the day and time when meals, snacks and beverages are taken, a description of the food or drink, methods of preparation and amount consumed in restaurants and the amount of consumed convenience and processed foods.

Seven days food records or diet diaries: A minimum of 7 days is required to assess protein and energy intake to stay within 10% of SE, but may not be adequate to assess nutrient intake when these are obtained from few foods, such as Vitamin C, as 36 days are required to obtain the same accuracy [4]. The advantage of a 7-day diary is that variations in food intake over a longer period are included. In order to accurately calculate protein catabolic rate (PCR), dietary protein intake (DPI) and dietary energy intake (DEI), Kloppenburg et al. [5] found that a 7-day period correlated better with the mean of three consecutive PCR measurements and average protein and energy intake.
intake compared with a single measurement during the same period. Dietary protein and energy intake vary considerably from day to day as a result of dialysis treatment sessions and associated disturbances in food intake. In this study, qualified dietitians instructed patients regarding accurate recording techniques using standard household measures to record day to day food intake. Patients were also contacted when their records needed further discussion.

**Appetite assessment**

A specially designed questionnaire can be helpful in addition to food diaries to calculate nutrient intake in a large number of patients during a longer period. In the recent HEMO Study patients (1901 at onset) completed dietary records during an assigned 2-day period (including a dialysis day) after receiving detailed instructions from specially trained dietitians. The follow-up period lasted 7 years [6]. Self-assessed appetite was evaluated with the Appetite and Diet Assessment Tool (ADAT) to monitor changes in appetite and dietary habits on both dialysis and non-dialysis days. Other dietary information affecting nutritional intake was also obtained. Further research is required to assess prospectively the predictive power of the ADAT in its ability to monitor and detect changes in dietary habits and appetite [6]. In another study an Appetite and Dietary Assessment Questionnaire (ADAQ) was developed by Lou et al. [7] to predict inadequate intake in a small number of patients (44) on chronic HD (CHD). Diet-diary assisted recalls (DDAR) were used to evaluate nutritional intake. Dialysis and non-dialysis day’s diet data and PCR differences were also studied. The relationship between ADAQ and protein–energy intakes calculated by DDAR was highly significant. The questionnaire was found to be simple and could be used as a screening tool to detect poor nutrition and correct factors that could lead to malnutrition.

### (B) Body mass index (BMI)

- Haemodialysis patients should maintain a BMI > 23.0 (Evidence level III)

**Rationale**

BMI is known to predict the clinical outcome of disease. BMI is dependent on muscle and fat mass and total body water content, however weight changes over a period of time can still be of clinical value and more so in the case of unplanned weight loss over a short period of time. When assessing BMI it should be remembered that a higher percentage of muscle mass is seen in young people, athletes and body builders and a higher percentage of fat mass in less mobile and elderly patients.

Several studies have shown that a BMI of 23 and higher reduces the risk of morbidity and mortality [8–12]. BMI and anthropometric measurements change with age and dialysis vintage in diabetic and non-diabetic patients [13–15]. In a retrospective analysis, Kopple et al. [8] investigated the relationship between BMI and the rate of mortality in 12 965 MHD patients. BMI was calculated using post-dialysis weight and the mean age of patients was 60.3 years. The National Health and Nutrition Evaluation Survey (NHANES) II data, representing men and women with normal weights, were compared with weights of MHD patients matching in height, gender and were divided in two age ranges, 25–54 and 55–74 years. The results showed that death rates in MHD patients with a BMI in the 10th, the 10–25th and 25–50th percentile were significantly higher compared with men with a BMI in the 50th percentile or higher. Woman show a similar improvement in death rates with increasing BMI. This study also showed that advancing age was strongly associated with odds of death with lower BMI. Thus BMI is a strong predictor of mortality in MHD patients over a 12-month period and that is an independent predictor of increasing mortality rates in patients below the 50th percentile. The 50th percentile corresponds with a BMI of at 23.6 for males and 24.3 for women (see Appendix) [1].

Data from a cohort of 1610 patients of the French Study Group Nutrition in Dialysis indicated that nutritionally stable and well-dialysed MHD patients with a BMI of 23.0 ± 4.5 and albumin concentrations within normal range had an increased survival rate of 89.7 ± 0.8% at 1 year and 78.4 ± 1.2% at 2 years [13,15]. From the Case Mix Adequacy Special Study of the USRDS with a national sample of 3607 MHD patients with a mean age of 58.8 years, Leavey et al. [10] concluded that BMI at baseline was a valuable independent predictor of mortality risk and persisted 5 years later. The prospective DOPPS provided baseline demographic, comorbidity and BMI data on 9,714 MHD patients in USA and Europe during 1996–2000 [11]. Multivariate survival analysis was used to evaluate the relationship between BMI and relative risk (RR) of mortality in MHD patients subdivided by continent, race, gender, tertiles of severity in illness (based on a score derived from comorbid conditions and serum albumin levels), age ranges (<45, 46–64 and >65 years), smoking and diabetic status. Results showed a lowering in the RR of mortality as BMI increased and this was statistically relevant but not for patients in the younger age group of <45 years who were also in the healthiest tertile of comorbidity. A BMI of <20 was consistently associated with the highest mortality risk [11,12]. Abbott et al. [12] also concluded that a higher BMI was associated with improved survival in 1675 cohort patients on MHD with a follow-up of 5 years, in a retrospective study of the USRDS Dialysis, Morbidity and Mortality Wave II Study (DMMS). Results showed that patients with a high BMI > 30 kg/m² had a 5-year survival of 39.8% vs 32.3% for patients with a lower BMI and this was statistically significant (P = 0.001).
Comparing SFT and bio impedance analysis (BIA): Oe et al. [19] evaluated body composition [lean body mass (LBM), body fat (BF) and total body water (TBW)] using SFT and BIA techniques in 20 stable MHD patients pre- and post-dialysis. These authors showed a good agreement between the two techniques ($R = 0.93, P < 0.005$) and proposed that BIA might be the preferred method, as BIA is not operator dependent and requires minimal training to assess fluid status. Kamimura et al. [20] also found that SFT measurements were comparable with BIA and remain interesting for routine body fat assessment. Ninety clinically stable MHD patients were studied; body fat measurements using SFT and BIA were similar ($13.5 \pm 6.2$ kg and $13.7 \pm 6.7$ kg). Further research is recommended to obtain references for body composition assessment that are simple to use in the routine care of MHD patients.

### (E) Normalized protein nitrogen appearance (nPNA)

- Normalized PNA should be measured in clinically stable haemodialysis patients and be above 1.0 g/kg ideal BW/day (Evidence level III) (see Guideline 3).

**Rationale**

Normalized PNA provides an independent and less time consuming assessment of dietary protein intake. Nitrogen balance, the difference between intake and losses, is zero in the steady state or slightly positive. Both net protein breakdown under fasting conditions and dietary protein requirements are strongly influenced by body mass. In order to normalize PNA it should be related to body weight of the patient. When determining nPNA, patients should be stable and neither anabolic nor catabolic [21]. The protein equivalent of total PNA can be estimated from interdialytic changes in urea nitrogen concentrations in serum and urine (see Appendix). A recent study in more than 50,000 US adult haemodialysis patients reported that mortality was lowest for patients having a nPNA between 1.0 and 1.4 g protein/kg BW/day; furthermore, when patients had a decreased nPNA after a 6-month follow-up, the 18-month subsequent mortality increased [22]. PNA should however not be used alone to evaluate nutritional status, but rather as one of several independent measures when evaluating nutritional status.

### (F) Serum albumin and serum prealbumin

- Serum albumin should be above 40 g/l by bromocresol green method (Evidence level III).
- For other albumin assessment methods the target values should be adapted to the above (Opinion).
- Serum prealbumin should be above 0.3 g/l (Evidence level III).

**Rationale**

Serum albumin and prealbumin are two important indicators of nutritional status. Serum albumin is a measure of total body protein mass and is strongly influenced by dietary and metabolic factors. Prealbumin, on the other hand, is a more sensitive indicator of recent changes in protein intake, as it is rapidly depleted during periods of negative nitrogen balance. The target values for serum albumin and prealbumin are given by a recent study [13] and should be adapted for other methods of albumin determination (Opinion).

### (C) Subjective global assessment (SGA)

- SGA should be used to identify severe malnutrition in haemodialysis patients (Evidence level III).

**Rationale**

SGA is based on a combination of subjective and objective features from the medical history and physical examination. A modified version of the SGA has been used in the Canada/United States Peritoneal Dialysis Study (CANUSA) and DOPPS studies (see Appendix). It was demonstrated that lower values of the mSGA were associated with a higher mortality risk [16]. The investigators concluded that in haemodialysis patients malnutrition, as indicated by lower values obtained with the mSGA, was associated with higher mortality risk [16]. In a prospective observational study, it was also shown that patients with the lowest SGA score had higher mortality and hospitalization rates [17]. In a direct comparison with the lowest SGA score had higher mortality and hospitalization rates [17]. In a direct comparison with the mSGA, was associated with a higher mortality risk [16]. The investigators concluded that in haemodialysis patients malnutrition, as indicated by low values obtained with the mSGA, was associated with higher mortality risk [16]. In a prospective observational study, it was also shown that patients with the lowest SGA score had higher mortality and hospitalization rates [17]. In a direct comparison with the determination of body nitrogen content by means of neutron activation analysis it was demonstrated that SGA was able to differentiate severely malnourished patients from those with normal nutrition, but appeared not to be a reliable predictor of the degree of malnutrition [18].

### (D) Anthropometry

- Anthropometry in MHD patients should be assessed immediately after dialysis (Opinion).
- Anthropometry (Mid Arm Circumference (MAC), Mid-Arm Muscle Circumference (MAMC) and four site Skin Fold Thickness (SFT) should be performed by the same individual on the non-fistula arm (Opinion).

**Rationale**

BMI, Four-site skin fold thickness (SFT), mid-arm-circumference (MAC) and mid-arm-muscle-circumference (MAMC) are anthropometric screening methods to assess fat and lean body mass and may detect a potential risk for Protein and Energy Wasting (PEW). These are easy to use, widely available and cost effective tools to help assess nutritional status of patients on MHD but fluid status influences calculations. Four-site SFT, MAC and MAMC: these anthropometric measurements are important for overall nutritional assessment. Measuring muscle mass, MAC and MAMC, is essential to assess muscle mass. It is necessary to perform skin fold thickness at four sites to obtain an accurate assessment of total body fat: triceps, biceps, sub-escapular and iliac crest. The Frisancho Tables (1984) and Durnin and Womersley (1974) equations are used to calculate lean body mass and body fat percentage from obtained details (see Appendix for methods).
Rationale

Serum albumin is recommended for routine measurement because a large body of literature is available, that defines normal serum albumin values and characterizes the clinical factors affecting serum albumin concentrations. Serum albumin, per se, is an indicator of visceral protein stores. During recent years the interactions between inflammation and malnutrition status became complex, as inflammation and dietary protein intake exert competing effects on serum albumin levels [23]. A number of publications demonstrate the relationship between serum albumin concentrations and outcome [24]. Hypoalbuminaemia is a predictor of future mortality [25–29] and cardiac disease [27] at the time of initiation of dialysis and at any time during dialysis treatment. Among 1411 patients enrolled in the HEMO study, those in the low albumin group had significantly greater prevalence of coronary heart disease [30]. Serum albumin should not fall below 40 g/l (measured by the bromocresol green method). Patients with a serum albumin level below 35 g/l have a relative mortality risk of 4 [31], or a 2-year survival of 20% as compared with a 2-year survival of 80% in those with a serum albumin greater than 40 g/l [23].

Serum albumin levels are not only affected by poor energy and protein intake, but also by other factors including inflammation, catabolic and anabolic processes, age, comorbidity, fluid overload (i.e. plasma volume) and urinary albumin losses [32,33]. Albumin synthesis is reduced during the acute phase response. The presence of acute or chronic inflammation limits the specificity of serum albumin as a nutritional marker. Measurements of serum albumin levels is inexpensive, easy to perform and widely available. Since there are currently more than fifty different methods for measuring serum albumin in laboratories, reference values should be known to all nephrologists especially when benchmarking is done in order to compare levels in between centres on the national or international level.

The available literature suggests that prealbumin, also called transthyretin, may have unique validity among the panel of available biochemical nutritional indicators. However, no formal guideline was developed for serum prealbumin so far. Although predicting outcome, more mechanistic understanding of its functions is mandatory. Beside issues of reproducibility, costs inhibited implementation of prealbumin so far. Serum prealbumin is a more sensitive indicator for the nutrition status than albumin due to its shorter half life [34,35]. Prealbumin levels correlate strongly with serum albumin and have shown to provide prognostic value independent of albumin [36]. Because albumin is markedly influenced by inflammation as negative acute phase reactant its levels change more rapidly than prealbumin [37]. Therefore prealbumin is a good indicator of liver anabolic protein synthesis. The half life of serum prealbumin is approximately two days instead of 20 days for albumin [34,35]. Serum prealbumin levels lower than 0.3 g/l predict a relative mortality risk of 2.64 [36]. The patients 2-year survival rate was 50% with a serum prealbumin level < 0.3 g/l and 90% in patients with a prealbumin level > 0.3 g/l. Another cohort of 130 patients observed for 10 years demonstrated that each 0.01 g/l increase in serum prealbumin at enrolment was associated with a 9% decrease in the relative risk of death [38].

(G) Serum cholesterol

- Serum total cholesterol should be measured and be above the minimal laboratory threshold value (Evidence level III).

Rationale and commentary

Serum cholesterol is a component of the lipid profile, recommended for routine measurement, to assess the cardiovascular risk of a given haemodialysis patient [39,40]. Low (<1.5 g/l) or declining serum cholesterol concentrations are predictive of increased mortality risk [31,34,41–45]. Hypcholesterolaemia is associated with chronic protein-energy deficits and/or the presence of comorbid conditions, including inflammation. Individuals with low, low–normal (1.5–1.8 g/l), or declining serum cholesterol levels should be investigated for possible nutritional deficits as well as for other comorbid conditions. The relationship between serum cholesterol and outcome has been described as either ‘J-shaped’ or ‘U-shaped’ with increasing risk for mortality as serum cholesterol falls below approximately 2 g/l or rises above 2–3 g/l. Low levels of cholesterol are confounded by inflammation [45] and are influenced by the same comorbid conditions that affect other nutritional markers (e.g. serum albumin). Predialysis serum cholesterol correlates with serum albumin, prealbumin, creatinine and age [46]. If a patient takes lipid lowering drugs, these should be taken into account in the total cholesterol values.

(H) Technical investigations

Rationale

For the assessment of malnutrition several technical tools are available such as bioelectrical impedance analysis (BIA), whole body dual energy X-ray absorptiometry (DXA), near infrared interactance (NIR) and in vivo neutron activation analysis. In addition the presence of malnutrition can be investigated by means of subjective global assessment. In vivo neutron activation analysis is considered the reference standard for the determination of protein
malnourishment. In a sex- and age-matched study, Allman et al. [47] demonstrated that haemodialysis patients manifested a significantly lower total body nitrogen content, suggesting protein depletion. This observation was confirmed by Rayner et al in a larger group showing that nitrogen levels were more decreased in males (13%) than in females (4%). Later studies demonstrated that a significant proportion of haemodialysis patients had total body nitrogen depletion, expressed as a nitrogen index < 80% being the ratio of measured nitrogen vs the predicted nitrogen for sex-, age- and height-matched controls [48–50]. This seems especially to be the case in older patients [48] and patients starting dialysis late i.e. at low levels of renal function [49]. Pollock et al. [48] demonstrated that patients with a nitrogen index of < 80% had a relative risk of dying of 4.1 compared with patients with a higher index whereas Cooper et al. [51] found a hazard ratio of mortality of 1.6 per 10% of decline in nitrogen index. Several studies demonstrated that compared with in vivo neutron activation analysis, nutritional state analysis by means of anthropometry underestimated the presence of protein malnutrition in haemodialysis patients [47,50,52]. Studies comparing in vivo neutron activation analysis with BIA, DXA or NIR are lacking.

DXA determines in a non-invasive way fat mass, fat-free mass and bone mineral mass and density from which body composition is computed. Thus, protein-energy nutritional status can be assessed. However, there are only limited data comparing DXA-determined body composition of haemodialysis patients with that of healthy subjects. Woodrow et al. [53] demonstrated that in comparison with control subjects patients on chronic haemodialysis have a significant reduction in lean tissue mass. In this study, the investigators also found similar reductions in fat-free mass with BIA, which were not found with skin-fold anthropometry. Data comparing nutritional status between haemodialysis patients and control subjects determined with BIA are scarce. Madore et al. [54] developed an impedance index with which they could demonstrate in a small group of haemodialysis patients that fat mass and lean body mass were significantly reduced in 50% of patients compared with the ideal value obtained from the NHANES II tables, suggesting the existence of malnutrition in these patients. Likewise, Maggiore et al. [55] demonstrated that haemodialysis patients after haemodialysis had lower body weight and a reduced phase angle compared with healthy controls. However, these investigators concluded that bioimpedance indexes were not reliable in detecting clinically overt lean body mass depletion albeit that phase angle was strongly related to patient survival [55]. Likewise, Woodrow et al. [56] found in patients with chronic renal failure compared with control subjects larger errors with BIA and skin fold anthropometry compared with DXA, suggesting that the latter technique is the preferred one.

**Suggestion for future research**

- Validation of the assessment of nutritional state by means of in vivo neutron activation analysis vs BIA, DXA and NIR.
- More frequent use of handgrip testing in clinical research studies.

**Guideline 2.2. Monitoring and follow-up of nutritional status**

- Nutritional status should be followed using the following assessment tools (Opinion):
  - (A) Dietary interviews
  - (B) Body weight
  - (C) nPNA, serum albumin and serum cholesterol
- The use of other technical investigations should be restricted to research purposes (Opinion).

(A) Dietary interviews

- Stable and well-nourished haemodialysis patients should be interviewed by a qualified dietitian every 6–12 months or every 3 months if they are over 50 years of age or on haemodialysis for more than 5 years (Evidence level III).
- Malnourished haemodialysis patients should undergo at least a 24-h dietary recall more frequently until improved (Opinion).

**Rationale**

Dietary interviews are the best way to detect in time a reduced food intake before other objective malnutrition parameters start changing. Depending on staffing constraints a 3-day dietary recording or a 24-h recall of previous day intake should be performed, evaluated and findings must be noted in the patient’s care plan. During the same appointment any new dietary information can be implemented. Patients may need to be seen sooner if abnormal monthly blood tests require dietary intervention. Patients may request to visit the dietitian more often to alter parts of their dietary regimens or changes in their personal circumstances may indicate the need for additional information. Establishing a telephone help line and/or access to internet facilities enables dietitians and patients to communicate more frequently.

The reasons for more frequent interviews in patients over 50 years of age and treated by haemodialysis for more than 5 years have been discussed in Guideline 1.
were independent factors ($R = -0.05$). There was a small but significant positive association between IDWG and serum albumin concentrations: 3.78 vs 3.83 g/dl ($P < 0.001$) in patients with a <3% and >4.5% dry weight gain, respectively. Testa et al. [59] also found that dietary protein and energy intake was higher in patients with a higher IDWG of 4.5 ±1.5% during the 3 days interval. Dietary protein, energy and sodium intake were assessed from 3-day diet diaries from 32 patients, for each patient for 1 year. This study suggests that a stable IDWG may be a clinical indicator of adequate protein and energy intake and that the extent of IDWG was not directly related to blood pressure even in hypertensive patients. Some patients are afraid of gaining more than 2 kg in between dialysis treatment and this may affect their nutritional intake. Nutritional counselling is therefore important and should establish which patients eat well compared with those who do not and have a lower IDWG. Patients with large weight gains should nevertheless be challenged to assess what proportion is nutritive and non-nutritive fluid consumption. Staff should be aware of this when assessing compliance with dietary and fluid intake when discussing ‘individual ideal IDWG’ of 2 kg or less as this may be inappropriate for some patients. A percentage of dry body weight gain of 4–4.5% seems acceptable in patients with an optimal nutritional intake and observing salt restriction.

**Recommendation for future research**

Further studies are required to evaluate what constitutes an ‘ideal’ IDWG for the well nourished and what percentage is acceptable for hypertensive and cardiovascularly unstable MHD patients.

**Table 1.** Significance of unplanned weight loss

<table>
<thead>
<tr>
<th>Unplanned weight loss in past 3–6 months (% body weight)</th>
<th>Significance</th>
</tr>
</thead>
<tbody>
<tr>
<td>&gt;10% of body weight</td>
<td>Clinically significant</td>
</tr>
<tr>
<td>5–10% of body weight</td>
<td>More than normal intra-individual variation (potentially significant) — early indicator of risk of malnutrition increased</td>
</tr>
<tr>
<td>&lt;5% of body weight</td>
<td>Within ‘normal’ intra-individual variation (small)</td>
</tr>
</tbody>
</table>

**Rationale**

It has been suggested that MHD patients should have been on MHD for 60 days as this can reflect ‘dry weight’ more accurately [12]. Ideal body weight (IBW) is the weight based on a range of BMI’s that yields the lowest morbidity and mortality rates. IBW may need to be adjusted in overweight and underweight patients.

Unintentional weight loss during the previous 3–6 months period is more accurate as a risk factor for protein–energy malnutrition than BMI. This weight loss may be categorized according to the British Association of Parenteral and Enteral Nutrition (BAPEN) Malnutrition Advisory Group as in Table 1 [57].

Therefore, a simple cut-off of >10% weight loss during the last 3–6 months can be recommended for the diagnosis of malnutrition.

Typically MHD patients are advised to keep IDWG between 2 and 2.5 kg. Current guidelines for daily fluid intake vary from 500 to 750 ml in addition to daily urine output. Thirst is dependent on dietary sodium (salt) intake and a high sodium intake will contribute to excessive IDWG and may not be the immediate result of food intake itself. Therefore, MHD patients must be advised to reduce their daily sodium (salt) intake to 5–6 g salt (Na 85–100 mmol). However, patients eating well also gain additional weight in between dialysis and this is due to the invisible fluid content of food. A ‘dry’ diet of 2100 Kcal can contain as much as 300–350 ml fluid and this adds to the daily fluid intake.

It has been suggested by Sherman et al. [58] that IDWG could reflect nutritional intake. In a study of 860 randomly selected patients, a relationship between IDWG and nPNA was noted, a higher protein (g/kg) intake was associated with a higher IDWG, confirmed by a correlation analysis that dry weight and nPCR
for protein/energy nutritional status in MHD patients. Hypoalbuminaemia in MHD patients does not necessarily indicate protein–energy malnutrition, which may also not correlate with changes in other nutritional parameters.

Normalized PNA is a valid estimate of protein intake, is well validated and simple to use in the clinical setting. It is important to monitor protein intake in MHD patients. However, there are limitations as well such as overestimation of dietary protein intake when the protein intake is <1 g/kg/day, possibly due to protein catabolism [60,61]. Normalizing PNA to body weight can be misleading in obese and volume overloaded patients. It is recommended that for individuals who are <90% or >115% of standardized body weight, the oedema-free adjusted body weight is used.

Serum total cholesterol is part of the routine lipid profile measured in 3–6 month intervals according to changes in clinical status and during lipid modifying interventions [39]. Similar arguments apply for serum cholesterol as for serum albumin. Serum cholesterol may not correlate with changes in nutrition but also with changes in other nutritional parameters e.g. with those pointing to an activated acute phase reaction.

Technical investigations are not recommended for routine follow up

Rationale

Changes in body composition reflecting nutritional status can be monitored with several techniques although the number of studies is limited and direct comparisons of these techniques with a gold standard are lacking. By means of in vivo neutron activation analysis a declining trend in the nitrogen index was found after 1 year in prevalent haemodialysis patients whereas the nitrogen index correlated with dietary caloric intake [48]. Pupim et al. [62] investigated nutritional parameters for 1 year in 50 incident haemodialysis patients including BIA every 3 months and DXA at the beginning and end of the year. BIA-derived fat mass as well as DXA-measured fat mass increased over time suggesting an improvement of nutritional status. This was not associated with a change in body mass of these patients, which was explained by a decrease in total body water. In prevalent chronic diabetic haemodialysis patients a decrease in DXA-determined fat mass was found after 1 year of treatment which was attributed to impaired nutritional status in these patients [63]. From these studies it can be concluded that technical tools can be used to monitor changes in body composition. Future research, however, should further clarify which method is the preferred one and with what intervals it should be applied.

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Guideline 3. Recommendations for protein and energy intake

Guideline 3.1. Recommended protein intake

- The dietary protein intake in clinically stable chronic haemodialysis patients should be at least 1.1 g protein/kg ideal body weight/day (Evidence level III).
- The achieved nPNA in a clinically stable chronic haemodialysis patient should be at least 1.0 g/ideal body weight/day (Evidence level III).

Rationale

The prevalence rate of protein–energy malnutrition in chronic haemodialysis patients ranges from 20% to 70% with an average of 40% [1–3]. A poor nutrient intake is the most frequent cause for malnutrition in MHD patients. Observational or interventional clinical trials have reported patients’ spontaneous intakes to be as low as 20–25 kcal/kg/day and/or 0.8–1.0 g/kg protein/day [4,5]. Although some patients may do well with slightly lower intakes than recommended, the general dialysis population should be advised to reach a minimal protein intake of 1.1 g protein/kg/day. Protein intake should be taken with a sufficient energy intake (e.g. 30–40 kcal/kg/day, see Recommendation 3.2) to guarantee an optimal metabolic balance.

Protein requirements. There has been controversy regarding the optimal protein intake in MHD patients since clinical studies are scarce and their duration is usually too short (on average less than 10 days) to obtain valid conclusions. In healthy adults, metabolic studies are expressed as a mean ± SD. However, when transferred to the general population, a mean experimental value of a minimal intake means that 50% of subjects will be covered whereas 50% will not be adequately covered by the proposed level. Since there is no method for identifying those patients who will not be in balance by eating this mean value, the World Health Organization defined a ‘population level’ by adding 2 SDs of the mean to the protein intake obtained through metabolic studies [6]. This ‘population level’ is therefore considered safe, since it will ensure that 97.5% of patients would get enough protein to balance their needs. Consequently, this also implies that almost half of the subjects will be counselled to achieve a protein intake above their individual needs. This is the reason why a given subject can remain in metabolic equilibrium, e.g. in adequate nutritional status when receiving a protein intake less than recommended.

For healthy young adults, the most recent recommendations have slightly increased the daily protein intake towards 0.8–0.85 g/kg body weight [6,7]. Furthermore, in a recent meta-analysis, it was not possible to recommend different values for elderly people, nor was it possible to find marked differences in requirement according to the nature of animal or vegetable protein [7]. Thus, a balanced intake of high quality animal protein and vegetable protein source should be proposed.

Protein requirements in the normal population chronic haemodialysis. During routine haemodialysis, protein requirements do not appear to be sufficient for the following reasons. First, the dialysis treatment induces a loss of nutrients (glucose, amino acids, vitamins and trace elements) through the dialysis filter, which may even be more important today in response to the use of more porous membranes and/or more efficient techniques such as haemofiltration [8,9]. Second, the dialysis procedure itself is a catabolic event responsible for protein catabolism (fragmentation of albumin, release of pro-inflammatory cytokines, role of heparin) [9–15]. For example, in response to the rapid decrease in plasma amino acid at the start of the haemodialysis session, muscle proteolysis occurs in order to maintain an adequate plasma and cellular amino acid concentration [16,17]. This catabolic event may lead to muscle wasting over the long term. Feeding patients during the dialysis session through regular meals, special liquid feeding or parenteral administration has been shown to revert this catabolic state and should be used as frequently as possible [11,17,18]. Some authors have hypothesized that, during the non-dialysis days, the catabolic stress may not be present or even be replaced by an anabolic response [19]. Nutrient intake may vary according to the dialysis schedule: food intake was greater by approximately 10% on non-dialysis days than on dialysis days [5], an observation not confirmed by Kloppenburg et al. [20]. During a standard three-weekly dialysis schedule, food intake was recently reported to be spontaneously reduced by 40% on the last day of the long interdialysis interval, probably in order to avoid fluid overload [21]. This last observation fits well with the previous report from Sherman et al. [22] showing that patients with a reduced interdialytic weight gain (<3% dry weight) had a mean nPNA of 0.94 g/kg BW/day, as compared with those who had an interdialytic weight gain >4% corresponding to a nPNA of 1.17 g/kg BW/day.

Research data in dialysis patients indicate that in most metabolic studies performed in adult chronic dialysis patients, a protein intake of 0.8–0.85 g/kg BW/day or less was constantly associated with a
negative metabolic balance [23–27]. When protein intake averaged 1.1 g/kg/day or more, most patients showed neutral or positive balance [23–27] but not all [28]. These observations have led many investigators to recommend a safety level of protein intake of 1.2 g/kg BW/day. After publication of previous nutritional guidelines in renal disease [29, 30], sporadic reports have challenged these recommendations, by reporting good nutritional status in patients eating less protein [31,32]. These observations may have been obtained in selected patients, and for the safety reasons detailed above, lower levels of protein intake should not be recommended for the general dialysis population.

**Protein intake and nutritional status in epidemiological studies in maintenance dialysis.** In a cross-sectional survey of more than 7400 haemodialysis patients, Aparicio et al. [4] showed that serum albumin reached a plateau of 39.3 g/l for a PNA of 1.2 g/kg/day, but no superior serum albumin values were observed in patients with greater nPNA. Additional data have recently been obtained from prospective epidemiological studies [33–35]. Okhawa et al. [33] reported in 127 MHD patients that body composition, as assessed by CT scan, was maintained constant with a level of protein intake of 0.9–1.1 g protein/kg/day, and that there was no clinical or biochemical benefit for the patients eating more than 1.1 g protein/kg/day. Kloppenburg and colleagues performed a randomized cross-over trial comparing two levels of protein intake (0.9 vs 1.1 g/kg BW/day) for 40 weeks each in 45 haemodialysis patients [36]. They did not observe significant changes in nutritional parameters between the two diets which were comparable in terms of energy intake (28–30 kcal/kg BW/day). In a secondary analysis of the HEMO study, serum albumin was shown to be positively associated with protein intake (assessed by equilibrated normalized PCR) only between 0.4 and 1.0 g/kg/day, without further benefit on serum albumin for a nPCR > 1.0 [37].

**Is a protein intake greater than 1.2 g/kg/day harmful in chronic haemodialysis?** Although larger protein intakes may not improve nutritional status, they may possibly be associated with better survival: in a 2-year prospective follow-up of more than 1600 chronic haemodialysis patients in France, higher nPNA was associated with higher survival by univariate analysis [34]. More recently, the same group reported increased survival in patients with an nPNA between 1.24 and 1.46 g/kg BW/day, as compared with the quartiles having an nPNA lower than 1.24. Survival was not further improved in the upper quartile of patients taking 1.46 g protein and above [35]. In another recent 1-year prospective study, Kalantar et al. [38] reported an inverse relationship between PNA (mean value, 1.13 ± 0.29 g/kg/day, range 0.5–2.15) and mortality or hospitalization rate in 122 patients adequately dialysed (Kt/V > 1.2).

**Protein intake and CKD mineral and bone disease.** Elevated protein intakes are not dissociable from an increase in dietary phosphate, which has led some investigators to warn against a potential increase in vascular calcification. Most clinical trials have specifically addressed the question of dietary phosphate restriction only in CKD stages 2 and 3, well before end-stage renal disease (stage 5), in an attempt to prevent secondary hyperparathyroidism [39]. Once dialysis treatment is started however, the relationship between dietary phosphate and hyperphosphataemia is less straightforward, since bone metabolism and intestinal absorption become the focus of complex interactions and new therapeutic interventions [40]. In 39 patients undergoing a 80-week randomized cross-over trial, Kloppenburg et al. [36] reported that two different protein intakes (0.94 vs 1.15 g/kg IBW/day, estimated from food reports, corresponding to a nPNA of 0.9 and 1.0 g/kg/day, respectively) were not associated with different serum phosphate levels (1.88 ± 0.40 vs 1.89 ± 0.39 mmol/l, respectively). Serum phosphate was markedly influenced by the dialysis dose, being lower in the greater dialysis dose group (1.77 ± 0.30 vs 2.01 ± 0.41 mmol/l for Kt/Vs of 1.26 ± 0.14 and 1.02 ± 0.08, respectively), underlining the predominant importance of the dialysis dose over the protein intake in controlling serum phosphate and the phosphocalcic product. Many individuals may have a high serum phosphate without eating a large quantity of proteins, possibly from a greater intestinal fractional absorption of phosphate and the influence of vitamin D therapy, and these patients may better benefit from oral phosphate binders than from a reduction in their protein intake. In contrast, low serum phosphate is frequently associated with low protein intake in patients undergoing regular 4-h or shorter haemodialysis sessions. Indeed, Lorenzo et al. [41] reported that patients with a serum phosphate < 4 mg/dl did eat 0.86 ± 0.3 g protein/kg BW/day whereas those with a serum phosphate > 4 mg/dl had a protein intake of 1.05 ± 0.4 g/kg/day. Finally, and most importantly, there is no prospective clinical trial to show that the vascular risk associated with elevated serum phosphorus or calcium phosphate product occurs in response to a high protein intake.

**Protein intake and frequency of haemodialysis.** The frequency of the dialysis sessions should be considered when analysing nutritional intake. Indeed, fear of overload or pulmonary oedema may significantly limit food intake during the interdialytic interval, particularly during the long 3-day period [21]. Switching patients to a daily haemodialysis program, either nocturnal or short 2-h, has been reported to augment protein intake up to 40%, an increase that was sustained over 1 year and associated with improved serum albumin in almost all pilot studies [42]. The reasons for this improved nutrient intake is probably due to the lifting of the fluid restriction and other general limitations of food intake, especially for those nutrients containing phosphate and/or potassium.
Protein intake and inflammation. Inflammation, which has been repeatedly reported in 20–50% of routine haemodialysis patients, may impair nutritional status by different mechanisms such as increased anorexia and/or protein catabolism [37,43]. Controversial debate occurs as to whether protein intake may reverse impaired nutritional status in the presence of chronic inflammation. In a randomized dietary intervention study, Leon et al. [44] showed that it was possible to increase serum albumin over 6 months in haemodialysis patients by simple dietary counselling, and this improvement also occurred when chronic inflammation was present. In an ancillary analysis of the HEMO study in more than 1000 MHD patients, Kaysen et al. [37] showed that serum albumin was independently influenced by either protein intake or inflammation status. Indeed, serum albumin correlated positively with protein intake (as assessed by ‘nPCR’) only for a protein intake <1.0 g/kg/day, with no further benefit above, and there was no inflammation impact on serum albumin for C reactive protein (CRP) values <13 mg/l [37]. These authors suggested that the independent effect of protein intake might positively impact on nutritional status of inflamed patients with a CRP >13 mg/l. A comparable observation has been reported by Chauveau and colleagues [35] in a recent prospective cohort of more than 400 MHD patients. These authors showed that the 2-year survival of patients with a serum CRP >10 mg/l was superior for the patients with a nPNA ≥1.2 g/kg/day. Thus, the relationship between chronic inflammation, dietary intake and nutritional status still remains unclear but may suggest that malnourished inflamed patients may benefit from increased protein intakes.

Thus from these studies, it seems that nutritional status does not much improve when protein intake is 1.0–1.2 g/kg/day or above, whereas there might be a sustained protective effect on morbi-mortality for protein intakes slightly above these nutritional recommendations. These hypotheses should be confirmed in larger specifically designed prospective studies. Practically however, if an individual dialysis patient eats slightly less (0.9–1.0 g/kg/day) than recommended and presents with a stable nutritional status, in absence of superimposed disease or catabolic event, and until further survival studies become available, his/her protein intakes may be maintained under the recommended level unless clinical and/or biochemical nutritional indices worsen.

Recommendation for further research
- Which PNA gives the best survival in chronic haemodialysis?
- Formula for normalizing PNA.
- Specific protein needs for malnourished HD patients (may differ from well-nourished patients).
- Effect of dialysis techniques on nutritional status (haemofiltration, daily dialysis, etc.) on appetite and their relationship with appetite regulatory factors (leptin, ghrelin).
- Effects of higher protein intake on malnourished inflamed patients.
- Relationship between protein intake, vascular calcification and bone metabolism.

Guideline 3.2. Recommended energy intake
- The recommended energy intake in a clinically stable chronic haemodialysis patient should be 30–40 kcal/kg IBW/day, adjusted to age, gender and to the best estimate of physical activity level (Evidence level III).
- Regular physical activity should be encouraged, and energy intake should be increased proportionally to the level of physical activity (Opinion).

Rationale

Energy metabolism in chronic kidney disease. Energy metabolism may be impaired during CKD, in response to metabolic disorders such as insulin resistance and impaired triglyceride utilization, carnitine deficiency, hyperparathyroidism, metabolic acidosis, chronic inflammation and the haemodialysis procedure itself [45]. However, except in severely sick patients, these abnormalities do not seem to greatly affect resting energy expenditure (REE) [45–50]. Indeed, even if some activities or treatments impact on energy metabolism in CKD, this will occur for only short periods of time in the entire nycthemere and the resulting overcost may not significantly alter the daily energy expenditure (DEE) [6,51]. Energy expenditure has even been shown to be reduced in CKD patients as compared with control subjects [51,52]. The main reason for altered energy metabolism seems therefore to be a predominant deficit in energy intake rather than an increase in energy expenditure. Indeed, many reports in MHD showed energy intake being as low as 20–22 kcal/kg BW/day [53–56]. When normalized by lean body mass, REE may be more elevated in MHD than in peritoneal dialysis for yet unexplained reasons [57], but this normalization does not reflect a general consensus until now [45,53].

How to estimate daily energy expenditure? Estimation of daily energy expenditure (DEE) has been performed by different research tools including indirect calorimetry (sequential or continuous over 24h or more), deuterated water, physical activity questionnaires, and Harris-Benedict or Schofield formulas [6,47,48,57–60]. Daily energy expenditure strongly depends on the active metabolic mass, e.g. lean body mass, but is independent on fat mass [48]. Since excess energy intake is rapidly stored in fat tissue in the body, the optimal daily energy intake (DEI) in a stable adult equals his/her daily energy expenditure. A detailed individual calculation of DEI firstly includes the estimation of resting energy expenditure (REE) also
called basal metabolic rate, strongly influenced by thermic conditions including ambient temperature, and thyroid function.

Resting energy expenditure (REE) can be estimated as follows:

### 3.2.1 By the use of the Schofield tables reported by the WHO [6]:

<table>
<thead>
<tr>
<th>Age (years)</th>
<th>Males</th>
<th>Females</th>
</tr>
</thead>
<tbody>
<tr>
<td>18 – 30</td>
<td>15.3 \times BW + 679</td>
<td>14.7 \times BW + 496</td>
</tr>
<tr>
<td>30 – 60</td>
<td>11.6 \times BW + 879</td>
<td>8.7 \times BW + 829</td>
</tr>
<tr>
<td>&gt; 60</td>
<td>13.5 \times BW + 487</td>
<td>10.5 \times BW + 596</td>
</tr>
</tbody>
</table>

where REE is expressed in kcal/day and body weight (BW) in kg.

### 3.2.2 By the use of Harris-Benedict equations as follows:

For men: \[ REE = 66 + (13.7 \times BW) + (5 \times H) - (6.8 \times A) \]

For women: \[ REE = 655.1 + (9.6 \times BW) + (1.8 \times H) - (4.7 \times A) \]

where REE is expressed in kcal/day, body weight (BW) in kg, height (H) in cm and age (A) in years.

### 3.2.3 By the use of Black equations [61] as follows:

For men: \[ REE = 259 \times BW^{0.48} \times H^{0.50} \times A^{-0.13} \]

For women: \[ REE = 230 \times BW^{0.48} \times H^{0.30} \times A^{-0.13} \]

where REE is expressed in kcal/day, body weight (BW) in kg, height (H) in m and age (A) in years.

The third important and highly variable determinant of daily energy expenditure (DEE) is physical activity. To obtain the optimal DEE, REE should be multiplied by an activity factor, which greatly depends on the type and the duration of professional and recreational activities. This factor is generally comprised between 1.3 and 2, with a mean of 1.5 in most publications [59]. From a recent analysis from 40 MHD patients, Kloppenburg et al. [58] reported that the energy requirement, the DEE is estimated for a 30-year old adult and then reduced by 7 and 10 kcal/year for age above 30 [63]. Blanc et al. [59] reported that the WHO recommendations led to a 10% overestimation of daily energy expenditure in elderly women (mean age, 75 year), underlining the need for further research in larger cohorts of patients. Thus, from the most recent publications in the field, it seems that energy requirements could be lower than previously reported.

Finally, there is a metabolic adaptation to a reduced energy intake, which includes a decrease in resting energy expenditure, both from a loss of active lean body mass, but also through an improved efficiency of energy metabolism, as recently showed by Friedlander et al. [64]. Thus, even though their energy intake does not reach the recommended values, malnourished patients may still benefit from a relative increase in their nutritional intake, from spontaneous or supplemental oral intake, or from oral or parenteral sources during the dialysis session [11,18,65].

How to estimate daily energy intake? Since energy intake in excess of expenditure is rapidly stored as fat in a stable well-nourished haemodialysis patient, the optimal energy intake equals his/her daily energy expenditure. In contrast to protein, estimation of energy intake can only be done by monitoring intake and not by collecting any fluid parameter. There are a lot of difficulties in performing diet collections, among which dietitian availability and patient training, knowledge and perception of exact food intake, time consumption and cost. Precision of food reports are limited and may artefactually underestimate patients’ true energy intake, the magnitude of underestimation being greater in patients with larger BMIs, in both men and women [58]. From a recent analysis from 40 MHD patients, Kloppenburg et al. [66] measured basal metabolic rate and obtained self reports of energy intake. Whereas in general, the daily energy expenditure cannot be lower than 1.2-1.3 \times REE (see above), these authors found that 60% of patients had an
energy intake report lower than 1.27 × REE. Since these patients did not present with symptoms of chronic malnutrition, the authors suggested that daily energy intake was notably underestimated. The under-reporting of energy intake could be improved by increasing the number of dietary interviews: it has been shown that at least four different 3-day dietary interviews separated each by 1 month were necessary to reduce the intraindividual variability of reports [67], the impact of dialysis or non-dialysis day schedule [5], but 5–7 days appear optimal [20]. In addition, a 7-day collection is more conform with reality of intakes, since there is in some patients an important spontaneous intake reduction the ‘seventh-day’, e.g. the last day of the long interdialytic interval, [21]. Training should be performed, including spouse and/or relatives to help identifying nutrient type and size of servings since patients are not able to clearly identify the different sources of nutrients [44]. Finally, it should be emphasized that energy intake cannot be derived from other reported food components. Indeed, for a same amount of protein intake, the variability of energy intake between patients is too large to draw any reliable relationship between protein and energy intake [68]. Thus, physical activity determination and reported dietary energy intake will be the best estimate of patients’ needs.

Is energy intake sufficient in MHD patients? After the publication of previous guidelines [29,30], there was some disagreement between the recommended values as compared with what was reported in observational studies [54–56]. Most epidemiological studies have reported energy intakes lower than recommended, and being as little as 20–25 kcal/kg BW/day. Thus, when clinical or biological indices of malnutrition are found in a given patient (see Guideline 2), a nutritional work-up should be rapidly performed. However, if there is no clear sign of ongoing malnutrition or (a) catabolic process or processes, a number of facts may partly explain the discrepancy between low reported energy intake and patient’s nutritional status. There could be a true inadequate energy intake that will lead to reduced physical activity, altered protein metabolism and muscle losses: this could be corrected by active nutritional support (see Guideline 5). Alternatively, individual energy expenditure was not correctly assessed, in case of a more reduced than estimated physical activity, which is frequent in chronically haemodialysed patients: thus the true energy needs are closer to 30 kcal/kg/day or may even be slightly less in elderly sedentary women (see above formulas for calculation). A third event is the potential underestimation of energy intake through the food reports or dietary interviews, an occurrence recently confirmed in healthy adults [69] as well as in renal failure patients [45,53,66,70].

Energy intake can be markedly increased in MHD patients by administering oral supplements [71,72]. Daily amounts of 500 kcal could be delivered quite easily and the benefits related to these interventions have been recently reported to be better than previously expected (see Guideline 5). Increasing the frequency of dialysis to daily sessions has allowed a generous increase in energy intake in recent reports [42,73]. These interventions have been associated with weight gain, both from fat and lean body mass.

In summary, estimating energy needs and energy intake is a skilled task, and recent findings have underlined the following points: (1) there is a trend for lowering the daily energy requirements in healthy adults, particularly in women; (2) energy expenditure should be more closely matched to physical activity, which may be very variable between individuals, and particularly reduced in dialysis patients; (3) almost all methods for monitoring daily energy intake, even when used by a trained staff, do underestimate actual energy intake. Whether this information applies to dialysis patients is not fully known and may be the scope for further research. However, these points partly explain why, in routine practice, patients may do acceptably well despite recorded energy intakes lower than previously recommended.

References


4. Recommendations for vitamins, minerals and trace elements administration in MHD patients

Due to insufficient evidence from clinical trials for recommending administration of vitamins, the following information only reflects the expert’s opinion and cannot be considered as a clinical guideline but a recommendation.

4.1. Vitamins

Abnormal renal metabolism, inadequate intake and/or gastrointestinal absorption and dialysis losses, account for vitamin deficiencies amongst dialysis patients. Losses are even greater with high-flux and high-efficiency dialysis. Vitamin deficiency progresses slowly depending on body stores, nutritional intake and chronic dialysis losses. Vitamin status in individual patients depends on age, gender, actual vitamin intake, previous supplementation, dialysis losses, residual renal function, time on dialysis and types of dialysers in addition to impaired metabolism. Ideally vitamin supplements should be tailored to individual needs. Overt clinical manifestations include depressed immune system, neuropathy and impaired amino acid and lipid metabolism, mild scurvy and other abnormalities. The most frequently observed vitamin disturbances concern water soluble vitamins and these may be supplemented daily or administered after dialysis, three times weekly, which promotes compliance.

In a recent prospective cohort study, the DOPPS evaluated the relative risk (RR) for hospitalization and mortality in 16,345 MHD patients from 308 randomly selected renal centres in Europe, Japan and USA [1]. There were large regional variations in the percentage of patients who received various multivitamin types of water soluble vitamins. In Europe, this ranged from 3.7% in the United Kingdom to 6.4% in Italy and 37.9% in Spain; it was 5.6% in Japan as compared with 71.9% in the US. Possible reasons for these large variations may be due to differences in cost, health insurance coverage, patient’s preferences and patients and medical staff health beliefs regarding efficacy as several short-term studies have in the past not shown benefits. The DOPPS evaluation showed a 16% reduction in the relative risk for mortality in MHD patients taking water soluble vitamins [2].

However, only a prospective randomized controlled trial would prove that water soluble vitamin supplementation improves outcomes. The authors meanwhile proposed that while awaiting further more robust evidence, prescription of water soluble vitamin supplements, being of minimal medical risk, could be proposed to the patients [1]. If administered, watersoluble vitamin supplements should be taken or infused at the end of the dialysis session. Patients should be discouraged to purchase regular vitamin...
and mineral supplements over the counter as requirements differ from those for healthy people, and some formulas include vitamins that are not recommended in maintenance dialysis.

4.1.1 Water-soluble vitamins

Thiamine (B1)

- **A daily supplement of 1.1–1.2 mg thiamin hydrochloride is recommended**

**Rationale.** Thiamine deficiency is responsible for beri-beri, a rare condition in MHD patients. Vitamin B1 deficiency may also be evoked in case of atypical neurological symptoms (Wernicke encephalitis). Thiamine is strongly removed during haemodialysis. Thiamine plasma concentration may not reflect its biological activity. Thiamine intake in MHD patients can range from 0.6 to 1.5 mg/day depending on individual food consumption, and is mainly contained in pork meat, beer and dried vegetables [3]. Patients with a poor nutritional intake, as may occur in the elderly, are most likely to benefit from supplementation. Thiamine has been administered in amounts up to 300 mg/week in patients undergoing high-flux haemodialysis [4]. Presently, all renal multivitamin formulas include thiamine, from 1.5 mg (Nephrovite®, Dialyvite3000®, Diatx®, USA, Renavit®, Germany), 3 mg (Renax®, USA) to 50 mg (Dialvit®, Switzerland) per tablet.

Riboflavin (B2)

- **A daily supplement of 1.1–1.3 mg is recommended**

**Rationale.** Although it is well cleared during haemodialysis, not tightly bound to proteins, riboflavin deficiency is uncommon. A supplement of 1.1–1.3 mg is equal to the recommended daily allowance of healthy people and is sufficient to supplement inadequate nutritional intake and dialysis losses [3]. Riboflavin is contained in milk, bread and cereals, lean meat and egg. Presently, all renal multivitamin formulas include riboflavin, from 1.7 mg (Nephrovite®, Dialyvite3000®, USA, Renavit®, Germany), 2 mg (Renax®, USA) to 10 mg (Dialvit®, Switzerland) per tablet.

Pyridoxine (B6)

- **A daily supplement of 10 mg as pyridoxine hydrochloride is recommended**

**Rationale.** There is evidence that plasma and red cell pyridoxine levels are low in MHD patients. Although the pyridoxine recommended dietary allowance in healthy adults is 1.3–1.7 mg, the use of erythropoietin (EPO) may increase requirements because of increased erythropoiesis. Some drugs and other substances interfere with pyridoxine metabolism, an additional cause for deficiency. A decreased level of pyridoxine may be associated with hyperhomocysteinaemia, but the benefit of supplementation is as yet unclear [3,5]. Pyridoxine is contained in yeast, cereal buds, green vegetables, egg yolk and meat. A supplement of 10 mg/day is recommended as this is the lowest pyridoxine hydrochloride dose that has consistently normalized pyridoxine deficiency and the transamination activation index of stable MHD patients. Pyridoxine supplementation given to correct hyperoxalaeia or hyperhomocysteinaemia is still a controversial issue. High doses of pyridoxine hydrochloride (200–600 mg daily) should be avoided as these have been associated with peripheral neuropathy. Presently, all renal multivitamin formulas include pyridoxine, 10 mg (Nephrovite®, Dialyvite3000®, USA, Renavit®, Germany), 15 mg (Renax®, USA), 40 mg (Dialvit®, Switzerland) and 50 mg (Diatx Zn®, USA) per tablet.

Ascorbic Acid (vitamin C)

- **A daily supplement of 75–90 mg is recommended.**

**Rationale.** Vegetables and fresh fruit are the main sources of vitamin C but these foods are often restricted or need to be avoided in a potassium restricted diet, resulting in an inadequate intake. In addition, vitamin C is inactivated by heat during cooking. Vitamin C is readily removed by dialysis as reported by Wang et al. [6]. Serum levels fell by 30–40% after a single dialysis session and losses from 80 to 280 mg per dialysis session have been reported [3]. Vitamin C deficiency contributes to a mild form of scurvy sometimes seen in MHD patients, may lead to abnormal amino acid metabolism and disturbances in folic acid metabolism. Although high-flux dialysis techniques increase vitamin C losses, Descombes et al. [4] reported normal plasma ascorbate values in patients receiving 500 mg vitamin C at the end of the dialysis session thrice weekly. Vitamin C supplements appear to improve functional iron deficiency and hence the response to EPO [7–9].

Vitamin C supplementation may help to relieve muscle cramps. In a double blind randomized trial, 60 MHD patients, divided into four groups, daily received either vitamin E (400 mg), vitamin C (250 mg), either vitamins or a placebo for 8 weeks [10]. Muscle cramps significantly improved in patients receiving both vitamins E and C (97%), vitamin E alone (54%), vitamin C (61%) as compared with only 7% of placebo-treated patients [10].

More recently, Deicher et al. [11] reported that, in MHD patients followed for 30 months, total vitamin C plasma levels were predictive of mortality, with a risk of dying at least three times greater in the plasma vitamin C tertiles <60 μmol/l, suggesting to keep patients plasma vitamin C levels above this target. Further studies are required to explore safety, to what extent patients are vitamin deficient, and whether other dialysis factors may increase removal. High doses of vitamin C (e.g. superior to 500–1000 mg daily) should however be avoided in MHD patients because of tissue oxalate deposition in response to increased
serum oxalates not cleared by the failing kidney. Presently, renal multivitamin formulas generally include vitamin C, e.g. 50 mg (Renax®, USA), 60 mg (Nephrovite®, Diatx Zn®, USA, Renavit®, Germany), 100 mg (Dialyvite3000®, USA) and 200 mg (Dialvit®, Switzerland) per tablet.

**Folic Acid (Folate, vitamin B9)**

- A daily supplement of 1 mg folic acid is recommended.

*Rationale.* In MHD patients, folic acid levels may be reduced in serum and red blood cells and induce megaloblastic anaemia. Folic acid is contained in yeast, liver, green vegetables, fruit and meat. Because of impaired intestinal absorption, ethanol or drug interaction and dialysate losses, particularly with high flux/high efficiency dialysis [12], it is prudent to prescribe 1 mg folic acid/day to prevent deficiency. This may be insufficient to lower elevated plasma homocysteine levels as the administration of 5–10 mg/day has shown a plasma homocysteine reduction by 30–50% [3]. Indeed, the National Kidney Foundation Task Force on Cardiovascular Disease issued a report with recommendations for treatment of hyperhomocysteinaemia [13]. It was recommended that MHD patients should receive daily 5 mg folic acid, 50 mg pyridoxine and 400 μg vitamin B12, to reduce serum homocysteine levels and protect against cardiovascular disease.

Presently, all renal multivitamin formulas include folic acid, 0.8 mg (Renavit®, Germany), 1 mg (Nephrovite®, Dialyvite®, USA), 3 mg (Dialyvite 3000®, USA, Dialvit®, Switzerland) and 5 mg (Diatx Zn®, USA) per tablet.

**B12 (cobalamin)**

- A daily supplement of 2.4 μg vitamin B12 is recommended.

*Rationale.* Vitamin B12 or cobalamin, combined with the gastric intrinsic factor, are necessary factors for an optimal folate metabolism, a normal non-megaloblastic erythropoiesis and to avoid nervous system demyelination observed in pernicious anaemia. Cobalamin is found in sufficient amounts in meat, liver, seafood, milk and egg yolk. Vitamin B12 undergoes an enterohepatic cycling. Most MHD patients present plasma levels of cobalamin in the normal range, whether they receive vitamin B12 supplements or not. Administration of vitamin B12 has been shown to improve or correct nerve conduction velocity in MHD patients having low vitamin B12 plasma levels [14]. Vitamin B12, when administered for 1 mg monthly, is also efficient in decreasing serum homocysteinaemia by ~10% [15]. Since there is no clear report of vitamin B12 toxicity even for high vitamin B12 doses, i.e. 2.5 mg three times weekly [16], and because some dialysis patients have an intake below the daily requirements, a daily supplement of vitamin B12 equal to the requirement, e.g. 2.4 μg/day, seems safe.

Presently, most but not all renal multivitamin formulas include vitamin B12, 6 μg (Nephrovite®, Dialyvite®, USA, Renavit®, Germany), 12 μg (Renax®, USA) and 1 mg (Dialyvite3000®, USA) per tablet.

**Niacin (vitamin B3, nicotinamide, nicotinic acid, vitamin PP)**

- A daily supplement of 14–16 mg niacin is recommended.

*Rationale.* Niacin is contained in meat, fish, dry vegetables, coffee and tea. A deficit in niacin store results in signs of pellagra, a dermatosis associated with diarrhea and dementia, as soon as 50–60 days after a complete dietary niacin removal. However, pellagra has never been reported in a chronic dialysis patient. Niacin undergoes a rapid metabolic clearance and does not seem to be cleared by dialysis. Pharmacological niacin doses improve lipid profile by increasing serum HDL and decreasing LDL cholesterol fraction and serum triglycerides. Since many MHD patients have limited intakes of food containing niacin, it is recommended to supplement patients with the required allowance of normal adults, e.g. 14–16 mg daily.

Recently, niacin, given at about 1000 mg daily has been reported to efficiently decrease serum phosphate by 20% in hyperphosphataemic MHD patients, by inhibiting intestinal phosphate transport [17]. A mild thrombopenia was recently reported as a side effect of this treatment dose in maintenance dialysis [18]. High doses of niacin have been alternatively proposed for controlling dyslipidaemia and reduce cardiovascular risk in non-renal patients [19]. Side effects of high-dose niacin (1000–1500 mg daily) include flushing and impaired glucose metabolism [20]. Thus, high doses of niacin should be prescribed with great caution in dialysis patients, since no long-term clinical trial has been performed in these patients.

Presently, many renal multivitamin formulas include niacin, 20 mg (Nephrovite®, Dialyvite®, Dialyvite3000®, Diatx Zn®, Renax®, USA, Renavit®, Germany) per tablet.

**Biotin (vitamin B8)**

- A daily supplement of 30 μg biotin is recommended.

*Rationale.* Major sources of biotin (vitamin B8) include yeast, egg yolk, liver, soybean, mushrooms and cauliflower. Biotin deficiency may be responsible for depression, somnolence, hyperesthesia, anorexia and dermatosis, symptoms often present to a certain extent in MHD patients. In renal patients, a decrease in intestinal biotin absorption has been reported, as well as a plasma biotin decrease during the dialysis session [3]. Furthermore, food intakes that are low in protein are also low in biotin and do not meet the minimal daily biotin requirement. An adequate biotin
intake has been proposed at 30\(\mu g/day\), and for the aforementioned reasons, it seems prudent to recommend this value also to MHD patients. Clearly, further studies are needed to better address the biotin needs in maintenance dialysis.

Presently, many renal multivitamin formulas include biotin, 150 \(\mu g\) (Nephrocaps\textsuperscript{14}, USA) and 300 \(\mu g\) (Dialyvite\textsuperscript{15}, Diatx Zn\textsuperscript{15}, Nephrovite\textsuperscript{15}, Renax\textsuperscript{15}, USA, Renavit\textsuperscript{15}, Germany) per tablet.

**Pantothenic acid (vitamin B5)**

- A daily supplement of 5\(mg\) pantothenic acid is recommended.

**Rationale.** Pantothenic acid is widely spread in many food including liver, kidney, fresh vegetables and egg yolk. It plays an important role in \(\beta\)-oxidation, free fatty acid and amino acid oxidation and protein acylation. To date, there is no clear information on pantothenic acid stores for MHD patients. Pantothenic acid is cleared by dialysis, and although no data are yet available, newer more efficient techniques might increase pantothenate losses. Since diets low in protein may not provide the adequate daily needs (5\(mg/day\)), it is recommended that MHD patients take a supplement of 5\(mg/day\). Further research is warranted on pantothenic acid dialysate losses and stores in dialysis patients.

Presently, many renal multivitamin formulas include pantothenic acid, 5\(mg\) (Nephrocaps\textsuperscript{14}, USA) and 10\(mg\) (Nephrovite\textsuperscript{15}, Dialyvite\textsuperscript{15}, Diatx Zn\textsuperscript{15}, Renax\textsuperscript{15}, USA, Renavit\textsuperscript{15}, Germany) per tablet.

4.1.2 Fat-soluble vitamins

Vitamin D is not considered in this section as its metabolism, effect and administration in MHD patients depend on phosphocalcic metabolism and bone status, and has been the focus of a recent set of guidelines [21].

**Vitamin A (retinol)**

- A daily intake of 700–900\(\mu g\) is recommended.
- Vitamin A supplements are not recommended.

**Rationale.** Vitamin A is found in dairy products, fish oil, liver, spinach and carrots. Vitamin A is necessary for night vision and epithelium maintenance. Serum plasma levels of vitamin A are elevated in patients with chronic kidney disease. Vitamin A is not removed during MHD and deficiencies are rare and mostly related to inadequate nutritional intake. Vitamin A toxicity includes hypercalcaemia, anaemia and hypertriglyceridaemia. In order to prevent vitamin A toxicity, supplements containing larger amounts than 700–900\(\mu g/day\) should not be given to MHD patients. Patients receiving total parenteral nutrition (TPN) may require vitamin A supplements, but not greater than 700–900\(\mu g/day\) [3,5].

**Vitamin E (alpha-tocopherol)**

- A daily supplement of 400–800\(IU\) is recommended in secondary prevention of cardiovascular events and for preventing recurrent muscle cramps.

**Rationale.** Vitamin E is a strong antioxidant and cell membrane protector. Vitamin E is mainly found in vegetable oils (corn, sunflower and soybean) and wheat germ. Vitamin E plasma levels are not influenced by the dialysis session, and no vitamin E is found in the spent dialysate. There is no decrease in vitamin E plasma levels in long-term MHD patients [22]. The potential benefits of vitamin E supplementation were addressed in a large randomized controlled trial (Secondary Prevention with Antioxidants of Cardiovascular disease in End-stage renal disease, SPACE). A total of 196 MHD patients with pre-existing cardiovascular disease were randomly assigned to either a treatment group (97 patients) receiving 800\(IU\) vitamin E or a control group (99 patients) receiving a placebo. Patients were followed up for a median of 519 days. The primary end point was myocardial infarction, ischaemic stroke, peripheral vascular disease, unstable angina. Sixteen percent of patients on vitamin E vs 33\% on placebo proceeded to a primary endpoint and 5.1\% on vitamin E vs 17.2\% on placebo suffered from myocardial infarction, both reductions being highly significant in favour of the use of vitamin E [23].

Vitamin E supplementation appears to be effective in reducing the incidence of leg cramps. Roca et al [24] compared the effect of quinine and vitamin E in 40 patients with a history of leg cramps who were randomized to either quinine 325\mg or vitamin E 400\IU taken daily at bedtime for 2 months. Quinine and vitamin E were equally effective in reducing cramps as compared with the wash-out period, but due to potential toxicity of quinine, vitamin E should be recommended as treatment of choice [24]. As shown previously for vitamin C in the Khajehdehi et al. trial [10], vitamin E (400\mg) daily alone or in combination with vitamin C (250\mg) daily was able to alleviate muscle cramps significantly when patients receiving both vitamins E and C (97\%) or vitamin E alone (54\%) whereas only 7\% of the placebo group patients improved [10].

Presently, some but not all renal multivitamin formulas include vitamin E, 30\IU (Dialyvite3000\textsuperscript{15}, USA) and 35\IU (Renax\textsuperscript{15}, USA) per tablet.

**Vitamin K**

- A daily intake of 90–120\(\mu g\) is recommended.
- There is no need for vitamin K supplementation, except in patients receiving long term antibiotic treatment or those with altered coagulant activity; a daily amount of 10\mg vitamin K may be temporarily administered.

**Rationale.** Vitamin K is contained in green leaves vegetables (cabbage, spinach) and cow milk.
Vitamin K undergoes intestinal reabsorption through enterohepatic cycling, which may be reduced during oral antibiotic administration. Vitamin K (K for Koagulation in German) is essential in promoting synthesis of II, VII, IX and X coagulation factors but also of some coagulation inhibitors such as factor C, S and Z. Vitamin K is a cofactor for the γ-carboxylation of glutamate in proteins (GLA-proteins) such as the matrix GLA-protein and osteocalcin, explaining a potential role of vitamin K deficiency in patients with bone fractures. In addition, high plasma vitamin K levels have been associated with soft tissue calcifications in MHD patients and elevated serum parathormone values were found in patients with a low plasma vitamin K level [25]. The daily recommended allowance for healthy individuals is 90–120 μg [26]. There is no evidence that MHD patients suffer from vitamin K deficiency. Patients who are treated with antibiotics for prolonged periods and who have a poor nutritional intake may benefit from a 10 mg/day vitamin K supplement. Patients receiving TPN may require 7.5 mg vitamin K per week [5].

4.2. Minerals

Phosphate (phosphorus)

- A daily intake of 800–1000 mg phosphate is recommended.
- Dietary education improves phosphate control.
- Dietary phosphate control should not compromise protein intake.

Rationale. Dietary phosphate intake should be restricted in MHD patients to avoid hyperphosphataemia leading to secondary hyperparathyroidism. The consequences and treatment of hyperphosphataemia are well known and have been reviewed recently in the context of the management of renal bone disease [21,27]. Foods with a high protein content may contain 12–16 mg phosphate per gram protein, with dairy products having the highest ratio. Thus, a protein intake of 80 g (optimal for a MHD patient weighing 70 kg) will bring about 1100 mg phosphate daily. Since 40–80% of the oral phosphate load will be absorbed, depending on vitamin D administration, the net phosphate gain for two days will be 800–1700 mg. Because one standard haemodialysis session can only clear 500–700 mg phosphate, this will result in a positive phosphate balance, an increase in calcium–phosphate product, and a greater cardiovascular risk [21]. However, compromising protein intake at the expense of phosphate restriction should be avoided. Foods high in protein but with the least amount of phosphate should preferably be prescribed through a detailed dietitian interview. Hyperphosphataemia should be treated by intensive counselling, by increasing phosphate binders and by reviewing the dialysis regimen as appropriate.

Calcium

- The total intake of elemental calcium should not exceed 2000 mg/day including calcium obtained from calcium-based phosphate binders.

Rationale. Calcium intake may be limited due to dietary phosphate restriction (milk and dairy products). Overall, a mean food calcium intake is comprised between 500 and 800 mg/day. However, other sources of calcium include calcium-based phosphate binders, and thus the total daily intake of calcium could be much greater, leading to a positive calcium balance, vascular calcifications and episodes of hypercalcaemia. For these reasons, the total amount of oral calcium intake including calcium-based phosphate binders should not exceed 2000 mg daily, and non-calcium phosphate binders should be used if hyperparathyroidism is not controlled. The consequences and treatment of altered phosphocalcic metabolism have been extensively reviewed in recent guidelines regarding the management of renal bone disease and metabolism [21].

Sodium and fluid

- A daily intake of no more than 80–100 mmol (2000–2300 mg) sodium or 5–6 g (75 mg/kg BW) per day of sodium chloride is recommended.
- Interdialytic weight gain (IDWG) should not exceed 4–4.5% of dry body weight.

Rationale. The importance of controlling interdialytic weight gain (IDWG) by restricting dietary sodium (and fluid intake) and the preference for using lower sodium dialysate, has been described in the Haemodynamic Instability Guideline 2.1.

With progressive loss of urine output, sodium and fluid restrictions are vital to control extra cellular volume, blood pressure and to prevent excessive IDWG in anuric and oliguric MHD patients. By reducing the sodium load from diet and dialysate, the lesser urge for patients to quench their
thirst improves compliance with fluid restriction and reduces IDWG. A reduction in sodium intake to 80–100 mmol/l (5–6 g salt) in addition to lowering the dialysate sodium concentration from 140 to 135 mmol/l appears to be sufficient to suppress thirst and hence excessive weight gain. This also benefits blood pressure control and might result in the withdrawal of antihypertensive treatment in some patients [30].

The majority of dietary sodium, 70–80%, is derived from salt and mono sodium glutamate added to food at home, in restaurants and food outlets or by food manufacturers. Examples of some convenience foods are: ready to eat meals, cured meat and fish products, canned and processed foods. The salt content of some staple foods such as breakfast cereals (i.e. cornflakes), bread, butter and margarine and sandwich fillings contribute significantly to dietary sodium intake.

In anuric patients, each 8 g NaCl (140 mmol Na+) requires 11 of fluid intake to maintain normal serum sodium. Dietary Na+ intake (mmol) may be calculated from average daily fluid weight gain (kg) × average serum Na+ concentration (mmol/l). An 80 kg dialysis patient with 4% IDWG, will have 12 g NaCl intake per day. Current guidelines for daily fluid intake vary from 500 to 1000 ml in addition to daily urine output to achieve an IDWG of 2–2.5 kg or 4–4.5% dry body weight. Some dialysis centres include the amount of ‘hidden’ fluid in food in fluid allowance prescriptions. Individual fluid allowances need to be adapted for patients living in warmer climates, during periods of hot weather, working in hot environments and as a result of clinical conditions (high fever). However, it is more efficient to carefully monitor salt rather than fluid intake, since as a response to salt intake, thirst will regulate the subsequent fluid ingested.

All foods that are liquid at room temperature (18–20°C) should be counted as fluid except oil and foods with a high fat or sugar content. Reducing sodium and fluid in addition to a potassium and phosphate restriction and ensuring that protein and energy intake is adequate, is difficult and a stepwise approach to educate the patient is most important. MHD patients must be advised to avoid those convenience foods that contain potassium chloride or other potassium containing additives to replace salt.

Potassium

- In patients with a pre-dialysis serum potassium greater than 6 mmol/l, a daily intake of potassium of 50–70 mmol (1950–2730 mg) or 1 mmol/kg IBW is recommended.

Rationale. Hyperkalaemia is a potential cause of sudden death in MHD patients. There are no warning signs and when pre-dialysis serum potassium levels approach 6 mmol/l, nutritional counselling to lower dietary potassium is indicated, in addition to Calcium Resonium® or Kayexalate®. However, other causes for hyperkalaemia should also be investigated and corrected such as metabolic acidosis together with a review of drug therapies that contribute to hyperkalaemia such as angiotensin-converting enzyme inhibitors, angiotensin receptor blockers, spironolactone, β-blockers, non-steroidal anti-inflammatory drugs and other contributing drug therapies. Tissue destruction (e.g. catabolism) as a result of trauma or weight loss releases potassium from intracellular space and results in hyperkalaemia in haemodialysis patients [31].

4.3. Trace elements

Iron (Fe)

- A daily intake of 8 mg Fe for men and 15 mg for women is recommended.
- Supplementary Fe should be given to all haemodialysis patients treated with an erythropoiesis-stimulating agent (ESA), to maintain adequate serum transferrin and serum ferritin levels, aimed to achieve a target haemoglobin (Hb) concentration >110 g/l or a haematocrit >33%, except for those receiving the iron intravenously.

Rationale. The Institute of Medicine (USA) published the Dietary Reference Intakes (DRI) in 2001 and recommended a daily iron allowance for adults of 8 mg for men and 15 mg for women [26]. Fe deficiency is common in MHD patients and is mainly due to blood losses during dialysis, frequent blood testing, blood remaining in dialysers and gastrointestinal bleeding. Iron absorption from food and oral supplements may be impaired due to increased gastric pH levels as a result of phosphate binder and antacid use. Oral Fe supplements should be taken between meals (at least 2 h after and 1 h before a main meal) to maximize Fe absorption and should not be taken with phosphate binders. Oral Fe supplements are known to cause adverse gastrointestinal effects and compliance with drug therapy may be compromised. Indeed, most MHD patients will receive Fe supplementation, intravenously or orally, as described in detail in the updated European Best Practice Guidelines for management of anaemia in patients with chronic renal failure [27].

Zinc (Zn)

- A daily nutritional intake of 8–12 mg of elemental zinc (Zn) for women and 10–15 mg for men is recommended.
- Routine zinc supplementation is not recommended.
- A zinc supplementation of 50 mg Zn element per day for 3–6 months should be considered in haemodialysis patients with a chronic inadequate protein/energy intake and symptoms evoking zinc deficiency (impaired taste or smell, skin fragility, impotence, peripheral neuropathy).
Rationale. Zinc deficiency is rare in western countries since zinc is absorbed in large quantities from protein rich foods such as red meat, fish and shellfish, milk and milk products, poultry and eggs. Zinc is albumin bound and plays an important role in protein, carbohydrate, energy, nucleic acid and lipid metabolism [32]. The Institute of Medicine (USA) recommends for healthy adults a daily zinc intake of 8 mg for women and 11 mg for men [26]. In the United Kingdom, recommendations are slightly different, 7 mg for women and 9 mg for men [33]. Matson et al. [34] and Kalantar-Zadeh et al. [35] recommend 12 mg of elemental zinc for women and 15 mg for men.

Early signs of deficiency include defects in rapidly dividing tissues such as skin, intestinal mucosa and immune response, decreased taste acuity with a loss in taste buds, impotence, glucose intolerance and hyperlipidaemia. Taste and smell impairment associated with chronic uremia contributes to anorexia leading to a reduced food intake that includes protein and may result in zinc deficiency [5]. Zinc deficiency in uremic patients may contribute to peripheral neuropathy [36]. Oral iron supplements, calcium-based phosphate binders and corticosteroids may promote zinc deficiency. Although improvements in taste, smell, appetite, wound healing, immune response and sexual function have been reported when zinc supplements were prescribed, results were not supportive in earlier studies. Most involved a small number of MHD patients, were of short duration whereas zinc concentration levels may vary due to different laboratory techniques. Zinc supplementation should be given for at least 3 months since shorter trials did not show expected improvements on taste [34] or immune system [36,37]. It was shown in a 3-month randomized crossover trial that zinc supplementation, 50 mg Zn element per day for 90 days significantly increased serum zinc level from low to normal and also increased nPCR and serum cholesterol [38,39]. In more observational reports, nerve conduction velocity improved with zinc supplementation [40] as well as sexual potency [41] but not all studies have confirmed this [42].

Zinc sulphate is a gastric irritant and should be taken with meals. Zn acetate, Zn aspartate and Zn chloride seem to be better tolerated even on an empty stomach [36,41]. Adding zinc to the haemodialysate may be considered if side effects associated with oral supplementation prohibit their use. During a randomized crossover study, serum zinc levels, taste acuity and nerve conduction velocity improved by adding zinc to dialysate during 12 weeks and achieving an increase in serum zinc from 10.1 ± 1.3 to 23.1 ± 0.7 μmol/l (N = 13.8 ± 1.9) [40]. Once this supplement was discontinued at the end of the 3 months supplementation, taste acuity reduced. Serum zinc levels lowered to baseline levels indicating that zinc supplementation should have been continued to maintain normal zinc levels.

Presently, some but not all renal multivitamin formulas include zinc, 15 mg (Dialyvite3000®, USA), 20 mg (Renax®, USA) and 50 mg (Dialyvite 3000-Zinc®, USA) per tablet.

Selenium (Se)

- A daily intake of 55 μg of selenium is recommended.
- Routine selenium supplementation is not recommended.
- A selenium supplementation for 3–6 months should be considered in haemodialysis patients with symptoms evoking selenium deficiency (cardiomyopathy, skeletal myopathy, thyroid dysfunction, haemolysis, dermatosis).

Rationale. Selenium is an essential trace element leading to an adequate glutathione peroxidase (GPX) activity that protects cells from lipid peroxidation. Thyroid function regulation depends on selenium. The recommended intake for healthy males and females is 55 μg/day [26]. In case of acute oxidative stress, selenium needs may increase up to 100–150 μg/day. Intestinal absorption is thought to be 50–65% [32]. The main sources of selenium are meat, fish, fat, vegetables and cereals. However, selenium content of food depends on the selenium content of local soil on which crops have grown or animals have grazed. A severe cardiomyopathy has been reported in the region of Keshan, China, where there is an extreme lack in selenium in earth and food, leading to very low selenium intake (<15 μg/day) and low serum selenium in humans. This cardiac disease is reversed by administering a selenium supplement. Other clinical symptoms of altered selenium metabolism include skeletal muscle dystrophy, haemolysis and dermatosis.

Low serum selenium in CKD and MHD patients are not uncommon. There is no recommendation for selenium supplementation for CKD patients but if prescribed, selenium levels should be monitored closely, as selenium is excreted by the kidney and not removed by dialysis [5]. Selenium supplementation might be helpful in partially improving thyroid function in MHD patients. In a randomized control trial, Napolitano et al. [43] administered selenium to stable MHD patients. Ten patients received sodium selenite supplements orally, 500 μg three times weekly for the first 3 months followed by 200 μg three times weekly for the next 3 months, whereas five patients received a placebo. Selenium supplementation was well tolerated and a significant increase in serum selenium was observed as well as an improvement in thyroid function tests (i.e. a reduction in TSH) and an improvement in immune parameters in patients receiving selenium. No side effect was reported [43,44]. In a small pilot trial, Richard et al. [45] administered selenium intravenously as sodium selenite in six MHD patients, 50 μg at the end of the dialysis session three times weekly for the first five weeks then 100 μg for the next 15 weeks. This treatment was able to increase serum selenium levels and restore glutathione peroxidase activity to normal.
Presently, some renal multivitamin formulas include selenium, 70 μg per tablet (Dialyvit3000® and Renax®, USA).

Table 2. Recommended dietary intake and supplements of vitamins and trace elements in adult haemodialysis patients (opinion)

<table>
<thead>
<tr>
<th>Vitamins</th>
<th>Daily recommendation</th>
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</thead>
<tbody>
<tr>
<td>Thiamine hydrochloride (B1)</td>
<td>1.1–1.2 mg supplement</td>
</tr>
<tr>
<td>Riboflavin (B2)</td>
<td>1.1–1.3 mg supplement</td>
</tr>
<tr>
<td>Pyridoxine hydrochloride (B6)</td>
<td>10 mg supplement</td>
</tr>
<tr>
<td>Ascorbic Acid (C)</td>
<td>75–90 mg supplement</td>
</tr>
<tr>
<td>Folic Acid (B9)</td>
<td>1 mg supplement</td>
</tr>
<tr>
<td>Cobalamin (B12)</td>
<td>2.4 μg supplement</td>
</tr>
<tr>
<td>Niacin (B3, nicotinamide, nicotinic acid, PP)</td>
<td>14–16 mg supplement</td>
</tr>
<tr>
<td>Biotin (B8)</td>
<td>30 μg supplement</td>
</tr>
<tr>
<td>Panthenic acid (B5)</td>
<td>5 mg supplement</td>
</tr>
<tr>
<td>Retinol (A)</td>
<td>700–900 μg intake</td>
</tr>
<tr>
<td>Alpha-tocopherol (E)</td>
<td>400–800IU supplementa</td>
</tr>
<tr>
<td>Vitamin K</td>
<td>90–120 μg intake</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Minerals and Trace elements</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Phosphorus</td>
<td>800–1000 mg intake</td>
</tr>
<tr>
<td>Calcium</td>
<td>2000 mg intake including calcium from phosphate binders</td>
</tr>
<tr>
<td>Sodium</td>
<td>2000–2300 mg intake</td>
</tr>
<tr>
<td>Potassium</td>
<td>50–70 mmol intake</td>
</tr>
<tr>
<td>Iron</td>
<td>8 mg (men) and 15 mg (women) intake</td>
</tr>
<tr>
<td>Zinc</td>
<td>10–15 mg (men) and 8–12 mg (women) intake (no supplement)</td>
</tr>
<tr>
<td>Selenium</td>
<td>55 μg intake (no supplement)</td>
</tr>
</tbody>
</table>

*SFor secondary prevention of cardiovascular events and muscle cramps.
*Only in hyperkalaemic patients, otherwise liberal intake.
*In case of EPO treatment, supplemental iron is to be added, either orally or parenterally.
*In case of malnutrition or symptoms of zinc deficiency, a trial of 50 mg Zn/day for 3–6 months could be considered (see text).
*In case of vitamin C deficiency, a trial of 50 mg Zn/day for 3–6 months could be considered (see text).

References

26. Trumbo P, Schlicker S, Yates AA et al. Dietary references intakes for energy, carbohydrate, fiber, fat, fatty acids,


Guideline 5. Treatment of malnutrition

5.1. Dietary intervention

- Malnourished haemodialysis patients should receive nutritional counselling (Evidence level III).
- In hospitalized patients counselling should be started within 3 days of referral. A daily follow-up should be performed when patients are at high nutritional risk and weekly when at low risk (Opinion).

Rationale and commentary

Regular dietary counselling is an important part of the overall nutritional management of MHD patients. A qualified dietitian is trained to apply specific counselling techniques and these are aimed at behavioural change strategies to empower the patient to make successful changes in his/her diet. A ‘renal’ diet is complex as the intake of several nutrients requires modification during the different stages of chronic kidney disease and once again when the mode of dialysis changes or the patient is transplanted.

Thus a MHD diet requires changes in the intake of protein and energy, sodium and fluid, potassium, calcium and phosphate and also in mineral and vitamin requirements. Early intervention during regular follow up may prevent nutritional inadequacies and could avert malnutrition.

Several investigators studied the effect of regular nutritional counselling regarding nutritional intake of MHD patients. Removing some of the reasons for inadequacies considerably improved intake without the need for nutritional supplements. Sehgal et al. [1] investigated barriers to protein nutrition among MHD patients in a cohort of 298 patients from 22 dialysis units. Four parameters were assessed: nutritional status (serum albumin and PNA); potential medical barriers (poor appetite, difficulty in chewing, inadequate dialysis, bioincompatible dialysis membranes and comorbidity); behavioural barriers (knowledge of the protein content of food and dietary non-compliance) and socio-economic barriers (expense of foods with a high protein content, lack of support with shopping and cooking). It was concluded that three medical factors (poor appetite, inadequate dialysis and comorbidity), two behaviour factors (lack of knowledge and low interdialytic weight gain, IDWG) and one socio-economic factor (need for help with shopping and cooking) were independently associated with poor nutritional intake of MHD patients. Leon et al. [2] showed that frequent nutritional counselling by trained dietitians and tactfully removing existing barriers to food optimizes dietary protein and energy intake that leads to improved serum albumin, even in the presence of chronic inflammation.
Akpele and coll. compared in a pilot study in 40 patients on haemodialysis for at least 6 months intensive dietary counselling vs the prescription of nutritional supplements [3]. The difference in rate of change in serum albumin (3.5 g/dl at onset) was measured [3]. The dietary goal was 1.2 g protein/kg IBW/day and 30–35 kcal/kg IBW/day. Twenty-six patients received nutritional supplements and 14 dietary counselling only. Patients receiving intensive counselling showed greater benefit than those receiving supplements. Indeed, albumin concentrations rose by 0.06 g/dl/month in the non-supplemented group vs 0.04 g/dl/month in the supplemented group [3]. A possible reason for this smaller change in the supplemented group was non-compliance and ageusia, anorexia, gastro intestinal side effects (diarrhoea), fear of weight gain, taste fatigue and preference for whole foods. Sharma et al. [4] surveyed 106 Indian MHD patients and found that patient’s diets changed over time on dialysis to a lower protein and energy intake than originally prescribed. Intakes were also compromised on dialysis days also adding to an increasingly poorer nutrient intake [4]. Dietary counselling and continuous monitoring therefore play an important role in malnutrition prevention. Steiber et al. [5] analysed nutritional intakes of CKD patients on admission in a general hospital. Less than 25% of patients with chronic renal failure (pre-dialysis, HD and CPD) achieved a 75% intake of the recommended diets. By knowing factors that can predict poor oral intake at referral, patients at risk could be identified earlier and their nutritional loss during hospitalization decreased. Similarly, it was shown that MHD patients only received 80% or less of recommended intake when hospitalized over 1 week [6]. Thus, it is now clear that malnutrition can start or deteriorate in hospital. Early aggressive intervention from the start of hospitalization should be initiated within 3 days of referral with a daily follow up for high and weekly follow up for low risk patients.

5.2. Oral supplements and enteral feeding

- Nutritional supplements should be prescribed if nutritional counselling does not achieve an increase in nutrient intake to a level that covers minimum recommendation (see Guideline 3) (Evidence level III).
- Products specifically formulated for dialysis patients should be prescribed in preference to standard supplements for non-renal patients (Evidence level III).
- Enteral tube [naso-gastric or percutaneous entero-gastrostomy (PEG)] feeding using disease specific formulas for dialysis patients should be prescribed if attempts to increase dietary intake with oral supplements fail and nutritional status does not improve (Evidence level IV).

Rationale

The development of several feeding methods such as PEG tube feeding and Intradialytic Parenteral Nutrition (IDPN) and the development of disease-specific commercial products for oral (flavoured), enteral (no added flavour) and parenteral feeding have greatly modified the application of nutritional support for MHD patients during the past years. Oral or enteral nutritional support is less expensive than parenteral nutrition. Standard products for oral and enteral nutritional support contain a mixture of whole protein and/or amino acids, glucose polymers, fat components and added vitamins, minerals (including phosphate) and trace elements. It is preferable to use specific formulated products for dialysis patients containing more protein and energy (1.5–2.0 kcal/ml) and less potassium and phosphate if patients are hyperphosphataemic and/or hyperka-laemic [7]. Some of the oral supplements contain a non-sweet glucose polymer without addition of other nutrients or flavours. These are presented as a powder or as a concentrated liquid and are suitable to be added to food and drinks to improve energy intake without significantly altering the taste.

Few clinical trials have been conducted about the provision of oral nutritional support to dialysis patients [7] and are limited in patient number and by short duration. Results have been affected due to early withdrawal from the trials when patients experienced side effects such as nausea or diarrhoea and when non-compliant with prescribed amount of nutritional supplement(s) [3]. Using essential amino acid tablets (15 tablets daily for 3 months), Eustace et al. [8] reported non-compliance as high as 50% at 3 months of oral supplement, underlining the necessity for more research on supplements specifically designed for MHD patients.

A recent systematic review and meta-analysis of 18 studies (five randomized and 13 non-randomized controlled trials) on the use of multi-nutrient oral supplements and tube feeding in MHD patients by Stratton et al. [7] showed that oral and enteral nutritional support improves protein and energy intake, increases serum albumin concentrations by 0.23 g/dl (P < 0.05) and improves total energy intake. There is still insufficient data on the clinical effect on MHD patients and specifically malnourished patients to determine from which regimen they would benefit most.

Clinical benefits of oral nutritional supplements. A number of studies have investigated the effect of oral supplements on the nutritional parameters of MHD patients [3,8–15]. Iversen et al. [15] and Eustace et al. [8] studied the effect of amino acid supplements on albumin changes in MHD patients. Providing 19 stable, well-dialysed and well-nourished MHD patients a daily protein supplement containing 7.8 g amino acids (4.8 g were essential) for 3 months in addition to their usual MHD diet,
no improvement in nutritional parameters was shown, although intracellular amino acid concentration improved significantly [15]. In a randomized double blind trial, Eustace et al. [8] studied the benefits of a supplement containing a daily total of 10.8 g essential amino acids (EAA) in 29 MHD and 19 peritoneal dialysis (PD) patients with a mean three months pre-study serum albumin of 3.8 g/dl or less. Patients were taking five tablets (e.g. 3.6 g EAA), three times daily with food for 3 months. Serum albumin concentrations increased by 0.22 ± 0.09 g/dl ($P = 0.02$) in MHD patients but did not change significantly in the PD group. Patients with lower serum albumin concentrations improved more. No improvement was seen in serum amino acid concentrations.

Non-compliance with prescribed supplements and gastrointestinal side effects can affect the outcome of oral nutritional support as shown by Akpele et al. [3]. In this pilot study of 41 MHD patients, 26 patients were given a commercial renal-specific nutritional supplement in addition to their regular dietary intake for an average of 6–7 months. The expected increase in protein and energy intake however was not achieved and several factors contributed to the lack of compliance such as taste impairment, anorexia and gastrointestinal side effects such as nausea, diarrhoea, fear of weight gain, taste fatigue and preference to whole food. These investigators concluded that patients may benefit from a choice in a selection of different nutritional products with different consistency and flavour to boost their protein and energy intake [3]. Changing the timing of the prescribed nutritional supplement can improve compliance. Cockram et al. [11] compared the gastrointestinal symptoms, bowel habits, routine blood chemistries, urea kinetics and nPNA in 79 patients over a three week period using three products: one standard and two special formulas (one with and one without a fructooligosaccharide – FOS) developed for renal patients. The investigators found that the two specialized formulas resulted in lower serum phosphate levels and a decrease in the calcium–phosphate product. Patients taking the product containing FOS had less gastrointestinal side effects and the products were well tolerated. In the first trial, Allman et al. randomly prescribed 100–150 g glucose polymer equivalent to 400–600 kcal/day to 9 patients in addition to their usual diet (protein intake: 1.16 ± 0.28 g/kg and energy: 30 ± 10 kcal/kg) for 6 months vs no supplement in the control group. The supplemented patients gained 3.1 ± 2.3 kg ($P < 0.005$), 1.8 kg as body fat and 1.3 kg as lean body mass indicated by changes in anthropometry. BMI rose from 21.3 to 22.9 and this weight gain was maintained 6 months after stopping the supplement [9]. In the second trial, Milano et al. [13] studied the effect of a 100 g glucose polymer supplement (380 kcal) daily in 21 MHD patients in addition to their usual diet. This amount increased their energy intake to at least 34 kcal/kg/day. After 6 months of supplement, their mean weight increased by 2.4 kg (range 0.6–6.3 kg) but assessed by anthropometry, this weight gain appeared to be predominantly fat. This gain was maintained for 6 months after cessation of the supplement [13].

Clinical benefits of enteral tube feeding. If the provision of oral supplements in addition to intensive counselling is unsuccessful, tube feeding should be proposed. Renal specific formulas may be used to maximize protein and energy supply, while controlling excessive amounts of fluid, phosphate, potassium and unnecessary vitamins usually contained in standard feed products.

Sayce et al. [16] analysed serum albumin and anthropometry in eight malnourished MHD outpatients who received PEG feeding for 3–15 months. Energy dense renal formula tube feeds were prescribed based on
individual requirements. The feed was administered as a bolus or by means of continuous pump feeding overnight. Two anuric patients required changes in their feeding regimens due to fluid overload. The amount of tube feed was reduced even further during the 3-day weekend interval when fluid overload was likely to occur. After 3 months, mean dry weight increased from a mean of 43.0 to 48.3 kg ($P = 0.01$). Mid upper arm circumference increased from 20.2 to 24.8 cm ($P = 0.02$) and triceps skinfold thickness from 17.7 to 19.8 mm ($P = 0.03$). Serum albumin rose from 29.5 to 36.5 g/l ($P = 0.01$). It was concluded that home enteral feeding using PEG access is effective and safe in improving and maintaining nutritional status in malnourished MHD patients. Frequent monitoring with provision of additional support throughout the feeding period is paramount.

Holley et al. [17] performed a retrospective analysis of a small cohort of 10 MHD patients (mean age 66 years) who received nasogastric or PEG enteral feeding for 1–36 months. Seven out of ten patients had suffered a cerebrovascular accident; two patients were in intensive care units. In five patients, tube feeding was supplementary to a normal nutrition and provided 50% of protein and energy of their requirements to achieve a protein intake of 1.0–1.3 g protein/kg BW and 30–35 kcal/kg per day. Serum albumin rose from 2.8 to 3.3 g/dl ($P = 0.04$) by the end of the feeding period, but no significant weight gain was reported. Eight out of ten patients had at least one episode of hypophosphataemia (<2.0 mg/dl in four of the eight patients) which was resolved by replacing the regimen from the renal to a standard formula and by using phosphate supplements [17]. Although retrospective and of limited size, these results should encourage well designed prospective studies in MHD patients.

**Recommendations for further research**

- What is the optimal composition of oral supplements (taste, concentration, electrolyte composition, vitamin and trace content)?
- What is the optimal schedule and delivery rate of oral and enteral supplements?
- What are the indications, optimal duration and complications of PEG in MHD patients?
- Is it possible to improve patient’s appetite with specific oral supplements?

### Rationale and commentary

There are many speculative reasons that intravenous nutrition may improve patient’s nutritional status. In the particular case of maintenance dialysis, the fact that patients will be referred three times weekly with vascular access allowing additional nutrient infusion theoretically simplifies applicability, delivery and compliance to parenteral nutrition. On the other hand, time to exposure for nutritional support is rather short (standard 10–15 h weekly) as compared with total parenteral nutritional support used in intensive care units or at home for patients with intestinal failure. Hence, non-renal nutritionists often question the efficacy of IDPN. In addition, IDPN is more expensive than any oral or enteral nutrition. The key questions are: do patients have a spontaneous intake great enough to supplement through a limited delivery related to the intermittent pattern of intradialytic parenteral nutrition (e.g. greater than 20 kcal and 0.8 g protein/kg IBW/day), and how will IDPN interfere with spontaneous patient’s intake throughout metabolic and appetite alterations?

Several retrospective analyses, prospective trials and reviews have addressed the various aspects of IDPN [18–28]. From a metabolic point of view, one haemodialysis session dramatically decreases the plasma amino acids and as a consequence, blunts intracellular muscle protein synthesis. In addition, in response to the rapid plasma amino acid decrease at the start of the haemodialysis session, muscle proteolysis occurs in order to maintain an adequate plasma and cellular amino acid concentration [29]. These events result in a clear catabolic state at the end of the dialysis session [30–32]. In the long term these catabolic modifications may lead to muscle wasting. Feeding patients by parenteral route during the dialysis session has been shown to revert this acute catabolic state by maintaining a normal plasma amino acid concentration [30]. Recently, Pupim et al. [33] reported that a brief 15 min cycling exercise at the beginning of the dialysis session dramatically improved the anabolic effect of the IDPN supplement.

However, it is less clear if these beneficial effects are associated with long-term improvement in patient’s nutritional status and morbi-mortality. Indeed, protein metabolism may be modified during the non-dialysis days, and to some extent, compensate for the dialysis-induced acute catabolic state. In more prolonged surveys, improvement in serum albumin [22,24] and patient’s spontaneous intake has been reported [23,24,27] but few studies were adequately designed and to date, evidence is low. Thus, long-term randomized studies should address the potential effect of IDPN on nutritional status and morbi-mortality of MHD patients. The ongoing Fines study, the largest prospective randomized controlled trial addressing the efficacy on oral and intradialytic nutritional support in malnourished MHD patients.
will provide evidence for indication and limits of nutritional support in these patients [18].

5.4. Anabolic agents

- In case of severe malnutrition resistant to optimal nutritional intervention, a course of androgens should be considered in MHD patients for three to 6 months (Evidence level II).
- Androgens should be administered weekly or bimonthly (Evidence level II).
- Patients should be monitored at regular intervals for side effects (hirsutism, voice change, priapism, alteration in plasma lipids, liver tests and prostatic markers) (Evidence level II).
- Patients with a known prostate cancer should not receive androgens (Evidence level II).

Rationale

In healthy adults the body protein mass is maintained at equilibrium by a subtle tuning between anabolism and catabolism that is regulated through independent signals. Among anabolic factors (which promote growth in childhood, and maintenance of protein mass in adults), growth hormone (GH) and insulin-like growth factor (IGF)-1 have been studied in-depth in many disorders, including CKD. Many acute administration studies and most mid-term (3–6 months) treatments with recombinant GH have reported beneficial metabolic, nutritional and body composition changes [34–45]. Long-term administration of recombinant GH has not been studied in CKD adults, and potential side-effects may occur, such as hyperglycaemia, hypertriglyceridaemia, and sodium retention. Recombinant GH treatment is only approved in short stature CKD children and helps to catch-up growth. However, GH is not currently approved in adult MHD patients. Recombinant IGF-1, which also exerts acute anabolic responses in malnourished dialysis patients [46], is only approved for the specific Laron nanism, a GH receptor deficient disease. Thus, except for pituitary insufficiency, a treatment by recombinant GH to improve nutritional status cannot yet be proposed to adult dialysis patients.

Androgens are well-known anabolic compounds. It should be emphasized that with age and CKD, a relative androgen insufficiency may be present, underlining a potential cause for muscle loss in males. Recently, it has been reported that non-renal male patients with coronary disease and low plasma testosterone levels did benefit from low-dose transdermal testosterone treatment, which improved their coronary symptoms [47]. In elderly patients without known kidney disease, a 6-month administration of low doses of testosterone to reach supranormal plasma levels significantly increased muscle mass and strength [48]. In kidney patients, androgens have been largely utilized before the era of erythropoietin to correct anaemia and reduce the number of blood transfusions. However, since the release of recombinant EPO in the 1990s, androgens were left apart and it is only since recently that their anabolic properties were rediscovered [48–54]. In a randomized controlled trial, Johansen et al. [52] assessed body composition and strength while administering nandrolone decanoate, 100 mg intramuscularly weekly for 6 months in 29 MHD patients. Body composition was monitored by dual energy X-ray absorptiometry and functional status by treadmill, walking, and stair-climbing times. Nandrolone decanoate induced a 4.5 kg lean body mass gain \((P < 0.01)\) and a fat loss of 2.4 kg \((P < 0.01)\) from baseline. There was a reduction in reported symptoms of fatigue and a decrease in walking and stair climbing times in the nandrolone group. No changes in serum cholesterol or triglycerides were reported in either group. Dose adjustment was done in two women who complained of acne and amenorrhea [52].

In elderly MHD patients receiving EPO, Gascon et al. [54] administered nandrolone decanoate, 200 mg intramuscularly weekly for 6 months in 14 patients who were withdrawn from EPO, whereas 19 patients continued to receive regular EPO treatment. Patients receiving nandrolone gained weight (from 68.2 ± 9.1 to 70.3 ± 8.6 kg; \(P < 0.05\)) and muscle mass (\(P < 0.05\)) [54]. Haemoglobin improved from 9.6 ± 1.0 to 11.0 ± 1.4 g/dl (\(P < 0.01\)) in the nandrolone group, whereas no change was observed in the EPO group who received 6000 ± 3900 IU EPO weekly. Serum albumin decreased from 4.0 ± 0.3 to 3.6 ± 0.5 g/dl (\(P < 0.05\)) in the EPO group, whereas it did not change in the nandrolone group. During nandrolone treatment, although serum triglycerides increased from 144 ± 78 to 180 ± 76 mg/dl (\(P < 0.05\)) and HDL-cholesterol decreased from 39 ± 13 to 32 ± 11 (\(P < 0.05\)), Lp(a), a strong predictor of cardiovascular risk, decreased from 26 ± 23 to 9 ± 8 mg/dl (\(P < 0.005\)) [54]. Thus, it is not clear in MHD patients if androgens impair lipid metabolism to the point of increasing long-term cardiovascular risk, which should be weighted against the risk of rapidly worsening malnutrition.

In another retrospective study by Pai and colleagues [50] in which five women received 25 mg nandrolone decanoate intramuscularly weekly for 3 months, no side effect was reported. Serum albumin significantly rose from 29 to 33 g/l (\(P < 0.05\)) in the study of Pai et al. [50] and from 32 to 38 g/l (\(P < 0.001\)) in the prospective randomized trial reported by Navarro and colleagues [51]. The dose of nandrolone administered in these studies ranged from 100 mg twice monthly for 3 months to 200 mg weekly for 6 months. Potential side effects include voice change and hirsutism in women, prostatic markers in men and abnormal liver tests and change in lipid metabolism in all and regular follow-up should be done accordingly [55,56]. Thus, in moderate...
amounts for 3–6 months, nandrolone improved body composition in MHD patients.

**Recommendation for further research**

- Larger randomized controlled trials of androgens administration should be performed in various degrees of malnutrition in MHD patients, in order to characterize a likely dose-response, the optimal duration and frequency of administration and to monitor potential side-effects.
- Test the efficacy of a combined intervention of androgens and exercise training on body composition and nutritional status of malnourished MHD patients.
- Measure the effects of a combined intervention by androgens and supplemental nutrition (either oral, enteral or parenteral) on body composition and nutritional status of malnourished MHD patients.

### 5.5. Other interventions: daily dialysis

- A 6–12 month trial of daily dialysis (either short daily or long nocturnal) should be considered as a rescue therapy in unstable patients undergoing difficult haemodialysis sessions with symptoms of malnutrition or malnourished patients with poor appetite after a negative nutritional work-out (Opinion).

**Rationale**

Since almost 10 years, daily haemodialysis pilot trials in Europe and North America have reported nutritional and metabolic effects in MHD patients [57–73]. Different schedules as well as durations have been proposed, such as six 2-h morning sessions to seven 8-h slow nocturnal sessions weekly. Interestingly, although nutrition was not the primary cause for enrolling patients in these programs, most reported unexpected improvements in appetite, clinical and biological nutritional parameters.

Table 3 reports the change in serum albumin before and at the end of daily dialysis programs in pilot maintenance dialysis adult patients trials (SDHD: short daily haemodialysis; NHHD: nocturnal home haemodialysis).

<table>
<thead>
<tr>
<th>Reference</th>
<th>Treatment</th>
<th>Nb Patients</th>
<th>S. albumin (g/l)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>At start</td>
</tr>
<tr>
<td>Woods et al. [67]</td>
<td>SDHD</td>
<td>72</td>
<td>39</td>
</tr>
<tr>
<td>Buoncristiani et al. [68]</td>
<td>SDHD</td>
<td>50</td>
<td>39</td>
</tr>
<tr>
<td>Pinciaroli [69]</td>
<td>SDHD</td>
<td>22</td>
<td>35</td>
</tr>
<tr>
<td>Kooistra and Vos [70]</td>
<td>SDHD</td>
<td>13</td>
<td>42.2</td>
</tr>
<tr>
<td>Galland et al. [61]</td>
<td>SDHD</td>
<td>10</td>
<td>39</td>
</tr>
<tr>
<td>Lugon et al. [71]</td>
<td>SDHD</td>
<td>5</td>
<td>40</td>
</tr>
<tr>
<td>Perras et al. [57]</td>
<td>NHHD</td>
<td>12</td>
<td>41.2</td>
</tr>
<tr>
<td>McPhatter et al. [72]</td>
<td>NHHD</td>
<td>9</td>
<td>34</td>
</tr>
<tr>
<td>O'Sullivan et al. [66]</td>
<td>NHHD</td>
<td>5</td>
<td>36.3</td>
</tr>
<tr>
<td>Cacho et al. [73]</td>
<td>NHHD</td>
<td>5</td>
<td>43.0</td>
</tr>
</tbody>
</table>

Whether those nutritional changes are beneficial over the long term is not known and should be the subject to future well-designed randomized trials. Indeed, one recent systematic review on the benefits of daily nocturnal haemodialysis, although clearly showing improved blood pressure and left ventricular hypertrophy, did report mixed effects on quality of life, anaemia control and phosphocalcic metabolism [65]. Authors asked for harder endpoint events such as mortality or cardiovascular morbidity before this strategy could be more largely diffused.

**References**


Guideline 6. Metabolic acidosis

- Mid-week predialysis serum bicarbonate levels should be maintained at 20–22 mmol/l (Evidence level III).
- In patients with venous predialysis bicarbonate persistently < 20 mmol/l, oral supplementation with sodium bicarbonate and/or increasing dialysate concentration to 40 mmol/l should be used to correct metabolic acidosis (Evidence level III).

Rationale

In two epidemiological studies a U-shape relationship between serum bicarbonate levels and mortality has been demonstrated in haemodialysis patients [1,2]. Lowrie et al. [1] reported from a retrospective analysis in over 12 000 haemodialysis patients an increased risk of dying if serum bicarbonate levels were < 17.5 or > 25 mmol/l. Recently, this was confirmed in the DOPPS study, in which a rise in mortality was present if predialysis bicarbonate level was < 17 or > 27 mmol/l [2]. In the latter study, it was demonstrated that serum bicarbonate levels between 20.1 and 21.0 mmol/l faced the lowest risk for mortality and levels of 21.1–22.0 the lowest risk of hospitalization. From these epidemiological data it can be concluded that predialysis serum bicarbonate levels of 20–22 mmol/l seem to be optimal.

Although in a number of studies the effect of metabolic acidosis on nutritional status in haemodialysis patients has been investigated, their outcome is inconclusive as only a small number of patients were investigated, the follow-up was short and few randomized prospective trials are available. In a single-blind crossover study design, Williams et al. [3] demonstrated that 27% of patients had metabolic acidosis defined as pH < 7.36 when treated with a 30 mmol/l bicarbonate dialysis solution. When the bicarbonate content of the dialysate was increased to 40 mmol/l, all patients had a pH > 7.36 which appeared to be associated with a rise in triceps skinfold thickness but no change in serum albumin or other nutritional parameters. When using a 40 mmol/l bicarbonate dialysis solution, it seems prudent to monitor post-dialysis venous bicarbonate to avoid post-dialytic alkalinaemia.

In two small prospective studies, it could be demonstrated in haemodialysis patients [4] and patients with chronic renal failure [5] that metabolic acidosis (serum bicarbonate levels < 21 mmol/l) could be corrected by oral sodium bicarbonate supplementation (serum bicarbonate levels in both studies after treatment > 24 mmol/l). This resulted in a rise in serum albumin levels, but no other changes in nutritional parameters. In both studies it was shown that after correction of metabolic acidosis nPNA decreased. Verove et al. [5] did not find a difference in daily protein intake. Likewise, Movilli et al. did not find a change in protein intake [6] or urea kinetics and
serum proteins [4] after the correction of metabolic acidosis. In a longitudinal observational study of 248 patients it was also observed that correction of metabolic acidosis by increasing the dialysate bicarbonate concentration to 39 mmol/l resulted in a fall in MPNA whereas serum albumin and SGA did not change [7]. Thus, several authors have concluded that in moderate to severe metabolic acidosis protein catabolism is present resulting in a rise in MPNA which then does not reflect daily protein intake only [4–8]. The existence of increased protein turnover in acidic haemodialysis patients was indeed shown by studies with radiolabelled leucine [9]. Likewise, it was demonstrated that net daily acid gain was higher in acidic haemodialysis patients [10]. Uribarri et al. [11] found in the HEMO study a negative correlation between serum total carbon dioxide levels and MPNA, and concluded that this could be attributed to a high protein intake in the patients with more severe metabolic acidosis as there were no signs of abnormal nutritional parameters in these patients. Thus, in patients with persistent metabolic acidosis protein intake may be assessed to see whether a high protein intake could contribute to the acidosis.

In one randomized prospective study, treatment of acidic haemodialysis patients with oral sodium bicarbonate and increasing the bicarbonate dialysate concentration to 40 mmol/l caused a rise in serum bicarbonate levels without any effect on serum albumin and other nutritional parameters [12]. In this study, however, oral supplementation and increasing dialysate bicarbonate only resulted in a rise of serum bicarbonate levels to 20 mmol/l, which could be too low to see positive effects on nutritional outcome. Kooman et al. [13] demonstrated in acidic haemodialysis patients that oral sodium bicarbonate supplementation resolved metabolic acidosis, caused a rise in serum branched-chain amino acids but did not affect nutritional parameters. Similar findings have been obtained in children [14].

In summary, it may be concluded that correction of metabolic acidosis to serum bicarbonate levels at around 20–22 mmol/l should be aimed for to reduce the risk of mortality and morbidity, to increase serum albumin levels and to reduce protein catabolism.

References


Appendices

Formulas (body weight, nPNA, dialysis dose, residual renal function)

Body weight: definitions

BW: body weight—should be assessed in patients wearing stocking feet and light indoor clothing with accurate scales, calibrated on a regular basis. Ask the patient to remove coat, jacket and heavy objects such as coins, keys, whichever is appropriate.

ABW: actual body weight—the patient’s present body weight at the time of the observation.

SBW: standard body weight—normal weight of healthy Americans of similar sex, age, height and skeletal frame size, obtained through the NHANES II Tables [1].

USB: usual body weight—the patient’s weight obtained through history or previous measurements, considered to be stable over time.

eBW: oedema free body weight, corresponding to ‘dry weight’—obtained post-dialysis in HD patients.
based on clinical judgement whether the patient still presents clinical oedema.

**AefBW:** adjusted oedema-free body weight—should be used in order to calculate the optimal dietary intake of protein and energy. It may avoid recommendations for too large intakes that may induce overproduction of waste products increasing uraemic symptoms. When patient’s body weight will improve towards standard body weight value, adjustment of body weight will not be necessary anymore.

\[
\text{AefBW} = \text{efBW} + (\text{SBW} - \text{efBW}) \times 0.25
\]

where SBW obtained from NHANES II Tables [1].

**Height**

Height should be measured as follows: ask the patient to remove shoes, to stand straight with feet together, buttocks, shoulder blades and head against the measuring device or wall and looking straight ahead. The measuring device will indicate length in meters/centimetres.

The practitioner should not depend on self-reporting as patients tend to overestimate height which decreases with advancing age.

### Estimating height in elderly and physically disabled patients [2]

If possible use recent documented (i.e. passport details) or self reported height, although patients tend to overestimate height which decreases with advancing age.

**Alternative height measurements**

**Length forearm (ulna), knee height and arm demispan.** In older people, the measurement of height to calculate BMI does not take into account bone loss (osteoporosis) that results in reduced height. Using alternative measurements such as knee height compensates for age-related changes.

Arm demispan can be used for people with curvatures of the spine, with infirmity and confusion.

**Length of forearm (ulna) (Fig. 1)**

- Ask the patient to bend the left arm if possible, palm across the chest, fingers pointing to opposite shoulder.
- Using a tape measure, measure the length in centimeters to the nearest 0.5 cm between the point of the elbow (olecranon) and the mid-point of the prominent bone of the wrist (styloid process).
- Use Table 1 to convert ulna length (cm) to height (m).

**Knee height (Fig. 2)**

- Measure left leg if possible.
- The patient should sit on a chair, without shoes, with knee at a right angle.
- Hold tape measure between 3rd and 4th finger with zero reading underneath fingers.
- Place your hand flat across the patient’s thigh, about 4 cm behind the front of the knee.
- Extend the tape measure straight down the side of the leg in line with the bony prominence at the ankle (lateral malleolus) to the base of the heel. Measure to the nearest 0.5 cm.
- Note the length and use Table 2 to convert knee height (cm) to height (m).

**Demispan (Fig. 3).** Demispan should not be used in patients with severe or obvious curvature of the spine (kyphosis or scoliosis).

### Table 1. Estimating height from ulna length (permission from MAG BAPEN, www.bapen.org.uk)

| Height (m) | Men (< 65 years) | 1.69 | 1.67 | 1.66 | 1.64 | 1.62 | 1.60 | 1.59 | 1.57 | 1.55 | 1.53 | 1.51 | 1.49 | 1.48 | 1.46 | 1.45 | 1.44 | 1.42 | 1.40 |
| Height (m) | Men (≥ 65 years) | 1.65 | 1.63 | 1.62 | 1.60 | 1.59 | 1.57 | 1.56 | 1.55 | 1.54 | 1.52 | 1.51 | 1.50 | 1.48 | 1.47 | 1.45 | 1.44 | 1.42 | 1.40 |
| Height (m) | Women (< 65 years) | 1.65 | 1.63 | 1.62 | 1.61 | 1.59 | 1.58 | 1.56 | 1.55 | 1.54 | 1.52 | 1.51 | 1.50 | 1.48 | 1.47 | 1.45 | 1.44 | 1.42 | 1.40 |
| Height (m) | Women (≥ 65 years) | 1.61 | 1.60 | 1.58 | 1.56 | 1.55 | 1.53 | 1.52 | 1.50 | 1.48 | 1.47 | 1.45 | 1.44 | 1.42 | 1.40 | 1.39 | 1.38 | 1.37 | 1.36 |
| Ulna length (cm) | | 25.0 | 24.5 | 24.0 | 23.5 | 23.0 | 22.5 | 22.0 | 21.5 | 21.0 | 20.5 | 20.0 | 19.5 | 19.0 | 18.5 | 18.0 | 17.5 | 17.0 | 16.5 |
| Height (m) | Women (< 65 years) | 1.65 | 1.63 | 1.62 | 1.61 | 1.59 | 1.58 | 1.56 | 1.55 | 1.54 | 1.52 | 1.51 | 1.50 | 1.48 | 1.47 | 1.45 | 1.44 | 1.42 | 1.40 |
| Height (m) | Women (≥ 65 years) | 1.61 | 1.60 | 1.58 | 1.56 | 1.55 | 1.53 | 1.52 | 1.50 | 1.48 | 1.47 | 1.45 | 1.44 | 1.42 | 1.40 | 1.39 | 1.38 | 1.37 | 1.36 |
For bed bound patients, those with severe disabilities and those with kyphosis or scoliosis, it is preferable to use forearm (ulna) length to estimate height.

- The patient should stand as this makes taking the measurement easier.
- Locate and mark the mid-point of the sternal notch (V at the base of the neck).
- Ask the patient to raise the right arm until it is horizontal with the shoulder (give assistance if necessary; make sure the wrist is straight).

Place a tape measure between the middle and ring finger of the patient’s right hand, with zero at the base of the fingers.

Extend the tape measure along the length of the arm to the mid-point of the sternal notch and note the measurement to the nearest 0.5 cm. Use Table 3 to convert demispan length to height (m).

Ideal body weight estimation (Tables 4 and 5).

Body mass index (BMI). BMI is calculated from the weight (kg) divided by the square of the height (m). BMI of maintenance dialysis patients should be maintained in the upper 50th percentile (BMI for men and women of at least approximately 23.6 and 24.0 kg/m²).

The World Health Organization describes the condition of low BMI as thinness which is divided into three grades:

- Grade 1: BMI 17.0–18.49 (mild thinness)
- Grade 2: BMI 16.0–16.99 (moderate thinness)
- Grade 3: BMI < 16.0 (severe thinness)

The Malnutrition Advisory Group (MAG), a standing committee of the British Association for Parenteral and Enteral Nutrition [2] has recommended

![Fig. 2. Estimating height from knee height (permission from MAG BAPEN, www.bapen.org.uk).](https://academic.oup.com/ndt/article-abstract/22/suppl_2/ii45/1871238/167282)

*Source: Malnutrition Advisory Group (MAG) British Association of Enteral and Parenteral Nutrition (BAPEN), 2003 (www.bapen.org).*

![Fig. 3. Estimating height using demispan (permission from MAG BAPEN, www.bapen.org.uk).](https://academic.oup.com/ndt/article-abstract/22/suppl_2/ii45/1871238/167282)

*Source: Malnutrition Advisory Group (MAG) British Association of Enteral and Parenteral Nutrition (BAPEN), 2003 (www.bapen.org).*

### Table 2. Estimating height (in meter) from knee height (permission from MAG BAPEN, www.bapen.org.uk)

<table>
<thead>
<tr>
<th>Gender</th>
<th>Age Group</th>
<th>Knee Height (cm)</th>
<th>Height (m)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Men</td>
<td>18–59 years</td>
<td>1.94 1.93 1.92 1.91 1.90 1.89</td>
<td>1.88 1.87 1.86</td>
</tr>
<tr>
<td>Men</td>
<td>60–90 years</td>
<td>1.94 1.93 1.92 1.91 1.90 1.89</td>
<td>1.88 1.87 1.86</td>
</tr>
<tr>
<td>Women</td>
<td>18–59 years</td>
<td>1.89 1.88 1.875 1.87 1.86 1.85</td>
<td>1.84 1.83 1.82</td>
</tr>
<tr>
<td>Women</td>
<td>60–90 years</td>
<td>1.86 1.85 1.84 1.835 1.83</td>
<td>1.82 1.81</td>
</tr>
<tr>
<td>Men</td>
<td>18–59 years</td>
<td>1.80 1.79 1.78 1.77 1.76 1.75</td>
<td>1.74 1.73 1.72</td>
</tr>
<tr>
<td>Men</td>
<td>60–90 years</td>
<td>1.79 1.78 1.77 1.76 1.74 1.73</td>
<td>1.72 1.71 1.70</td>
</tr>
<tr>
<td>Knee Height (cm)</td>
<td>57.5 57 56.5 56</td>
<td>55.5 55</td>
<td>54.5 54 53.5 53</td>
</tr>
<tr>
<td>Women</td>
<td>18–59 years</td>
<td>1.75 1.74 1.735 1.73 1.72</td>
<td>1.71 1.70</td>
</tr>
<tr>
<td>Women</td>
<td>60–90 years</td>
<td>1.72 1.71 1.70 1.69 1.68 1.67</td>
<td>1.66 1.65</td>
</tr>
<tr>
<td>Men</td>
<td>18–59 years</td>
<td>1.66 1.65 1.64 1.63 1.62 1.61</td>
<td>1.60 1.59 1.58</td>
</tr>
<tr>
<td>Men</td>
<td>60–90 years</td>
<td>1.63 1.62 1.61 1.60 1.59 1.58</td>
<td>1.57 1.56 1.55</td>
</tr>
<tr>
<td>Knee Height (cm)</td>
<td>50 49.5 49</td>
<td>48.5 48 47.5 47</td>
<td>46.5 46 45.5 45</td>
</tr>
<tr>
<td>Women</td>
<td>18–59 years</td>
<td>1.61 1.60 1.59 1.585 1.58</td>
<td>1.57 1.56 1.55</td>
</tr>
<tr>
<td>Women</td>
<td>60–90 years</td>
<td>1.58 1.57 1.56 1.55 1.54</td>
<td>1.53 1.52 1.51</td>
</tr>
</tbody>
</table>
nutritional measurements should be height, weight and recent weight loss. The BMI categories are:
- BMI < 18.5 chronic protein-energy undernutrition probable
- BMI 18.5–20.0 chronic protein–energy undernutrition possible
- BMI > 20.0 chronic protein-energy undernutrition unlikely

**Classification of BMI. Significance: (normal individuals) from Wiggins [3]**

<table>
<thead>
<tr>
<th>BMI</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt;16</td>
<td>Severely underweight</td>
</tr>
<tr>
<td>&lt;18.5</td>
<td>Associated with health problems</td>
</tr>
<tr>
<td>18.5–24.9</td>
<td>May be associated with health problems for some people</td>
</tr>
<tr>
<td>25–29.9</td>
<td>Within normal weight range</td>
</tr>
<tr>
<td>30–34.9</td>
<td>Overweight</td>
</tr>
<tr>
<td>35–39.9</td>
<td>Obesity class I</td>
</tr>
<tr>
<td>&gt;40</td>
<td>Obesity class III</td>
</tr>
</tbody>
</table>

**Table 3. Estimating height (in meter) using demispan (permission from MAG BAPEN, www.bapen.org.uk)**

<table>
<thead>
<tr>
<th>Age</th>
<th>Height (cm)</th>
<th>Small frame</th>
<th>Medium frame</th>
<th>Large frame</th>
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</thead>
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<tr>
<td>25–54 years</td>
<td>157</td>
<td>64</td>
<td>82</td>
<td>97</td>
</tr>
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<td>160</td>
<td>61</td>
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<td>97</td>
</tr>
<tr>
<td></td>
<td>163</td>
<td>66</td>
<td>84</td>
<td>97</td>
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<tr>
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<td>165</td>
<td>66</td>
<td>79</td>
<td>72</td>
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<tr>
<td></td>
<td>168</td>
<td>67</td>
<td>84</td>
<td>74</td>
</tr>
<tr>
<td></td>
<td>170</td>
<td>71</td>
<td>84</td>
<td>78</td>
</tr>
<tr>
<td></td>
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<td>79</td>
<td>93</td>
<td>88</td>
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<tr>
<td></td>
<td>188</td>
<td>80</td>
<td>92</td>
<td>95</td>
</tr>
</tbody>
</table>

**Table 4. Fiftieth (50th) Percentile of Standard Body Weight for Men (NHANES 1 and II); reproduced with permission from Frisancho et al. The American Journal of Clinical Nutrition [1].**

<table>
<thead>
<tr>
<th>Age</th>
<th>Weight (kg)</th>
<th>Height (cm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>25–54 years</td>
<td>157</td>
<td>64</td>
</tr>
<tr>
<td></td>
<td>160</td>
<td>61</td>
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<td>79</td>
</tr>
<tr>
<td></td>
<td>188</td>
<td>80</td>
</tr>
</tbody>
</table>

**Table 5. Fiftieth (50th) Percentile of Standard Body Weight for Women (NHANES 1 and II); reproduced with permission from Frisancho et al. The American Journal of Clinical Nutrition [1].**

<table>
<thead>
<tr>
<th>Age</th>
<th>Weight (kg)</th>
<th>Height (cm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>25–54 years</td>
<td>147</td>
<td>52</td>
</tr>
<tr>
<td></td>
<td>150</td>
<td>53</td>
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<td></td>
<td>170</td>
<td>59</td>
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<tr>
<td></td>
<td>173</td>
<td>62</td>
</tr>
<tr>
<td></td>
<td>175</td>
<td>63*</td>
</tr>
<tr>
<td></td>
<td>178</td>
<td>64*</td>
</tr>
</tbody>
</table>

**Calculating BMI in amputees [3].** IBW needs to be adjusted by taking into account the weight of body segment(s) that is/are amputated. Adjustments of body weight can be made from knowledge of missing limbs segments.

Upper limb: whole arm 5% (upper arm 2.7% and fore arm 1.6%, hand 0.7%)
Lower limb: whole leg 16% (thigh 10.1%, lower leg 4.4%, foot 1.5%)
To measure full body weight equation

Estimated full body weight (kg) = \( \frac{\text{measured weight}}{\left(100 - \% \text{ weight of amputation}\right)} \times 100 \)

Change in body weight over the previous 6 months
Unintentional weight loss over the previous 3–6 months is categorized as
- 10% of body weight: clinically significant,
- 5–10% of body weight: more than normal intra-individual variation,
- <5% of body weight: within ‘normal’ intra-individual variation.
A cut-off of >10% weight loss during the last 6 months is recommended to be used in the diagnosis of malnutrition [4].

**Body surface.** Body surface area should be estimated according to Gehan and George method [5]

\[ \text{BSA} = 0.235 \times \frac{\text{BW}^{0.51456} \times \text{BH}^{0.42246}}{\text{C}_{15}} \]

**Body water.** Total body water (TBW), necessary for the calculation of the correct dialysis dose, can be estimated by the Watson formulas (2):

- \[ V_{\text{male (L)}} = 2.447 + 0.3362 \times \text{BW (kg)} + 0.1074 \times \text{BH (cm)} - 0.09516 \times \text{age (years)} \]
- \[ V_{\text{female (L)}} = -2.097 + 0.2466 \times \text{BW (kg)} + 0.1069 \times \text{BH (cm)} \]

**Normalized protein equivalent of total nitrogen appearance (nPNA).** By the use of protein nitrogen appearance (PNA), formerly called protein catabolic rate (PCR), the dietary protein intake can be estimated in patients with neutral nitrogen balance (i.e. in patients, who are neither anabolic nor catabolic). In order to normalize PNA, it should be related to the body weight of the patient, leading to nPNA.

In order to optimize the diet of patients with renal disease, the dietary protein intake has to be controlled. In stable patients (non-catabolic, non-anabolic) nPNA reflects the dietary protein intake and can be calculated based on UNA (urea nitrogen appearance in the urine and/or in the dialysate, respectively) for the following reasons:

1. In patients with a neutral nitrogen balance, the nitrogen intake is identical with the loss of nitrogen (that is the total nitrogen appearance: TNA)
2. As nitrogen in protein accounts for 16% of the protein's weight, the protein equivalent of nitrogen appearance (PNA) can be calculated by total nitrogen appearance (TNA) as:

\[ \text{PNA} = 6.25 \times \text{TNA} \]

3. There is a linear relationship between TNA and UNA. The ratio between UNA and TNA, however, depends on the dietary intake of protein as well as the status and treatment of the patient [2]
4. In order to normalize PNA to body weight, the K/DOQI Nutrition Work Group recommends the use of the following formula [4]:

\[ \text{nPNA} = \frac{\text{PNA}}{\text{AEFBW}} \]

where AEFBW is the adjusted, oedema free body weight, see above

nPNA is calculated by using spKt/V and \( C_0 \). Different formulas are used for different days of the week (5) in a two-BUN, single-pool, variable-volume model:

- **Beginning of the week** \( \text{PNA} = C_0/[36.3 + (5.48)(\text{spKt/V}) + (53.5)/(\text{spKt/V})] + 0.168 \)
- **Midweek** \( \text{PNA} = C_0/[25.8 + (1.15)(\text{spKt/V}) + (56.4)/(\text{spKt/V})] + 0.168 \)
- **End of week** \( \text{PNA} = C_0/[16.3 + (4.3)(\text{spKt/V}) + (56.6)/(\text{spKt/V})] + 0.168 \)

where \( C_0 = \text{predialysis blood urea nitrogen (BUN)} \)

\[ \text{spKt/V} = \text{single pool Kt/V} \]

In patients with considerable residual renal function, \( C_0 \) should be replaced by \( C_0' \):

\[ C_0' = C_0[1 + (0.79 + 3.08/(\text{Kt/V})) \text{Kr}/\text{V}] \]

where \( \text{Kr} \) is residual renal clearance in ml/min [4].

**Residual renal function: glomerular filtration rate (GFR).** Residual renal function should be expressed as equivalent of glomerular filtration rate (GFR). One accepted method in both PD and HD patients is to estimate GFR from the mean of urea and creatinine clearance [6], normalized to 1.73 m² using the Gehan & George method [5] for calculating body surface area (BSA).

The collection time in haemodialysis patients is identical with the interval between two dialysis sessions. The plasma concentration of urea and creatinine used in the following formula are determined at the beginning and end of the collection. As in pre-ESRD and peritoneal dialysis patients, the bladder must be empty at the beginning of the collection (i.e. at the end of the dialysis) and must be completely emptied at the end of the collection (i.e. before the next dialysis starts):

\[ \text{GFR} = \left( \frac{\text{U}_{\text{urea}}}{\text{preP}_{\text{urea}} + \text{postP}_{\text{urea}}} + \frac{\text{U}_{\text{creat}}}{\text{preP}_{\text{creat}} + \text{postP}_{\text{creat}}} \right) \times \frac{\text{U}_{\text{vol}}}{\text{t}} \times \left( \frac{1.73}{\text{BSA}} \right) \]

derived from:

\[ \text{GFR} = 0.5 \times \left( \frac{\text{U}_{\text{urea}}}{0.5 \times (\text{preP}_{\text{urea}} + \text{postP}_{\text{urea}})} \right) + \left( \frac{\text{U}_{\text{creat}}}{0.5 \times (\text{preP}_{\text{creat}} + \text{postP}_{\text{creat}})} \right) \times \frac{\text{U}_{\text{vol}}}{\text{t}} \times \left( \frac{1.73}{\text{BSA}} \right) \]

with \( 0.5 \times (\text{preP}_{\text{urea}} + \text{postP}_{\text{urea}}) = \text{average concentration of urea in the plasma during dialysis sessions} \)

\( 0.5 \times (\text{preP}_{\text{creat}} + \text{postP}_{\text{creat}}) = \text{average concentration of creatinine in the plasma between dialysis} \)

\( \text{preP}_{\text{urea}}, \text{plasma urea concentration before dialysis at end of collection} \)

\( \text{postP}_{\text{urea}}, \text{plasma urea concentration after dialysis at beginning of collection} \)

\( \text{preP}_{\text{creat}}, \text{plasma creatinine concentration before dialysis at end of collection} \)

\( \text{postP}_{\text{creat}}, \text{plasma creatinine concentration before dialysis at end of collection} \)
after dialysis at beginning of collection; \( U_{\text{urea}}, \) urine urea concentration; \( U_{\text{creat}}, \) urine creatinine concentration; \( U_{\text{vol}}, \) urine volume; \( t, \) time of collection between dialysis sessions.

In order to be more precise, the post-dialysis concentrations should be replaced by the post-rebound concentrations, which can be calculated as follows [7]:

Post-rebound concentration for urea:
\[
\text{rebound} = \text{pre} \times \left( \frac{\text{post}}{\text{pre}} \right)^{\frac{\text{td}}{\text{td} + 35}}
\]

where \( \text{td}, \) dialysis time in minutes; \( \text{pre}, \) concentration before dialysis before collection; \( \text{post}, \) concentration immediately after dialysis.

Post-rebound concentration for creatinine:
\[
\text{rebound} = \text{pre} \times \left( \frac{\text{post}}{\text{pre}} \right)^{\frac{\text{td}}{\text{td} + 70}}
\]

where \( \text{td}, \) dialysis time in minutes; \( \text{pre}, \) concentration before dialysis before collection; \( \text{post}, \) concentration immediately after dialysis.

Dialysis dose

For MHD it is recommended to calculate the equilibrated \( K_t/V \) (e\( K_t/V \)) instead of the single pool \( K_t/V \) (sp\( K_t/V \)) from pre- and post-HD blood samples taken under standard conditions (see below). The e\( K_t/V \) takes into account the urea rebound post dialysis resulting from redistribution of peripheral pools.

Haemodialysis dose should be quantified as equilibrated \( K_t/V \) (e\( K_t/V \)) based on the regional blood flow two-pool model [8,9]:
\[
e\text{Kt/V} = \text{spKt/V} - (0.6 \times \text{spKtV/T}) + 0.03 \text{ (with an arteriovenous access)}
\]

The value for single pool \( K_t/V \) (sp\( K_t/V \)) should be derived from the urea kinetic model or alternatively from the natural logarithm equation to estimate sp\( K_t/V \) [10]:
\[
\text{spKt/V} = -\ln(R - 0.008 \times t) + (4 - 3.5 \times R) \\
\times dBW/BW
\]

where \( R, \) post-HD/pre-HD BUN ratio; \( T, \) treatment time in hours; dBW, intradialytic weight lost (corresponding to ultrafiltration); BW, end session body weight.

Standard conditions for blood sampling

Methods procedure derived from the K/DOQI guidelines [4,11].

(a) Pre-dialysis blood sampling procedure:

for arteriovenous graft or fistula: obtain the blood from arterial needle before connecting the tube or flushing the needle, avoid dilution of the sample by saline and/or heparin

(b) Post-dialysis blood sampling procedure:

As recirculation of dialysated blood in the arterial line or rebound of urea can significantly influence the value of urea, the sampling technique must be performed in a standardized way leading to reproducible results. In following the techniques explained below, the sample will be drawn after possible recirculation but before rebound of urea from peripheral compounds.

1. Turn off dialysate flow or reduce to minimum, decrease ultrafiltration rate to 50 ml/h
2. Decrease blood flow to 50–100 ml/min for 15 s proceed with slow pump or stop pump technique

Slow flow sampling technique
3. draw blood sample with pump running at 50–100 ml/min
4. stop blood pump and complete disconnection procedure

Stop pump sampling technique
5. stop the blood pump
6. clamp arterial and venous blood lines; clamp arterial needle tubing
7. sample blood either from arterial sampling port nearest to patient or from the arterial needle tubing after disconnection from the arterial blood line
8. blood is returned to the patient, complete disconnection procedure

LABORATORY METHODS

Serum Albumin

The gold-standard for determining serum albumin levels are immunological methods. Serum albumin levels determined by established methods like the bromcresol green (BCG) or bromcresol purple (BCP) method differ from those obtained by immunological methods due to limitations in methodology [12].

Comparing the values of albumin in plasma and serum, measured by the same method, identical values are found. However, differences are found between the bromcresol green and the bromcresol purple method. Uremic toxins as well as certain medications (e.g. phenylbutazon, clofibric acid) [13] influence the measurement.

In general, bromcresol green methods overestimate albumin levels [14] compared with bromcresol purple [15] and nephelometry [16]. Bromcresol purple, on the other hand, generally underestimates albumin values [12,17]. Thus the evaluation of the...
**Subjective Global Assessment Rating Form**

<table>
<thead>
<tr>
<th>A. History</th>
<th>Rating</th>
<th>Mild-Moderate</th>
<th>Normal</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Weight changes</td>
<td>over past 2 weeks</td>
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<td></td>
</tr>
<tr>
<td>over past 6 months</td>
<td>increasing weight</td>
<td></td>
<td></td>
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<tr>
<td>___ &lt; 5% weight change (or gain)</td>
<td>stable weight</td>
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<tr>
<td>___ 5 to 10% weight lost</td>
<td>ongoing weight lost</td>
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<td>___ &gt; 10% weight lost</td>
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<tr>
<td>overall: ___ usual intake</td>
<td>1 2 3 4 5 6 7</td>
<td></td>
</tr>
<tr>
<td>___ &lt; usual and decreasing</td>
<td></td>
<td></td>
</tr>
<tr>
<td>duration: ___ weeks</td>
<td></td>
<td></td>
</tr>
<tr>
<td>type of ___ suboptimal solids</td>
<td>___ full liquid intake</td>
<td></td>
</tr>
<tr>
<td>change: ___ hypocaloric fluids</td>
<td>___ unable to eat</td>
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<table>
<thead>
<tr>
<th>3. Gastrointestinal symptoms</th>
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<th></th>
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<tbody>
<tr>
<td>___ none</td>
<td>___ anorexia</td>
<td>___ nausea</td>
</tr>
<tr>
<td>___ vomiting</td>
<td>___ diarrhea</td>
<td></td>
</tr>
<tr>
<td>duration: ___ weeks</td>
<td></td>
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</table>

<table>
<thead>
<tr>
<th>B. Physical examination</th>
<th>Rating</th>
<th>Mild-Moderate</th>
<th>Normal</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>1 2 3 4 5 6 7</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>1 2 3 4 5 6 7</td>
<td></td>
<td></td>
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</tbody>
</table>

<table>
<thead>
<tr>
<th>C. Overall SGA classification</th>
<th>Final Rating</th>
</tr>
</thead>
<tbody>
<tr>
<td>1. Normal or well nourished</td>
<td>Rating of 6 to 7 in most categories or significant and sustained improvement</td>
</tr>
<tr>
<td>2. Mild to moderately malnourished</td>
<td>3 to 5 rating is indicated in most categories</td>
</tr>
<tr>
<td>3. Severely malnourished</td>
<td>1 to 2 ratings in most categories</td>
</tr>
</tbody>
</table>


serum albumin level must take into account the different normal ranges of the applied laboratory methods.

**Bicarbonate**

Plasma bicarbonate is estimated from total CO\(_2\) measurements. It is thus mandatory to use fresh blood samples because CO\(_2\) might be lost leading to significant underestimation of plasma bicarbonate [18]. One must also keep in mind that total CO\(_2\) levels assessed by electrode-based methods are an average 4 mmol/l higher compared with plasma bicarbonate concentrations determined by enzymatic assays [19].
The plasma bicarbonate, calculated from pCO₂ by using the Henderson-Hasselbach equation, gives:
\[ \text{cHCO}_3^- (\text{mmol/l}) = 0.0307 \times 10^{\text{pH} - 6.1} \]

**C-reactive protein**

C-reactive Protein (CRP) has been used as a marker of inflammation for many years. In this respect, levels in the range of 5–300 mg/l have been of interest, which are detected by the common laboratory methods. Standard immunological methods like immunonephelometry and immunoturbidimetry should detect levels at least above 5 mg/l.

New assays allow the detection of even lower CRP levels in the range of 0.1–10 mg/l. These so-called ‘high sensitive-CRP-assays’ (hs-CRP) are needed to assess the risk of atherosclerosis [20–22], as already only slightly elevated CRP levels below 5 mg/l have been found to be associated with increased risk of cardiovascular disease [23].

**TECHNICAL ASSESSMENT**

**Subjective global assessment (SGA)**

The SGA was developed for use in assessing the nutrition of general surgery patients [24]. It is recommended also for patients on dialysis, because it is a valid clinical assessment of nutritional status and is strongly associated with patient survival.

Remind that the overall SGA classification is not simply a numerical store. It does strongly depend on the clinical judgement of the examiner. He has to consider whether the patient’s status is improving or deteriorating, this information may lead to different ‘scores’ given in each section.

The SGA is based on history and physical examination [24]. It focuses on gastrointestinal symptoms (anorexia, nausea, vomiting, diarrhoea), weight loss in the preceding 6 months, and visual assessment of subcutaneous tissue and muscle mass. Scores are subjectively rated on a four-point or seven point scale [4]. The use of the seven point scale is recommended because of its greater sensitivity and its use in large epidemiological studies such as the CANUSA study [25].

<table>
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<th>Women (y)</th>
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<td>11.1</td>
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**Anthropometry**

Skinfold thickness (SFT) should be measured by someone experienced in the use of skinfold calipers. The measurement is taken at four different sites (biceps, triceps, subscapular and suprailiac). For each skinfold, the mean of three measurements is taken.

Severe malnutrition: 1 or 2,
Mild malnutrition to normal nutritional state: 6 to 7,
Moderate to mild malnutrition: 3 to 5,
Mild malnutrition to normal nutritional state: 6 to 7.

SGA was found to be related to other markers of nutritional status in dialysed [26] and non-dialysed uraemic subjects [27] as well as to mortality [28].
Table 7. Mid-arm muscle circumference (MAMC) for adult men and women from USA (from NHANES I study [33]); measurements made in the right arm. Reproduced with permission from Bishop CW, Bowen PE, Ritchey SJ. The American Journal of Clinical Nutrition [33].

<table>
<thead>
<tr>
<th>Age (years)</th>
<th>50th Percentile (cm)</th>
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<tbody>
<tr>
<td>Men</td>
<td></td>
</tr>
<tr>
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</tr>
<tr>
<td>25–34</td>
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<td>35–44</td>
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<td>65–74</td>
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<td>Women</td>
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<td>55–64</td>
<td>22.6</td>
</tr>
<tr>
<td>65–74</td>
<td>22.5</td>
</tr>
</tbody>
</table>


Mid-arm muscle circumference (MAMC, Table 7) should be calculated as the MAC in cm minus (triceps skinfold thickness × π). There are no agreed cut off points for MAC, TSF or MAMC for the diagnosis of malnutrition in either the normal population or patients with chronic renal failure. Frisancho’s tables provide standards for mid-arm muscle circumference in normal subjects [29,30] whilst norms have also been published for the dialysis population [31].

Handgrip strength

Muscle strength is best evaluated by muscle dynanometry of the handgrip strength which has been related to protein stores assessed by neutron activation analysis in non-uraemic subjects [34]. In pre-ESRD patients handgrip strength was strongly related to lean body mass determined by DEXA, anthropometry, and creatinine kinetics and the strongest factor related to malnutrition defined by SGA [27]. In dialysis patients handgrip strength was reduced in malnourished patients determined by SGA [35].

References