Mobile D-loops are a preferred substrate for the Bloom’s syndrome helicase

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ABSTRACT
The Bloom’s syndrome helicase, BLM, is a member of the highly conserved RecQ family, and possesses both DNA unwinding and DNA strand annealing activities. BLM also promotes branch migration of Holliday junctions. One role for BLM is to act in conjunction with topoisomerase III to process homologous recombination (HR) intermediates containing a double Holliday junction by a process termed dissolution. However, several lines of evidence suggest that BLM may also act early in one or more of the recombination pathways to eliminate illegitimate or aberrantly paired DNA joint molecules. We have investigated whether BLM can disrupt DNA displacement loops (D-loops), which represent the initial strand invasion step of HR. We show that mobile D-loops created by the RecA recombinase are a highly preferred substrate for BLM with the invading strand being displaced from the duplex. We have identified structural features of the D-loop that determine the efficiency with which BLM promotes D-loop dissociation. We discuss these results in the context of models for the role of BLM as an ‘anti-recombinase’.

INTRODUCTION
Bloom’s syndrome (BS) is an extremely rare genetic disorder, characterized by proportional dwarfism, a sun-sensitive facial erythema, male sterility and female sub-fertility and non-insulin-dependent diabetes mellitus in young patients. The most striking feature, however, is the high incidence of malignancies of most types (1). Cells carrying mutations in the BS gene (designated \textit{BLM}) display decreased plating efficiency and various signs of genomic instability; namely, chromosomal rearrangements, an elevated frequency of homologous recombination (HR), and sensitivity to certain DNA damaging agents and replication inhibitors [(2) and references therein]. The hallmark feature of these cells, which is used in the diagnosis of the disorder, is an elevated frequency of sister-chromatid exchanges (SCEs) (3).

The BS gene product, BLM, is a DNA helicase belonging to the highly conserved RecQ family (4). BLM is a DNA structure-specific helicase that displays 3'\textendash-5' directionality (5). The RecQ family of helicases includes four additional members in humans: WRN, the gene defective in Werner’s syndrome (WS) (6,7); RECQ4, associated with some cases of Rothmund–Thomson syndrome (8) and RAPADILINO (9); RECQ1 (10,11); and RECQ5 (12). Unicellular organisms generally contain only one representative of the family, such as the founding member in \textit{Escherichia coli}, RecQ (13), and Sgs1 in the budding yeast, \textit{Saccharomyces cerevisiae} (14,15).

Although the precise function of any RecQ helicase is yet to be established, several lines of evidence suggest that they play roles in HR. HR is important for the repair of DNA double strand breaks (DSBs), for the restoration of productive DNA synthesis following disruption of DNA replication forks, and for meiosis, where it is essential for creation of genetic diversity and for faithful chromosome segregation. Although the biochemical details of HR are not fully worked out, several proteins are required to process DSBs to create a 3' ssDNA tail that can invade a homologous duplex to create a three-stranded, branched structure called a displacement loop (D-loop). The key catalytic factor in D-loop formation is the Rad51 recombinase. It polymerizes on the ssDNA tail generated by resection of the DSB to create an extended filament in which the search for homology is conducted. This function requires the action of several accessory factors in yeast, including the Rad50/Mre11/Xrs2 complex, replication protein A (RPA), Rad52, Rad54 and the Rad51-related proteins, Rad55 and Rad57. In mammals, this basic process is conserved, although it requires some additional factors; most notably, the BRCA2 protein that is associated with some cases of hereditary breast and ovarian cancer (16,17).

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RecQ helicase mutants frequently display an elevated rate of HR, which in BS cells is manifested as an increase in the frequency of SCEs and inter-homologue recombination (3). BLM-defective cells of mouse or chicken origin show an elevated frequency of gene targeting by HR, as well as loss of heterozygosity (18–20), which can arise when crossing-over events occur between recombining homologous chromosomes. Similarly, E.coli recQ mutants (21) and S.cerevisiae sgs1 mutants (22) show excessive illegitimate recombination, suggestive of a failure to suppress inappropriate recombination occurring between non-identical sequences. The presence of a constitutively high level of RAD51-containing nuclear foci in BS cells, even outside of S-phase where they normally exist, also indicates the presence of a constitutively high number of sites of ongoing or incomplete HR events (23,24). Moreover, RecQ helicases show direct physical interactions with HR proteins; e.g. BLM and Sgs1 each bind to the Rad51 protein in their respective organisms through a conserved C-terminal interaction domain (23).

There is evidence of roles for RecQ helicases both early and late in the HR pathway. Rockmill et al. (25) showed that Sgs1 is required for the suppression of meiotic crossovers by decreasing the number of sites of ongoing or incomplete HR events. They also act late, this time alongside topoisomerase IIIα, to resolve recombination intermediates. In this study, we set out to test the hypothesis that the BLM helicase can perform an anti-recombinase role through the disruption of the D-loop recombination intermediate. For this, we specifically utilized RecA-generated mobile (or migratable) D-loop structures. We report that BLM is highly efficient at displacing the invading strand of mobile D-loops, and have identified structural features of the D-loop that direct BLM to act in this manner.

**MATERIALS AND METHODS**

**Proteins**
Recombinant BLM was purified as described previously (30).

**Oligonucleotides**
Oligonucleotide sequences used in this study are listed in Table 1. Additionally, the X12 four-way junction, X12 replication fork and OX-1T G-quadruplex substrates described by Mohaghegh et al. (31) were also generated. The oligonucleotides were either custom synthesized by Sigma–Genosys (Sigma–Aldrich House, Haverhill, UK), or provided by the oligonucleotide synthesis service of Cancer Research UK.

<table>
<thead>
<tr>
<th>Oligonucleotide</th>
<th>Description</th>
<th>Length</th>
<th>Sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>DL3</td>
<td>3’ Invading with 5’ tail (44)</td>
<td>90</td>
<td>5’-AATTCTCATT TTACTTACCG GACGCTATTA GCAGTGCCAGATTTGATCT- GAAAGTTCACCATATGCGGTGAATCGACAGATGCGT-3’</td>
</tr>
<tr>
<td>DL33</td>
<td>3’ Invading with a 25 nt reverse polarity region on the tail</td>
<td>90</td>
<td>3’-AATTCTCATT TTACTTACCG GACGCTATTA GCAGTGCCAGATTTGATCT- GAAAGTTCACCATATGCGGTGAATCGACAGATGCGT-3’</td>
</tr>
<tr>
<td>DLm</td>
<td>Fully complementary invading strand with no tail</td>
<td>54</td>
<td>5’-GCAAGTTGACTGAGAATGCGCCATATGCGGTGAATCGACAGATGCGT-3’</td>
</tr>
<tr>
<td>DL5</td>
<td>5’ Invading with 3’ tail</td>
<td>90</td>
<td>5’-GCAAGTTGACTGAGAATGCGCCATATGCGGTGAATCGACAGATGCGT-3’</td>
</tr>
<tr>
<td>DL55</td>
<td>5’ Invading with a 25 nt long reverse polarity region on the tail</td>
<td>90</td>
<td>5’-GCAAGTTGACTGAGAATGCGCCATATGCGGTGAATCGACAGATGCGT-3’</td>
</tr>
<tr>
<td>G/T2.1</td>
<td>3’ Invading with 5’ tail; 1x G/T mismatch</td>
<td>90</td>
<td>5’-GCAAGTTGACTGAGAATGCGCCATATGCGGTGAATCGACAGATGCGT-3’</td>
</tr>
<tr>
<td>G/T2.2</td>
<td>3’ Invading with 5’ tail; 2x G/T mismatches</td>
<td>90</td>
<td>5’-GCAAGTTGACTGAGAATGCGCCATATGCGGTGAATCGACAGATGCGT-3’</td>
</tr>
<tr>
<td>G/T2.4</td>
<td>3’ Invading with 5’ tail; 4x G/T mismatches</td>
<td>90</td>
<td>5’-GCAAGTTGACTGAGAATGCGCCATATGCGGTGAATCGACAGATGCGT-3’</td>
</tr>
<tr>
<td>pUCop</td>
<td>Reverse complement of pUC18 nucleotides 440–470 and 525–555 with scrambled sequences in between to form a bubble when annealed to pUCbottom</td>
<td>116</td>
<td>5’-GCAATGCGGCC CTGATGCGG ATTTTCTCCT TACGCTTCG AGTTGATCT- GCAAGGCTGAGGCTGGCCATACGATATCTCAT- AG TTAAGGCCAG CCCGAC-3’</td>
</tr>
<tr>
<td>pUCbottom</td>
<td>pUC18 440–555 nt</td>
<td>116</td>
<td>5’-GCAATGCGGCC CTGATGCGG ATTTTCTCCT TACGCTTCG AGTTGATCT- GCAAGGCTGAGGCTGGCCATACGATATCTCAT- AG TTAAGGCCAG CCCGAC-3’</td>
</tr>
</tbody>
</table>
Reverse polarity oligonucleotides were synthesized using 5'-CE phosphoramidites and 5'-supports (Glen Research, Sterling, VA) by the CR-UK oligonucleotide synthesis service. All oligonucleotides were purified on a 10% denaturing polyacrylamide gel before use. Labelling was carried out using either T4 polynucleotide kinase (New England Biolabs, Beverly, MA) and [γ-32P]ATP (Amersham Biosciences, Chalfont St Giles, UK), or terminal deoxynucleotidyl transferase (New England Biolabs) and [α-32P]dATP (Amersham) according to the manufacturer’s instructions. Activity of the labelled oligonucleotides was measured and specific activity was calculated.

**Construction of substrates**

The oligonucleotide-based D-loops were prepared by the sequential annealing of the constituent oligonucleotides, as described by Opresko et al. (32). The X12 four-way junction, replication fork and OX-1T G4 substrates were generated as described previously (31). The plasmid-based D-loop substrates were generated as described by McIlwraith et al. (33). Briefly, the invading oligonucleotide (Table 1) was coated with RecA (New England Biolabs), for 5 min, and then supercoiled pUC18 plasmid was added to initiate strand invasion. The strand invasion reaction was allowed to progress for 3 min. The DNA was then deproteinized using proteinase K in the presence of SDS for 30 min. Substrates were purified essentially as described previously (31), but using 5% polyacrylamide gels to achieve better recovery of large D-loop molecules. The purified substrates were stored in 10 mM Tris–HCl (pH 7.5) and 10 mM MgCl2 (for D-loops and standard B-form DNA substrates) or 100 mM NaCl, 10 mM Tris–HCl (pH 8.0), 1 mM EDTA (for G4 DNA). Activity was measured and, using the specific activity of the corresponding oligonucleotide, the substrate concentration was calculated.

**Helicase assays**

Helicase assays were carried out in a 10 μl reaction volume containing 1x helicase buffer [33 mM Tris-acetate (pH 7.8); 1 mM Mg-acetate; 66 mM Na-acetate; 0.1 mg/ml BSA; 1 mM DTT; 1 mM ATP], 56 pm substrate and the indicated amount of enzyme for 15 min at 37°C, unless otherwise stated. The reaction products were separated by non-denaturing PAGE. The gels were dried, and radiolabelled DNA was visualized using a Storm 840 PhosphorImager. Band intensities were quantified using ImageQuant software (Amersham). Quantitative results were expressed as percent unwinding normalized to activities in the untreated and denatured controls, as described previously (31). Enzyme concentrations required for 50% unwinding, as well as inhibitor concentrations required for 50% inhibition (IC50 values), were determined after fitting a sigmoid curve with the

$$Y = \frac{\text{Baseline}}{1 + 10^{\text{HillSlope} \cdot \log X}}$$

non-linear regression model in the SPSS software package.

**RESULTS**

We set out to test the hypothesis that one mechanism by which BLM acts as an antirecombinase is through its ability to disrupt the DNA D-loop that forms at the initial strand invasion step of HR. In this way, BLM could abort HR reactions that are deemed inappropriate; e.g. because the recombining sequences are not identical. Previous studies indicated that RecQ family helicases are able to disrupt oligonucleotide-based mimics of a D-loop created by the annealing of three partially complementary oligonucleotides (32,34,35). We considered it unlikely, however, that these D-loop mimics represent an ideal model for the D-loop structure generated in vivo for several reasons. First, oligonucleotide-based D-loops are ‘static’ and are not capable of branch migration along the DNA, in contrast to genuine ‘mobile’ D-loops created enzymatically by the RecA recombinase or one of its eukaryotic orthologues. Second, the unwinding of oligonucleotide-based D-loops generates products that can be reannealed by the ssDNA annealing activity of BLM (36,37), potentially generating artifactual products that could never arise when mobile D-loops are employed. Third, oligonucleotide-based D-loops cannot be generated in negatively-supercoiled molecules, unlike mobile D-loops, and do not, therefore, model the in vivo situation. Fourth, oligonucleotide-based D-loops present recognition features for helicases that are not evident in mobile D-loops or in vivo; in particular, two free dsDNA ends.

We considered the possibility that oligonucleotide-based D-loops might be disrupted by a mechanism different from that employed for the disruption of mobile D-loops. We anticipated that the physiologically-relevant mechanism of disruption would entail the expulsion of the invading ssDNA from the recipient duplex in a single step. If such a mechanism were not obviously in evidence during disruption of oligonucleotide-based D-loops, then this would suggest that studies of D-loop unwinding by helicases should focus exclusively on mobile D-loop substrates. Indirect evidence to suggest that this proposal might have some credence came from our analysis of the data presented previously by van Brabant et al. (34). This analysis suggested that the ‘invading’ oligonucleotide was not simply being expelled from the duplex in a single step because reaction intermediates were evident in the data presented by these authors (34). Using substrate oDL3 (see Materials and Methods and Figure 1), we confirmed the results of van Brabant et al. (34) indicating that BLM can disrupt oligonucleotide-based D-loops (Figure 2). Moreover, we also observed at least two reaction intermediates that contained the 32P-labelled oligonucleotide, but had a slower mobility on PAGE than did the free ‘invading’ oligonucleotide. These intermediate species are designated A and B on Figure 2. From the time-course experiments presented in Figure 2a it is clear that the oligonucleotide-based D-loop substrate was unwound in a multi-step manner. Consistently, we found that species A appeared first over time, and was then converted to species B. Subsequently, species B gave rise to the radiolabelled single-stranded oligonucleotide. Using another substrate, this time with a 5’ invading end (oDL5; Figure 1), the pattern of appearance of products was different from that seen with oDL3. In this case, the single-stranded product was generated preferentially (Figure 2b), but again species A appeared as the first product. Species B was also observed; however, the timing of its appearance did not obviously precede that of the free single-stranded oligonucleotide. Species B had a mobility identical to that of a partially
unwound intermediate comprising the bottom strand of the duplex annealed to the invading oligonucleotide (data not shown). These results would indicate that the unwinding of an oligonucleotide-based, static D-loop can proceed via the denaturation of the duplex portion of the substrate (equivalent to the recipient duplex in strand invasion during HR) to create a partially unwound intermediate that can be denatured in a second phase by BLM to yield the free oligonucleotide. This order of events is clearly not representative of the physiological process of D-loop disruption.

Generation of ‘mobile’ D-loops

Given the above considerations, we set out to generate mobile D-loops employing the established method of McIlwraith et al. (33). In this, RecA is used to catalyse the invasion of an 90mer single-stranded oligonucleotide into a negatively-supercoiled pUC18 plasmid DNA that contains a 54 bp region of sequence identity with the 3’ end of the oligonucleotide (Table 1 and Figure 1). The 5’ region of the oligonucleotide used was not complementary to the pUC18 and can form a 36 nt single-stranded tail protruding from the D-loop. By minor modification of the sequence of the oligonucleotide, it was a straightforward task to create D-loops incorporating one or more single base mismatches between the oligonucleotide and the recipient plasmid. This permitted us to analyse the influence of MMR factors on the ability of BLM to process the D-loop structure (see Discussion).

BLM catalyses efficient D-loop disruption

Using a conventional D-loop (pDL3), in which the 3’ end of the oligonucleotide was base paired with the pUC18 plasmid and the 32P-labelled 5’ tail was protruding, we tested whether
BLM could disrupt the D-loop by displacing the invading strand. Using either a titration of increasing BLM concentration, or a time course at a fixed BLM concentration, we found that BLM could catalyse displacement of the 5’ labelled oligonucleotide from the D-loop (Figure 3). Based on our previous analyses of the unwinding of other DNA substrates by BLM, it appeared that the D-loop was very efficiently disrupted by BLM. To study this formally, we undertook additional analyses. First, we preformed a head-to-head comparison of the ability of BLM to disrupt the D-loop and two other DNA structures that have been shown previously to be efficient substrates for BLM; a four-way junction modelling the Holliday junction and a G-quadruplex structure created from sequences of the Oxytricha telomere repeat region. G-quadruplexes are acknowledged as the best substrate found to date not only for BLM, but also for most other RecQ helicases analysed (31,38–40). As shown in Figure 4, the BLM concentration required to disrupt the D-loop and the G-quadruplex were very similar (50% unwinding at 7.38 and 5.84 nM, respectively). In contrast, disruption of the four-way junction DNA consistently required a higher BLM concentration. Moreover, the extent of unwinding of this substrate peaked at a value below 100% and declined at high BLM concentrations. This phenomenon has been described previously and reflects the known DNA strand annealing activity of BLM (36,37) (discussed more in detail below).

The above data suggested that the D-loop is very efficiently disrupted by BLM, and that it may rank alongside the G-quadruplex as a preferred substrate for BLM. To analyse this issue more quantitatively, competition experiments were performed. For this, increasing concentrations of unlabelled (cold) competitor DNA were added to helicase assays to determine the concentration of competitor DNA that reduced the level of unwinding of the D-loop, G-quadruplex or four-way junction DNA to 50% of control levels seen in the absence of competitor. The cold competitor DNAs analysed were an 80 nt oligonucleotide of unrelated sequences to any of the substrates, and the four-way junction and Oxytricha telomeric G-quadruplex described above. Unwinding of the four-way junction DNA was inhibited by the non-specific oligonucleotide competitor at concentrations of oligonucleotide ~10-fold below those required to inhibit unwinding of the G-quadruplex or D-loop substrates (IC50 values of 2.73 nM for the four-way junction compared to 29.88 and 29.8 nM, respectively, for the G-quadruplex and D-loop; Figure 5a and b). Consistent with the G-quadruplex and D-loop molecules being significantly better substrates for BLM than was the four-way junction, competition experiments using cold four-way junction revealed efficient inhibition of unwinding of the labelled four-way junction substrate, but not of the G-quadruplex or D-loop substrates at the concentrations of competitor used (Figure 5c and d). If, as indicated by the above results, the D-loop is comparable to the G-quadruplex in being a highly efficient substrate for BLM, it would be predicted that cold G-quadruplex competitor DNA would inhibit unwinding of the labelled G-quadruplex essentially as efficiently as it would inhibit unwinding of the D-loop. We found, indeed, that this was the case, although it should be noted that the transition in the sigmoidal inhibition curve was sharper for the D-loop substrate than for the G-quadruplex substrate (Figure 5e and f). Taken together, these data indicate that BLM can disrupt a mobile D-loop with very high efficiency.

**Structural features of D-loops that are recognized by BLM**

The D-loop substrate described above mimics an HR event in which the 3’ end of the ssDNA has invaded the homologous duplex, and the 5’ end is left protruding from the D-loop (designated pDL3; Figure 1). According to our current understanding, this is the situation that occurs during HR in vivo, because it permits priming of DNA synthesis from the 3’ end of the invading DNA. To address the key structural features of the D-loop that direct efficient BLM-mediated unwinding, we created two additional mobile D-loops: one with no protruding tail (pDLm) and one in which the 5’ end of the oligonucleotide had invaded and the 3’ end was protruding (pDL5). Analysis of the kinetics of unwinding of these three D-loop substrates indicated that the pDL5 molecule was unwound at a
significantly faster rate than either the pDLm or pDL3 substrates (Figure 6). These data indicate that, while a protruding tail is not essential for BLM-mediated D-loop disruption, the orientation of the invading DNA must somehow influence the efficiency of D-loop unwinding.

While a protruding tail may not be an essential element for unwinding of a D-loop, it appeared possible that the 3′ tail of pDL5 provided an additional recognition feature for BLM that would explain why this substrate was unwound more efficiently than was pDL3 or pDLm. To analyze this, we created two additional D-loop substrates in which the invading oligonucleotide contained a 25 nt stretch of reverse polarity (Figure 1). In pDL33, the oligonucleotide contained a 3′ invading end and a 3′ terminus on the protruding tail. In pDL55, the oligonucleotide had two 5′ ends (see Materials and Methods and Figure 1). The data in Figure 6 show that pDL55 was unwound as rapidly as pDL5, and at a significantly faster rate than was pDL33 (which was comparable in efficiency to pDL3). Taken together, these data indicate that neither the presence nor the polarity of the protruding tail are important features in determining the efficiency by which BLM disrupts mobile D-loops.

DISCUSSION

We have shown that the BLM helicase can disrupt both mobile D-loops generated enzymatically by a bona fide recombination initiating protein, and static D-loops created from oligonucleotides. Mobile D-loops are disrupted with high efficiency by BLM and rank alongside G-quadruplexes as the preferred substrate for the unwinding activity of BLM. We have also identified structural features of mobile D-loops that influence BLM helicase activity and have presented evidence that mobile and static D-loops are disrupted via different mechanisms.

There are a number of possible explanations for why mobile D-loops are such good substrates for BLM. First, and perhaps most logically, D-loops may represent one of the major functionally relevant in vivo substrates for BLM. This possibility has been proposed previously by others on the basis of genetic analyses in model organisms. For example, the data of Rockmill et al. (25) are consistent with a role for the yeast BLM orthologue, Sgs1, in disruption of an early, strand-invasion intermediate in meiotic recombination. Moreover, during a study of the role of the Drosophila BLM orthologue in DNA DSB repair, Adams et al. (41) provided evidence that DmBLM was involved in the process of synthesis-dependent strand annealing, which essentially requires the propagation of a D-loop intermediate along DNA concomitant with DNA synthesis. A second possible reason for the D-loop appearing to be such a good substrate is that it is not influenced by the single stranded DNA (ssDNA) annealing activity of BLM. The unwinding of more widely studied helicase substrates, such as forked duplexes and 3′-tailed duplexes, generates products that can be re-annealed to re-create the starting substrate. As a result, there is competition between the forward (helicase) and reverse (annealing) reactions as the concentrations of substrate and products change over time. This creates a characteristic bell shaped curve if the concentration of ssDNA product is plotted against BLM.
concentration or time (Figure 4b) (36). Hence, there can be an underestimation of the true efficiency with which BLM unwinds any substrate where the products are susceptible to re-annealing. This issue is irrelevant where mobile D-loop substrates are concerned, because BLM cannot re-generate the initial substrate from the reaction products as it lacks an ability to catalyse DNA strand invasion. Simple re-annealing would not be sufficient to re-generate the D-loop. Moreover, the equally efficiently unwound G-quadruplex substrate is not subject to re-annealing, as the re-generation of the quadruplex would require specific incubation conditions not provided during a helicase reaction. Although this ‘resistance’ to strand annealing may at least partially explain why D-loops and G-quadruplexes appear to be such good substrates for BLM, we would argue that the competition experiments described both in our study and previously during the study of G-quadruplexes by Maizels and co-workers (38,39) indicate that these DNA structures are intrinsically good substrates for BLM.

There are several features of D-loops that could influence the mechanism by which BLM effects their disruption. These include the distortion of the duplex in the region where strand invasion has occurred, the 3-stranded nature of the D-loop, and the presence of an ssDNA tail emerging from the D-loop. The finding that the ‘tail-less’ pDLm substrate can be unwound by BLM argues strongly against the tail providing a fundamentally important recognition feature. Of course, in vivo, although there is ssDNA invading a homologous duplex, this ssDNA is continuous and would not terminate with a

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**Diagram Description**

- **Diagram a:** Shows a comparison of oligonucleotide unwinding with and without BLM. The graph displays inhibition of unwinding as a function of oligonucleotide concentration.
- **Diagram b:** depicts the inhibition of pDL3, G4, and 4-way junction with varying concentrations of oligonucleotides.
- **Diagram c:** illustrates the unwinding of different DNA structures with BLM, comparing pDL and G4 substrates.
- **Diagram d:** presents the inhibition of pDL3 and G4 with varying concentrations of 4-way junction.

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*Figure 4b* (36): Shows the concentration or time dependence of unwinding with BLM.
BLM can distinguish between different D-loops depending on whether the ssDNA has a 5' or a 3' invading end. As reported previously using oligonucleotide-based D-loops (34), BLM prefers to unwind D-loops with a 5' invading terminus, and we have confirmed this using mobile D-loops. What we have shown additionally through the analysis of the reverse polarity oligonucleotides is that this preference is not determined by the presence of a 3' terminus on the tail of the oligonucleotide, since a substrate with two 5' ends is unwound as efficiently as is a conventional 5' invading oligonucleotide with a 3' tail.

The human orthologue of RecA, RAD51, interacts physically with BLM (23). RAD51 differs from RecA in that it promotes strand invasion from a 5' ssDNA end more efficiently than it does from a 3' ssDNA end. Since 5' end invasions are unproductive because they cannot be extended by DNA polymerases, some mechanisms must exist in vivo to limit their occurrence. Recently, the BRCA2 homologue of *Ustilago maydis* (42) has been shown to control the polarity of strand invasion. We suggest that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might induce a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invasions through its preferential unwinding of this form of D-loop. We propose that BLM might impose a second barrier to unproductive 5' strand invas...
generated from the oligonucleotide-based D-loops is not a genuine reaction intermediate but instead represents the product of a ssDNA annealing reaction, analysis of the order of appearance of the products would argue otherwise. These products clearly appear at earlier time points than does the free ssDNA oligonucleotide. A similar conclusion can be derived from an analysis of the data in Figure 5 of van Brabant et al. (34).

Figure 6. Analysis of the recognition features important for efficient unwinding of plasmid-based D-loops. (a) Head-to-head comparison of the efficiency of unwinding of the different plasmid-based D-loops in a time-course study in the presence of 0.52 nM BLM. Samples were taken at the time points indicated above the lanes. (b) Quantification of data from (a). Symbols used were: red square, pDL5; green diamond, pDL55; blue double-triangle, pDLm; purple triangle, pDL3; cyan circle, pDL33.
The studies described herein were initially aimed at testing the hypothesis that BLM and MMR factors co-operate to disrupt aberrant HR reactions. While our data are consistent with such a role for BLM, we obtained no evidence for a combined role for BLM and MMR proteins in D-loop disruption through studying the efficiency by which BLM and the MMR proteins together disrupt D-loops containing one, two or four G:T type mismatches (Table 1; C. Z. Bachrati, R. H. Borts and I. D. Hickson, unpublished data). This does not, in any way, argue against a key role for MMR proteins in ‘quality controlling’ HR processes; there is ample evidence to indicate such a role. The reason(s) why we failed to identify a role for MMR proteins at the D-loop stage of HR may reflect any of several possibilities. First, they may not play any role during this step of HR, but instead act later. Second, we may not have placed the mismatches in the appropriate location(s) in the D-loop. Third, factors in addition to BLM and MMR proteins are required to form a complex in order to promote abortion of mismatched D-loops. Further analyses will be required to analyse whether any of these possibilities is true.

In summary, we have presented evidence that mobile D-loops are a highly preferred substrate for the BLM helicase. One phenotype of BLM-deficient cells is an elevated frequency of recombination between homologous sequences. Moreover, cells from lower organisms deficient in BLM orthologues display hyper-recombination and a failure to suppress homologous recombination. We propose that a plausible explanation, at least in part, for these phenotypes is that BLM (and perhaps other RecQ helicases) can prevent D-loop structures from being converted into mature recombinants. Clearly, it would not be advantageous for BLM to effect this role indiscriminately in vivo; otherwise, all recombination reactions would be abortive. The identification of factors that modulate the function of BLM in the process of HR will likely identify factors that are crucial for the maintenance of genome stability and suppression of tumorigenesis in man.

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