Aminoacylation and translational quality control strategy employed by leucyl-tRNA synthetase from a human pathogen with genetic code ambiguity

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ABSTRACT

Aminoacyl-tRNA synthetases should ensure high accuracy in tRNA aminoacylation. However, the absence of significant structural differences between amino acids always poses a direct challenge for some aminoacyl-tRNA synthetases, such as leucyl-tRNA synthetase (LeuRS), which require editing function to remove mis-activated amino acids. In the cytoplasm of the human pathogen Candida albicans, the CUG codon is translated as both Ser and Leu by a uniquely evolved CatRNA\textsuperscript{Ser}(CAG). Its cytoplasmic LeuRS (CaLeuRS) is a crucial component for CUG codon ambiguity and harbors only one CUG codon at position 919. Comparison of the activity of CaLeuRS-Ser\textsuperscript{919} and CaLeuRS-Leu\textsuperscript{919} revealed yeast LeuRSs have a relaxed tRNA recognition capacity. We also studied the mis-activation and editing of non-cognate amino acids by CaLeuRS. Interestingly, we found that CaLeuRS is naturally deficient in tRNA-dependent pre-transfer editing for non-cognate norvaline while displaying a weak tRNA-dependent pre-transfer editing capacity for non-cognate α-amino butyric acid. We also demonstrated that post-transfer editing of CaLeuRS is not tRNA\textsuperscript{Leu} species-specific. In addition, other eukaryotic but not archaeal or bacterial LeuRSs were found to recognize CatRNA\textsuperscript{Ser}(CAG). Overall, we systematically studied the aminoacylation and editing properties of CaLeuRS and established a characteristic LeuRS model with naturally deficient tRNA-dependent pre-transfer editing, which increases LeuRS types with unique editing patterns.

INTRODUCTION

Aminoacyl-tRNA synthetases (aaRSs) are essential components required to establish the genetic code during protein biosynthesis by coupling specific amino acids with their cognate tRNAs in a two-step aminoacylation reaction (1,2). This process requires amino acid activation by condensation with ATP to form the aminoacyl-adenylate (aa-AMP) and pyrophosphate; the activated amino acid is then transferred to the cognate tRNA to yield the aminoacyl-tRNA (aa-tRNA), which is then transferred to the protein biosynthesis machinery as a building block (1). Aminoacylation of tRNA requires adequate efficiency and accuracy, which requires tightly regulated control of the speed of the aa-tRNA production for the ribosome and the risk of generation of aberrant aa-tRNA pairs (3–5). Transfer RNA always harbors various identity determinants and/or anti-determinants, facilitating selection of the correct tRNA from a large pool of tRNA species (6). However, the specificity of aaRS is greatly challenged by the presence of various types of amino acids and their analogues and the fact that amino acids differ only in the side-chain. AaRSs that do not show an overall selectivity above 1 in 3000 are predicted to require some form of proofreading (editing) mechanism to maintain sufficient accuracy during aa-tRNA synthesis (5,7,8). Editing activity has evolved in half of the currently identified aaRSs to remove any aberrantly produced aa-AMP (pre-transfer editing) and/or aa-tRNA (post-transfer editing). This is an essential checkpoint that ensures translational fidelity (5). Pre-transfer editing can be further divided into tRNA-independent and tRNA-dependent pre-transfer editing. In tRNA-independent pre-transfer editing, the non-cognate aa-AMP is hydrolyzed into the amino acid and AMP molecules without the presence of cognate tRNA, whereas in tRNA-dependent pre-transfer editing, aa-AMP hydrolysis is triggered by the addition of the cognate tRNA (9–11).
Mis-translation due to the impairment or loss of editing activity can lead to ambiguity of the proteome, having a seriously negative effect on the cellular function of most organisms and causing neuron-degeneration in a mouse model (12).

Leucyl-tRNA synthetase (LeuRS) is a large multidomain class Ia aaRS with both aminoacylation activity to generate tRNALeu and editing activity to clear non-cognate aa-AMP and aa-tRNA (13). It can be divided into bacterial and archaeal/eukaryotic types based on primary sequence and domain location (14). Both types of LeuRSs usually consist of a Rossmann-fold domain (for amino acid activation and aminoacylation), an α-helix bundle, a C-terminal domain (for tRNA binding) and a CP1 domain (for editing) (15–17). Extensive studies of various LeuRS species all found that non-cognate norvaline (Nva) is the most significantly mis-activated amino acid among all the non-cognate amino acids tested, including Ile, Val, Met and α-amino butyric acid (ABA). For instance, compared with cognate Leu, Nva is mis-activated by Aquifex aeolicus LeuRS (AaLeuRS) (9), Saccharomyces cerevisiae LeuRS (ScLeuRS) (18), human cytoplasmic LeuRS (hLeuRS) (19), Mycoplasma mobile LeuRS (MmLeuRS) (20), human mitochondrial LeuRS (hmtLeuRS) (unpublished data) 72-, 105-, 100-, 122- and 180-fold less efficiently, respectively. Nva is a non-proteinogenic amino acid differing from Leu only by the absence of a side-chain methyl group. Nva is naturally present in vivo and is a by-product of the Leu biosynthesis pathway (21). Its synthesis is predominantly related to an imbalance in the synthesis of the branched-chain amino acids under pyruvate-high conditions. In addition, Nva significantly accumulates immediately after a shift from aerobic culture conditions to oxygen limitation at high glucose concentrations (22). Therefore, the amount of Nva is dynamic and varies according to the environment. The incorporation of Nva in proteins at Leu codons has been clearly demonstrated. It has been reported to be a natural component of an antifungal peptide of Bacillus subtilis (23) and can be intentionally inserted into heterologous proteins by culturing Escherichia coli in the presence of Nva (US patent, Nov 7, 1989, 4879223). Accompanied by conditions of an elevated ratio of available Nva to Leu in the medium, increasing mis-incorporation of Nva at Leu codons has been observed in recombinant human hemoglobin produced in E. coli as a result of mis-aminoacylation of tRNALeu by E. coli LeuRS (EcLeuRS) (24). It is proposed that Nva replacement may disrupt the correct folding and assembly of hemoglobin and other proteins (24). All this evidence suggests that Nva mis-activation by LeuRS is a non-artificial event that occurs in vivo, and that mis-activated Nva-tRNALeu can be accommodated and used by the ribosome. Therefore, editing of Nva by LeuRS seems to be essential for the correct functioning of organisms.

Based on significant mis-activation of Nva, editing catalyzed by LeuRS (with a functional CP1 domain) has been shown to be one of the most interesting editing mechanisms. This process is predominantly mediated by three diverse pathways (tRNA-independent, tRNA-dependent pre-transfer and post-transfer editing) (10). Both types of LeuRS critically depend on the editing active site embedded in the CP1 domain to perform post-transfer editing (15–17, 25). However, MmLeuRS harbors only tRNA-independent pre-transfer editing activity owing to its natural lack of the CP1 domain (20). Another example of a unique LeuRS is hmtLeuRS, which possesses a degenerate editing active site in the CP1 domain as well as defunct post-transfer editing (26) and tRNA-dependent pre-transfer editing activities (unpublished data). Combining site-directed mutagenesis and AMP formation methodology, the contribution of different pathways to the overall editing process can be quantified (9, 10, 19). Strikingly, there are quantitative and species-specific differences in the contribution of a specific pathway to the total editing activity of a LeuRS (9, 10, 19). To evaluate the significance of each mechanism, we have attempted to generate LeuRSs lacking one or more editing mechanisms; to date, two types have been successfully established. One type contains LeuRSs with abolished post-transfer editing activity, obtained by introducing mutations at key residues (e.g. EcLeuRS-T252R, AaLeuRS-T273R, AcLeuRS-D373A, ScLeuRS-D419A, hLeuRS-D399A, Giardia lamblia (GI) LeuRS-D444A (9, 10, 18, 19, 27) or by the inclusion of a small molecule inhibitor (AN2690) of the CP1 editing domain (10). The second type includes LeuRSs for which both the post-transfer editing and tRNA-dependent pre-transfer editing activities (EcLeuRS-Y330D, AaLeuRS-Y358D) have been abolished (10). Our aim was to determine whether a LeuRS with defective tRNA-dependent pre-transfer editing activity but intact post-transfer editing would produce mis-charged tRNAs. However, extensive efforts to establish such a LeuRS model failed.

The protein biosynthesis machinery of Candida albicans is of great interest, not only because it is a human pathogen but also in its cytoplasm, the universal Leu codon CUG is translated as both Ser (97%) and Leu (3%) (28, 29). This genetic code alteration is mediated by a uniquely evolved tRNA, which bears a CAG anti-codon [C. albicans tRNAser(CAG), CatRNAser] and can be aminoacylated either with Ser or C. albicans seryl-tRNA synthetase (CaSerRS) or with Leu by leucyl-tRNA synthetase (CaLeuRS) (29). Therefore, the proteome of C. albicans is ambiguous with some proteins exhibiting differences in primary sequences. For example, a key player in CUG reassignment, CaSerRS, has two isoforms (SerRS-Leu197 and SerRS-Ser197). The residue at position 197 is located at the SerRS dimer interface, and replacement of Ser by Leu at this site induces a local structural rearrangement, leading to a slightly higher (27%) activity of SerRS-Leu197 compared with SerRS-Ser197 (30). These data indicate that distribution of the CUG codon and its ambiguity is not random and has potential significance. CaLeuRS is another critical molecule in the CUG reassignment in C. albicans, which charges CatRNAser with Leu to produce Leu-CatRNAser. CaLeuRS comprises 1098 residues and has a molecular mass of 126 kDa. A single CUG codon is present at position 919 of CaLeuRS, which is located at the C-terminal domain. Thus, CaLeuRS should also have
two isoforms, CaLeuRS-Ser\textsuperscript{919} and CaLeuRS-Leu\textsuperscript{919}. Based on the decoding rule of \textit{C. albicans} (28,29), CaLeuRS exists mainly as CaLeuRS-Ser\textsuperscript{919} (~97%), and this was used here as the wild-type form.

In this study, we compared the activity of two LeuRS isoforms and analyzed the cross-species tRNA\textsuperscript{Leu} recognition and editing capacity of CaLeuRS. Interestingly, we showed that CaLeuRS is naturally deficient in tRNA-dependent pre-transfer editing activity but with obvious tRNA-independent pre-transfer editing and efficient post-transfer editing of Nva. However, it harbored a measurable level of tRNA-dependent pre-transfer editing of ABA when specific tRNA was present, although editing of ABA seemed not to be a necessity, as the rejection of ABA was efficient at the aminocyclation active site. Furthermore, post-transfer editing of CaLeuRS was not tRNA\textsuperscript{Leu} species-specific but was functional for mischarged CatRNA\textsuperscript{Ser}(CAG), being recognized by other eukaryotic LeuRSs.

**MATERIALS AND METHODS**

**Materials**

L-leucine (Leu), L-norvaline (Nva), L-isoleucine (Ile), L-valine (Val), L-methionine (Met), L-serine (Ser), ABA, dithiothreitol (DTT), ATP, CTP, GTP, UTP, 5’-GMP, tetrasodium pyrophosphate, inorganic pyrophosphate, ATP, Tris–HCl, MgCl\textsubscript{2}, NaCl, yeast total tRNA and yeast tRNALeu species-specific but was functional for mischarged CatRNA\textsuperscript{Ser}(CAG), being recognized by other eukaryotic LeuRSs.

**Gene cloning, mutagenesis and protein expression**

The \textit{C. albicans} genome was kindly provided by Prof. Jiang-Ye Chen of our institute and was used as the template for amplifying genes encoding \textit{CaLeuRS}, \textit{C. albicans} SerRS (\textit{CaSerRS}) and \textit{C. albicans} mitochondrial LeuRS (\textit{CmtLeuRS}). Gene sequences of \textit{CaLeuRS}, \textit{CaSerRS} and \textit{CmtLeuRS} were obtained from the Candida Genome Database (http://www.candidagenome.org/). \textit{CaLeuRS}, \textit{CaSerRS} and \textit{CmtLeuRS} genes were cloned into pET28a at the \textit{NheI} and \textit{XhoI} sites with N-terminal His\textsubscript{6}-tag (the mitochondrial targeting sequence of \textit{CmtLeuRS} had been removed). Plasmids containing EcLeuRS (10), SrLeuRS (18) and \textit{Pyrococcus horikoshii} LeuRS (PhLeuRS) (31) were constructed previously. The \textit{E. coli} tRNA\textsuperscript{mG37} methyltransferase (TrmD) gene was amplified from the \textit{E. coli} genome and inserted between the EcoRI and XhoI sites of pET28a. The plasmid expressing \textit{E. coli} tRNA nucleotidyltransferase (CCase) was provided by Dr. Gilbert Eriani (Strasbourg, CNRS, France). Mutation at Asp\textsuperscript{422} of the CaLeuRS gene was performed with the KOD-plus mutagenesis kit according to the manufacturer’s instructions. Asp\textsuperscript{422} corresponds to Asp\textsuperscript{373}, Asp\textsuperscript{419}, Asp\textsuperscript{444} and Asp\textsuperscript{399} of \textit{AaLeuRS}, \textit{ScLeuRS}, \textit{GiLeuRS} and \textit{hclLeuRS}, respectively, which are crucial for post-transfer editing of these LeuRSs (9,10,18,19,27). The CTG and TCG codons at position 919 in the CaLeuRS gene were used to over-express the gene encoding CaLeuRS-Leu\textsuperscript{919} and CaLeuRS-Ser\textsuperscript{919}, respectively. All constructs were confirmed by DNA sequencing. \textit{E. coli} BL21 (DE3) was transformed with various constructs. A single colony of each of the transformants was chosen and cultured in 500 ml of 2 x YT medium at 37°C. When the cells reached mid-log phase (A\textsubscript{600} = 0.6), expression of the recombinant proteins was induced by the addition of 0.2 mM isopropyl-1-thio-β-D-galactopyranoside for 8 h at 22°C. Protein purification was performed according to a previously described method (32).

**tRNA gene cloning, transcription and methylation**

CatRNA\textsuperscript{Leu}(UAA) and CatRNA\textsuperscript{Ser}(CAG) genes were cloned between the \textit{PstI} and \textit{EcoRI} sites of pTrec99b with an N-terminal T7 promoter. Detailed T7 in vitro run-off transcription of CatRNA\textsuperscript{Leu} and CatRNA\textsuperscript{Ser} has been described previously (33). The amino acid accepting activities of CatRNA\textsuperscript{Leu}(UAA) or CatRNA\textsuperscript{Ser}(CAG) are 1390 and 1208 pmol/A\textsubscript{260}, respectively. The methyl group of m\textsuperscript{1}G37 of CatRNA\textsuperscript{Ser} is a critical element for recognition by LeuRS (29). The purified CatRNA\textsuperscript{Ser} transcript was methylated at position G37 with \textit{E. coli} TrmD (34) in a mixture containing 0.1 M Tris–HCl (pH 8.0), 1 mM DTT, 0.1 mM EDTA, 6 mM MgCl\textsubscript{2}, 24 mM NH\textsubscript{4}Cl, 7.5 μg of bovine serum albumin, 5 μM CatRNA\textsuperscript{Ser} transcript, 100 μM S-adenosylmethionine, 1 U/μl RNase inhibitor and 10 μM TrmD at 37°C for 1.5 h. Approximately 45% of transcripts were methylated in this reaction as estimated in a control experiment with \textsuperscript{3}H-labeled S-adenosylmethionine. m\textsuperscript{1}G37-CatRNA\textsuperscript{Ser} was ethanol-precipitated at −20°C after phenol/chloroform extraction (twice) and dissolved in 5 mM MgCl\textsubscript{2}. All CatRNA\textsuperscript{Ser} used in this study refers to m\textsuperscript{1}G37-CatRNA\textsuperscript{Ser}.

\textsuperscript{32}P-labeling of CatRNA\textsuperscript{Leu} or CatRNA\textsuperscript{Ser}

\textsuperscript{32}P-labeling of CatRNA\textsuperscript{Leu} or CatRNA\textsuperscript{Ser} was performed at 37°C in a mixture containing 60 mM Tris–HCl (pH 8.0), 12 mM MgCl\textsubscript{2}, 15 μM CatRNA\textsuperscript{Leu} or CatRNA\textsuperscript{Ser}, 0.5 mM DTT, 15 μM ATP, 50 μM tetrasodium pyrophosphate,
was dissolved in 5 mM MgCl₂.

mals were centrifuged (10 000 g, 5 min). The supernatant was loaded and separated by thin layer chromatography (TLC) according to the procedure described for mis- aminoacylation. Post-transfer editing of pre-formed Nva-

Aminoacylation of CatRNA⁹⁺ with Leu was performed in a reaction mixture containing 60 mM Tris–HCl (pH 7.5), 10 mM MgCl₂, 2 mM DTT, 4 mM ATP, 10 mM CatRNA⁹⁺, 20 μM [³²P]ATP and 20 nM CaLeuRS. The kinetics of amino acid activation were measured in the presence of Leu (3–1000 μM) or Nva (0.3–50 mM) or ABA (3–940 mM). Samples of the reaction mixture were removed at specific time-points, added to 200 μl of quenching solution containing 2% activated charcoal, 3.5% HClO₄ and 50 mM tetrasodium pyrophosphate and mixed by vortexing for 20 s. The solution was filtered through a Whatman GF/C filter, followed by washing with 20 ml of 10 mM tetrasodium pyrophosphate solution and 10 ml of 100% ethanol. The filters were dried, and [³²P]ATP was counted using a scintillation counter (Beckman Coulter).

Aminoacylation of CatRNA⁹⁺ with Leu was performed in a reaction mixture containing 60 mM Tris–HCl (pH 7.5), 10 mM MgCl₂, 2 mM DTT, 4 mM ATP, 10 mM CatRNA⁹⁺, 20 μM [³²P]ATP and 20 nM CaLeuRS at 30°C. The kinetics of CaLeuRS aminoacylation were measured in the presence of CatRNA⁹⁺ (0.6–15.8 μM) or transcribed or over-expressed CtRNA⁹⁺ (0.6–10 μM) or yeast total tRNA (0.2–6 μM) or over-expressed htrNA⁻⁹⁺ (0.2–6 μM) or transcribed htrNA⁻⁹⁺ (0.6–10 μM).

Aminoacylation of htrNA⁻⁹⁺ with Nva or ABA was carried out at 30°C in a reaction mixture containing 60 mM Tris–HCl (pH 7.5), 10 mM MgCl₂, 2 mM DTT, 4 mM ATP, 5 μM °cold' CatRNA⁻⁹⁺, 1 μM [³²P]CatRNA⁻⁹⁺, 20 mM Nva or 376 mM ABA and 1 μM CaLeuRS or CaLeuRS-D422A. Samples at specific time-points were taken for ethanol precipitation with NaAc (pH 5.2) at -20°C overnight. The precipitated samples were centrifuged (10000 g) at 4°C for 30 min, dried at room temperature for 30 min and digested with 6 μl of nuclease S₁ (25 U) for 2 h at 37°C. After treatment with nuclease S₁, aminoacyl-[³²P]tRNA should produce aminoacyl-[³²P]AMP and free [³²P]tRNA should produce [³²P]AMP. Samples (2 μl) of the digestion mixture were loaded and separated by thin layer chromatography (TLC) in 0.1 M NH₄Ac and 5% acetic acid. Known amounts of [³²P]AMP were diluted and loaded onto the TLC plate for the purposes of quantification. The plates were visualized by phosphorimaging, and the data were analyzed using Multi-Gauge Version 3.0 software (FUJIFILM). Measurement of Nva-[³²P]CatRNA⁻⁹⁺ synthesis by CaLeuRS and CaLeuRS-D422A was performed using a similar procedure, except [³²P]CatRNA⁻⁹⁺ was used as a substrate. Mis-aminoacylation of [³²P]CatRNA⁻⁹⁺ with Leu by various LeuRSs was carried out in a reaction mixture containing 60 mM Tris–HCl (pH 7.5), 10 mM MgCl₂, 2 mM DTT, 4 mM ATP, 5 μM °cold' CatRNA⁻⁹⁺, 1 μM [³²P]CatRNA⁻⁹⁺, 40 μM Leu and 1 μM CaLeuRS, CaLeuRS-D422A, CmLeuRS, EеЛeУRS, ScLeuRS, hLeuRS and PhLeuRS at 37°C.

In vitro activity assays

APT-PPi exchange measurement was carried out at 30°C in a reaction mixture containing 60 mM Tris–HCl (pH 7.5), 10 mM MgCl₂, 2 mM DTT, 4 mM ATP, 2 mM [³²P]tetrasodium pyrophosphate, 1 mM Leu or 50 mM non-cognate ABA, Nva, Val, Ile, Met, Ser and 20 mM CaLeuRS. The kinetics of amino acid activation were measured in the presence of Leu (3–1000 μM) or Nva (0.3–50 mM) or ABA (3–940 mM). Samples of the reaction mixture were removed at specific time-points, added to 200 μl of quenching solution containing 2% activated charcoal, 3.5% HClO₄ and 50 mM tetrasodium pyrophosphate and mixed by vortexing for 20 s. The solution was filtered through a Whatman GF/C filter, followed by washing with 20 ml of 10 mM tetrasodium pyrophosphate solution and 10 ml of 100% ethanol. The filters were dried, and [³²P]ATP was counted using a scintillation counter (Beckman Coulter).

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Mism-aminoacylation of [³²P]CatRNA⁹⁺ with Nva or ABA was carried out at 30°C in a reaction mixture containing 60 mM Tris–HCl (pH 7.5), 10 mM MgCl₂, 2 mM DTT, 4 mM ATP, 5 μM °cold' CatRNA⁻⁹⁺, 1 μM [³²P]CatRNA⁻⁹⁺, 20 mM Nva or 376 mM ABA and 1 μM CaLeuRS or CaLeuRS-D422A. Samples at specific time-points were taken for ethanol precipitation with NaAc (pH 5.2) at -20°C overnight. The precipitated samples were centrifuged (10000 g) at 4°C for 30 min, dried at room temperature for 30 min and digested with 6 μl of nuclease S₁ (25 U) for 2 h at 37°C. After treatment with nuclease S₁, aminoacyl-[³²P]tRNA should produce aminoacyl-[³²P]AMP and free [³²P]tRNA should produce [³²P]AMP. Samples (2 μl) of the digestion mixture were loaded and separated by thin layer chromatography (TLC) in 0.1 M NH₄Ac and 5% acetic acid. Known amounts of [³²P]AMP were diluted and loaded onto the TLC plate for the purposes of quantification. The plates were visualized by phosphorimaging, and the data were analyzed using Multi-Gauge Version 3.0 software (FUJIFILM). Measurement of Nva-[³²P]CatRNA⁻⁹⁺ synthesis by CaLeuRS and CaLeuRS-D422A was performed using a similar procedure, except [³²P]CatRNA⁻⁹⁺ was used as a substrate. Mis-aminoacylation of [³²P]CatRNA⁻⁹⁺ with Leu by various LeuRSs was carried out in a reaction mixture containing 60 mM tris–HCl (pH 7.5), 10 mM MgCl₂, 2 mM DTT, 4 mM ATP, 5 μM °cold' CatRNA⁻⁹⁺, 1 μM [³²P]CatRNA⁻⁹⁺, 40 μM Leu and 1 μM CaLeuRS, CaLeuRS-D422A, CmLeuRS, EеЛeУRS, ScLeuRS, hLeuRS and PhLeuRS at 37°C.

Preparation of Nva-[³²P]CatRNA⁹⁺ or Nva-[³²P]CatRNA⁻⁹⁺ was carried out with editing-deficient ScLeuRS-D419A or CaLeuRS-D422A, respectively, in a reaction mixture, which was identical to that used for mis-aminoacylation. Post-transfer editing of pre-formed Nva-

Results

CaLeuRS-Leu⁹⁺ is more active than CaLeuRS-Ser⁹⁺

Determination of the crystal structure of the PhLeuRS-

The AMP formation assay was carried out at 30°C in a reaction mixture containing 60 mM Tris–HCl (pH 7.5), 10 mM MgCl₂, 5 mM DTT, 10 U/ml PPiase, 15 mM Nva or 350 mM ABA, 3 mM [³²P]AMP and 0.2 μM CaLeuRS in the presence or absence of 15 μM transcribed CatRNA⁹⁺, or 5 μM transcribed or over-expressed EcRNA⁹⁺ or htrNA⁻⁹⁺. Samples (1.5 μl) were quenched in 6 μl of 200 mM NaAc (pH 5.0). The quenched aliquots (1.5 μl of each sample) were spotted onto polyethyleneimine cellulose plates pre-washed with water. Separation of Nva/ABA-[³²P]AMP or [³²P]AMP and [³²P]ATP was performed in 0.1 M NH₄Ac and 5% acetic acid. Quantification of [³²P]AMP was achieved by densitometry in comparison with [³²P]ATP samples of known concentrations.

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Subsequent comparisons of the aminoacylation kinetics of CaLeuRS-Ser919 and CaLeuRS-Leu919 revealed that CaLeuRS-Leu919 displayed a higher $K_m$ (2.91 ± 0.37 mM) and a higher $k_{cat}$ (0.62 ± 0.08 s$^{-1}$) compared with the values determined for CaLeuRS-Ser919 ($K_m$: 1.87 ± 0.23 mM, $k_{cat}$: 0.31 ± 0.05 s$^{-1}$). These data indicated that CaLeuRS-Ser919 has a stronger binding affinity for transcribed Ca$tRNA_{Leu(UAA)}$ during aminoacylation (Table 1). Based on the structure, we suggested that the presence of Ser in this helix may facilitate binding with the variable stem-loop element of Ca$tRNA_{Leu(UAA)}$. The catalytic efficiency of CaLeuRS-Leu919 (213.06 s$^{-1}$M$^{-1}$) is ~30% higher than that of CaLeuRS-Ser919 (165.78 s$^{-1}$M$^{-1}$). This phenomenon is similar to that observed in the case of CaSerRS, for which no differences were observed in the amino acid activation of CaSerRS-Leu$^{197}$ and CaSerRS-Ser$^{197}$, whereas CaSerRS-Leu$^{197}$ showed a slightly (27%) higher activity than CaSerRS-Ser$^{197}$(30).

According to the decoding rule of C. albicans, CaLeuRS is present in the cytoplasm mainly in the form of CaLeuRS-Ser919. Thus, in the following study, we used CaLeuRS-Ser$^{197}$ as the wild-type CaLeuRS.

Yeast LeuRSs efficiently recognized bacterial, yeast and human tRNA$^{14}$Leu

Species-specific charging of tRNA is common for some aaRS systems. The aaRSs from higher organisms often have the capacity to charge tRNA from lower species, whereas aaRSs from lower organisms fail to aminoacylate tRNA from higher ones. It is unclear whether yeast LeuRS is able to recognize various tRNA$^{14}$Leu from other species. In this study, we investigated the tRNA$^{14}$Leu recognition capacity in detail using CaLeuRS as a model system.

The CaRNA$^{14}$Leu gene could not be over-expressed in E. coli and was obtained by T7 in vitro transcription. We also obtained transcribed and over-expressed EctRNA$^{14}$Leu and hctRNA$^{14}$Leu to reveal any potential role of base modification in recognition. Moreover, as transcribed S. cerevisiae tRNA$^{14}$Leu without modification showed no Leu accepting activity (data not shown), commercial S. cerevisiae yeast total tRNA was used.

CaLeuRS recognized all the available tRNAs. It aminoacylated transcribed or over-expressed EctRNA$^{14}$Leu with similar $k_{cat}$ values (0.474 ± 0.026 and

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**Table 1.** Aminoacylation kinetic parameters of CaLeuRS-Ser$^{197}$ and CaLeuRS-Leu$^{199}$ for the Ca$tRNA_{Leu}$ transcripta

<table>
<thead>
<tr>
<th>Enzyme</th>
<th>$K_m$ (μM)</th>
<th>$k_{cat}$ (s$^{-1}$)</th>
<th>$k_{cat}/K_m$ (s$^{-1}$M$^{-1}$)</th>
<th>Relative $k_{cat}/K_m$</th>
<th>$k_{cat}$/K$^{-1}$M$^{-1}$</th>
</tr>
</thead>
<tbody>
<tr>
<td>CaLeuRS-Ser$^{197}$</td>
<td>1.87 ± 0.23</td>
<td>0.31 ± 0.05</td>
<td>165.78</td>
<td>1.0</td>
<td>213.06</td>
</tr>
<tr>
<td>CaLeuRS-Leu$^{199}$</td>
<td>2.91 ± 0.37</td>
<td>0.62 ± 0.08</td>
<td>165.78</td>
<td>1.3</td>
<td>213.06</td>
</tr>
</tbody>
</table>

aThe results are the average of three independent repeats with standard deviations indicated.
0.555 ± 0.061 s⁻¹, respectively), although the \( K_m \) for transcribed \( \text{EctRNA}^{\text{Leu}} \) (2.68 ± 0.39 mM) was nearly 4-fold greater than that for over-expressed \( \text{EctRNA}^{\text{Leu}} \) (0.74 ± 0.08 mM). Interestingly, \( \text{CaLeuRS} \) efficiently charged both transcribed and over-expressed \( \text{hctRNA}^{\text{Leu}} \), which was from a higher organism. A similar recognition pattern as seen with the two \( \text{EctRNA}^{\text{Leu}} \)'s was also observed, with comparable \( k_{\text{cat}} \) values but a smaller \( K_m \) for over-expressed \( \text{hctRNA}^{\text{Leu}} \), indicating base modification was important for tRNA recognition. Additionally, \( \text{CaLeuRS} \) obviously charged yeast total tRNA with \( K_m \) and \( k_{\text{cat}} \) values of 0.39 ± 0.05 mM and 0.174 ± 0.019 s⁻¹, respectively, and with the greatest catalytic efficiency (1486.05 s⁻¹mM⁻¹) for over-expressed \( \text{hctRNA}^{\text{Leu}} \) among all the tested tRNAs (Table 2).

Owing to recognition ability of \( \text{CaLeuRS} \) for \( \text{hctRNA}^{\text{Leu}} \), we further explored the capacity of \( \text{ScLeuRS} \) to aminoacylate bacterial and human tRNA\(^{\text{Leu}} \) as well as yeast tRNA. \( \text{ScLeuRS} \) aminoacylated yeast total tRNA with \( K_m \) and \( k_{\text{cat}} \) values of 0.332 ± 0.037 mM and 0.188 ± 0.025 s⁻¹, respectively. However, its \( k_{\text{cat}} \) values for over-expressed \( \text{EctRNA}^{\text{Leu}} \) or \( \text{hctRNA}^{\text{Leu}} \) increased >10-fold (2.09 ± 0.16 and 2.19 ± 0.22 s⁻¹, respectively), although the \( K_m \) values differed from each other remarkably (2.19 ± 0.47 mM for over-expressed \( \text{EctRNA}^{\text{Leu}} \) and 0.111 ± 0.027 mM for over-expressed \( \text{hctRNA}^{\text{Leu}} \)). These data demonstrated that over-expressed \( \text{hctRNA}^{\text{Leu}} \) was the best aminoacylation substrate for \( \text{ScLeuRS} \) (catalytic efficiency 19729.73 s⁻¹mM⁻¹) and furthermore suggested that base modification was important during recognition or catalysis. \( \text{ScLeuRS} \) recognized transcribed \( \text{EctRNA}^{\text{Leu}} \) with a similar \( K_m \) (1.71 ± 0.22 mM) but a sharply decreased \( k_{\text{cat}} \) (0.134 ± 0.011 s⁻¹) compared with the values of over-expressed \( \text{EctRNA}^{\text{Leu}} \). It also recognized transcribed \( \text{hctRNA}^{\text{Leu}} \) with an increased \( K_m \) (0.926 ± 0.170 mM) and a decreased \( k_{\text{cat}} \) (0.887 ± 0.114 s⁻¹) compared with the values for over-expressed \( \text{hctRNA}^{\text{Leu}} \) (Table 2).

Overall, both \( \text{CaLeuRS} \) and \( \text{ScLeuRS} \) recognized bacterial, yeast and human tRNA\(^{\text{Leu}} \). Interestingly, recognition of CatRNA\(^{\text{Leu}} \) by hcLeuRS was negligible (Supplementary Figure S1A). Furthermore, EcLeuRS failed to acylate CatRNA\(^{\text{Leu}} \) (Supplementary Figure S1B). These results were unexpected because it is widely accepted that aaRSs from higher organisms are able to aminoacylate tRNAs from lower organisms.

### Table 2. Aminoacylation kinetic parameters of \( \text{CaLeuRS} \) and \( \text{ScLeuRS} \) for various tRNAs

<table>
<thead>
<tr>
<th>Enzyme</th>
<th>tRNA</th>
<th>( K_m ) (µM)</th>
<th>( k_{\text{cat}} ) (s⁻¹)</th>
<th>( k_{\text{cat}}/K_m ) (s⁻¹mM⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td>( \text{CaLeuRS} )</td>
<td>( \text{OE}^{\text{E}}\text{-EctRNA}^{\text{Leu}} )</td>
<td>0.74 ± 0.08</td>
<td>0.474 ± 0.026</td>
<td>640.54</td>
</tr>
<tr>
<td></td>
<td>( \text{TS}^{\text{E}}\text{-EctRNA}^{\text{Leu}} )</td>
<td>2.68 ± 0.39</td>
<td>0.555 ± 0.061</td>
<td>207.09</td>
</tr>
<tr>
<td></td>
<td>yeast total tRNA</td>
<td>0.39 ± 0.05</td>
<td>0.174 ± 0.019</td>
<td>446.15</td>
</tr>
<tr>
<td></td>
<td>( \text{OE}-\text{hctRNA}^{\text{Leu}} )</td>
<td>0.853 ± 0.05</td>
<td>0.639 ± 0.055</td>
<td>1486.05</td>
</tr>
<tr>
<td></td>
<td>( \text{TS}-\text{hctRNA}^{\text{Leu}} )</td>
<td>1.30 ± 0.23</td>
<td>0.719 ± 0.094</td>
<td>553.08</td>
</tr>
<tr>
<td>( \text{ScLeuRS} )</td>
<td>( \text{OE}-\text{EctRNA}^{\text{Leu}} )</td>
<td>2.19 ± 0.47</td>
<td>2.09 ± 0.16</td>
<td>954.34</td>
</tr>
<tr>
<td></td>
<td>( \text{TS}-\text{EctRNA}^{\text{Leu}} )</td>
<td>1.71 ± 0.22</td>
<td>0.134 ± 0.011</td>
<td>78.36</td>
</tr>
<tr>
<td></td>
<td>yeast total tRNA</td>
<td>0.332 ± 0.037</td>
<td>0.188 ± 0.025</td>
<td>566.27</td>
</tr>
<tr>
<td></td>
<td>( \text{OE}-\text{hctRNA}^{\text{Leu}} )</td>
<td>0.111 ± 0.027</td>
<td>2.19 ± 0.22</td>
<td>19729.73</td>
</tr>
<tr>
<td></td>
<td>( \text{TS}-\text{hctRNA}^{\text{Leu}} )</td>
<td>0.926 ± 0.170</td>
<td>0.887 ± 0.114</td>
<td>957.88</td>
</tr>
</tbody>
</table>

*The results are the average of three independent repeats with standard deviations indicated.

\( \text{OE} \), over-expressed.

\( \text{TS} \), transcribed.

### Amino acid activation capacity of \( \text{CaLeuRS} \)

Various LeuRSs have been shown to mis-activate a series of non-cognate amino acids. To investigate mis-activation of non-cognate amino acids by \( \text{CaLeuRS} \), we included ABA, Nva, Val, Ile, Met, Ser in the ATP-PPi exchange reaction. The data clearly showed that \( \text{CaLeuRS} \) significantly mis-activated Nva; furthermore, ABA was also mis-activated to an obvious level compared with the control reaction conducted in the absence of amino acids. In contrast, mis-activation of Val, Ile, Met and Ser was comparable with that of the control reaction conducted in the absence of amino acids (Figure 2). To further define the quantitative discrimination capacity of the aminoacylation active site of \( \text{CaLeuRS} \), we measured the activation kinetics for cognate Leu and non-cognate Nva and ABA of \( \text{CaLeuRS} \). \( \text{CaLeuRS} \) gave much higher \( K_m \) values for Nva (5487 ± 645 mM) and ABA (12087 ± 1698 mM); however, the \( k_{\text{cat}} \) values were comparable with that for Leu, equating to discriminator factors for Nva and ABA of 220 and 3462, respectively (Table 3). These results indicated that Nva is a real challenge for \( \text{CaLeuRS} \) and that removal of Nva-AMP and/or Nva-tRNA\(^{\text{Leu}} \) is required to maintain the translational quality control. However, the discrimination against ABA was below the proposed threshold of 1/3000, indicating that editing of ABA may not be necessary.

### \( \text{CaLeuRS} \) exhibited little tRNA-dependent pre-transfer editing for Nva

The hydrolysis of Nva-AMP or Nva-tRNA\(^{\text{Leu}} \) may be separately or simultaneously catalyzed by \( \text{CaLeuRS} \). Editing leads to the net consumption of ATP (yielding AMP) due to repetitive cycles of synthesis-hydrolysis of the non-cognate products. This is the basis of the TLC-based AMP formation methodology, in which the editing capacity is measured by monitoring the quantity...
of AMP produced (9–11,36,37). In the presence of tRNA and non-cognate amino acid, the TLC assay measures the global editing activity, including the tRNA-independent and tRNA-dependent pre-transfer editing in addition to the post-transfer editing. In the absence of tRNA, but with non-cognate amino acid, AMP is produced only by the lack of availability of over-expressed CatRNALeu.

### Table 3. Amino acid activation kinetics of CaLeuRS for various amino acids

<table>
<thead>
<tr>
<th>Amino acid</th>
<th>$k_{\text{cat}}$ (s$^{-1}$)</th>
<th>$k_{\text{cat}}/K_m$ (s$^{-1}$ mM$^{-1}$)</th>
<th>Discrimination factor</th>
</tr>
</thead>
<tbody>
<tr>
<td>Leu</td>
<td>40.11 ± 3.84</td>
<td>93.05 ± 10.25</td>
<td>2319.87</td>
</tr>
<tr>
<td>Nva</td>
<td>5487 ± 645</td>
<td>57.77 ± 6.35</td>
<td>10.53</td>
</tr>
<tr>
<td>ABA</td>
<td>120387 ± 1698</td>
<td>80.28 ± 10.75</td>
<td>0.67</td>
</tr>
</tbody>
</table>

*aThe results are the average of three independent repeats with standard deviations indicated.
*bDiscrimination factor corresponds to the loss of catalytic efficiency relative to Leu.
*Discrimination factor corresponds to the loss of catalytic efficiency relative to Leu.

Figure 2. Amino acid activation properties of CaLeuRS. Activation of Leu (black square), ABA (black up-pointing triangle), Nva (black down-pointing triangle), Val (black diamond), Ile (white circle), Met (white square) and Ser (white up-pointing triangle) by CaLeuRS, Leu and non-cognate amino acids were used at final concentrations of 1 and 50 mM, respectively. A control reaction without any amino acids (black circle) was included. The insert compares the activation of ABA and Nva to Leu.

### Table 4. $k_{\text{obs}}$ values of CaLeuRSs for editing Nva with various tRNAs

<table>
<thead>
<tr>
<th>Enzyme</th>
<th>tRNA</th>
<th>$k_{\text{obs}}$ (s$^{-1}$)*</th>
</tr>
</thead>
<tbody>
<tr>
<td>CoLeuRS</td>
<td>No tRNA</td>
<td>0.25 ± 0.03</td>
</tr>
<tr>
<td>CatRNALeu</td>
<td>0.28 ± 0.04</td>
<td></td>
</tr>
<tr>
<td>TS-EctRNALeu</td>
<td>0.40 ± 0.06</td>
<td></td>
</tr>
<tr>
<td>OE-EctRNALeu</td>
<td>0.51 ± 0.04</td>
<td></td>
</tr>
<tr>
<td>TS-hctRNALeu</td>
<td>0.33 ± 0.02</td>
<td></td>
</tr>
<tr>
<td>OE-hctRNALeu</td>
<td>0.38 ± 0.05</td>
<td></td>
</tr>
<tr>
<td>CoLeuRS-D422A</td>
<td>No tRNA</td>
<td>0.22 ± 0.06</td>
</tr>
<tr>
<td>OE-hctRNALeu</td>
<td>0.25 ± 0.04</td>
<td></td>
</tr>
<tr>
<td>CoLeuRS-Leu919</td>
<td>No tRNA</td>
<td>0.22 ± 0.02</td>
</tr>
<tr>
<td>CatRNALeu</td>
<td>0.25 ± 0.05</td>
<td></td>
</tr>
</tbody>
</table>

*aThe results are the average of three independent repeats with standard deviations indicated.
*bTS, transcribed.
*cOE, over-expressed.

The absence of significant stimulation of editing of Nva by CaLeuRSs with various tRNALeu's prompted us to investigate its post-transfer editing capability. Usually, the post-transfer editing ability of various LeuRSs is monitored by hydrolysis of Ile- or Met-tRNALeu, which are easily available and thus available radioactive Ile or Met using a LeuRS mutant without post-transfer editing capability. Because we focused on the Nva-editing properties of CaLeuRS and Nva labeled with radioactive isotope was not commercially available, the 3’ end of CatRNALeu was first labeled with [z-32P]ATP by E. coli CCase, and then Nva-[32P]CatRNALeu was prepared by editing-deficient
ScLeuRS-D419A (13,18). Hydrolytic analysis clearly showed that CaLeuRS edited Nva-[32P]tRNALeu when compared with the control experiment conducted in the absence of the enzyme (Figure 3A and B). To confirm the post-transfer editing reaction catalyzed by CaLeuRS, we mutated the conserved and post-transfer editing-essential Asp422 to generate CaLeuRS-D422A. Asp422 corresponds to Asp373, Asp419, Asp444 and Asp399 of AaLeuRS, ScLeuRS, GlLeuRS and hcLeuRS, respectively, which are crucial to post-transfer editing by these LeuRSs (9,10,18,19,27). Indeed, CaLeuRS-D422A did not hydrolyze Nva-[32P]tRNALeu and was deficient in post-transfer editing, indicating that this mutation inactivated the CP1 domain of CaLeuRS (Figure 3A and B). Further mis-aminoacylation of [32P]AMP showed that a significant amount of Nva-[32P]tRNALeu was formed by CaLeuRS-D422A; however, a negligible amount of mis-charged [32P]tRNA^{Leu} was formed by CaLeuRS (Figure 3C and D). Overall, these data showed that CaLeuRS harbored an obvious and efficient capability for post-transfer editing of Nva-[32P]tRNALeu, the loss of which caused accumulation of mis-charged tRNALeu. No further accumulation of AMP after the addition of tRNALeu in the AMP formation assay (Table 4) suggested that post-transfer editing of mis-charged tRNALeu contributed little to the total editing. To further explore the absence of tRNA-dependent pre-transfer editing of Nva by CaLeuRS, we tested the AMP formation of CaLeuRS-D422A in the presence of Nva with or without over-produced hctRNALeu. The data showed that over-produced hctRNA^{Leu} with modified bases did not stimulate further AMP production after abolishing post-transfer editing (k_{obs} 0.22 ± 0.06 versus 0.25 ± 0.04 s^{-1}), confirming the lack of tRNA-dependent pre-transfer editing of Nva by CaLeuRS (Table 4).

**CaLeuRS inhibited synthesis of Nva-tRNA^{Ser} by non-tRNA species-specific post-transfer editing**

The ability of CaLeuRS to efficiently mis-activate non-cognate Nva and recognize non-cognate CatRNA^{Ser} raises the interesting question of how to prevent the formation of Nva-CatRNA^{Ser}. To test for the presence of post-transfer editing activity that hydrolyzes potentially synthesized Nva-CatRNA^{Ser}, mis-aminoacylation of [32P]CatRNA^{Ser} with Nva by CaLeuRS was compared with that of the post-transfer editing-deficient CaLeuRS-D422A. The data clearly showed that, mutation of Asp422 resulted in significant synthesis of Nva-[32P]CatRNA^{Ser} by the mutant, in contrast to wild-type enzyme, which generated negligible amounts of Nva-[32P]CatRNA^{Ser}, indicating that CaLeuRS used post-transfer editing to prevent Nva-CatRNA^{Ser} synthesis (Figure 4A and B).
We then prepared Nva-CatRNASer for use in hydrolysis assays to more directly monitor the post-transfer editing activity. Obvious hydrolysis of Nva-[^32P]CatRNASer was mediated by CatLeuRS but not CatLeuRS-D422A (Figure 4C and D). Above all, these data showed that the post-transfer editing by CatLeuRS was not only CatRNALeu specific but also efficient for CatRNASer to inhibit synthesis of both Nva-CatRNALeu and Nva-CatRNASer.

CatLeuRS possessed weak tRNA-dependent pre-transfer editing capacity for ABA

ABA was selected to test whether CaLeuRS possessed any tRNA-dependent pre-transfer editing of other non-cognate amino acids because it was obviously activated by CaLeuRS. CatRNA^Leu transcript, transcribed or over-produced EctRNA^Leu and hctRNA^Leu were used to trigger editing of ABA by CaLeuRS (Table 5). The data showed that over-produced hctRNA^Leu obviously stimulated editing by increasing the k_{obs} 5-fold [(23.19 ± 3.62) \times 10^{-3} \text{s}^{-1}] compared with that in the absence of tRNA [(4.69 ± 0.51) \times 10^{-3} \text{s}^{-1}]. Over-expressed EctRNA^Leu led only to an ~3-fold increase in k_{obs} [(14.58 ± 2.14) \times 10^{-3} \text{s}^{-1}]. However, CatRNA^Leu, EctRNA^Leu and hctRNA^Leu transcripts had little effect on the rate of ABA-editing (Table 5). These data implied that editing of ABA was tRNA modification-dependent.

### Table 5. k_{obs} values of CaLeuRSs for editing ABA with various tRNAs

<table>
<thead>
<tr>
<th>Enzyme</th>
<th>tRNA</th>
<th>k_{obs} (x10^3) (s^{-1})</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>No tRNA</td>
<td>4.69 ± 0.51</td>
</tr>
<tr>
<td>CaLeuRS</td>
<td>CatRNA^Leu</td>
<td>4.90 ± 0.37</td>
</tr>
<tr>
<td></td>
<td>TS-EctRNA^Leu</td>
<td>4.78 ± 0.26</td>
</tr>
<tr>
<td></td>
<td>OE-EctRNA^Leu</td>
<td>14.58 ± 2.14</td>
</tr>
<tr>
<td></td>
<td>TS-hctRNA^Leu</td>
<td>10.93 ± 0.85</td>
</tr>
<tr>
<td></td>
<td>OE-hctRNA^Leu</td>
<td>23.19 ± 3.62</td>
</tr>
<tr>
<td>CaLeuRS-D422A</td>
<td>OE-hctRNA^Leu</td>
<td>12.16 ± 1.98</td>
</tr>
</tbody>
</table>

*aThe results are the average of three independent repeats with standard deviations indicated.
*bTS, transcribed.
*cOE, over-expressed.

As over-produced hctRNA^Leu was the most efficient stimulator of ABA-editing, we measured AMP formation by the editing-deficient CaLeuRS-D422A mutant in the presence of ABA with over-produced hctRNA^Leu. Mutation of Asp^{422}, which abolished post-transfer editing, apparently decreased the rate of AMP formation with a k_{obs} of (12.16 ± 1.98) \times 10^{-3} \text{s}^{-1}. Therefore, with over-produced hctRNA^Leu, post-transfer editing of ABA by CaLeuRS accounted for 47.6% of the total editing [(23.19 – 12.16)/23.19], whereas tRNA-independent and
tRNA-dependent pre-transfer editing of ABA only accounted for 20.2% (4.69/23.19) and 32.2% [(12.16 - 4.69)/23.19], respectively, of the total editing of ABA by CaLeuRS.

We further performed aminoacylation of \([^{32}P]CatRNA_{Leu}\) by CaLeuRS and CaLeuRS-D422A with saturating concentrations of ABA. The data showed that defective post-transfer editing resulted in the generation of significantly more ABA-\([^{32}P]CatRNA_{Leu}\), however, surprisingly, even CaLeuRS yielded a significant amount of ABA-\([^{32}P]CatRNA_{Leu}\) (Figure 3C and D). These data implied that editing of ABA by CaLeuRS was not sufficient to prevent the synthesis of ABA-\([^{32}P]CatRNA_{Leu}\) in the presence of saturating ABA concentrations. This paradox between ABA mis-aminoacylation and charging accuracy may be solved by fine discrimination against ABA at the aminoacylation active site (Table 3).

These results revealed that CaLeuRS exhibits a weak level of tRNA-dependent pre-transfer editing activity for ABA. In addition, the total ABA-editing capacity is not sufficient to avoid the formation of mis-charged tRNA_{Leu}, which is different from Nva-editing capacity.

ScLeuRS, like CaLeuRS, also exhibited little tRNA-dependent pre-transfer editing capacity for Nva

The natural deficiency in tRNA-dependent pre-transfer editing of Nva by CaLeuRS prompted us to investigate whether it is a common characteristic of other yeast LeuRSs. Therefore, we assayed the Nva-included AMP formation catalyzed by ScLeuRS in the absence or presence various tRNA_{Leu}s. The data showed that over-produced hctRNA_{Leu} obviously stimulated editing (k_{obs} of 0.64 ± 0.04 s\(^{-1}\)) compared with that observed in the absence of tRNA (0.10 ± 0.02 s\(^{-1}\)). However, transcribed CatRNA_{Leu}, EctRNA_{Leu}, hctRNA_{Leu} and over-expressed EctRNA_{Leu} failed to trigger further editing by ScLeuRS (Table 6). The formation of AMP stimulated by tRNA_{Leu} should be derived from tRNA-dependent pre-transfer editing and/or post-transfer editing. To distinguish between these two pathways, the conserved Asp\(^{419}\) was mutated to generate ScLeuRS-D419A, which has been shown to be defective in post-transfer editing and is homologous with Asp\(^{422}\) of CaLeuRS (13,18). Assay of the Nva-included AMP formation by ScLeuRS-D419A, which has been shown to be defective in post-transfer editing and is homologous with Asp\(^{422}\) of CaLeuRS (13,18).

<table>
<thead>
<tr>
<th>Enzyme</th>
<th>tRNA</th>
<th>k_{obs} (s(^{-1}))^a</th>
</tr>
</thead>
<tbody>
<tr>
<td>ScLeuRS</td>
<td>no tRNA</td>
<td>0.10 ± 0.02</td>
</tr>
<tr>
<td></td>
<td>TS(^{-})-EctRNA_{Leu}</td>
<td>0.10 ± 0.02</td>
</tr>
<tr>
<td></td>
<td>OE(^{-})-EctRNA_{Leu}</td>
<td>0.12 ± 0.03</td>
</tr>
<tr>
<td></td>
<td>CatRNA_{Leu}</td>
<td>0.10 ± 0.03</td>
</tr>
<tr>
<td></td>
<td>TS-hctRNA_{Leu}</td>
<td>0.10 ± 0.01</td>
</tr>
<tr>
<td></td>
<td>OE-hctRNA_{Leu}</td>
<td>0.64 ± 0.04</td>
</tr>
<tr>
<td>ScLeuRS-D419A</td>
<td>no tRNA</td>
<td>0.094 ± 0.001</td>
</tr>
<tr>
<td></td>
<td>OE-hctRNA_{Leu}</td>
<td>0.100 ± 0.010</td>
</tr>
</tbody>
</table>

^aThe results are the average of three independent repeats with standard deviations indicated.
^TS, transcribed.
^OE, over-expressed.

**DISCUSSION**

Insertion of Ser or Leu at CUG codons might not be incidental

In *C. albicans* and most other CUG clade species, a mutant tRNA\(^{Ser}\)(CAG) has evolved to decode the Leu CUG codon both as Ser and Leu (28,29). This peculiarity is derived from its combined tRNA\(^{Leu}\) and tRNA\(^{Ser}\) identity elements (38). This mRNA is mainly aminocylated by SerRS and charged by LeuRS to a small extent (29). Both biochemical and structural data have revealed that ambiguity at the single CUG codon of SerRS induces local structural rearrangement, leading to a slightly increased activity (27%) of CaSerRS-Leu\(^{197}\) compared with the wild-type CaSerRS-Ser\(^{197}\) (30). Furthermore, genetic studies showed that increased Leu incorporation across all the CUG codons of *C. albicans* had no visible effect on the growth phenotype but had an impressive impact on cell morphology (39). Therefore, it was proposed that CUG decoding ambiguity has a potential regulatory role in protein structure and/or function (30). CaLeuRS is another crucial player in this genetic code alteration and also contains only one CUG codon at position 919. This site is located at the C-terminal domain of LeuRS, which has been shown to be responsible for binding the variable loop of tRNA\(^{Leu}\) and involved in the aminoacylation activity; however, this
domain is not strictly conserved among archaeal/eukaryotic LeuRSs (Figure 1A and B). Here, we revealed that both CaLeuRS-Leu$^{919}$ and CaLeuRS-Ser$^{919}$ catalyzed Leu activation and aminoacylation, but the former was more active (30%) than the latter, indicating that the conformation of the 919-containing $\alpha_2$9 helix might be finely controlled by the introduction of either Ser or Leu. This phenomenon was also observed in another crucial player in the CUG decoding alteration pathway, CaSerRS (30).

We suggested that insertion of either Ser or Leu at the CUG codon was not random and incidental. The relative amounts of CaLeuRS-Ser$^{919}$/CaLeuRS-Leu$^{919}$ should be strictly regulated by an unidentified but precise molecular mechanism in vivo. Whether the fine balance of CaLeuRS-Ser$^{919}$/CaLeuRS-Leu$^{919}$ is critical for decoding other Leu codons and correlates with the ratio of CaSerRS-Ser$^{197}$/CaSerRS-Leu$^{197}$ requires further investigation.

Yeast LeuRS exhibited a relaxed tRNA recognition capacity

In tRNA aminoacylation, species-specific charging, where a tRNA from one taxonomic domain is not aminoacylated by an aaRS from another, is widespread. This may be as a result of the co-evolution of synthetase/tRNA pairs by the addition of species-specific elements. For instance, human tyrosyl-tRNA synthetase does not recognize bacterial tRNA$^{Tyr}$, and E. coli tyrosyl-tRNA synthetase is unable to charge eukaryotic tRNA$^{Tyr}$ (40), and there is no cross-recognition of E. coli and human tRNA$^{Gly}$ by the respective glycyl-tRNA synthetases (41). Similarly, E. coli isoleucyl-tRNA synthetase is unable to charge eukaryotic tRNA$^{Ile}$ (42). Yeast ArgRS charges E. coli tRNA$^{Arg}$; however, E. coli ArgRS acylates only its cognate E. coli tRNA (43). Human cysteinyl-tRNA synthetase charges bacterial tRNACys, but E. coli cysteinyl-tRNA synthetase is non-functional in aminoacylating human tRNA$^{Cys}$ (44). Here, we showed that both hcLeuRS and EcLeuRS failed to aminoacylate CaTmLeu; however, both CaLeuRS and ScLeuRS readily aminoacylated bacterial, yeast and even human tRNA$^{Leu}$. These results showed that yeast LeuRSs exhibit a more relaxed recognition specificity compared with other LeuRSs. Indeed, CaTmRNA$^{Ser}$ itself harbors only tRNA$^{Leu}$ recognition elements in the anticodon loop with other parts being crucial for SerRS recognition. In addition, G33 is also unfavorable for LeuRS; even in this adverse state, CaLeuRS aminoacylates it in vivo (29). Comparison between transcribed and overexpressed tRNA$^{Leu}$ showed that base modification of tRNA$^{Leu}$ plays an important role in both binding and catalysis.
**CaLeuRS was deficient in tRNA-dependent pre-transfer editing but exhibited efficient post-transfer editing for Nva**

Nva is inherently mis-activated by various LeuRSs to a significant level that requires editing for translational accuracy (9,18–20). With an elevated ratio of Nva to Leu, Nva can escape the safeguarding of EcLeuRS and replace Leu in proteins rich in Leu codons, indicating that Nva-tRNA^{Leu}_{Leu} can escape further checking by the ribosome and pose a direct threat to the accuracy of newly synthesized proteins (24). From the viewpoint of editing, some LeuRSs with degenerated (e.g. hmtLeuRS) or deleted CP1 (e.g. MmLeuRS) domains are exceptional examples, which use alternative pathways (efficient discrimination at the active site) for translational quality control (hmtLeuRS) (26) or do not edit mis-aminoacylation product to produce proteome ambiguity (MmLeuRS) (20). However, all LeuRSs with functional CP1 domains studied so far display tRNA-independent, tRNA-dependent pre-transfer editing and post-transfer editing for Nva. Through inactivation of CP1 or utilization of LeuRS inhibitors, post-transfer editing has been successfully isolated (9,10,18,19,27). Similarly, by mutating a crucial Tyr residue to Asp in EcLeuRS and AaLeuRS, both tRNA-dependent pre-transfer and post-transfer editing are inactivated (10,33). Interestingly, this study identified that CaLeuRS itself is naturally defective in tRNA-dependent pre-transfer editing for Nva. With CtrNA^{Leu}, no tRNA-dependent pre-transfer editing was identified. In contrast, weak tRNA-dependent pre-transfer editing for ABA in the presence of specific tRNA^{Leu} was observed, despite the indication that ABA-editing might not be necessary in vivo based on fine discrimination at the active site. Similarly, ScLeuRS did not mediate tRNA-dependent pre-transfer editing. These results indicate that the capacity for tRNA-dependent pre-transfer editing for Nva has been lost by CaLeuRS (also ScLeuRS), and that ABA is also rarely induced. The reason for this deficiency in tRNA-dependent pre-transfer editing and the pathway by which this deficiency was introduced remains to be elucidated.

Post-transfer editing contributed little or negligibly to the total Nva-editing since addition of any tRNAs in the AMP formation assays did not significantly induce additional AMP. Thus, the produced AMP was mainly derived from tRNA-independent pre-transfer editing. However, the energy-saving post-transfer editing pathway critically controls the accuracy of aminoacylation. Mutation at the conserved Asp^{422} of CaLeuRS led to a LeuRS with abolished post-transfer editing capacity; consequently, Nva-tRNA^{Leu}_{Leu} was synthesized. Similarly, ScLeuRS did not synthesize Ile-tRNA^{Leu}_{Leu}; however, ScLeuRS-D419A readily generated significant amounts of Ile-tRNA^{Leu}_{Leu} (13,18). Using these unique CaLeuRS and ScLeuRS models devoid of tRNA-dependent pre-transfer editing capacity, we concluded that the post-transfer editing pathway is the most economic but efficient editing mechanism for LeuRS. Consistent with other LeuRS models and even other aaRS systems, once post-transfer editing is impaired, the mis-charged tRNA is unavoidably accumulated (9–11,18,19,27,45,46).

Our results also revealed that post-transfer editing by CaLeuRS is not tRNA-species specific, as Nva-tRNA^{Ser}_{Ser} was also a substrate. Indeed, based on the poor discrimination against Nva in the active site, Nva-tRNA^{Ser}_{Ser} is possibly synthesized but should be removed. Otherwise, the CUG codon might be decoded as Ser, Leu and Nva in vivo. It has been proposed that the acceptor end of the tRNA switches from a hairpin conformation to a helical conformation for editing by class I aaRSs, whereas the reverse change in conformation occurs at the acceptor end of the tRNA for editing by class II aaRSs (47). Notably, CtrNA^{Ser} corresponds to a class II SerRS; however, results here showed that CtrNA^{Ser} could switch from a hairpin to a helical conformation for editing by a class I LeuRS.

**Eukaryotic LeuRSs recognized CatRNA^{Ser}**

In addition, we revealed that other eukaryotic LeuRSs could efficiently aminoacylate CatRNA^{Ser}, implying that the introduction or evolution of this type of tRNA in other eukaryotic systems would reprogram or disambiguate the genetic code, leading to proteome chaos. In other words, a specific eukaryotic genetic code could be artificially reprogrammed by expression of this tRNA^{Ser}. Indeed, CatRNA^{Ser} has been shown to be efficiently produced, processed and aminoacylated in *S. cerevisiae*, with its expression triggering a stress response, blocking mating and re-defining the gene expression model of *S. cerevisiae* (48). Notably, archaeal LeuRS is in the same group with eukaryotic LeuRS according to primary or higher structure (14) and only differs at the C-terminal tRNA binding domain, indicating that this domain in eukaryotic LeuRSs is a key element for recognition of CatRNA^{Ser}. This observation is consistent with the structural and biochemical results showing that the C-terminal domain of archaeal LeuRS specifically contacts the variable loop but not the anti-codon loop of archaeal tRNA^{Leu} (16,49). However, the anti-codon loop, which is a key recognition element in both CatRNA^{Ser} (28,29,38) and yeast tRNA^{Leu} (49), is likely to be bound by the C-terminal domain of eukaryotic LeuRS. This proposal requires confirmation from eukaryotic LeuRS-tRNA^{Leu}/tRNA^{Ser} structures.

**Concluding remarks**

Translational machinery of human pathogen *C. albicans* is of particular interest because its CUG codon in the genome is decoded as both Ser and Leu by a unique CatRNA^{Ser} leading to proteome ambiguity (28,29). One of the most crucial components in this decoding process is CaLeuRS, which catalyzes two successive steps, aminoacylation and editing reactions, which together are essential for ensuring high specificity of tRNA charging. In aminoacylation, we showed that Leu isofrom was more active than Ser isofrom of CaLeuRS in charging CatRNA^{Leu}, implying the existence of an in vivo mechanism regulated by balance of CaLeuRS-Leu^{919} and CaLeuRS-Ser^{919}. In addition, as a yeast LeuRS model,
CaLeuRS recognized tRNA\textsuperscript{Leu} from bacteria, yeast and higher eukaryotes. In editing, CaLeuRS efficiently mis-activated non-cognate Nva. One of the most interesting findings was that CaLeuRS lacked tRNA-dependent pre-transfer editing for Nva, which has been well investigated for bacterial and human LeuRSs (10,19). Instead, CaLeuRS prevented insertion of Nva into proteome mainly via post-transfer editing no matter whether Nva has been loaded onto either tRNA\textsuperscript{Leu} or tRNA\textsuperscript{Ser}. Collectively, we further improved our understanding of mechanism and significance of genetic code ambiguity in \textit{C. alibicans} and revealed interesting properties of both aminoacylation and editing reactions by CaLeuRS. Furthermore, the capacity of eukaryotic LeuRSs at aminoacylation CatRNA\textsuperscript{Ser} suggests the possibility of re-constructing proteome of other eukaryotes by simply introducing this unique tRNA\textsuperscript{Ser}.

**SUPPLEMENTARY DATA**

Supplementary Data are available at NAR Online.

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