Remarkable acceleration of a DNA/RNA inter-strand functionality transfer reaction to modify a cytosine residue: the proximity effect via complexation with a metal cation

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ABSTRACT

Modified nucleosides in natural RNA molecules are essential for their functions. Non-natural nucleoside analogues have been introduced into RNA to manipulate its structure and function. We have recently developed a new strategy for the in situ modification of RNA based on the functionality transfer reaction between an oligodeoxynucleotide probe and an RNA substrate. 2′-Deoxy-6-thioguanosine (6-thio-dG) was used as the platform to anchor the transfer group. In this study, a pyridinyl vinyl ketone moiety was newly designed as the transfer group with the expectation that a metal cation would form a chelate complex with the pyridinyl-2-keto group. It was demonstrated that the (E)-pyridinyl vinyl keto group was efficiently and specifically transferred to the 4-amino group of the opposing cytosine in RNA in the presence of NiCl₂ with more than 200-fold accelerated rate compared with the previous system with the use of the diketo transfer group. Detailed mechanistic studies suggested that NiCl₂ forms a bridging complex between the pyridinyl keto moiety and the N7 of the purine residue neighboring the cytosine residue of the RNA substrate to bring the groups in close proximity.

INTRODUCTION

Genome science has advanced rapidly over the past two decades, and the remarkable achievements from this period are best exemplified by the discoveries of the important roles of non-coding DNA and RNA. In particular, knowledge of the post-transcriptional chemical modification of non-coding RNA, a widely observed phenomenon, has been considerably broadened, and more than 100 chemical modifications have been determined (1–8). Of particular interest, a chemical modification of mRNA may alter genetic information via ribonucleic acid (RNA) editing by deamination of cytidine to uridine and adenosine to inosine (9,10). 5-Methylcytosine (11,12) and N₆-methyladenosine (13) have been recently identified in RNA and have attracted attention because of their potential functions in diverse biological processes (14). Endogenous and exogenous chemical entities may also modify RNA and have significant effects, such as inhibition of translation by anti-cancer chemotherapy agents (15) and defective protein synthesis resulting from oxidation with reactive oxygen species (16–18). As advanced sequencing technology has become available, there is an urgent need for chemical tools that are suitable for a variety of investigations on structure and function of RNA. These include site-specific alkylation (19–21), photo-affinity labeling (22,23), site-specific chemical labeling of long RNA molecules (24), chemically modified ribozymes (25) and random acylation of the 2′-hydroxyl group of RNA for profiling (26), among others (27). To attain biological functions through the modification of RNA, it is desirable for a method to be utilized in situ.

Recently, we developed a functionality transfer reaction for the site-specific modification of RNA using an ODN (oligodeoxynucleotide) probe incorporating 6-thio-functionalized-6-thio-2′-deoxyguanosine (1) (Figure 1A). The 2-vinyliden-1,3-diketo moiety was first determined to be a transfer group, and the transfer was accomplished by a sequential reaction of a Michael addition by the 4-amino group of the RNA substrate to bring the groups in close proximity.

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Figure 1. (A) Conceptual illustration of the inter-strand functionality transfer from the ODN probe to the RNA substrate within the hybridized complex. (B) The 4-NH₂ group of cytidine in RNA participates in a Michael addition to the vinyl group of the 2-vinylidene-1,3-diketo moiety, and the following elimination of 6-thio-dG accomplishes an S to N functionality transfer. (C) Design of the pyridinyl vinyl keto group as a new transfer group in anticipation of activation through the formation of a metal chelation complex.

thio-dG (Figure 1B) (28). Subsequently, it was found that the transfer reaction was enhanced with selectively for the 2-amino group of the guanine base at alkaline pH or in the presence of NiCl₂ at neutral pH (29,30). This method was applied to the site-specific labeling of RNA (31) and O₆-methyl guanosine-containing DNA (32). The functionality transfer ODN probe (FT-ODN) is advantageous for RNA modification in that the driving force to initiate the transfer reaction is duplex formation with the target RNA. This feature represents a contrast to other methods that use activation stimuli such as photo-irradiation (21) or oxidation (33). However, it was observed that the 2-vinylidene-1,3-diketo moiety is not sufficiently stable or reactive as a transfer group for the modification of the cytosine residue. In this study, to meet the conflicting requirements of stability and reactivity, a pyridinyl vinyl ketone moiety (Pyk) was designed as a new transfer group with the anticipation that activation would occur through complexation with a metal cation, as illustrated by 3. This activation mechanism resembles the activation of DNA alkylating agents that have a quinolin-8(5H)-one substructure (34–36). This FT-ODN functionalized with an S₆-vinyl pyridinyl ketone moiety showed higher stability in a buffer solution and also demonstrated, in the presence of NiCl₂, a remarkably efficient transfer reaction with high selectivity for the 4-amino group of the opposing cytidine in RNA with more than 200-fold accelerated rate. A detailed mechanistic study revealed that the transfer reaction occurred specifically with (E)-geometry of the thiovinyl moiety and that NiCl₂ effectively brings the two reactants in close proximity by forming a bridging complex between the FT-ODN and N7 of the purine residue neighboring the cytidine of the target RNA. 

In this manuscript, we describe in detail the design, synthesis and evaluation of the transfer reaction, as well as a mechanistic study on the effect of metal complexation.

MATERIALS AND METHODS

Reagents and solvents were purchased from commercial suppliers and were used without purification. ¹H-, ¹³C-NMR spectra were recorded on Bruker Avance-III. 2D-NMR spectra were measured on Varian Inova 500. IR spectra were recorded on Perkin Elmer Spectrum One FT-IR spectrometer. ESI-HRMS spectra were measured on Applied Biosystems Mariner Biospectrometry Workstation using neuropeptides, angiotensin I, bradykinin and picolinic acid as internal standards. MALDI-TOFMS spectra were recorded on BRUKER DALTONICS microflex-KS Linear mass spectrometer.

Synthesis of the precursors for pyridinyl vinyl keto transfer groups (5–7)

1-(Pyridin-2-yl)prop-2-yn-1-one (5). A solution of ethynylmagnesium bromide (0.5 M in THF, 20 mL, 10.0 mmol) was added into a solution of 2-cyanopyridine (517 mg, 4.97 mmol) in dry THF (tetrahydrofuran) (3 ml) under argon atmosphere at 0°C. The mixture was warmed to room temperature during 45 min, and again cooled at 0°C. The reaction mixture was quenched with 1% aqueous H₂SO₄ and then neutralized with saturated aqueous NaHCO₃. Precipitates were filtered off through a Celite pad, and the filtrate was extracted with AcOEt. The organic layers were washed with brine, dried over Na₂SO₄ and evaporated.

Chart 1. The precursors used for attaching pyridinyl keto transfer groups to 6-thio-2′-deoxyguanosine in ODN probe.

| ODN1: | 5′ CTTC-⁵G-TTCCTTTTCT |
| FT-ODN1: | 5′ CTTC-(Pyk)⁵G-TTCCTTTTCT |
| ODN2: | 5′ CTTC-⁵G-M²TTCCTTTTCT |
| M² = dA, dG, dC, T |
| FT-ODN2: | 5′ CTTC-⁵G-H⁵TTCCTTTTCT |
| M³ = dA, dG, dC, T |
| RNA1(X): | 5′ agaaagagaa-C-X-aaag |
| X = rA, rG, rC, U |
| RNA2(X): | 5′ agaaagagag⁻²-c-H²aaag |
| N², N³ = rA, rG, rC, U, 7-deaza-rG |

Chart 2. Sequences of ODNs and RNAs used in this study.
The residue was chromatographed on a silica gel column (KANTO 60N, 10 g. Hex-AcOEt = 5:1. v/v) to give 5 as colorless needles (45 mg) in 7% yield. mp: 97–99°C (lit (37) 100–105°C, decomposition). IR (solid) cm⁻¹: 3149, 2088, 1653, 1584. ¹H-NMR (500 MHz, CDCl₃) δ ppm: 8.76 (1H, ddd, J = 4.7, 1.7, 1.0 Hz), 8.07 (1H, ddd, J = 7.8, 1.2, 1.0 Hz), 7.95 (1H, ddd, J = 7.8, 1.7, 1.7 Hz), 7.63 (1H, ddd, J = 7.7, 4.7, 1.2 Hz), 3.97 (1H, s). ¹³C-NMR (125 MHz, CDCl₃) δ ppm: 179.0, 153.5, 150.8, 138.4, 129.3, 123.8, 84.7, 81.3. ESI-HRMS (m/z): calcd for C₉H₇INO⁺ [M+H]+, 259.9567; found 259.9556.

1. A solution of ethynylmagnesium bromide (0.5 M in 780 mg, 38%) and 5-ethynylpicolinonitrile (38) (80 mg, 0.625 mmol) in THF (1.5 mL) at 0°C under argon atmosphere at 0°C. The mixture was warmed to room temperature for 30 min, and then again cooled at 0°C. The reaction mixture was quenched with 10% aqueous HI and stirred for 10 min, then neutralized with saturated aqueous NaHCO₃. Precipitates were filtered off through a Celite pad and the filtrate was extracted with AcOEt. The organic layer was washed with brine, dried over Na₂SO₄ and evaporated. The residue was chromatographed on a silica gel column (KANTO 60 N, 50 g, Hex-AcOEt = 5:1, v/v) to give 6 (780 mg, 38%) and 7 (69 mg, 3%) as a viscous oil.

(E)‑6: IR (film) cm⁻¹: 1673, 1559, 1310, 1285, 1229. ¹H-NMR (500 MHz, CDCl₃) δ ppm: 8.71 (1H, ddd, J = 4.8, 1.7, 1.0 Hz), 8.57 (1H, d, J = 15.0 Hz), 8.18 (1H, d, J = 15.0 Hz), 8.06 (1H, ddd, J = 7.8, 1.2, 1.0 Hz), 7.94 (1H, ddd, J = 7.8, 7.6, 1.7 Hz), 7.58 (1H, ddd, J = 7.6, 4.8, 1.2 Hz). ¹³C-NMR (125 MHz, CDCl₃) δ ppm: 187.6, 153.3, 150.2, 141.3, 138.5 128.6, 123.9, 101.8. ESI-HRMS (m/z): calcd for C₉H₇INO⁺ [M+H]+, 259.9567; found 259.9556.

(Z)‑7: IR (film) cm⁻¹: 1678, 1569, 1325, 1266, 1230. ¹H-NMR (500 MHz, CDCl₃) δ ppm: 8.70 (1H, ddd, J = 4.8, 1.7, 1.0 Hz), 8.62 (1H, d, J = 8.8), 8.10 (1H, d, J = 7.8, 1.0, 1.2 Hz), 7.96 (1H, ddd, J = 7.8, 7.6, 1.7 Hz), 7.70 (1H, d, J = 8.8 Hz), 7.58 (1H, ddd, J = 7.6, 4.8, 1.2 Hz). ¹³C-NMR (125 MHz, CDCl₃) δ ppm: 190.0, 154.1, 150.0, 138.5, 133.0, 128.7, 123.6, 95.6. ESI-HRMS (m/z): calcd for C₉H₇INO⁺ [M+H]+, 259.9567; found 259.9556.

(E)‑I-(5-Ethynylpyridin-2-yl)-3-iodoprop-2-en-1-one (8). A solution of ethynylmagnesium bromide in THF (0.5 M solution, 2.5 mL, 1.25 mmol) was added into a solution of 5-ethynylpicolinonitrile (38) (80 mg, 0.625 mmol) in THF (1.5 mL) at 0°C under argon atmosphere. After stirring at room temperature for 1 h 10 min, 20% aqueous HI (13 mL) were added to the mixture at room temperature, and then the mixture was stirred for an additional 10 min at the same temperature. Saturated aqueous NaHCO₃ (20 mL) was added to the reaction mixture and the inorganic precipitate was removed by filtration through a Celite pad. The filtrate was extracted with AcOEt (30 mL×2). The combined organic layers were washed with brine (30 mL), dried over Na₂SO₄, filtered and concentrated under reduced pressure to give a crude product, which was purified by flash column chromatography (FUJI SYLISIA FL60D, 15 g, Hex-AcOEt = 20:1, v/v) to give 8 as a yellow foam (17 mg, 0.0608 mmol, 10%). IR (film) cm⁻¹: 3238, 3068, 2927, 2108, 1662, 1581, 1559, 1552, 1371, 1316, 1201, 1016.

A general procedure for preparation of the functionality transfer reaction (FT-ODN). The ODNs incorporating 6-thio-2'-deoxyguanosine (6-thio-dG) and determination of the (E)- and (Z)-isomer ratio by ¹H-NMR. Pyridinyl keto derivatives (5–7) were reacted with TBS-protected 6-thio-dG (9) (37) in MeOH in the presence of TEA at room temperature (Scheme 2). The ratio of the (E)- and (Z)-isomers of the S²-vinylated product (10 or 11) was estimated by the integration of the vinyl protons in the NMR spectrum (Supplementary Figure S2).
modified RNA1 was analyzed by MS/MS to confirm that the target rC was modified by the transfer reaction (Supplementary Figure S4). To determine the structure of the modified cytidine, the corresponding DNA substrate was used. The modified DNA substrate was purified, reduced with NaBH4, HPLC purification and hydrolyzed with bacterial alkaline phosphatase (BAP), nuclease P1 and venom phosphodiesterase (VPDE). The lysates were analyzed by HPLC (Supplementary Figure S5). The authentic sample was synthesized separately (Supplementary Scheme S3) and used for confirmation by HPLC co-injection.

Kinetic analysis of the functionality transfer reaction using RNA1 (rC) and (E)-FT-ODN1. The reaction was performed at 15, 20, 25, 30 and 35°C with different concentrations of NiCl2 (0, 1, 2, 3, 4, 5, 15 μM), and the modified RNA was quantitatively analyzed by HPLC. The reaction was analyzed as a first-order reaction using an initial duplex concentration of 4.5 μM as the reactive duplex formed with (E)-FT-ODN1, and the remaining concentration (0.5 μM) as the nonreactive duplex formed with (Z)-FT-ODN1. The activation energy (Ea) was obtained by Arrhenius plots, and kinetic parameters were obtained from the Eyring equation.

Molecular modeling of the complex of 5'-functionalized ODN, target RNA and NiCl2. The OPLS 2005 parameters installed in MacroModel were modified to calculate the NiCl2 complex in water using the model complex structure optimized by ub3lyp/6-31G(d). An octahedral NiCl2 complex was assumed to coordinate with the nitrogen and oxygen atoms of the pyridyl keto transfer moiety, the N7 atom of adenine or guanine and one water molecule. During optimization, the NiCl2 complex structure was constrained by setting bond lengths within a range of values. Therefore, the simulated structures illustrate only candidate complexes leading to the transfer reaction.

RESULTS AND DISCUSSION
Selective synthesis of the 6-thio-dG monomer functionalized with the (E)- or (Z)-pyridinyl vinyl keto transfer moiety
Pyridinyl keto transfer groups (Pyk, 5-8) were used to modify the ODN probe incorporating 6-thio-2'-deoxyguanosine. Ethynyl derivative 5 was synthesized using 2-cyanopyridine and ethynyl magnesium bromide. A mixture of (E)- and (Z)-iodovinyl derivatives (6-7) were obtained by an analogous reaction, except that the reaction mixture was treated with aqueous HI solution and each isomer was separated by column chromatography. The 5-ethynylpyridine derivative 8 was synthesized in a similar fashion from 2-cyano-5-ethynylpyridine, which was prepared from the corresponding 5-bromo derivative. In a preliminary study, ODN1 containing 6-thio-dG was functionalized using the ethynyl keto pyridine derivative (5) to prepare FT-ODN1, and it was tested for the transfer reaction with RNA1 (Scheme 1). The reaction proceeded rapidly, but the formation of the modified RNA reached a plateau of ~30% yield within 1 h (Supplementary Figure S1). The (E)- and (Z)-stereochemistry of the transfer unit was suspected to influence reactivity; however, determination of the E/Z ratio of the synthesized FT-ODN1 was not possible by HPLC or gel electrophoresis. The 1H-NMR signals of the vinyl keto pyridine moiety are in the same region of nucleobases. Therefore, the E/Z ratio was measured in a model reaction using a 6-thio-dG nucleoside in organic solvents (Scheme 2). (E)- and (Z)-iodovinyl derivatives (6 and 7, respectively) were purified and then reacted with TBS-protected 6-thio-dG (9) in methanol in the presence of triethylamine (Scheme 2). The (E)- and (Z)-ratio (10 / 11 ratio) was determined by 1H-NMR (Supplementary Figure S2). The ethynylketopyridine derivative (5) that was used to prepare FT-ODN1 in a preliminary study produced 10 and 11 in a 56:44 ratio. The pure (E)-isomer (6) gave 10 and 11 in a 92:8 ratio with selectivity for the (E)-isomer; however, the reaction with the pure (Z)-isomer (7) resulted in only a 35:65 ratio in favor of the (Z)-isomer.

The functionality transfer reaction
ODN1 containing 6-thio-dG was functionalized with 6 to produce (E)-FT-ODN1 as the sole product (Figure 2A). Assuming that the modification of 6-thio-dG in DNA in aqueous solution occurs in a similar fashion as in organic solvents, FT-ODN1 was thought to be prepared in a ratio of ~92:8 in favor of the (E)-isomer and is thus identified as (E)-FT-ODN1. Alternatively, FT-ODN1 was assumed to form in a 35:65 ratio in favor of the (Z)-isomer after reaction with 7 and is represented as (Z)-FT-ODN1. These isomers were used without separation for the transfer reaction.

The transfer reaction shown in Scheme 1 was performed at 37°C using 5 μM of RNA1 and 7.5 μM of FT-ODN1 in 50 mM HEPES buffer containing 100 mM NaCl at pH 7, and progress was analyzed by HPLC. Figure 2B shows an example of an HPLC trace revealing a 55% transfer yield for the reaction between RNA1(rC) and (E)-FT-ODN1. The yields of modified RNA1 obtained under dif-

**Scheme 1.** The functionality transfer reaction from FT-ODN1 to RNA1 (rC).

**Scheme 2.** Model experiments to determine (E)- and (Z)-stereochemistry of the transfer unit of 6-thio-dG.
different conditions are plotted against time in Figure 3A. In the presence of 5 μM NiCl2, the transfer reaction with (E)-FT-ODN1 proceeded rapidly and was completed before 10 min to produce modified RNA2 in ~90% yield (solid line in Figure 3A). The transfer yield at 10 min was improved from ca. 1% to 90% compared with the previous system with the use of the diketo transfer group,28 meaning more than 200-fold rate acceleration. Remarkably, the reaction yield with (Z)-FT-ODN1 only reached a plateau of ~40% yield in contrast (dotted line in Figure 3A). Assuming that the (E)-isomer represents ~92% of (E)-FT-ODN1 and 35% of (Z)-FT-ODN1, these results can be interpreted as a result of the high reactivity of the (E)-isomer and the lack of isomerization from the Z-isomer to the E-isomer during the transfer reaction. It should also be noted that addition of NiCl2 significantly increased the reaction rate for (E)-FT-ODN1 (continuous line versus dashed line in Figure 3A). Figure 3B summarizes the transfer yields with (E)-FT-ODN1 at 10 min. EDTA completely inhibited the reaction, most likely by trapping metal cations in the buffer, suggesting that metal cations in the buffer also contribute to activation to some extent. A high selectivity for rC was clearly shown in Figure 3B. The site-specific labeling of rC in RNA has been demonstrated by using an acetylene-functionalized pyridinyl keto unit (8) for a Cu-catalyzed click reaction with azido derivatives (Biotin-N3 and FAM-N3) (31) (Supplementary Scheme S2 and Supplementary Figure S3).

A comparison of the effect of other metal cations on activation is displayed in Figure 4A, showing an order of Ni²⁺ > Co²⁺ > Cu²⁺ > Zn²⁺ > Ca²⁺, Mg²⁺, Mn²⁺, and Fe²⁺. The rate enhancement by NiCl2 was dependent on the concentration, and a maximum was reached at a concentration equimolar to the RNA substrate (Figure 4B inset). Thus, the initial design in which the pyridin vinyl keto unit is activated by forming a chelate complex with NiCl2 appeared to be validated by these results. In contradiction to the observed activation by NiCl2, the degradation half-life of (E)-FT-ODN1 in the buffer (10–12 h) was not affected by NiCl2; i.e. the intrinsic reactivity of (E)-FT-ODN1 was not increased by NiCl2. The results posed two questions: the difference in reactivity of the (E)-or (Z)-isomers in the transfer group, and the mechanism for activation by NiCl2.

Mechanistic study of the activation of the transfer reaction by NiCl2

As part of a mechanistic study, the modification site was determined to be the cytosine residue by UPLC/MS/MS (Supplementary Figures S4). The N4-alkylation of rC was confirmed by HPLC analysis of the hydrolysates of the modified product using the DNA substrate corresponding to RNA1 (Supplementary Figures S5 and S6). To obtain further insight into the effect of NiCl2 on kinetics, the reaction was performed at different temperatures in the presence of different concentrations of NiCl2. As the transfer reaction occurs through an intra-complex interaction, the rate constants were obtained by analyzing the reaction as first-order. The Ea values were obtained by Arrhenius plots, and kinetic parameters were calculated from the Eyring equation, and are expressed in a bar graph (Figure 5).

The ΔG‡ value decreased with increasing NiCl2 concentration in accord with the fact that NiCl2 increased the transfer yield (Figure 4B). The entropy of activation (−TΔS‡) was a major component of the ΔG‡ value in the
absence of NiCl₂ (NiCl₂, 0 μM). Interestingly, although the transfer reaction was enhanced by NiCl₂, the enthalpy of activation (ΔH°) conversely increased (see also Supplementary Figures S7 and Table S2). The −TΔS° value decreased significantly in the presence of NiCl₂, with an accompanying increase of the ΔH° value. At a high concentration of NiCl₂, the −TΔS° value was substantially diminished, and the ΔH° value became a major component of the ΔG° value. In spite of increases in the ΔH° value, larger decreases of the −TΔS° value reduced the ΔG° value, favorably affecting the reaction. These kinetic data accord with the conflicting properties of (E)-FT-DON1 in that NiCl₂ increased the reaction rate of the transfer moiety without affecting the intrinsic reactivity of the transfer moiety in the buffer. Considering that the functionality transfer reaction proceeds through a Michael addition of the 4-NH₂ group and subsequent β-elimination of the 6-thio-dG residue (Scheme 3), a large −TΔS° value for the reaction in the absence of NiCl₂ suggests that the 1,4-addition step is most likely a rate-determining step. On the other hand, the small −TΔS° value and large ΔH° value for the reaction at high NiCl₂ concentrations may be an indication that the bond-breaking β-elimination is a rate-determining step.

To satisfy these interpretations, we noted that N7 of the purine residue is a binding site for NiCl₂ in addition to the pyridinyl keto unit (40). NiCl₂ forms a particularly stable complex with the N7 of guanine in an extra-helical or terminal position (41). Thus, it was supposed that Ni²⁺ forms complexes with both the pyridinyl ketone of (E)-FT-DON1 and N7 of the purine residue, either at the 5′ or 3′ side of the opposing rC of the RNA1 substrate. Such a complex may form an inter-strand bridge between (E)-FT-ONNI and RNA1 and bring the 4-NH₂ group of rC in RNA1 in proximity with the thio-vinyl reactive site as shown in Supplementary Figure 6A and 6B. Molecular modeling also suggested that such a Ni²⁺-bridged complex formation with (Z)-FT-DON1 requires distortion of the duplex structure (Supplementary Figure S8). Thus, by predicting a bridging complex with NiCl₂ as shown in Figure 6, it may be reasonably explained why only the (E)-transfer group is highly reactive and how NiCl₂ activates the transfer reaction without decreasing the stability of FT-ODN. To confirm a significant role of the purine residue, we next investigated effects of base pairs flanking the target rC. The results are summarized in Figure 7A. Flanking bases composed of rA and rG at both 5′ and 3′ sides were the most effective. Efficiency was retained with rG at the 3′ side (N3′-N3 = UG and CG) or rA at the 3′ side (N3′-N3 = UA and CA). In contrast, the transfer reaction remarkably did not occur when the flanking bases were composed of U and rC at both 5′ and 3′ sides (N5-N5 = UU, UC, CU, CC). 7-Deaza-rG (deG) was used as a control residue, as it lacks an N7 binding site for NiCl₂. Replacement of rG with deG diminished the transfer reaction (Figure 7B), clearly underscoring the importance of N7. These results have strongly suggested that the formation of a bridging complex with NiCl₂, N7 of the purine residue and the (E)-pyridinyl keto unit as shown in Figure 6 is responsible for activation by NiCl₂.

In conclusion, the highly efficient and site-specific modification of rC in RNA has been successfully demonstrated by a functionality transfer reaction using the (E)-pyridinyl vinyl keto transfer group. A remarkable acceleration in the transfer reaction is achieved by the enforced proximity effect mediated by a bridged complex formed between NiCl₂, the (E)-pyridinyl vinyl keto moiety and N7 of purine residues flanking the opposing rC. The transfer rate of the (E)-pyridinyl vinyl keto moiety has exhibited more than 200-fold increased reaction rate compared with the diketo transfer group. Notably, NiCl₂ enhances the transfer reaction without increasing the intrinsic reactivity of the transfer group that is ascribable to the instability of the ODN probe, and a variety of organic functional groups can be

![Scheme 3](Image 1)

1,4-Michael addition and following β-elimination leading to S- to N transfer reaction.

![Figure 6](Image 2)

(A) A hypothetical complex of Ni²⁺ with the pyridinylketo unit and N7 of an adenine residue at the 5′ side. (B) Molecular modeling of the complex in the ODN1/RNA1 duplex.
easily attached on the specific site of RNA under mild conditions. With regard to its application, it is interesting to investigate the effect on deamination activity and template effect by the specific modification of a single cytidine such as in the large apolipoprotein B mRNA. The search of compounds that may have an impact on the structure and function of RNA is now ongoing.

SUPPLEMENTARY DATA
Supplementary Data are available at NAR online.

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REFERENCES


