

Hematopoietic stem cell and progenitor defects in *Sca-1*/*Ly-6A*-null mice

Caryn Y. Ito, Carol Y. J. Li, Alan Bernstein, John E. Dick, and William L. Stanford

Despite its wide use as a marker for hematopoietic stem cells (HSCs), the function of stem cell antigen-1 (*Sca-1*) (also known as lymphocyte activation protein-6A [*Ly-6A*]) in hematopoiesis remains poorly defined. We have previously established that *Sca-1*^{-/-} T cells develop normally, although they are hyperresponsive to antigen. Here, we report detailed analysis of hematopoiesis in *Sca-1*-deficient animals. The differentiation potential of *Sca-1*-null bone marrow was determined from examination of the most mature precursors (culture colony-forming units [CFU-Cs]) to less committed progenitors (spleen CFUs [CFU-Ss]) to long-term repopulating HSCs. *Sca-1*-null

mice are mildly thrombocytopenic with a concomitant decrease in megakaryocytes and their precursors. Bone marrow cells derived from *Sca-1*^{-/-} mice also have decreased multipotential granulocyte, erythroid, macrophage, and megakaryocyte CFU (GEMM-CFU) and CFU-S progenitor activity. Competitive repopulation assays demonstrated that *Sca-1*^{-/-} HSCs are at a competitive disadvantage compared with wild-type HSCs. To further analyze the potential of *Sca-1*^{-/-} HSCs, serial transplantations were performed. While secondary repopulations using wild-type bone marrow completely repopulated *Sca-1*^{-/-} mice, *Sca-1*^{-/-} bone marrow failed to rescue one third of lethally irradiated wild-

type mice receiving secondary bone marrow transplants from irradiation-induced anemia and contributed poorly to the surviving transplant recipients. These data strongly suggest that *Sca-1* is required for regulating HSC self-renewal and the development of committed progenitor cells, megakaryocytes, and platelets. Thus, our studies conclusively demonstrate that *Sca-1*, in addition to being a marker of HSCs, regulates the developmental program of HSCs and specific progenitor populations. (*Blood*. 2003; 101:517-523)

© 2003 by The American Society of Hematology

Introduction

Hematopoiesis proceeds through the differentiation of stem cells into a cascade of committed progenitors and, finally, into all of the terminally differentiated cell lineages of the blood system. At least in C57BL/6 and other *Ly-6.2* mouse strains, the cell surface phenotype of the hematopoietic stem cells (HSCs) and primitive progenitors has been well elucidated. One of the markers whose expression defines the HSCs and various developmental intermediates is the glycosyl phosphatidylinositol-anchored protein (GPI-AP) stem cell antigen-1 (*Sca-1*). Single lineage-negative (*Lin*⁻) *Thy-1*^{lo}*Sca-1*⁺*Kit*⁺*CD38*⁺*CD34*⁻ cells fulfill the criteria of HSCs, as they are unable to give rise to short-term repopulating cells but are able to provide long-term repopulation of lethally irradiated mice, including secondary and tertiary transplantations.^{1,2} *Sca-1* is regulated in a complex fashion during hematopoietic differentiation. As HSCs differentiate into common lymphoid progenitors (CLPs) that give rise to T, B, and natural killer (NK) cells but not myeloid lineages, *Sca-1* and *Kit* expression decreases while interleukin 7-receptor (*IL-7R*) expression increases.³ *Sca-1* and another *Ly-6* gene family member, *Sca-2*, are up-regulated on prothymocytes, which seed the thymic cortex.⁴ *Sca-1* expression is turned off at an early stage of thymocyte differentiation, and *Sca-1*

is re-expressed by mature single-positive medullary thymocytes and peripheral T cells.⁵ Transgenic overexpression of *Sca-1* during all stages of thymocyte development arrests development at the *CD3*⁻*CD4*⁻*CD8*⁻*CD44*⁺*CD25*⁺ stage, the stage in the thymic cortex at which *Sca-1* expression normally ceases.⁵ Concurrently with differentiation of the HSCs into common myeloid progenitors (CMPs), *Sca-1* expression is down-regulated.⁶ *Sca-1* expression is activated in a proportion of spleen colony-forming unit (CFU-S) and culture-CFU (CFU-C) progenitors.⁷

All functional analysis of *Sca-1* has been performed in primary lymphocytes or lymphoid cell lines. The *Sca-1* protein (or lymphocyte activation protein-6A [*Ly-6A*]) was originally identified as an antigen up-regulated on activated lymphocytes.⁸ GPI-APs are believed to facilitate cell signaling by creating glycosphingolipid cholesterol "rafts" that concentrate or exclude specific signaling molecules.^{9,10} Consistent with this mechanism, *Src* family kinase members have been shown to associate with *Sca-1*.¹¹ Several members of the *Ly-6* gene family, which share similar structural motifs as well as primary sequence, have identified cognate ligands, such as the *Ly-6* protein *CD59*, which protects cells from complement-mediated lysis by binding the α -chain of complement

From the Programme in Cancer/Blood, Hospital for Sick Children, Toronto, ON, Canada; the Programme in Development and Fetal Health and the Programme in Molecular Biology and Cancer, Samuel Lunenfeld Research Institute, Mount Sinai Hospital, Toronto, ON, Canada; and the Department of Molecular and Medical Genetics and the Institute of Biomaterials and Biomedical Engineering, University of Toronto, ON, Canada.

Submitted June 20, 2002; accepted August 14, 2002. Prepublished online as *Blood* First Edition Paper, August 29, 2002; DOI 10.1182/blood-2002-06-1918.

Supported by grants from the Canadian Institutes of Health Research, Ottawa, ON, Canada; the Leukemia and Lymphoma Society, New York, NY; and the Terry Fox Foundation, Vancouver, BC, Canada. C.Y.I. was supported by a

National Research Service Award (NRSA) training fellowship (National Institutes of Health [NIH]); and W.L.S. is the Karyn Glick Memorial Special Fellow of the Leukemia and Lymphoma Society of America and the Canadian Research Chair in Stem Cell Biology and Functional Genomics.

Reprints: William L. Stanford, Institute of Biomaterials and Biomedical Engineering, University of Toronto, 4 Taddle Creek Rd, Rm 407 Rosebrugh Bldg, Toronto, ON M5S 3G9 Canada; e-mail: william.stanford@utoronto.ca.

The publication costs of this article were defrayed in part by page charge payment. Therefore, and solely to indicate this fact, this article is hereby marked "advertisement" in accordance with 18 U.S.C. section 1734.

© 2003 by The American Society of Hematology

8 and the "b" domain of complement 9 (Ninomiya and Sims¹²). CD59 does not simply act to block the complement cascade, but induces a signaling cascade, which includes activation of the Src family kinase protein Hck, the adapter Src homology and collagen protein (Shc), and Syk following ligand binding.¹³ Thus, it has long been believed that Sca-1 function is dependent upon interaction with its cognate ligand. Consistent with the expectation that Sca-1 functions as a receptor, an antibody to a 66-kDa GPI-AP immunoprecipitates Sca-1¹⁴; however, the identity of the purported 66-kDa ligand has yet to be published.

The absence of an identified ligand has made determination of the role of Sca-1 in cell signaling difficult. Simulation of ligand binding by cross-linking Sca-1 with anti-Sca-1 antibodies has generated conflicting results.¹⁵⁻¹⁸ However, antibody cross-linking assays are artificial and probably do not represent physiologic ligand-receptor interactions. Therefore, we undertook a genetic approach to determine the function of Sca-1.¹⁹ Homologous recombination was used to ablate the Sca-1 gene in 129 ES cells. Homozygous mutant mice were produced at normal Mendelian frequencies, demonstrating that Sca-1 is not required for development. However, T cells from Sca-1-deficient mice demonstrate significantly higher and more prolonged proliferation in response to stimulation through the T-cell-receptor complex as compared with T cells from *Sca-1*^{+/+} littermates. Although the mechanism driving hyperproliferation has not been determined, increased cytokine production, skewed thymocyte development, and overall expression of Src family kinases have been ruled out (Stanford et al,¹⁹ and W.L.S., unpublished observations, 1998). Consistent with these results, overexpression of Sca-1 in T cells inhibits CD4⁺ T-cell proliferation induced by T-cell-receptor activation.²⁰ Together, these results suggest that Sca-1 acts to down-modulate an active immune response. Furthermore, we have recently demonstrated that Sca-1 is required for the immunosuppressive function of CD4⁺CD8⁻ T cells and the induction of major histocompatibility complex (MHC)-mismatched transplantation tolerance.²¹

To analyze the developmental potential of *Sca-1*^{-/-} HSCs and progenitors, we generated congenic Sca-1-null mice on the C57BL/6 (Ly-6.2) background. HSCs from *Sca-1*^{-/-} mice had impaired repopulation potential, as demonstrated by a competitive disadvantage compared with wild-type HSCs. Moreover, the lower engraftment of secondary transplants suggests a defect in *Sca-1*^{-/-} HSC self-renewal. In addition to HSC deficiencies in Sca-1-deficient mice, specific cell lineages and progenitor subpopulations were also affected. The fact that not all lineages were equally compromised suggests that Sca-1 performs complex and specific functions in hematopoiesis.

Materials and methods

Animals

All mice were maintained in a conventional mouse facility at Mount Sinai Hospital (Toronto, ON, Canada). For all primary assays, 8- to 12-week-old mice were used. The C57BL/6J-*Kit*-white-spotted viable (*Kit*^{W^v}) (*W*^v/+) and C57BL/6J-*Gpi1*^{aa} mice were obtained from The Jackson Laboratory (Bar Harbor, ME) and bred in-house. *W*^v/+ genotyping was inferred by coat-color analysis or polymerase chain reaction (PCR) analysis. *Sca-1*^{+/+} mice (discussed in "Results") were used as controls for all experiments except competitive and secondary repopulations, in which C57BL/6J-*Gpi1*^{aa} mice were used. Clonogenic assays demonstrated no differences between *Sca-1*^{+/+} and C57BL/6J-*Gpi1*^{aa} control mice.

Antibodies

Purified rat antimouse monoclonal antibodies (mAbs) for flow cytometry were all purchased from BD PharMingen (San Diego, CA). The following mAbs, conjugated with either fluorescein isothiocyanate (FITC) or phycoerythrin (PE), were used: anti-CD3ε, anti-macrophage adhesion molecule-1 (anti-MAC-1) (CD11b), anti-CD41 (integrin αIIb), anti-B220 (CD45R), anti-CD61 (integrin β3), anti-Thy-1, anti-Kit, anti-IL-7Rα, anti-Sca-1, and anti-TER-119. In some experiments, allophycocyanin-conjugated anti-Gr-1 (Ly-6G) and anti-Kit were used. The appropriate conjugated rat antimouse mAbs were used as negative controls.

Cytokines

Purified thrombopoietin (recombinant human thrombopoietin [rhTPO]), rhIL-6, and rhIL-11 were purchased from PeproTech (Rocky Hill, NJ). Purified rhIL-7 was purchased from Stem Cell Technologies (Vancouver, BC, Canada), and recombinant mouse IL-3 (mIL-3) was purchased from BD PharMingen.

Flow cytometry

Single cell suspensions were prepared from bone marrow (tibial and femoral). The bone marrow was subjected to red blood cell (RBC) lysis with ammonium chloride, washed once in Iscove Dulbecco modified Eagle medium (DMEM), counted, and resuspended in phosphate-buffered saline (PBS) plus 2% fetal bovine serum (FBS) (staining medium [SM]) at 5 × 10⁶ cells per milliliter. Then, 10⁶ cells were incubated with mAbs for 30 minutes, washed twice in SM, and resuspended in 0.2 mL SM for analysis by means of a Becton Dickinson FACScalibur (Mountain View, CA).

Glucose phosphate isomerase 1 (Gpi1) isoenzyme analysis

The samples were subjected to electrophoresis on cellulose acetate plates (Helena Laboratories, Beaumont, TX), and the Gpi1 isoenzyme bands were visualized as previously described.²² The contribution of mutant cells to the samples was visually estimated by comparing them with a set of standard mixtures of Gpi1-AA and Gpi1-BB cells.

Hematologic analysis

Peripheral blood (50 to 100 μL) from tail bleeds was collected into EDTA (ethylenediaminetetraacetic acid)-coated capillary pipettes (Drummond, Broomall, PA) and transferred to Eppendorf (Hamburg, Germany) tubes. Complete blood counts and differential counts were performed by means of a Coulter Ac-T Differential Hematology Analyzer with veterinary software (Beckman-Coulter, Fullerton, CA).

Bleeding time assay

Six- to 8-week-old mice were anesthetized with the use of 0.15 mg ketamine per gram body weight to reduce influences of heart rate and activity on bleeding time. Approximately a 3-mm portion of the tail was cleanly severed with the use of scissors. The tail was immersed in physiologic saline at 37°C. The time required for the small stream of blood to stop was recorded as the bleeding time.

Culture CFUs

For each experiment, 3 animals of each genotype were used. Each experiment was performed between 3 and 8 times. Single cell suspensions were prepared from bone marrow (tibial and femoral). Cells were counted and diluted to 10⁶ cells per milliliter in Iscove modified Dulbecco medium (IMDM) with 1% FBS. For each assay, 3 aliquots of 10⁴ cells were mixed with the appropriate methylcellulose media, plated, and grown in humidified chambers at 37°C, 5% CO₂. To assay megakaryocyte CFU precursors (CFU-Mk), cells were grown in methylcellulose/collagen medium (Stem Cell Technologies) containing 50 ng/mL rhTPO, 10 ng/mL mIL-3, 20 ng/mL rhIL-6, and 50 ng/mL rhIL-11, and visualized by acetylcholinesterase activity. All other CFU-C colonies were grown in methylcellulose containing IL-3, IL-6, Steel factor (SLF), and erythropoietin (EPO)

Table 1. Peripheral blood analysis of wild-type and *Sca-1*-null mice

Hematologic parameter	<i>Sca-1</i> ^{+/+}	<i>Sca-1</i> ^{-/-}
WBC count, × 10 ³ /μL	12.5 ± 0.2	12.2 ± 0.1
RBC count, × 10 ⁶ /μL	11.1 ± 0.1	11.1 ± 0.1
Plt count, × 10 ³ /μL	1021.2 ± 25.2*	615.6 ± 18.5*
Hgb level, g/dL	17.4 ± 0.1	17.6 ± 0.1
Hct, %	54.6 ± 0.2	54.2 ± 0.3
Percentage of WBC count		
Lymphocytes, %	90.57 ± 0.14†	92.20 ± 0.33†
Monocytes, %	6.65 ± 0.47†	5.69 ± 0.36†
Granulocytes, %	2.78 ± 0.16†	2.11 ± 0.23†

Analysis was performed on 12-week-old mice, 15 for each genotype. The peripheral blood values were determined by means of a Becton-Dickinson veterinary hematologic analyzer.

WBCs indicates white blood cells; RBCs, red blood cells; Plts, platelets; and Hgb, hemoglobin.

*Highly significant differences; $P < .01$.

†Significant differences; $P < .05$.

(M3434; Stem Cell Technologies). Erythroid CFU (CFU-E) precursors were assayed after 2 days by staining in situ with benzidine (Sigma, St Louis, MO) to detect hemoglobin. After 7 to 10 days, erythroid burst-forming units (BFU-Es) and CFU-Cs were counted by colony morphology. Representative colonies were occasionally analyzed by Wright-Giemsa to confirm colony identity.

Spleen CFUs

The results shown are combined from 3 separate experiments. For each experiment, we combined the bone marrow from 3 mice of the same genotype. Single cell suspensions were prepared from bone marrow flushed from both tibiae and femora and diluted to 10⁵ cells per milliliter. Then, 10⁴ or 2 × 10⁴ cells were injected into the tail vein of sublethally irradiated (8.0 Gy, ¹³⁷C source) mice. For each experiment, 6 recipients were used for each donor genotype. Recipients were humanely killed on day 12 of transplantation, and their spleens were fixed with Bouin solution. Macroscopic colonies were counted. Spleens from irradiated mice that did not receive cells contained on average less than 1 CFU-S colony.

Transplantation assays

For each repopulation experiment, 6 C57BL/6J-*Gpi1*^{awa} and 6 C57BL/6J-*Gpi1*^{b/b} (either *Sca-1*^{-/-} or *Sca-1*^{+/+} used in some control experiments) bone marrow donor mice were treated with 150 μg 5-fluorouracil (5-FU) (Sigma) per gram body weight. At 2 days later, bone marrow (tibial and femoral) was flushed from each donor, and the bone marrow cells were combined according to *Gpi1* haplotype. Whole bone marrow cells were then subjected to iso-osmotic Percoll (Pharmacia, Piscataway, NJ) centrifugation. The density-separated (1.077 g/mL) donor bone marrow cells (or "buffy coats") were then mixed at the following ratios for competitive repopulation: 100% *Gpi1*^{awa} to 0% *Gpi1*^{b/b}, 67.7% *Gpi1*^{awa} to 33.3% *Gpi1*^{b/b}, 33.3% *Gpi1*^{awa} to 67.7% *Gpi1*^{b/b}, and 0% *Gpi1*^{awa} to 100% *Gpi1*^{b/b}. Bone marrow recipients (12 for each experiment) were treated with a single-dose, total body lethal irradiation (9.5 Gy, ¹³⁷C source). Then, 5 × 10⁶ cells of each separated bone marrow mixture (a total of 4 groups) were injected into the tail vein of 3 recipients. Tail bleeds were performed at 2, 4, 8, and 16 weeks for analysis of bone marrow contribution by *Gpi1* chimerism. In some experiments, blood samples were obtained at 32 and 52 weeks. No significant differences were observed between 16 and 52 weeks. Bone marrow was isolated from recipients at either 16 or 52 weeks, analyzed for *Gpi1* chimerism, and used for secondary repopulations. Bone marrow was harvested from 2 groups of 16-week-long primary bone marrow recipients for secondary transplantation: (1) wild-type (*Gpi1*^{awa}) irradiated mice were repopulated with *Sca-1*^{-/-} (*Gpi1*^{b/b}) bone marrow; and (2) *Sca-1*^{-/-} (*Gpi1*^{b/b}) irradiated mice were repopulated with wild-type (*Gpi1*^{awa}) bone marrow. For each primary recipient, 5 × 10⁶ Percoll-separated bone marrow cells were injected into the tail veins of 3 lethally irradiated mice of the same genotype (and *Gpi1* isotype) as the primary recipient.

Results

Hematologic analysis

To minimize genetic background effects on the analysis of the role of *Sca-1* in hematopoiesis, we backcrossed the *Sca-1*-targeted null allele onto the C57BL/6 background for 10 generations. Theoretically, the entire genetic background of these animals is derived from the C57BL/6 strain with the exception of approximately 20 cM flanking the *Ly-6* locus, which is derived from the 129 ES cell line used for gene targeting.¹⁹ The backcross-10 mice were bred to generate *Sca-1*^{+/+} and *Sca-1*^{-/-} progeny, which were subsequently independently bred. Hematopoietic analysis of *Sca-1*^{-/-} mice began with automated peripheral blood counts (Table 1). The hematocrits, hemoglobin levels, and total RBC and WBC counts were similar in the *Sca-1*^{+/+} and *Sca-1*^{-/-} mice. Yet the percentages in the peripheral WBC counts were significantly altered in *Sca-1*^{-/-} mice ($P < .05$). The percentage of peripheral blood lymphocytes was increased, with a concomitant decrease in both monocytes and granulocytes. These results are consistent with our previous report that lymphocytes derived from *Sca-1*^{-/-} mice are hyperproliferative in response to antigen.¹⁹ Unexpectedly, peripheral blood analysis revealed that *Sca-1*^{-/-} mice have 31% fewer platelets ($P < .01$). Next, flow cytometric analysis was performed to compare the number and percentages of bone marrow lineages in *Sca-1*^{+/+} and *Sca-1*^{-/-} mice. Bone marrow cellularity was found to be normal in *Sca-1*^{-/-} mice as were the proportions of monocytes, granulocytes, erythroblasts, and lymphocytes; however, bone marrow from *Sca-1*^{-/-} mice exhibited a marked reduction in megakaryocytes (by greater than 75%), consistent with the decrease in peripheral blood platelets (Table 2; $P < .01$). In addition, histologic examination of adult spleens also showed a reduction in megakaryocytes (data not shown).

To determine if the defective megakaryopoiesis resulted in a physiologic defect, bleeding times in response to tail cuts were measured on *Sca-1*^{+/+} and *Sca-1*^{-/-} mice. A 69% increase in average bleeding time was observed for *Sca-1*^{-/-} mice; however, the results were not statistically significant owing to the large variation in bleeding times in both wild-type and mutant mice (data not shown). The limited increase in bleeding time is consistent with the mild thrombocytopenia observed in *Sca-1*^{-/-} mice.

Precursor analysis

Colony-forming assays were performed to analyze the role of *Sca-1* in the development of myeloid-committed precursors. The most primitive in vitro colony-forming cell (granulocyte, erythroid, macrophage, megakaryocyte CFU [CFU-GEMM]) was reduced by more than 60%. As expected by the 4-fold decrease in bone marrow megakaryocytes, there was also a substantial decrease in megakaryocyte precursors (megakaryocyte CFUs [CFU-Mk's]) in *Sca-1*^{-/-}

Table 2. Comparison of myeloid lineages in wild-type and *Sca-1*-null bone marrow

Lineage	<i>Sca-1</i> ^{+/+} , %	<i>Sca-1</i> ^{-/-} , %
Erythroblast	14.87 ± 2.45	14.79 ± 2.31
Granulocyte	48.87 ± 2.01	46.04 ± 2.01
Megakaryocyte	7.42 ± 0.36*	1.82 ± 1.49*
Monocyte	2.90 ± 0.43	2.67 ± 0.31

Data were obtained by flow cytometric analysis of myeloid lineages with the use of the following lineage-specific markers: TER119⁺ (erythroblast), CD11b⁺Gr-1⁺ (granulocyte), CD41⁺CD61⁺ (megakaryocyte), and CD11b⁺Gr-1⁻ (monocyte).

*Highly significant differences; $P < .01$.

bone marrow (Table 3). Consistent with unaltered percentages of RBCs and TER-119⁺ cells in the periphery and bone marrow, *Sca-1*^{-/-} bone marrow contained normal percentages of erythroid (both BFU-E and CFU-E) progenitors. Although peripheral blood and bone marrow granulocyte and monocyte numbers are modestly decreased in *Sca-1*^{-/-} mice, CFU-GM, CFU-M, and CFU-G precursors are all significantly increased in mutant marrow. The CFU-spleen day-12 (CFU-S₁₂) assay is an in vivo clonogenic method that measures a primitive progenitor. As shown in Figure 1, *Sca-1*^{-/-} bone marrow has a 38% decrease in CFU-S₁₂ progenitors.

Hematopoietic stem cell analysis

Competitive bone marrow repopulations were performed to measure HSC activity in *Sca-1*^{-/-} mice. First, lethally irradiated mice received 5×10^6 medium-density *Sca-1*^{+/+} and *Sca-1*^{-/-} bone marrow cells that had been mixed at specific ratios. Wild-type and mutant cells were distinguished by expression of the glucose phosphate isomerase 1 (Gpi1) isoenzyme expressed by all cells, including erythrocytes. Wild-type bone marrow cells (from the C57BL/6J-*Gpi1*^{wa} strain) express the Gpi1AA isoenzyme, whereas control *Sca-1*^{+/+} and *Sca-1*^{-/-} bone marrow cells express the Gpi1BB isoenzyme. When mixtures of wild-type Gpi1AA and Gpi1BB bone marrow were used to repopulate lethally irradiated wild-type Gpi1AA mice, approximately the same percentage of input donor cells was recovered from the bone marrow of long-term repopulated animals (Figure 2, square dotted line), closely resembling the expected values depicted in the solid line. In contrast, when mixtures of wild-type Gpi1AA and *Sca-1*^{-/-} bone marrow cells were used to repopulate lethally irradiated Gpi1AA mice, a significantly greater proportion of Gpi1AA cells were recovered from the bone marrow of long-term repopulated mice (Figure 2, dashed line). For example, mice receiving transplants of bone marrow at a ratio of 2:1 mutant-to-wild-type cells showed reconstitution by wild-type cells at more than twice the expected frequency. In addition, Gpi1 analysis of peripheral blood from mice that have received transplants suggested that *Sca-1*^{-/-} bone marrow had impaired short-term repopulating activity (data not shown). Yet no differences in Gpi1 ratios were observed in 4-month and 12-month repopulated mice. Surprisingly, the Gpi1AA mice repopulated with 100% *Sca-1*^{-/-} Gpi1BB bone marrow (BM), showed only an average of 80% Gpi1BB bone marrow. Thus, some endogenous wild-type Gpi1AA HSCs recolonized the bone marrow despite the transplantation of 5×10^6 *Sca-1*^{-/-} bone marrow cells.

Reciprocal experiments were conducted in which *Sca-1*^{-/-} mice were used as host animals for the bone marrow transplants.

Table 3. Altered populations of committed progenitors, *Sca-1*-null mice

Progenitor	<i>Sca-1</i> ^{+/+}	<i>Sca-1</i> ^{-/-}
CFU-Es	338.7 ± 27.3	367.6 ± 18.7
BFU-Es	39.4 ± 6.5	35.0 ± 9.6
CFU-GEMMs	26.1 ± 2.8*	16.3 ± 2.2*
CFU-Mk's	46.3 ± 5.6*	19.7 ± 2.4*
CFU-GMs	86.7 ± 8.9*	127.1 ± 8.4*
CFU-Gs	135.0 ± 4.6	126.3 ± 7.0
CFU-Ms	198.9 ± 6.1*	220.4 ± 4.6*

Data are the average of 6 experiments using 3 animals from each genotype per experiment. Analysis was performed on 8- to 12-week-old mice for each genotype. Numbers are shown per 10^5 input bone marrow cells.

CFU-E indicates erythroid colony-forming unit; BFU-E, erythroid burst-forming unit; CFU-GEMM, granulocyte, erythroid, macrophage, megakaryocyte CFU; CFU-GM, granulocyte, macrophage CFU; CFU-M, macrophage CFU; CFU-G, granulocyte CFU; and CFU-Mk, megakaryocyte CFU.

*Highly significant differences; $P < .01$.

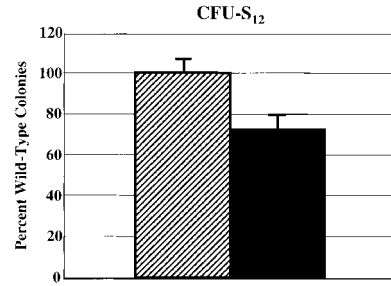


Figure 1. Reduction of CFU-S₁₂ precursors in *Sca-1*-null mice. CFU-S analysis of *Sca-1*^{-/-} bone marrow demonstrated a 38% reduction (4.4 CFU-S/10⁴ *Sca-1*^{+/+} cells [▨] versus 3.2 CFU-S/10⁴ *Sca-1*^{-/-} cells [■]; $P < .01$) in CFU-S₁₂ compared with wild-type bone marrow. The results are shown as the percentage of colonies from wild-type bone marrow from 3 separate experiments.

Again, a significantly higher percentage of wild-type cells repopulated *Sca-1*^{-/-} mice than were used for the transplant (Figure 2, round dotted line). For example, *Sca-1*^{-/-} mice that received transplants consisting of 33% wild-type and 67% *Sca-1*^{-/-} bone marrow cells demonstrated, on average, repopulation by 60% wild-type and 40% mutant cells. In contrast to the incomplete engraftment of Gpi1AA mice by *Sca-1*^{-/-} bone marrow, transplantations of 100% wild-type bone marrow cells into *Sca-1*^{-/-} mice demonstrated complete engraftment by wild-type cells.

Serial transplantations

HSC self-renewal activity was analyzed by using serial transplantation of *Sca-1*^{-/-} bone marrow into wild-type Gpi1AA mice and of wild-type Gpi1AA bone marrow into *Sca-1*^{-/-} mice (Figure 3). As shown in Figures 2 and 3, transplantation of 5×10^6 wild-type bone marrow cells into lethally irradiated *Sca-1*^{-/-} mice demonstrated complete reconstitution by wild-type cells. At 4 months after transplantation, bone marrow was harvested for Gpi1 isoenzyme analysis, and the remaining cells were subjected to gradient centrifugation. For each primary

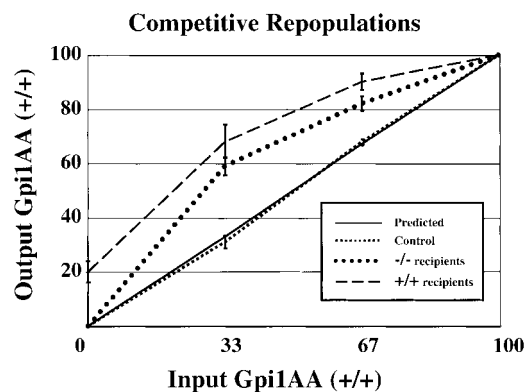


Figure 2. Competitive disadvantage for *Sca-1*^{-/-} bone marrow compared with *Sca-1*^{+/+} bone marrow. *Sca-1*^{+/+} (Gpi1AA) and *Sca-1*^{-/-} (Gpi1BB) bone marrows were mixed at specific ratios and used to repopulate lethally irradiated mice. The x-axis represents input *Sca-1*^{+/+} cells (Gpi1AA), and the y-axis represents output *Sca-1*^{+/+} (Gpi1AA) cells. If there is no difference in the ability of HSCs to repopulate lethally irradiated mice, the output percentage of Gpi1AA cells will be the same as the input percentage (solid line, or predicted values). In control experiments, *Sca-1*^{+/+} Gpi1AA bone marrow was mixed with *Sca-1*^{+/+} Gpi1BB bone marrow and used to repopulate lethally irradiated Gpi1AA mice. These results (square dotted line) reflect the expected result that there is no competitive advantage for either bone marrow population. The experimental results (dashed and round dotted lines) demonstrate that *Sca-1*^{-/-} bone marrow has a competitive disadvantage compared with *Sca-1*^{+/+} cells. The dashed and round dotted lines represent *Sca-1*^{+/+} Gpi1AA mice or *Sca-1*^{-/-} mice, respectively, as transplant recipients.

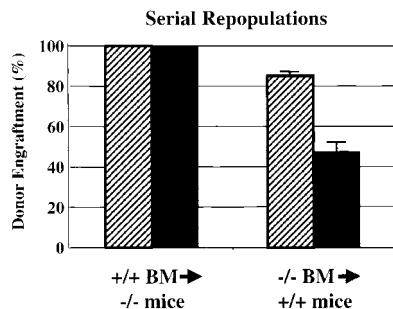


Figure 3. Serial bone marrow transplantations. Serial bone marrow transplantations demonstrate a significant defect in HSC competition and self-renewal by *Sca-1*^{-/-} mice. *Sca-1*^{+/+} BM was able to completely repopulate *Sca-1*^{-/-} mice in both primary and secondary recipients. However, when *Sca-1*^{-/-} bone marrow was used to repopulate lethally irradiated *Sca-1*^{+/+} mice, endogenous *Sca-1*^{+/+} HSCs were able to compete against donor stem cells in the primary repopulations (▨). In secondary transplantations using bone marrow from primary recipients to repopulate lethally irradiated *Sca-1*^{+/+} mice (■), the donor bone marrow could not rescue one-third of the recipients from irradiation-induced anemia. Of the remaining recipients, the *Sca-1*^{-/-} bone marrow contributed approximately 45% to the recipient bone marrow 4 months following repopulation. Three of 9 secondary recipients died.

recipient, 5×10^6 medium-density bone marrow cells were injected into the tail veins of 3 lethally irradiated *Sca-1*^{-/-} mice. All of the secondary recipients demonstrated complete reconstitution of wild-type cells into *Sca-1*^{-/-} mice by 4 months after transplantation. However, repopulation of wild-type mice with the use of *Sca-1*^{-/-} bone marrow demonstrated remarkably different results. As shown in Figures 2 and 3, primary wild-type mice that received transplants of *Sca-1*^{-/-} bone marrow exhibited poor donor engraftment. Secondary transplants from these animals into lethally irradiated wild-type mice resulted in about 45% engraftment by *Sca-1*^{-/-} bone marrow in the 6 recipients that survived transplantation. Three of the 9 secondary transplant recipients died within 17 days after transplantation. The moribund animals demonstrated hematocrits of less than 20% measured a week after transplantation.

Loss of *Sca-1* and diminished *Kit* signaling reduces embryonic survival

The receptor tyrosine kinase *c-Kit* encoded by the white-spotted (*W*) locus²³ is required for hematopoietic development and HSC self-renewal. The white-spotted viable (*W*^v) allele of *c-Kit* is a homozygous viable mutation that results in a threonine-to-methionine substitution at position 660 in the kinase domain resulting in reduced kinase activity.²⁴ To assess the functional capabilities of *Sca-1*^{-/-} HSCs in situ (ie, without ex vivo manipulation and transplantation), *Sca-1*^{-/-} mice were bred with *W*^v/*+* mice to generate *Sca-1*^{-/-};*W*^v/*+* mutants, which were then intercrossed. While *W*^v/*+* intercrosses generated 22% *W*^v/*W*^v homozygous pups, which exhibit mild anemia and impaired HSC competition, CFU-S activity, and decreased CFU-C precursors, *Sca-1*^{-/-};*W*^v/*+* intercrosses generated only 5% *Sca-1*^{-/-};*W*^v/*W*^v compound homozygote pups, indicating that most *Sca-1*^{-/-};*W*^v/*W*^v compound homozygotes die in utero ($P < .0001$; Figure 4). Preliminary analysis of embryonic day -14 litters from *Sca-1*^{-/-};*W*^v/*+* intercrosses indicate that compound homozygotes are very anemic and have dramatically reduced CFU-Cs, suggesting that the cause of embryonic lethality is due to a stem cell or very early progenitor defect (C.Y.J.L., N. Ciliberti, and W.L.S., unpublished data, 2002).

Discussion

The data presented here demonstrate that *Sca-1* plays an important role in regulating the repopulating ability of HSCs and the development of committed progenitor cells, megakaryocytes, and platelets. The decrease in CFU-S and CFU-GEMM progenitors may be attributable to deficient stem cell self-renewal and differentiation observed by means of the repopulation assays. The simplest explanation for the dramatic reduction in *Sca-1*^{-/-} HSC-repopulating ability between primary and secondary BM transplantations is that *Sca-1* plays an important role in HSC self-renewal, at least in response to hematologic stress caused by lethal irradiation and transplantation. This explanation is consistent with the regulation of *Sca-1* by interferons^{25,26} and tumor necrosis factor- α (TNF- α),²⁷ which are known to be induced in response to (1) cellular stress; (2) poor competition of *Sca-1*^{-/-} HSCs with wild-type stem cells in primary transplantations but not a complete lack of competition (Figure 2); and (3) the failure of *Sca-1*^{-/-};*W*^v/*W*^v compound homozygotes to survive (Figure 4).

Under homeostatic conditions (ie, *Sca-1*-null mice), *Sca-1* is not required for stem cell maintenance, which is consistent with a role for *Sca-1* as a coregulator of cell signaling; in this case, however, a coregulator of the signals that mediate stem cell self-renewal. A postulated molecular role of *Sca-1* and other GPI-anchored proteins is to modulate signaling by Src family kinases, receptor tyrosine kinases, and other signaling molecules in lipid rafts. *Sca-1* and the receptor tyrosine kinase *c-Kit* are coexpressed on HSCs, CLPs, early myeloid progenitors, and mast cells.^{1,3} While null *Kit* mutations lead to embryonic lethality owing to severe anemia caused by decreased stem cells and progenitors, the partial loss of function-*W* alleles, such as *W*^v/*W*^v and *W*⁴¹/*W*⁴¹, has impaired HSC competition and CFU-S activity, decreased CFU-Cs, and lead to mild anemia, mast cell deficiencies, and mild thrombocytopenia.^{28,29} Thus, the HSC and myeloid progenitor phenotype of *Sca-1*^{-/-} mice raises the intriguing possibility that *Sca-1* may play a role in modulating *Kit* signaling. The observation

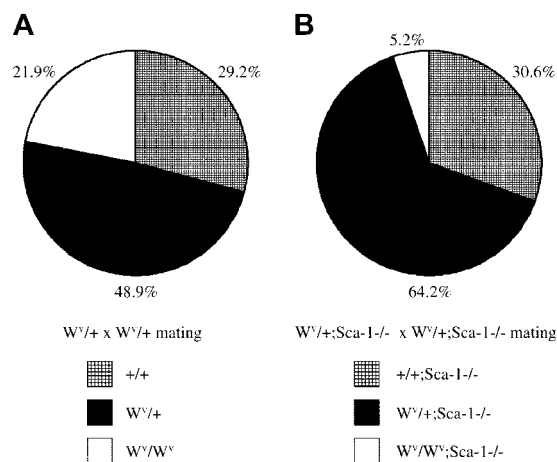


Figure 4. Embryonic lethality in *Sca-1*^{-/-};*W*^v/*W*^v compound homozygotes. (A) The ratio of wild-type, *W*^v/*+*, and *W*^v/*W*^v pups from *W*^v/*+* heterozygous intercrosses. A total of 178 pups were genotyped from 4 breeding pairs. Homozygous *W*^v/*W*^v pups were born at a slightly lower-than-expected, but not statistically significant, Mendelian ratio for a nonlethal mutation ($P > .1$). (B) The ratio of *Sca-1*^{-/-}, *W*^v/*+*;*Sca-1*^{-/-}, and *W*^v/*W*^v;*Sca-1*^{-/-} pups from *W*^v/*+*;*Sca-1*^{-/-} heterozygous intercrosses. A total of 288 pups were genotyped from 9 breeding pairs. Compound homozygous *W*^v/*W*^v;*Sca-1*^{-/-} pups were observed at an 84% reduction compared with *W*^v/*W*^v pups from *W*^v/*+* heterozygous intercrosses, demonstrating that most *Sca-1*^{-/-};*W*^v/*W*^v compound homozygotes die in utero ($P < .0001$).

of severe anemia and embryonic lethality in midgestational *Sca-1^{-/-};W^v/W^v* compound homozygote embryos does not discriminate between the possibility that Sca-1 acts in the Kit signaling pathway or that the lack of Sca-1 signaling and reduced Kit signaling are 2 independent genetic insults to fetal liver HSCs that together are incompatible with cell or population survival. We have initiated more complex mating strategies together with clonogenic assays as well as analysis of Kit signaling in *Sca-1^{-/-}* mast cells to distinguish between these 2 possibilities.

In addition to biochemical association of Sca-1 and Src family kinases,¹¹ there is genetic evidence that Sca-1 and other GPI-APs are negative regulators of Src family kinases. For example, the lymphoid phenotypes of Sca-1 and Src family kinase gene-targeted mice are diametrically opposite; *Sca-1^{-/-}* T cells are hyperresponsive to T-cell receptor (TCR) activation,¹⁹ while several Src family kinase mutants demonstrate poor TCR activation.³⁰ Interestingly, although *Fyn*-null peripheral T cells demonstrate normal TCR activation, they are refractory to activation via the GPI-anchored Thy-1 and Ly-6C.³¹ Finally, the potential interaction of Sca-1 and Kit may be mediated indirectly via Src family kinase signaling, which is required for ligand-mediated internalization of Kit.³² Unfortunately, HSC analysis of Src family kinase mutant strains has been limited; for example, competitive repopulation and serial transplantations have not been reported. However, the availability of mice with defined mutations in Src family kinases, Kit, or other signaling molecules that affect hematopoiesis will facilitate the molecular analysis of Sca-1 signaling in hematopoiesis and the molecular control of HSC self-renewal and commitment.

Significantly, our finding that Sca-1 plays a functional role in HSC self-renewal is consistent with the pathogenesis of paroxysmal nocturnal hemoglobinuria (PNH), an acquired disease resulting from a clonal expansion of HSCs that harbor a somatic null mutation in the X-linked phosphatidylinositol glycan-class A (*Pig-A*) gene, which catalyzes the first step in GPI biosynthesis.³³ PNH is characterized by complement-mediated intravascular hemolysis and eventual bone marrow failure. Gene-targeting experiments have shown that the lack of the GPI-anchored complement inhibitors CD55 (decay-accelerating factor [DAF]) and CD59 are the primary cause for the complement-mediated hemolysis affecting PNH patients.^{34,35} However, the mechanism enabling the PNH stem cell to expand within the bone marrow is not due to a loss of GPI-APs. GPI-deficient clones encoding *PIG-A* mutations have been detected in healthy individuals in trace amounts without the tendency to expand.³⁶ Furthermore, mice chimeric for a germ line-targeted mutation in *Pig-A* actually lose *Pig-A^{-/-}* cells over time,³⁷ demonstrating that *Pig-A^{-/-}* HSCs compete poorly with wild-type HSCs under homeostatic conditions, which is consistent with the role of Sca-1 that we report here.

In addition to HSCs, Sca-1 expression has been reported on muscle, bone, and mammary gland stem and/or primitive progenitors.³⁸⁻⁴¹ Consistent with the function of Sca-1 in HSCs, we have recently found that *Sca-1^{-/-}* mice develop age-related osteoporosis caused by a cell-autonomous defect in mesenchymal stem cell/primitive progenitor self-renewal (M. Bonyadi, S. D. Waldman, D. Liu, J. E. Aubin, M. D. Grynopas, W.L.S., manuscript submitted, 2002). The ligand/receptor signaling threshold (LIST) model of self-renewal versus differentiation suggests that a threshold level of signaling-competent cytokine-receptor complexes must be activated to propagate a self-renewal, rather than a differentiation, response.⁴² Thus, if stem cells receive self-renewal signals below a certain threshold, both daughter cells differentiate, leading to a reduction in the stem cell pool. The LIST model has recently been shown to account for gp130-mediated embryonic stem cell self-renewal in response to leukemia-inhibitory factor (LIF) and hyper-IL-6.⁴³ We suggest that coreceptors such as Sca-1 mediate self-renewal of stem cells by either increasing the number of functional cytokine-receptor complexes or altering the signaling properties of these complexes via the association of Sca-1 in lipid rafts and secondary signaling molecules sequestered in lipid rafts. Furthermore, we propose that adult stem cells use organ/stem cell-specific primary signaling molecules such as tyrosine kinases, whereas common cosignaling molecules, such as Sca-1, are conserved across many tissues and function to modify the balance between self-renewal and differentiation, especially under conditions of stress such as tissue regeneration. Thus, the absence of Sca-1 lowers the number of functional cytokine-receptor complexes or reduces the signaling capacity of these complexes, leading to a decrease in the stem cell pool. Consistent with this hypothesis, our preliminary biochemical analysis of Sca-1 signaling in mast cells demonstrates that *Sca-1^{-/-}* cells have altered levels of tyrosine-phosphorylated proteins in response to proliferation signals (N. Ciliberti, M. Ohishi, and W.L.S., unpublished results, 2002). We are now investigating other tissues to determine if Sca-1 can be used to isolate stem cells from these tissues; examining the regeneration potential of these organ systems in the *Sca-1^{-/-}* mice; and analyzing potential Sca-1 signaling pathways that may be used to manipulate stem cells for therapeutic and tissue-engineering ends.

Acknowledgments

The authors would like to thank Dwayne Barber, Jason Cohn, Norman Iscoves, Tammy Reid, and Peter Zandstra for critical reading and helpful discussions. This work is dedicated to the memory of Ms Karyn Glick.

References

- Osawa M, Hanada K, Hamada H, Nakauchi H. Long-term lymphohematopoietic reconstitution by a single CD34-low/negative hematopoietic stem cell. *Science*. 1996;273:242-245.
- Zhao Y, Lin Y, Zhan Y, et al. Murine hematopoietic stem cell characterization and its regulation in BM transplantation. *Blood*. 2000;96:3016-3022.
- Kondo M, Weissman IL, Akashi K. Identification of clonogenic common lymphoid progenitors in mouse bone marrow. *Cell*. 1997;91:661-672.
- Spangrude GJ, Klein J, Heimfeld S, Aihara Y, Weissman IL. Two monoclonal antibodies identify thymic-repopulating cells in mouse bone marrow. *J Immunol*. 1989;142:425-430.
- Bamezai A, Palliser D, Berezovskaya A, et al. Regulated expression of Ly-6A.2 is important for T cell development. *J Immunol*. 1995;154:4233-4239.
- Akashi K, Traver D, Miyamoto T, Weissman IL. A clonogenic common myeloid progenitor that gives rise to all myeloid lineages. *Nature*. 2000;404:193-197.
- Trevisan M, Iscove NN. Phenotypic analysis of murine long-term hemopoietic reconstituting cells quantitated competitively in vivo and comparison with more advanced colony-forming progeny. *J Exp Med*. 1995;181:93-103.
- Yotoku M, Grossberg AL, Pressman D. A cell surface antigenic determinant present on mouse plasmacytes and only half of mouse thymocytes. *J Immunol*. 1974;112:1774-1781.
- Horejsi V, Drbal K, Cebeceuer M, et al. GPI-microdomains: a role in signalling via immunoreceptors. *Immunol Today*. 1999;20:356-361.
- Simons K, Toomre D. Lipid rafts and signal transduction. *Nat Rev Mol Cell Biol*. 2000;1:31-39.
- Stefanova I, Horejsi V, Ansotegui IJ, Knapp W, Stockinger H. GPI-anchored cell-surface molecules complexed to protein tyrosine kinases. *Science*. 1991;254:1016-1019.
- Ninomiya H, Sims PJ. The human complement regulatory protein CD59 binds to the alpha-chain of C8 and to the "b" domain of C9. *J Biol Chem*. 1992;267:13675-13680.

13. Murray EW, Robbins SM. Antibody cross-linking of the glycosylphosphatidylinositol-linked protein CD59 on hematopoietic cells induces signaling pathways resembling activation by complement. *J Biol Chem.* 1998;273:25279-25284.
14. English A, Kosoy R, Pawlinski R, Bamezai A. A monoclonal antibody against the 66-kDa protein in mouse spleen and thymus inhibits Ly-6A.2-dependent cell-cell adhesion. *J Immunol.* 2000;165:3763-3771.
15. Malek TR, Ortega G, Chan C, Kroczek RA, Shevach EM. Role of Ly-6 in lymphocyte activation, II: induction of T cell activation by monoclonal anti-Ly-6 antibodies. *J Exp Med.* 1986;164:709-722.
16. Rock KL, Yeh ET, Gramm CF, Haber SI, Reiser H, Benacerraf B. TAP, a novel T cell-activating protein involved in the stimulation of MHC-restricted T lymphocytes. *J Exp Med.* 1986;163:315-333.
17. Codias EK, Rutter JE, Fleming TJ, Malek TR. Down-regulation of IL-2 production by activation of T cells through Ly-6A/E. *J Immunol.* 1990;145:1407-1414.
18. Marmor MD, Bachmann MF, Ohashi PS, Malek TR, Julius M. Immobilization of glycosylphosphatidylinositol-anchored proteins inhibits T cell growth but not function. *Int Immunol.* 1999;11:1381-1393.
19. Stanford WL, Haque S, Alexander R, et al. Altered proliferative response by T lymphocytes of Ly-6A (Sca-1) null mice. *J Exp Med.* 1997;186:705-717.
20. Henderson SC, Kamdar MM, Bamezai A. Ly-6A.2 expression regulates antigen-specific CD4+ T cell proliferation and cytokine production. *J Immunol.* 2002;168:118-126.
21. Zhang Z-X, Stanford WL, Zhang L. Ly-6A is critical for double negative T cell-mediated antigen-specific regulation. *Eur J Immunol.* 2002;32:1584-1592.
22. Nagy A, Rossant J. Production of completely ES cell derived fetuses. In: Joyner A, ed. *Gene Targeting: A Practical Approach.* New York, NY: IRL Press; 1993:147-179.
23. Chabot B, Stephenson DA, Chapman VM, Besmer P, Bernstein A. The proto-oncogene c-kit encoding a transmembrane tyrosine kinase receptor maps to the mouse W locus. *Nature.* 1988;335:88-89.
24. Nocka K, Tan JC, Chiu E, et al. Molecular bases of dominant negative and loss of function mutations at the murine c-kit/white spotting locus: W37, Wv, W41 and W. *EMBO J.* 1990;9:1805-1813.
25. Dumont FJ, Coker LZ. Interferon-alpha/beta enhances the expression of Ly-6A antigens on T cells in vivo and in vitro. *Eur J Immunol.* 1986;16:735-740.
26. Dumont FJ, Dijkmans R, Palfree RGE, Boltz RD, Coker LZ. Selective up-regulation by interferon-gamma of surface molecules of the Ly-6 complex on resting T cells: the Ly-6A/E and TAP antigens are preferentially enhanced. *Eur J Immunol.* 1987;17:1183-1191.
27. Malek TR, Danis KM, Codias EK. Tumor necrosis factor synergistically acts with IFN-gamma to regulate Ly-6A/E expression in T lymphocytes, thymocytes and bone marrow cells. *J Immunol.* 1989;142:1929-1936.
28. Russell ES. Hereditary anemias of the mouse: a review for geneticists. *Adv Genet.* 1979;20:357-459.
29. Ebbe S, Carpenter D, Yee T. Megakaryocytopenia in W/Wv mice is accompanied by an increase in size within ploidy groups and acceleration of maturation. *Blood.* 1989;74:94-98.
30. Lowell CA, Soriano P. Knockouts of Src-family kinases: stiff bones, wimpy T cells, and bad memories. *Genes Dev.* 1996;10:1845-1857.
31. Lancki DW, Qian D, Fields P, Gajewski T, Fitch FW. Differential requirements for protein tyrosine kinase Fyn in the functional activation of antigen-specific T lymphocyte clones through the TCR or Thy-1. *J Immunol.* 1995;154:4363-4370.
32. Broudy VC, Lin NL, Liles WC, et al. Signaling via Src family kinases is required for normal internalization of the receptor c-Kit. *Blood.* 1999;94:1979-1986.
33. Takeda J, Miyata T, Kawagoe K, et al. Deficiency of the GPI anchor caused by a somatic mutation of the PIG-A gene in paroxysmal nocturnal hemoglobinuria. *Cell.* 1993;73:703-711.
34. Sun X, Funk CD, Deng C, Sahu A, Lambris JD, Song WC. Role of decay-accelerating factor in regulating complement activation on the erythrocyte surface as revealed by gene targeting. *Proc Natl Acad Sci U S A.* 1999;96:628-633.
35. Holt DS, Botto M, Bygrave AE, Hanna SM, Walport MJ, Morgan BP. Targeted deletion of the CD59 gene causes spontaneous intravascular hemolysis and hemoglobinuria. *Blood.* 2001;98:442-449.
36. Araten DJ, Nafa K, Pakdeesuan K, Luzzatto L. Clonal populations of hematopoietic cells with paroxysmal nocturnal hemoglobinuria genotype and phenotype are present in normal individuals. *Proc Natl Acad Sci U S A.* 1999;96:5209-5214.
37. Rosti V, Tremml G, Soares V, Pandolfi PP, Luzzatto L, Bessler M. Murine embryonic stem cells without pig-a gene activity are competent for hematopoiesis with the PNH phenotype but not for clonal expansion. *J Clin Invest.* 1997;100:1028-1036.
38. Gussoni E, Soneoka Y, Strickland CD, et al. Dystrophin expression in the mdx mouse restored by stem cell transplantation. *Nature.* 1999;401:390-394.
39. Lee JY, Qu-Petersen Z, Cao B, et al. Clonal isolation of muscle-derived cells capable of enhancing muscle regeneration and bone healing. *J Cell Biol.* 2000;150:1085-1100.
40. Welm BE, Tepera SB, Venezia T, Graubert TA, Rosen JM, Goodell MA. Sca-1(pos) cells in the mouse mammary gland represent an enriched progenitor cell population. *Dev Biol.* 2002;245:42-56.
41. Short B, Brouard N, Occhiodoro-Scott T, Simmons PJ. Prospective isolation of stromal stem cells from mouse compact bone. *Keystone Symposium on Stem Cells: Origin, Fates and Functions.* March 17-23, 2002:99 (abstract 411).
42. Zandstra PW, Lauffenburger DA, Eaves CJ. A ligand-receptor signaling threshold model of stem cell differentiation control: a biologically conserved mechanism applicable to hematopoiesis. *Blood.* 2000;96:1215-1222.
43. Viswanathan S, Benatar T, Rose-John S, Lauffenburger DA, Zandstra PW. Ligand/receptor signaling threshold (LIST) model accounts for gp130-mediated embryonic stem cell self-renewal responses to LIF and HIL-6. *Stem Cells.* 2002;20:119-138.