

## Map kinase signaling pathways and hematologic malignancies

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**Mitogen-activated protein (Map) kinases are widely expressed serine-threonine kinases that mediate important regulatory signals in the cell. Three major groups of Map kinases exist: the p38 Map kinase family, the extracellular signal-regulated kinase (Erk) family, and the c-Jun NH<sub>2</sub>-terminal kinase (JNK) family. The members of the different Map kinase groups participate in the generation of various cellular responses, including gene transcription, induction of cell death or main-**

**tenance of cell survival, malignant transformation, and regulation of cell-cycle progression. Depending on the specific family isoform involved and the cellular context, Map kinase pathways can mediate signals that either promote or suppress the growth of malignant hematopoietic cells. Over the last few years, extensive work by several groups has established that Map kinase pathways play critical roles in the pathogenesis of various hematologic malignancies, pro-**

**viding new molecular targets for future therapeutic approaches. In this review, the involvement of various Map kinase pathways in the pathophysiology of hematologic malignancies is summarized and the clinical implications of the recent advances in the field are discussed. (Blood. 2003;101:4667-4679)**

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### Introduction

The different members of the superfamily of mitogen-activated protein (Map) kinases participate in signaling cascades conserved through evolution, which regulate important biologic activities. Three major groups of Map kinases (MAPKs) exist: the p38 Map kinase family, the extracellular signal-regulated kinase (Erk) family, and the c-Jun NH<sub>2</sub>-terminal kinase (JNK) kinase family<sup>1-12</sup> (Table 1). The p38 Map kinase family is composed of 4 different isoforms (p38 $\alpha$ , p38 $\beta$ , p38 $\gamma$ , and p38 $\delta$ ) that share significant structural homology,<sup>1,4,7,10,13-24</sup> while the JNK kinase group includes 3 members, JNK1, JNK2, and JNK3.<sup>2-5,25-28</sup> The Erk family of kinases includes Erk1 and Erk2<sup>2,29,30</sup> (Table 1). Erk1 and Erk2 are structurally and functionally similar kinases,<sup>2,25,26</sup> while Erk3<sup>31</sup> is a Map kinase that shares significant homology with Erk2 but also has distinct functions that differentiate it from the 2 classic Erk kinases (Erk1 and Erk2).<sup>2</sup> Furthermore, a subfamily of Erk3-related Map kinase genes, composed of 2 functional genes, MAPK6 and MAPK4, and several pseudogenes, has been recently identified.<sup>32</sup> Similarly, 2 other recently identified kinases, Erk5/BMK1 and Erk7, share some structural homology with the classic Erk kinases, but appear to also have distinct upstream and downstream effectors, and they are not classified in the classic Erk kinase group<sup>2,33-35</sup> (Table 1). Finally, a more recently cloned kinase, Erk8,<sup>36</sup> shares substantial homology with Erk7, but appears to have some differences in its substrate specificity (Table 1).

Map kinase signaling cascades are activated by a variety of different cellular stimuli and mediate diverse responses. Accumulating evidence indicates that an important function of Map kinases is the generation of signals of critical value to the control of normal and malignant hematopoiesis by cytokines and growth factors. The Erk pathway is activated in response to several cytokines and growth factors, and primarily mediates mitogenic and antiapoptotic signals.<sup>1-3,10,12</sup> Members of the p38 family of Map kinases are primarily activated by stress stimuli, but are also activated during

engagement of various cytokine receptors by their ligands.<sup>2-4,7,37</sup> The function of the p38 kinases is required for the generation of various activities, including regulation of apoptosis and cell-cycle arrest, induction of cell differentiation, as well as cytokine production and inflammation.<sup>2-4,7,38</sup> The JNK pathway is also activated in response to stress and growth factors, and, similarly, mediates signals that regulate apoptosis, cytokine production, and cell-cycle progression.<sup>2-5,10</sup>

In general, Map kinase pathways are activated by various stimuli to regulate, among other things, production of various cytokines and growth factors. Subsequently, they participate in signaling pathways activated by such cytokines or growth factors to mediate generation of specific biologic responses. Thus, for a given cytokine-dependent response, the function of Map kinases may be critical at 2 steps. The first step involves the biosynthesis and production of the cytokine in response to a stimulus, while the second step involves participation of Map kinases in signaling cascades activated by the cytokine to induce its biologic effects on target cells (Figure 1).

### Regulation of activation of Map kinase signaling cascades

In order for the different Map kinases to be activated by various stimuli, there is a requirement for dual phosphorylation on threonine (Thr) and tyrosine (Tyr) residues present in specific motifs (ThrXaaTyr) for each kinase group.<sup>1-12</sup> Such dual phosphorylation motifs are located in the activation loops of the different Map kinases. The distinct motifs for the different groups include: Thr-Gly-Tyr for the p38 kinases, Thr-Pro-Tyr for JNK kinases, and Thr-Glu-Tyr for the classic Erk kinases (Erk1 and Erk2) and for Erk5.<sup>1-12</sup> The phosphorylation of the different members of the Map

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Submitted December 3, 2002; accepted January 31, 2003. Prepublished online as *Blood* First Edition Paper, March 6, 2003; DOI 10.1182/blood-2002-12-3647.

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**Table 1. Protein members of the different mitogen-activated protein (Map) kinase groups**

Erk kinases	p38 kinases	JNK kinases	Other/unclassified
Erk1	p38 $\alpha$	JNK1	Erk3
Erk2	p38 $\beta$	JNK2	Erk5
—	p38 $\gamma$	JNK3	Erk7
—	p38 $\delta$	—	Erk8

— indicates not applicable.

kinase groups is regulated by upstream dual-specificity kinases, which are capable of phosphorylating Map kinases on both serine/threonine, as well as on tyrosine residues.<sup>1-12</sup> These kinases are called Map kinase kinases (MAPKKs or Mekks) and exhibit relative specificity for the substrate Map kinase proteins that they phosphorylate. Mkk1 and Mkk2 phosphorylate Erk kinases in the Thr-Glu-Tyr motif; Mkk3, Mkk4, and Mkk6 phosphorylate p38 kinases in the Thr-Gly-Tyr motif, while Mkk4 and Mkk7 regulate dual phosphorylation of JNK kinases in the Thr-Pro-Tyr motif (Table 2). Another member of the MAPKK family of kinases is Mkk5, which selectively phosphorylates the Thr-Glu-Tyr motif in Erk5/BMK-1.<sup>1-12,38</sup> Interestingly, MAPKK proteins also exhibit relative specificity for the different isoforms that they target within each group of Map kinases. For instance, among the different p38 subtypes, Mkk6 functions as a common activator for p38 $\alpha$ , p38 $\beta$ , and p38 $\gamma$ , while Mkk3 activates p38 $\alpha$  and p38 $\gamma$ , but not p38 $\beta$ .<sup>39</sup>

The activation of MAPK kinases (MAPKKs) is regulated by other upstream serine-threonine kinases, called MAPKK kinases (MAPKKKs), which phosphorylate the MAPK kinases (MAPKKs) on specific serine residues.<sup>1-12</sup> Thus, a series of phosphorylation events result in consecutive activation of serine-threonine and dual specificity kinases, ultimately leading to induction of the kinase domains of Map kinases and engagement of downstream pathways<sup>1-12</sup> (Figure 2). Several serine-threonine kinases have been implicated in acting as MAPKKKs for the 3 different groups of Map kinase pathways. In the case of the Erk pathway, the family of Raf kinases, as well as Mos and Tpl2, act as MAPKKKs.<sup>1-4,40-42</sup> On the other hand, the different MAPKKs that regulate activation of the JNK and p38 kinases include mixed-lineage kinases (Mlk1, Mlk2, Mlk3, Dlk, and Lzk), Mekk kinases (Mekk1, Mekk2, Mekk3, and Mekk4), Tak1, Ask1, Ask2, and Tpl-2.<sup>1-7,42-60</sup> It should be noted that, although transfection assays had demonstrated that Tpl-2 is a component of the JNK pathway, knock-out studies have indicated that it is a regulator of the Erk pathway.<sup>61</sup>

Activation of the MAPKKKs or MAPKKs occurs downstream from small G-proteins, whose function is regulated by guanine exchange factor (GEF) proteins (Figure 1). The small G-proteins that regulate activation of various Map kinase family members include Ras for the Erk pathway<sup>1-3,10,12,62</sup> and members of the Rho family of proteins (Rac1, Cdc42, RhoA, and RhoB) for the p38 and JNK pathways.<sup>4-10,63-68</sup> Thus, initial activation of GEFs leads to activation of GTPases and downstream initiation of distinct kinase cascades that regulate activation of different Map kinases (Figure 2). A summary of the distinct pathways that are activated downstream of various GTPases, ultimately leading to MAPK activation, is shown in Figure 3.

## Cellular responses mediated by Map kinases

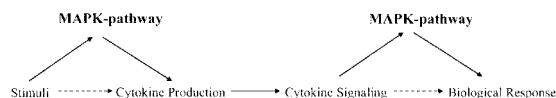
The activation of different Map kinase signaling cascades by various stimuli is required for induction of various important

cellular biologic responses, including phosphorylation of transcription factors and transcriptional regulation, nuclear chromatin remodeling and immediate gene induction, cytokine production, as well as regulation of apoptosis and cell-cycle progression. Such biologic activities vary with the specific family of Map kinases activated and the distinct stimulus inducing such activation. In general, the Ras/Erk pathway mediates primarily cell growth and survival signals, but under certain circumstances also promotes induction of cell differentiation.<sup>1-3,10-12</sup> On the other hand, the stress-activated p38 and JNK kinase pathways mediate primarily proapoptotic and growth inhibitory signals, as well as proinflammatory responses.<sup>1-12</sup> However, activation of the p38 pathway may also induce antiapoptotic, proliferative, and cell survival signals under certain conditions, depending on the tissue and specific isoform involved.<sup>5</sup>

Thus, multiple and divergent cellular functions are controlled by the activation of various components of Map kinase signaling cascades in response to a large number of stimuli. This raises questions regarding how the specificity of Map kinase-regulated responses occurs. An important issue revolves around the identity of cellular mechanisms that are in place to prevent cross-talk between the various cascades and to maintain the veracity and specificity of different signals generated by distinct Map kinase groups.<sup>69</sup> Extensive studies over the last few years have identified mechanisms that may account for Map kinase specificity.<sup>69-71</sup> It is likely that the events that define such specificity involve physical interactions of Map kinases with other proteins.<sup>69-71</sup> It has been established that Map kinases utilize for their interactions with other proteins the common docking site and the Glu-Asp (ED) site, while the proteins that interact with Map kinases have in their structure D-domains or Phe-Xaa-Phe-Pro (FXFP) domains.<sup>69-71</sup> Recently, the resolution of the crystal structure of p38 Map kinase bound to the docking domains of MEF2A and Mkk3b<sup>72</sup> defined the nature of the interaction between p38 and its substrates and provided a model on the structural basis of substrate specificity that may apply to the other Map kinase groups as well.<sup>69</sup> Thus, it appears that despite the extensive Map kinase networks and the multiple Map kinase members involved, specificity for Map kinase responses relies upon distinct interactions of different effector proteins with specific domains within the structure of Map kinases.

## Map kinases and cytokines that regulate hematopoiesis

It is well established that various cytokines and growth factors that regulate normal hematopoietic cell proliferation and differentiation activate Map kinase signaling pathways to generate their effects. Among the different hematopoietic growth factors, erythropoietin has been shown to activate, in responsive cell lines or primary hematopoietic progenitors, members of all different Map kinase groups, including Erk, p38, and JNK.<sup>73-80</sup> Similarly, another cytokine whose function is required for normal erythropoiesis, stem cell factor (SCF), is capable of inducing activation of all different Map kinase groups under certain conditions.<sup>77,79,81</sup> In



**Figure 1. Dual roles for Map kinase pathways in the induction of cytokine responses.** Regulation of cytokine production and participation in cytokine-dependent signaling cascades.

**Table 2. Ability of different MAP kinase kinases (MAPKKs) to phosphorylate/activate Map kinases from different groups**

	Mkk1	Mkk2	Mkk3	Mkk4	Mkk5	Mkk6	Mkk7
Motif	Thr-Glu-Tyr	Thr-Glu-Tyr	Thr-Gly-Tyr	Thr-Gly-Tyr, Thr-Pro-Tyr	Thr-Glu-Tyr	Thr-Gly-Tyr	Thr-Pro-Tyr
Erk	+	+	-	-	-	-	-
p38	-	-	+	+	-	+	-
JNK	-	-	-	+	-	-	+
Erk5	-	-	-	-	+	-	-

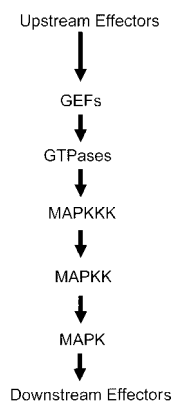
Mkk indicates Map kinase kinase; Erk, extracellular signal-regulated kinase; JNK, c-Jun NH<sub>2</sub>-terminal kinase; +, specified Map kinase is phosphorylated/activated by the specified Mkk; and -, specified Map kinase is not a known substrate for the specified Mkk.

addition, interleukin-3 (IL-3), granulocyte colony-stimulating factor (G-CSF), macrophage CSF (M-CSF), granulocyte-macrophage CSF (GM-CSF), and thrombopoietin, also activate Map kinase pathways.<sup>73,75,81-87</sup> Thus, it appears that essentially all known hematopoietic growth factors utilize Map kinase pathways for the generation of signals that potentially regulate normal hematopoiesis. In certain cases, the activation of a specific Map kinase pathway may be synergistically induced by a combination of growth factors to promote hematopoietic cell survival and growth. For instance, erythropoietin and stem cell factor synergistically activate Erk1/2 in purified human erythroid progenitors, and such synergistic activation of the classic Erk pathway appears to correlate with induction of cell growth.<sup>79</sup> On the other hand, certain Map kinase pathways also play critical roles in production of hematopoietic growth factors. As an example, targeted disruption of the p38  $\alpha$  gene in mice generally leads to embryonic lethality between days 11.5 and 12.5, but those mice who survive past this stage have normal morphology except that they are anemic because of diminished erythropoietin gene expression.<sup>88</sup> Thus, the p38 $\alpha$  Map kinase appears to be critical for developmental erythropoiesis via regulation of erythropoietin expression.<sup>88</sup> On the other hand, activation of p38 is necessary for erythropoietin-dependent differentiation of erythroid cells.<sup>78</sup> Such a dual function of the p38 pathway in erythropoiesis suggests the existence of Map kinase-regulated circuits, in which the function of a given Map kinase may control developmental expression of a certain hormone or growth factor and, at the same time, regulate its signaling capacity (Figure 1). However, it should be emphasized that defective erythropoiesis is not the primary consequence of targeted disruption of the p38 $\alpha$  gene in mice, and the major phenotype appears to be embryonic lethality related to placental developmental defects.<sup>89-91</sup>

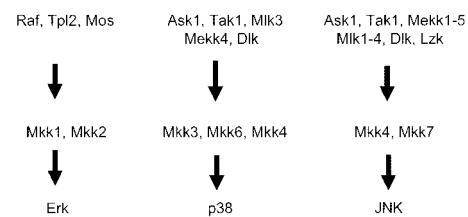
In addition to their activation by hematopoietic growth factors, Map kinases are activated by cytokines that negatively regulate normal human hematopoiesis. Among the several cytokines that have been shown to suppress hematopoiesis in vitro and in vivo, the

type I interferons (IFN $\alpha$ , IFN $\beta$ , and IFN $\omega$ )<sup>92,93</sup> are probably the most extensively studied. It has been known for many years that these cytokines are potent inhibitors of the growth of hematopoietic progenitors of all different lineages, including erythroid progenitors (burst-forming unit erythroid [BFU-E], colony-forming unit erythroid [CFU-E]), myeloid progenitors (CFU granulocyte-macrophage [CFU-GM]), megakaryocytic progenitors (CFU megakaryocyte [CFU-MK]) and mixed lineage progenitors (CFU granulocyte erythroid macrophage mixed [CFU-GEMM]).<sup>94-100</sup> Both the Erk1/2<sup>101,102</sup> and p38 pathways<sup>103,104</sup> are activated in response to treatment of human cell lines with type I interferons IFN $\alpha$  or IFN $\beta$ . Moreover, the p38 Map kinase cascade plays a critical role in type I interferon signaling, as it is required for regulation of type I IFN-dependent gene transcription, without modifying activation of signal transducer and activator of transcription (Stat) proteins.<sup>103,104</sup> Recent studies have shown that IFN $\alpha$  and IFN $\beta$  induce activation of the  $\alpha$  and  $\beta$  isoforms of the p38 Map kinase and its downstream effector Map kinase activated protein kinase-2 (MapKapK-2) in primary human erythroid progenitors.<sup>105</sup> Furthermore, treatment of normal bone marrow cells with the p38 pharmacologic inhibitors SB203580 and SB202190 reverses the suppressive effects of type I IFNs on normal human hematopoiesis.<sup>105</sup> On the other hand the Mek kinase inhibitor PD98059, which blocks activation of Erk kinases but not p38, had no effects on such hematopoietic suppression.<sup>105</sup> Thus, the p38 Map kinase pathway is required for the generation of the suppressive effects of type I interferons on normal human hematopoiesis, while the Erk pathway plays no role in the induction of such effects.<sup>105</sup>

The requirement of the p38 pathway for the generation of the antiproliferative effects of type I interferons ignited further studies, aimed to evaluate the role of this pathway in the generation of the antiproliferative effects of other well known myelosuppressive cytokines. Such studies demonstrated that, in addition to type I interferons, p38 $\alpha$  and p38 $\beta$  are also activated in primitive human hematopoietic progenitors in response to transforming growth factor  $\beta$  (TGF $\beta$ ),<sup>105</sup> tumor necrosis factor  $\alpha$  (TNF $\alpha$ ),<sup>106</sup> as well as type II interferon (IFN $\gamma$ )<sup>106</sup> treatment. Importantly, pharmacologic inhibitors of the p38 Map kinase reversed the inhibitory effects of all these different myelosuppressive cytokines on normal human hematopoiesis in vitro.<sup>105,106</sup> The results of these studies strongly suggested that the p38 Map kinase pathway acts as a common



**Figure 2. Schematic generic overview of the sequence of events leading to activation of Map kinase pathways.**



**Figure 3. MAPKKK and MAPKK proteins that regulate activation of the Erk, p38, and JNK Map kinase pathways.**

signaling mediator for growth inhibitory signals generated by different myelosuppressive cytokines (Figure 4). Such a role for p38 in normal hematopoiesis may have implications in the pathogenesis of certain bone marrow failure syndromes, in which suppression of normal hematopoiesis results from overproduction of myelosuppressive cytokines. Such a hypothesis was recently directly tested in studies using bone marrows from patients with idiopathic aplastic anemia, an acquired bone marrow failure syndrome caused by cytokine overproduction by activated immune cells.<sup>107,108</sup> Addition of pharmacologic inhibitors of p38 in aplastic anemia bone marrows enhanced erythroid (BFU-E) and myeloid (CFU-GM) hematopoietic colony formation *in vitro*,<sup>106</sup> raising the possibility that drugs that block activation of the p38 pathway may prove useful in the future treatment of aplastic anemia and other bone marrow failure syndromes.<sup>106</sup>

In general, the available evidence to date suggests that different Map kinase pathways play important roles in signaling for hematopoietic growth factors as well as for myelosuppressive cytokines. Such engagement of Map kinases in signaling for various cytokines appears to exhibit regulatory effects on normal human bone marrow hematopoiesis and has prompted extensive studies to define whether Map kinases participate in the regulation of the abnormal hyperproliferative signals seen in malignant hematopoiesis, as discussed below.

## Map kinases in human leukemias

### The Erk pathway in leukemias

The best characterized Map kinase signaling cascade in acute and chronic human leukemias is the Raf/Mek/Erk signaling cascade.<sup>12</sup> As indicated earlier in this review, the activation of Erk kinases is regulated via protein signaling cascades downstream of Ras, involving sequential engagement of Raf→Mkk1/2(Mek1/2)→Erk1/2 (Figure 3). It is of interest that a target protein for the Erk pathway is the acute myelogenous leukemia gene product (AML1, also called CBFA2 or PEPB2 alpha B), a transcription factor with transforming capacity that is involved in myeloid hematopoietic differentiation.<sup>109</sup> AML1 is phosphorylated by Erk in 2 serine sites within its proline-, serine-, and threonine-rich regions, and such phosphorylation is required for its transforming activity.<sup>109</sup>

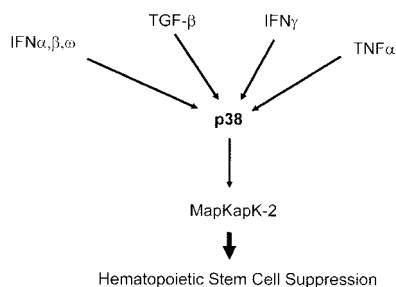
There is direct evidence that the Raf/Mek/Erk pathway promotes growth and prevents apoptosis of hematopoietic cells. Expression of a constitutively active mutant form of Mek1 can abrogate the requirement of human and murine hematopoietic cell lines to hematopoietic growth factors,<sup>109</sup> while such an abrogation of cytokine-dependence is associated with enhanced Map kinase phosphorylation/activation.<sup>110</sup> Similarly, expression of a constitu-

tively active form of Raf kinase in hematopoietic cell lines results in Mek1 and Erk1/Erk2 activation and growth factor independence.<sup>111</sup> Furthermore, concomitant expression of Bcl-2 and a conditionally active form of the Mek1 protein result in a synergistic induction of cytokine independence and protection of cells from apoptosis, strongly suggesting that the Mek/Erk pathway and Bcl-2 act synergistically to induce leukemogenesis.<sup>112</sup>

Recent studies have shown that Erk1/2 and their upstream effectors Mek1/2 are constitutively activated in primary human acute myelogenous leukemia (AML) cells<sup>113-115</sup> and that such a constitutive activation correlates with down-regulated expression of the PAC1 phosphatase.<sup>114</sup> Furthermore, the Erk protein was found to be overexpressed in the majority of acute leukemia cases studied,<sup>114</sup> suggesting that both overexpression and abnormal constitutive activation of Erk kinases are contributing to abnormal cell growth in acute leukemias. Similarly, another study demonstrated activation of the Mek/Erk pathway and its downstream targets, the CREB-1, ATF-1, and c-Myc transcriptional activators, in 9 of 14 acute myelogenous leukemia (AML) cell lines and 2 of 5 chronic myelogenous leukemia (CML) cell lines studied.<sup>116</sup> Others, however, failed to demonstrate a dependency of myeloid leukemia growth to Erk activation in certain myeloid leukemia cell lines examined.<sup>117</sup>

Independently of results observed with cell lines, which may reflect adaptive changes resulting from long-term culture of the leukemia cells, it is clearly established that in the majority of primary acute leukemia cases, the Erk pathway is constitutively activated and mediates mitogenic signals.<sup>113-115,118</sup> In a large study, constitutive MAPK kinase phosphorylation was detected in 138 (74%) of 186 freshly isolated leukemic blasts from AML patients.<sup>115</sup> Treatment of leukemia blasts with the Mek inhibitors PD98059 or PD184352 inhibited growth and induced apoptosis, while it also sensitized the cells to chemotherapy (Ara-C)-induced cytotoxicity.<sup>115</sup> Other studies have also demonstrated synergistic effects of chemotherapeutic agents and drugs that inhibit Map kinase activation in inducing growth suppression and apoptosis of acute leukemia cells.<sup>119,120</sup> Interestingly, such potentiating effects of Mek inhibition on chemotherapy-induced cytotoxicity appear to be sequence-dependent, with Mek kinase inhibitors being effective only when administered after the chemotherapeutic agents.<sup>115,120</sup> The demonstration of such synergistic effects of Mek kinase inhibitors with chemotherapeutic agents<sup>115,120,121</sup> and radiotherapy<sup>121</sup> has provided important models for the future clinical development of such inhibitors for the treatment of acute leukemias and other hematologic malignancies as well.

N-Ras or K-Ras mutations, which lead to constitutive activation of this GTPase and downstream activation of the Raf/Mek/Erk pathway, occur in a significant number of acute myeloid and acute lymphoblastic leukemias (ALLs) (20%-30%).<sup>122-124</sup> However, the constitutive activation of the Mek/Erk pathway in acute leukemias does not appear to correlate with the presence of N-Ras mutations,<sup>125</sup> suggesting that, in addition to Ras, other yet unidentified small G-proteins may also be abnormally activated and contribute to the activation of the Erk pathway in leukemic cells. Alternatively, in the absence of Ras mutations, autocrine or paracrine growth factor/cytokine pathways may be regulating activation on normal Ras and downstream engagement of the Mek/Erk cascade in acute myeloid leukemia cells. Interestingly, in a recent phase 1 clinical trial of the R11577 farnesyltransferase inhibitor in adults with poor-risk acute leukemias it was found that in approximately 36% of pretreatment patient bone marrows the phosphorylated/activated form of Erk was detectable.<sup>126</sup> No N-Ras mutations were



**Figure 4.** Myelosuppressive cytokines that utilize the p38 Map kinase pathway to suppress the growth of human primitive hematopoietic progenitors.

found in any of the 35 patients enrolled in that study, while in half of the cases in which Erk was phosphorylated/activated, such phosphorylation disappeared after one cycle of treatment.<sup>126</sup>

Altogether, these studies have established that the Raf/Erk kinase pathway plays a role in the pathophysiology of acute leukemias. However, other Erk-independent mitogenic mechanisms are apparently also involved, as constitutive activation of Mek1/2 and/or Erk1/2 is not detectable in all cases of acute leukemia. Independently of the roles that other signaling pathways may play in the pathophysiology of acute leukemias, pharmacologic targeting of the Erk pathway may be an attractive clinical-translational approach. Also, as there is recent evidence demonstrating synergistic effects of Mek/Erk pharmacologic inhibitors with Bcl-2 inhibitors in AML cells,<sup>127</sup> efforts to develop clinical trials combining such inhibitors for the treatment of acute leukemias may be warranted.

Chronic myelogenous leukemia (CML) results from oncogenic transformation of hematopoietic stem cells by the product of the *bcr-abl* oncogene, which is generated by the reciprocal translocation between chromosomes 9 and 22, resulting in the fusion of the *bcr* gene to the *c-abl* gene.<sup>128-132</sup> The function of the Erk signaling cascade has been implicated in transformation by the BCR-ABL proto-oncogene,<sup>131-133</sup> while inhibition of activation of this cascade in BCR-ABL-expressing cell lines results in cell death and correlates with the induction of apoptosis by various agents, including STI571.<sup>134-136</sup> Furthermore, Mek/Erk pharmacologic inhibitors appear to exhibit synergistic effects with STI571 in the induction of apoptosis of BCR-ABL-expressing cells.<sup>136</sup> Thus, it is possible that combined administration of Mek inhibitors and STI571 may prove useful in the development of future treatment approaches for CML patients and/or Philadelphia chromosome-positive (Ph<sup>+</sup>) acute lymphoblastic leukemia (ALL) patients. However, it should be pointed out that in contrast to acute leukemia, in which constitutive activation of Mek/Erk is seen in the majority of primary leukemic blasts, such an activation was not observed in 14 primary CML cases reported in one study.<sup>113</sup> Thus, more extensive studies to better understand the role of the Erk pathway in the pathogenesis and growth of CML cells may be required prior to the development of clinical trials using Mek inhibitors for the treatment of chronic myelogenous leukemia.

Other studies have examined the role that the Erk pathway plays in the maintenance of survival of malignant lymphocytes from patients with chronic lymphocytic leukemia (CLL).<sup>137,138</sup> In CLL cells, there is no constitutive activation of Erk1 or Erk2, but such activation can be induced by phorbol-ester treatment of the cells<sup>137</sup> or by engagement of the B-cell antigen receptor.<sup>139</sup> However, pharmacologic inhibition of the Erk pathway in malignant lymphocytes does not induce apoptosis.<sup>137,138</sup> Thus, Erk kinases do not appear to play a role in maintaining cell survival in CLL cells, and the defective apoptosis seen in these cells appears to be primarily mediated by activation of the phosphatidylinositol 3'-kinase (PI-3'K) and protein kinase C  $\delta$  (PKC- $\delta$ ).<sup>137,138</sup>

A leukemia in whose pathophysiology the Ras/Raf/Mek/Erk pathway appears to play a key role is natural killer (NK) large granular lymphocyte leukemia. In a recent study, it was found that constitutively active Erk was detectable in the peripheral blood of 11 of 11 patients with NK cell leukemia studied.<sup>140</sup> Of these patients, 2 had the aggressive form of the disease, while 9 had chronic leukemia. Treatment of the cells with Mek inhibitors (PD98059 and U0126) or overexpression of a dominant-negative form of Mek-1 resulted in apoptosis.<sup>140</sup> The constitutive activation of Mek-Erk in these cells was apparently downstream of Ras, as

treatment of the cells with a farnesyl transferase inhibitor or ectopic expression of a dominant-negative form of Ras inhibited Erk phosphorylation and induced cell death.<sup>140</sup> Thus, this relatively rare form of leukemia may be another disease in which Mek inhibitors, alone or in combination with other agents, may prove to be clinically efficacious.

### The p38 and JNK pathways in leukemias

Although the role of Erk kinases in mediating mitogenic and antiapoptotic signals in acute leukemia cells is well defined, the roles that members of the p38 and JNK kinase groups play in regulation of growth of acute leukemia blasts are not well established. A recent report demonstrated a relationship between constitutive activity of JNK in leukemic blasts and treatment failure in acute myelogenous leukemia.<sup>141</sup> In that study, a biochemical analysis of 67 primary adult acute myelogenous leukemia samples demonstrated a correlation between JNK expression/activation in the leukemic cells and hyperleukocytosis at presentation of disease.<sup>141</sup> Importantly, a correlation between JNK kinase activity and increased multidrug resistance-associated protein efflux was observed.<sup>141</sup> This raises the intriguing possibility that targeting JNK and/or its substrate, c-jun, may be a novel clinical approach to overcome multidrug resistance in AML. However, the development of such approaches is unlikely to be straightforward, as the JNK pathway is also activated by various chemotherapeutic agents, and its function may also be required for the induction of apoptosis by certain agents that are used in the treatment of AML.<sup>142-144</sup> As there are 3 different known JNK isoforms, it is possible that the JNK isoforms that mediate chemotherapy-dependent apoptosis are different from the ones involved in the promotion of multidrug resistance in AML cells. Thus, studies to precisely identify the isoforms involved in each of the 2 JNK-dependent responses in AML cells may provide valuable information for the design of new isoform-specific inhibitors with clinical translational potential.

There are no reports directly demonstrating a role for constitutive activation of the p38 Map kinase pathway in the pathophysiology of acute leukemias. A recent study showed that p38 and its downstream effector MapKapK-2 are activated during treatment of the NB-4 acute promyelocytic leukemia (APL) cell line with all-*trans* retinoic acid (ATRA).<sup>145</sup> Interestingly, pharmacologic inhibition of the p38 Map kinase using the SB203580 or SB202190 inhibitors was found to strongly enhance all-*trans* retinoic acid-dependent induction of APL cell differentiation and all-*trans* retinoic acid-dependent growth inhibition.<sup>145</sup> On the other hand, the Mek inhibitor PD98059 was found to block the induction of differentiation of NB-4 cells<sup>145</sup> or HL-60 cells<sup>146,147</sup> in response to all-*trans* retinoic acid, consistent with a positive role for the Mek/Erk pathway in the induction of differentiation of acute promyelocytic leukemia cells. These findings indicate that the p38 Map kinase pathway plays a negative role in the induction of all-*trans* retinoic acid responses in acute promyelocytic leukemia and raise the possibility that combined use of all-*trans* retinoic acid with pharmacologic inhibitors of p38 may prove more effective than ATRA alone in inducing differentiation of APL blasts *in vivo*. Similar mechanisms of synergy with pharmacologic inhibitors of p38 may also be applicable for other drugs used in the treatment of acute promyelocytic leukemia. Another agent, which is approved for the treatment of ATRA refractory acute promyelocytic leukemia, is arsenic trioxide (As<sub>2</sub>O<sub>3</sub>).<sup>148</sup> In a similar manner to what was observed with all-*trans* retinoic acid, treatment of NB-4 APL cells with arsenic trioxide also resulted in activation of the p38/

MapKapK-2 pathway, while pharmacologic inhibition of p38 further enhanced arsenic trioxide–induced apoptosis and growth inhibition of APL cells.<sup>149</sup> It is of interest that the activation of p38 and its upstream regulator Rac1 by arsenic trioxide is also inducible in an NB-4 variant cell line that is resistant to the effects of all-*trans* retinoic acid.<sup>149</sup> This indicates that different upstream regulatory mechanisms mediate activation of p38 in response to ATRA or  $As_2O_3$ .<sup>149</sup> However, in both cases pharmacologic inhibition of p38 promotes the induction of antileukemic responses, suggesting common downstream regulatory mechanisms.<sup>145,149</sup>

Other studies have shown that, under certain circumstances, the p38 pathway can cooperate with the Erk pathway to mediate cytokine-induced proliferation of AML cells.<sup>150</sup> In particular, it was shown that both p38 and Erk are required for optimal proliferation of the OCI-AML5 cell line in response to G-CSF and Flt3-ligand.<sup>150</sup> Thus, depending on the specific circumstances, the p38 pathway can either antagonize the Erk pathway or work synergistically with it to promote growth factor–dependent proliferation of AML blasts.

The functional roles that the p38 and JNK Map kinase pathways may play in the pathogenesis and pathophysiology of chronic leukemias have also been extensively studied. It is of particular interest that the RhoGEF domain of Bcr has the capacity to activate p38 and that the function of p38 is required for Bcr-regulated activation of nuclear factor- $\kappa$ B (NF- $\kappa$ B).<sup>151</sup> Thus, in addition to transformation via the tyrosine kinase activity of BCR-ABL, the RhoGEF domain of Bcr may be contributing to transformation, and p38 activity may mediate such effects.<sup>151</sup>

Other studies have shown that the function of the p38 pathway is essential for the suppression of growth of chronic myelogenous leukemia cells by interferon- $\alpha$  (IFN $\alpha$ ).<sup>152</sup> It has been demonstrated that IFN $\alpha$  selectively activates p38 in a type I IFN–sensitive BCR-ABL–expressing cell line (KT-1), but not in the BCR-ABL–expressing K562 cell line, which is refractory to the antiproliferative effects of IFN $\alpha$ .<sup>152</sup> Concomitant treatment of KT-1 cells with the p38-specific inhibitor SB203580 or the SB202190 inhibitor reverses the growth inhibitory effects of IFN $\alpha$  on these cells, while the Mek inhibitor PD98059 has no effects.<sup>152</sup> Most importantly, IFN $\alpha$  treatment of isolated peripheral blood granulocytes from CML patients induces phosphorylation/activation of p38 in vitro and addition of pharmacologic inhibitors of p38 in CML bone marrow cultures reverses the suppressive effects of IFN $\alpha$  on leukemic CFU-GM progenitor colony formation.<sup>152</sup> Thus, activation of the p38 signaling cascade appears to be essential for the generation of the antileukemic effects of IFN $\alpha$  in chronic myelogenous leukemia cells in vitro and possibly in vivo. It should also be pointed out that differentiation of BCR-ABL–expressing K562 cells during treatment with butyrate results in phosphorylation/activation of p38 and down-regulation of Erk kinase activity.<sup>153</sup> Such phosphorylation of p38 appears to be essential for the differentiation of such cells, as it is blocked by pharmacologic inhibition of p38 kinase activity.<sup>153</sup> On the other hand, inhibition of Erk activity using the U0126 selective Mek kinase inhibitor enhances such differentiation, demonstrating that Erk and p38 exhibit opposing functions in the induction of differentiation of BCR-ABL–expressing cells.<sup>153</sup>

BCR-ABL has been shown to activate the JNK kinase pathway,<sup>154</sup> while expression of the c-jun gene in BCR-ABL–transformed cells correlates with JNK kinase activity.<sup>155</sup> Dominant-negative mutants of c-Jun impair the transforming activity of BCR-ABL, indicating a requirement for the JNK pathway in BCR-ABL–mediated transformation of hematopoietic cells.<sup>155</sup> The

JNK-mediated transforming activity of BCR-ABL has also been shown to be inhibited by a JNK cytoplasmic inhibitor, the JNK interacting protein-1 (JIP-1).<sup>156</sup> A more recent study has demonstrated that disruption of the JNK ortholog Mapk8 (JNK1) in mice results in defective transformation of pre-B cells by BCR-ABL, in vitro and in vivo.<sup>157</sup> This study also established that failure of BCR-ABL–transformed cells to survive in the absence of JNK is because of decreased expression of Bcl2, as the effect of JNK deficiency could be reversed by transgenic Bcl2 expression.<sup>157</sup> Thus, the JNK pathway plays an important role in BCR-ABL–mediated transformation via regulation of c-jun activity, indicating that selective inhibitors of this pathway may be of clinical relevance in the treatment of chronic myelogenous leukemia.

The roles of the JNK and p38 pathways in the pathophysiology of other leukemias are not well known. There is some evidence implicating constitutive JNK activation in the pathogenesis of human lymphotropic virus (HTLV-1) tumorigenesis and indirectly implying a role for this pathway in the pathogenesis of adult T-cell leukemia.<sup>158</sup> Consistent with this, other studies have shown that a MAPKK kinase, Mlk-3, is involved in Tax-mediated NF- $\kappa$ B activation.<sup>155</sup> The Tax protein of HTLV-1 is an oncoprotein that transactivates various genes, which play key roles in HTLV-1 replication and pathogenesis.<sup>159</sup> The fact that Mlk-3 regulates Tax-mediated activation of the transcription factor NF- $\kappa$ B further supports a putative involvement of the JNK pathway in the pathogenesis of adult T-cell leukemia and raises the possibility that the JNK kinase may be an appropriate molecular therapeutic target.

Efforts have also been made to address the potential roles of the JNK and p38 pathways in the pathophysiology of chronic lymphocytic leukemia (CLL). It has been demonstrated that, in chronic lymphocytic leukemia cells, cross-linking of the B-cell antigen receptor induces activation of Erk, but not JNK or p38,<sup>139</sup> suggesting that these kinases do not play important roles in CLL cell proliferation.<sup>139</sup> Consistent with this, other studies have shown that pharmacologic inhibition of the p38 pathway does not induce apoptosis of CLL cells.<sup>138</sup> Interestingly, a recent study demonstrated that the induction of apoptosis of CLL cells by the chimeric anti-CD20 antibody rituximab is p38-dependent.<sup>160</sup> p38 and its downstream effector, MapKapK-2, were found to be activated during culture of isolated CLL cells with anti-CD20, while treatment with the p38 pharmacologic inhibitor SB203580 resulted in induction of apoptosis of the malignant lymphocytes.<sup>160</sup> Cross-linking of rituximab to CLL cells also induced strong phosphorylation of Erk and JNK kinases. However, the Mek inhibitor U0126 had no inhibitory effects on anti-CD20–induced apoptosis despite the fact that it blocked Erk activation, strongly suggesting that Erk is not required for its antileukemic activity.<sup>160</sup> Thus, the p38 pathway appears to play an important and specific role in the generation of the antileukemic effects of rituximab in chronic lymphocytic leukemia.

### Map kinases in lymphomas

The difficulty in working with primary tumor samples from patients suffering from lymphomas has been a limiting factor in efforts to uncover the roles that Map kinases may play in the pathogenesis and pathophysiology of these malignancies. So far, most of the evidence on the putative roles that Map kinases may play in the pathogenesis of lymphomas is based on work with lymphoma-derived cell lines. Tumor necrosis factor  $\alpha$  (TNF $\alpha$ ) has been shown to induce autocrine regulation of the growth of several lymphoma or leukemia cell lines, alone or in combination with other growth factors.<sup>161-163</sup> It has also been previously established

that the p38 pathway is required for the regulation of TNF $\alpha$  production.<sup>4</sup> Furthermore, the downstream effector of p38, Map kinase-activated protein kinase-2 (MapKapK-2), mediates such regulatory effects of the p38 pathway on TNF $\alpha$  production. This has been shown in studies using mice with targeted disruption of the MapKapK-2 gene, which have shown that p38/MapKapK-2 is required for TNF $\alpha$  protein biosynthesis in response to lipopolysaccharide (LPS), without affecting TNF $\alpha$  gene mRNA transcription.<sup>164</sup> Recently, it was demonstrated that the TNF $\alpha$ -inducible proliferation of certain non-Hodgkin lymphoma (NHL) and leukemia cell lines is dependent on activation of the p38 Map kinase in response to engagement of the TNF $\alpha$  receptor in malignant cells.<sup>164</sup> Thus, p38 may both regulate TNF $\alpha$  production and also, upon its activation by TNF $\alpha$ , mediate signals that regulate growth of lymphoma/leukemia cells. However, it is difficult to extrapolate from these studies using cell lines whether such effects also occur in primary lymphoma cells. Other studies have implicated the Raf-1/Mek/Erk and PI-3' kinase pathways in the suppression of Fas-induced apoptosis in lymphoma cells, as evidenced by experiments using the PD98059 Mek inhibitor and the LY294002 PI-3' kinase inhibitor.<sup>165</sup> It has been shown that both of these pharmacologic inhibitors lead to accumulation of Erk in the cytosol of lymphoma cells, suggesting a cross-talk between the 2 pathways.<sup>165</sup> Interestingly, such an induction of Fas-dependent apoptosis was selectively seen in lymphoma cells, but not normal thymocytes, that express low Raf-1 levels,<sup>161</sup> suggesting divergent Map kinase-dependent regulatory effects on apoptosis in malignant lymphoma cells versus normal cells.

The p38 Map kinase pathway has been implicated in the regulation of interleukin-10 (IL-10) production in Burkitt lymphoma cell lines.<sup>166</sup> This cytokine normally regulates growth and differentiation of B cells.<sup>167</sup> It has been demonstrated that the Epstein-Barr virus latent membrane protein 1 (LMP1) induces expression of IL-10 in Burkitt lymphoma cell lines in a p38 Map kinase-dependent manner.<sup>166</sup> Such an effect was selectively seen in Burkitt cell lines, but not in several other non-Hodgkin lymphoma (NHL) and Hodgkin disease-derived cell lines that were analyzed.<sup>166</sup> A recent report has also demonstrated that Epstein-Barr virus latent membrane protein 2A (LMP2A) activates both Erk and JNK kinases, and that LMP2A-inducible phosphorylation of c-Jun is Erk kinase-dependent.<sup>168</sup> Altogether, these studies suggest that Map kinase pathways play roles in the pathogenesis of Epstein-Barr virus-related lymphomas and raise the prospect that selective pharmacologic inhibitors of Map kinases may find clinical applications in the treatment of these lymphoma types in the future.

Map kinase pathways may also play roles in growth factor loops that promote cell proliferation of the malignant cells in Hodgkin disease. There is now strong evidence implicating aberrant expression of c-Jun and JunB, which are downstream effectors of Map kinase pathways, in the proliferation of malignant Hodgkin lymphoma cells.<sup>169</sup> In one study, constitutively activated AP-1 and significant overexpression of c-Jun and JunB were found in all primary tumor cells from patients with Hodgkin disease, while a similar activation of AP-1 was detected in anaplastic large-cell lymphoma cases, but not other types of lymphoma.<sup>169</sup> Interestingly, activated AP-1 was found to support the growth of Hodgkin lymphoma cells, but in anaplastic large-cell lymphoma cells it was mediating antiapoptotic effects.<sup>169</sup> Finally, other more recent studies have shown that all different Map kinase groups, Erk, p38, and JNK, are activated in Hodgkin disease cell lines in response to receptor activator of NF- $\kappa$ B ligand (RANKL),<sup>170</sup> a factor that is involved in the regulation of cytokine/chemokine secretion in

Reed-Sternberg cells via autocrine mechanisms.<sup>171</sup> Although there is no direct evidence to date implicating such activation of Map kinases in the growth of Reed-Sternberg cells, it is possible that they are involved in such activities, and future studies in that direction may uncover interesting and potentially important information.

## Map kinases in multiple myeloma

There is accumulating evidence that several hematopoietic growth factors are present in the bone marrow microenvironment and regulate survival and proliferation of the malignant plasma cells in multiple myeloma.<sup>172</sup> The most important myeloma growth factor is interleukin-6 (IL-6), a cytokine that is produced by myeloma cells in an autocrine or paracrine manner and promotes their survival *in vitro* and *in vivo*.<sup>173-181</sup> In addition, insulin-like growth factor 1 (IGF-1),<sup>182-184</sup> granulocytic colony-stimulating factor (G-CSF),<sup>172-175</sup> and interleukin-10<sup>172-175</sup> also act as growth factors for primary multiple myeloma cells and/or certain cell lines *in vitro*. Thus, various signaling networks are activated by different cytokines, primarily interleukin-6, to regulate the growth of malignant myeloma cells. Because of this, the pathways activated by such cytokines are of critical importance in the pathogenesis of the disease.

Interleukin-6 activates multiple signaling cascades, including the Jak-Stat, the Ras/Raf/Mek/Erk, and the PI-3' kinase/Akt pathways.<sup>172-175</sup> As interleukin-6 is a critical factor for the growth of multiple myeloma cells, it is not surprising that the function of the Raf/Mek/Erk cascade mediates signals that promote malignant myeloma cell proliferation.<sup>185</sup> It has been demonstrated that treatment of dependent myeloma cell lines with interleukin-6 induces tyrosine phosphorylation of Shc and its association with the Ras-guanine exchange factor Sos-1, resulting in downstream activation of the Mek/Erk pathway.<sup>185</sup> Importantly, using an antisense approach, it was established that Erk activation is essential for interleukin-6-dependent myeloma cell proliferation, while the phosphorylation of Stat1 and Stat3 are unrelated to multiple myeloma cell growth.<sup>185</sup> Thus, the Raf/Mek/Erk pathway is required for the proliferation of malignant plasma cells in cases in which such cells depend for their growth on interleukin-6.

As Ras is a well-known upstream effector of Raf, it is also likely that the Raf/Mek/Erk pathway contributes to the malignant phenotype in patients with multiple myeloma whose malignant cells express Ras mutations, leading to constitutive Ras activation. Such mutations usually occur in patients with advanced stage disease and involve N-Ras and K-Ras mutations.<sup>172,186,187</sup> However, there is also evidence that, in at least one interleukin-6-independent multiple myeloma cell line with constitutively activated K-Ras, the proliferation of cells was not blocked by the pharmacologic inhibition of the Mek/Erk pathway with PD98059.<sup>188</sup> Thus, although mutations that activate Ras may mediate proliferative signals via Mek/Erk activation in multiple myeloma cells, it is possible that other Ras-dependent, Mek/Erk-independent pathways are activated in certain cases and contribute to the malignant phenotype.

There is accumulating evidence that in addition to interleukin-6, other growth factors also play important roles in the growth of malignant myeloma cells. One such factor is insulin-like growth factor 1 (IGF-1). Recent studies have demonstrated that IGF-1 activates the PI-3K/Akt/FKHRL-1 and Mek/Erk pathways in multiple myeloma cell lines.<sup>189-191</sup> The IGF-1-induced multiple

myeloma cell proliferation is dependent on the activation of PI-3' kinase-dependent pathways, as treatment of cells with the PI-3' kinase inhibitor LY294002 abolishes the IGF-1-dependent proliferative response.<sup>189</sup> On the other hand, the Mek inhibitor PD98059 had no significant effects on the IGF-1-dependent promotion of cell growth of malignant myeloma cells in one study.<sup>189</sup> In studies using growth factor-independent cell lines, it was also demonstrated that the growth of multiple myeloma cells is dependent on PI-3' kinase activation, as reflected by the inhibition of growth of such cells by the PI-3' kinase inhibitors LY294002 and wortmannin.<sup>190</sup> In these studies, Erk inhibition with the PD98059 inhibitor had minimal effects on myeloma cell growth. Based on these facts, a proposed model for the functional contribution of the different signaling pathways in malignant myeloma cell proliferation is that the Mek/Erk pathway is required for IL-6-dependent myeloma cell proliferation. However, this pathway plays a minimal or no role in IGF-1-dependent cell proliferation, in which case the responses are primarily PI-3' kinase/Akt-dependent (Figure 5). It is of interest that some cross-talk between the IGF-1-inducible PI-3' kinase pathway and the Mek/Erk pathway in myeloma cells exists, as evidenced by the inhibition of Mek1/2 and Erk phosphorylation by LY294002.<sup>189</sup> On the other hand such a cross-talk was not seen in other studies using cell lines in which constitutive activation of PI-3'K and Map kinase pathways was found,<sup>187</sup> suggesting that this cross-talk selectively occurs during IGF-1-dependent activation of these pathways. It should also be pointed out that interleukin-6 activates PI-3' kinase and AKT, and the function of PI-3' kinase is required for multiple myeloma cell proliferation but not inhibition of IL-6-dependent Erk activation,<sup>191</sup> indicating that there is no cross-talk between the PI-3K/AKT and Mek/Erk pathways in response to IL-6 induction<sup>189</sup> (Figure 5).

It is well established that vascular endothelial growth factor (VEGF) promotes angiogenesis in various models.<sup>192</sup> It has been previously demonstrated that VEGF is produced in the bone marrows of severe combined immunodeficiency (SCID)-hu mice with multiple myeloma to promote angiogenesis.<sup>193</sup> This finding has strongly suggested that this protein may play an important role in the pathogenesis of multiple myeloma. This has prompted studies to understand the mechanisms of signal transduction of VEGF and the biologic relevance of such signals in multiple myeloma. Recently, it was demonstrated that VEGF activates the Raf/Mek/Erk signaling cascade in a myeloma cell line, as well as in malignant cells from patients with multiple myeloma.<sup>194</sup> Such VEGF activation of the Erk pathway was PKC-independent and was essential for VEGF-induced myeloma cell proliferation.<sup>194</sup> VEGF was also found to induce migration of myeloma cells in an Erk-independent

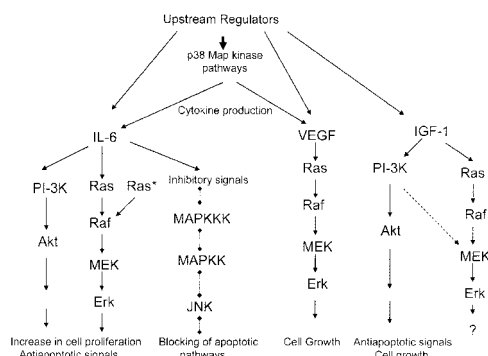
manner, while such a migration was blocked by treatment with a pharmacologic inhibitor of PKC.<sup>194</sup> Thus, in addition to mediating IL-6-dependent cell-proliferative signals, the Mek/Erk pathway promotes VEGF-dependent proliferation, but not migration, of multiple myeloma cells.

Although there is no evidence that the p38 pathway directly promotes the growth of multiple myeloma cells in response to growth factors, there is evidence that it may be doing so indirectly. A recent study demonstrated that a specific p38 Map kinase inhibitor (VX-745) inhibits IL-6 and VEGF secretion in bone marrow stromal cells.<sup>195</sup> Furthermore, it was shown that p38 inhibition blocks proliferation of multiple myeloma cells.<sup>195</sup> Such an effect appears to be the result of inhibition of IL-6 secretion induced by adherence of malignant plasmacytes to bone marrow stromal cells, which is a major mechanism by which these cells develop resistance to chemotherapy.<sup>195</sup> These results strongly suggest that the p38 Map kinase pathway is required for paracrine secretion of IL-6 in multiple myeloma bone marrows and raise the possibility that targeting the p38 Map kinase pathway may have therapeutic implications in the treatment of multiple myeloma.<sup>195</sup>

Map kinase pathways also play important roles in the regulation of apoptosis in malignant myeloma cells. Previous studies have demonstrated that ionizing radiation induces apoptosis of multiple myeloma cells, which is associated with activation of the JNK kinase signaling pathway.<sup>196</sup> On the other hand, corticosteroid (dexamethasone)-induced apoptosis has been found to be JNK-independent, and is associated with a decrease in Erk kinase and p70 S6 kinase activities.<sup>196</sup> It has also been demonstrated that, in response to ionizing irradiation, the JNK kinase associates with the retinoblastoma protein (Rb) and induces its phosphorylation, providing a mechanism by which apoptosis occurs in a JNK-dependent manner in myeloma cells.<sup>197</sup>

In addition to promoting cell proliferation, interleukin-6 inhibits Fas-induced apoptosis of multiple myeloma cells.<sup>198,199</sup> Such effects of interleukin-6 appear to result from inhibition of activation of the JNK kinase pathway.<sup>198,199</sup> Although the precise mechanisms by which interleukin-6 blocks activation of JNK remain to be established, these studies have provided evidence for an additional important mechanism that contributes to the malignant phenotype in multiple myeloma. It is likely that, in addition to inhibition of JNK activation, induction of the PI-3' kinase/Akt and Mek/Erk pathways by interleukin-6 and/or insulin-like growth factor 1 also contributes to the generation of an antiapoptotic state *in vitro*<sup>188-190</sup> and possibly *in vivo*, providing multiple potential therapeutic targets for this disease.

Various new pharmacologic agents are currently under clinical development for the treatment of multiple myeloma,<sup>200</sup> including agents that either block growth-promoting signaling cascades or trigger apoptotic signals in malignant plasma cells. Among the agents shown to induce apoptosis of myeloma cells is the farnesyl transferase inhibitor R115777.<sup>201</sup> Although the precise mechanisms by which such effects are induced remain to be defined, this drug blocks the interleukin-6-dependent phosphorylation of Stat3 and Erk1/2 in multiple myeloma cells, suggesting that it may work, in part, via interruption of IL-6-regulated pathways.<sup>201</sup> A very promising agent in the treatment of multiple myeloma is the proteasome inhibitor PS-341.<sup>202-205</sup> This agent has entered clinical trials and has already been shown to exhibit significant clinical activity in the treatment of patients with refractory multiple myeloma.<sup>205-207</sup> PS-341 induces apoptosis and inhibits the growth of drug-resistant multiple myeloma cells, as well as the binding of multiple myeloma cells to the bone marrow microenvironment.<sup>202-204</sup> Treatment of target cells with PS-341 induces several biochemical changes, including



**Figure 5. Schematic overview of the activation of different Map kinase pathways to regulate malignant plasma cell proliferation and/or inhibit apoptosis of multiple myeloma cells.** \* indicates constitutively activated.



activation of the JNK pathway<sup>204</sup> and inhibition of Erk1/Erk2 kinase-dependent signaling.<sup>202</sup> The precise contributions of different Map kinase pathways to the effects of PS-341 remain to be determined, but it is likely that stimulation of JNK activity and Mek/Erk inhibition contribute to PS-341-induced apoptosis.

The demonstration that Map kinase pathways play important roles in the growth of myeloma cells has provided important insights that may lead to future efforts to combine Map kinase inhibitors with other novel therapeutic agents that inhibit myeloma cell growth. A recent study demonstrated that combined treatment of myeloma cells with the Mek1/2 inhibitor PD184352 and UCN-01, a drug that inhibits Chk1 and abrogates the G<sub>2</sub>M checkpoint in the cell cycle, results in synergistic induction of mitochondrial damage and apoptosis. Such a combined use of these inhibitors also resulted in inhibition of growth of drug-resistant myeloma cells through an interleukin-6-independent mechanism.<sup>208</sup> These findings strongly suggest that simultaneous disruption of the Mek/Erk pathway and the cell cycle may be a more effective approach to target myeloma cells than Mek inhibitors alone, and they have provided an important model for the potential combined use of these inhibitors in clinical trials.<sup>208</sup>

## Conclusions and future directions

Dramatic advances have occurred over the last few years in the research field of Map kinases in hematologic malignancies. The realization that Map kinase-dependent signaling cascades play important roles in the regulation of apoptosis and growth of malignant hematopoietic cells has led to extensive studies, aimed to characterize the precise mechanisms that are responsible for such effects. It is now clear that the Raf/Mek/Erk pathway participates in the generation of mitogenic responses in essentially all hematologic malignancies, including acute and chronic leukemias, lymphomas, and multiple myeloma. On the other hand, the regulatory effects of the JNK and p38 Map kinase pathways vary, depending on the specific cellular type and possibly the distinct isoforms involved. Importantly, the JNK and p38 pathways appear to also mediate signals responsible for sensitivity or resistance to the effects of various pharmacologic and biologic agents currently in use for the treatment of various hematologic malignancies.

The acquired knowledge from all of these efforts has led to the development of specific pharmacologic inhibitors, some of which are now under evaluation in ongoing clinical trials. It is likely that over the next few years we will observe an exponential growth in the number of translational clinical research efforts, resulting from the rapidly accumulating new information. The remarkable scientific and clinical advances in the field of chronic myelogenous leukemia, which ultimately led to the introduction of STI571<sup>209,210</sup> in the treatment of the disease, further increase the enthusiasm toward such efforts. The STI571 paradigm underscores the impor-

tance of understanding in detail the molecular and signaling mechanisms responsible for kinase-dependent malignant cell proliferation and has provided an important model for the development of small molecules that target other kinases involved in the pathogenesis of hematologic malignancies.

The evidence accumulated so far, on the roles that Map kinases play in hematologic malignancies, points toward certain clinical and basic research directions that may be of importance in the development of new therapeutic approaches. Clearly, further characterization of the upstream regulatory signals and the downstream effectors for the different Map kinases will help us to better understand the ways in which Map kinases generate their effects. Also, further delineation of the multiple kinases involved in the different Map kinase group cascades may provide more specific targets for translational approaches. There is also a need to better characterize the functional roles that distinct isoforms within each Map kinase group play in the pathogenesis of various hematologic malignancies. Despite the substantial structural homology among the isoforms in each group, there is evidence that distinct isoforms also exhibit different properties and in some cases may mediate opposing biologic responses. Efforts to define the precise biologic activities mediated by such distinct isoforms in different hematologic malignancies, and to design selective pharmacologic inhibitors that target their distinct kinase domains, may prove valuable in the future. In the same context, very little is known about the functional roles that Erk 3, Erk5, and Erk7 Map kinases may play in the regulation of malignant hematopoiesis. It is possible that these kinases, which do not belong in any of the classic Map kinase groups, also contribute to the pathogenesis of certain hematologic malignancies, and studies in that direction are warranted.

Independently of any new information that will arise from future basic science research efforts, there is already ample evidence to support the further development of clinical studies using pharmacologic inhibitors of known Map kinase pathways. For instance, the design of trials using Mek inhibitors, alone or in combination with cell-cycle inhibitors, for the treatment of leukemias and multiple myeloma is strongly supported from the available preclinical data. Similarly, studies of pharmacologic inhibitors of Bcl-2 in leukemias would be important and may lead to the future design of trials combining Mek inhibitors with Bcl-2 inhibitors for the treatment of refractory acute leukemias. Beyond studies with Mek kinase inhibitors, studies using drugs that block p38 Map kinases are likely to be initiated in multiple myeloma patients, based on very recent studies indicating that p38 promotes multiple myeloma cell growth via paracrine secretion of IL-6 and VEGF. It is also likely that studies of pharmacologic inhibitors of p38, in combination with all-*trans* retinoic acid and/or arsenic trioxide, will be developed for the treatment of acute promyelocytic leukemia, based on already described synergistic interactions. Finally, clinical trials to evaluate the combined use of Mek/Erk inhibitors with traditional chemotherapy for a variety of hematologic neoplasias would be appropriate, and their development is likely in the future.

## References

- Schaeffer HJ, Weber MJ. Mitogen-activated protein kinases: specific messages from ubiquitous messengers. *Mol Cell Biol*. 1999;19:2435-2444.
- English J, Pearson G, Wilsbacher J, et al. New insights into the control of MAP kinase pathways. *Exp Cell Res*. 1999;253:255-270.
- Chang L, Karin M. Mammalian MAP kinase signaling cascades. *Nature*. 2001;410:37-40.
- Dong C, Davis RJ, Flavell RA. MAP kinases in the immune response. *Annu Rev Immunol*. 2002; 20:55-72.
- Davis RJ. Signal transduction by the JNK group of MAP kinases. *Cell*. 2000;103:239-252.
- Rincon M, Flavell RA, Davis RJ. Signal transduction by MAP kinases in T lymphocytes. *Oncogene*. 2001;20:2490-2497.
- Platanias LC. The p38 Map kinase pathway and its role in interferon signaling. *Pharmacol Ther*. In press.
- Hazzalin CA, Mahadevan LC. MAPK-regulated transcription: a continuously variable gene switch? *Nat Rev Mol Cell Biol*. 2002;3:30-40.
- English JM, Cobb MH. Pharmacological inhibitors of MAPK pathways. *Trends Pharmacol Sci*. 2002; 23:40-45.
- Robinson MJ, Cobb MH. Mitogen-activated protein kinase pathways. *Curr Opin Cell Biol*. 1997;9: 180-186.
- Kyriakis JM, Avruch J. Sounding the alarm: protein

- kinase cascades activated by stress and inflammation. *J Biol Chem*. 1996;271:24313-24316.
12. Lee JT Jr, McCubrey JA. The Raf/MEK/ERK signal transduction cascade as a target for chemotherapeutic intervention in leukemia. *Leukemia*. 2002;16:486-507.
  13. Lee JC, Laydon JT, McDonnell PC, et al. A protein kinase involved in the regulation of inflammatory cytokine biosynthesis. *Nature*. 1994;372:739-746.
  14. Han J, Lee JD, Bibbs L, Ulevitch RJ. A MAP kinase targeted by endotoxin and hyperosmolarity in mammalian cells. *Science*. 1994;265:808-811.
  15. Rouse J, Cohen P, Trigon S, et al. A novel kinase cascade triggered by stress and heat shock that stimulates MAPKAP kinase-2 and phosphorylation of small heat shock proteins. *Cell*. 1994;78:1027-1037.
  16. Jiang Y, Chen C, Lie Z, et al. Characterization of the structure and function of a new mitogen-activated protein kinase (p38 $\beta$ ). *J Biol Chem*. 1996;271:17920-17926.
  17. Kumar S, McDonnell PC, Gum RJ, Hand AT, Lee JC, Young PR. Novel homologues of CSBP/p38 Map kinase: activation, substrate specificity and sensitivity to inhibition by pyridinyl imidazoles. *Biochem Biophys Res Commun*. 1997;235:533-538.
  18. Enslin H, Raingeaud J, Davis RJ. Selective activation of p38 mitogen-activated protein (MAP) kinase isoforms by the MAP kinase kinases MKK3 and MKK6. *J Biol Chem*. 1998;273:1741-1748.
  19. Goebert M, Cuenda A, Craxton M, Jakes R, Cohen P. Activation of the novel stress-activated protein kinase SAPK4 by cytokines and cellular stresses is mediated by SKK3 (MKK6); comparison of its substrate specificity with that of other SAP kinases. *EMBO J*. 1997;16:3563-3571.
  20. Stein B, Yand MX, Young DB, et al. P38-2, a novel mitogen-activated protein kinase with distinct properties. *J Biol Chem*. 1997;272:19509-19517.
  21. Lechner C, Zahalka MA, Giot J-F, Moller NPH, Ullrich A. Erk6, a mitogen activated protein kinase involved in C2C12 myoblast differentiation. *Proc Natl Acad Sci U S A*. 1996;93:4355-4359.
  22. Li Z, Jiang Y, Ulevitch RJ, Han J. The primary structure of p38 $\gamma$ : a new member of the p38 group of MAP kinases. *Biochem Biophys Res Commun*. 1996;228:334-340.
  23. Mertens S, Craxton M, Goedert M. SAP kinase-3, a new member of the family of mammalian stress-activated protein kinases. *FEBS Lett*. 1996;383:273-276.
  24. Jiang Y, Gram H, Zhao M, et al. Characterization of the structure and function of the fourth member of p38 mitogen-activated protein kinases group, p38 delta. *J Biol Chem*. 1997;272:30122-30128.
  25. Derijard B, Hibi M, Wu IH, et al. JNK1: a protein kinase stimulated by UV light and Ha-Ras that binds and phosphorylates the c-Jun activation domain. *Cell*. 1994;76:1025-1037.
  26. Sanchez I, Hughes RT, Mayer BJ, et al. Role of SAPK/ERK kinase-1 in the stress activated pathway regulating transcription factor c-Jun. *Nature*. 1994;372:794-798.
  27. Gupta S, Barrett T, Whitmarsh AJ, et al. Selective interaction of JNK protein kinase isoforms with transcription factors. *EMBO J*. 1996;15:2760-2770.
  28. Sluss HK, Barrett T, Derijard B, Davis RJ. Signal transduction by tumor necrosis factor mediated by JNK protein kinases. *Mol Cell Biol*. 1994;14:8376-8384.
  29. Boulton TG, Yancopoulos GD, Gregory JS, et al. An insulin-stimulated protein kinase similar to yeast kinases involved in cell cycle control. *Science*. 1990;249:64-67.
  30. Boulton TG, Nye SH, Robbins DJ, et al. ERKs: a family of protein-serine/threonine kinases that are activated and tyrosine phosphorylated in response to insulin and NGF. *Cell*. 1991;65:663-675.
  31. Zhu AX, Zhao Yi, Moller DE, Flier JS. Cloning and characterization of p97MAPK, a novel human homolog of rat ERK-3. *Mol Cell Biol*. 1994;14:8202-8211.
  32. Turgeon B, Lang BF, Meloche S. The protein kinase ERK3 is encoded by a single functional gene: genomic analysis of the ERK3 gene family. *Genomics*. 2002;80:673-680.
  33. Zhou G, Bao ZQ, Dixon JE. Components of a new human protein kinase signal transduction pathway. *J Biol Chem*. 1995;270:12665-12669.
  34. Lee JD, Ulevitch RJ, Han J. Primary structure of BMK1: a new mammalian MAP kinase. *Biochem Biophys Res Commun*. 1995;213:715-724.
  35. Abe MK, Kuo W, Hershenson MB, Rosner MR. Extracellular signal-regulated kinase 7 (ERK7), a novel ERK with a C-terminal domain that regulates its activity, its cellular localization, and cell growth. *Mol Cell Biol*. 1999;19:1301-1312.
  36. Abe MK, Saelzler MP, Espinosa R III, et al. Erk8, a new member of the mitogen-activated protein kinase family. *J Biol Chem*. 2002;277:16733-16743.
  37. Verma A, Platanias LC. Signaling via the Type I interferon receptor in chronic myelogenous leukemia cells. *Leuk Lymphoma*. 2002;43:703-709.
  38. Young PR. Specific inhibitors of the p38 MAP kinase. In: Gutkind JS, ed. *Signaling Networks and Cell Cycle Control: The Molecular Basis of Cancer and Other Diseases*. Totowa, NJ: Humana Press; 2000:483-500.
  39. Enslin H, Raingeaud J, Davis RJ. Selective activation of p38 mitogen-activated protein (MAP) kinase isoforms by the MAP kinase kinases MKK3 and MKK6. *J Biol Chem*. 1998;273:1741-1748.
  40. Kyriakis JM, App H, Zhang XF, et al. Raf-1 activates MAP kinase-kinase. *Nature*. 1992;358:417-421.
  41. Dent P, Haser W, Haystead TAJ, Vincent LA, Roberts TM, Sturgill TW. Activation of the mitogen-activated protein kinase kinase by v-Raf in NIH 3T3 cells and in vitro. *Science*. 1992;257:1404-1407.
  42. Blank JL, Gerwins P, Elliott EM, Sather S, Johnson GL. Molecular cloning of mitogen-activated protein/ERK kinase kinases (MEKK) 2 and 3: regulation of sequential phosphorylation pathways involving mitogen-activated protein kinase and c-Jun kinase. *J Biol Chem*. 1996;271:5361-5368.
  43. Hirai S, Noda K, Moriguchi T, et al. Differential activation of two JNK activators, MKK7 and SEK1, by MKN28-derived nonreceptor serine/threonine kinase/mixed lineage kinase 2. *J Biol Chem*. 1998;273:7406-7412.
  44. Dorow DS, Devereux L, Tu GF, et al. Complete nucleotide sequence, expression, and chromosomal localisation of human mixed-lineage kinase 2. *Eur J Biochem*. 1995;234:492-500.
  45. Rana A, Gallo K, Godowski P, et al. The mixed lineage kinase SPRK phosphorylates and activates the stress-activated protein kinase activator, SEK-1. *J Biol Chem*. 1996;271:19025-19028.
  46. Tibbles LA, Ing YL, Kiefer F, et al. Mlk-3 activates the SAPK/JNK and p38/RK pathways via SEK1 and MKK3/6. *EMBO J*. 1996;15:7026-7035.
  47. Teramoto H, Coso OA, Miyata H, Igishi T, Miki T, Gutkind JS. Signaling from the small GTP-binding proteins Rac1 and Cdc42 to the c-Jun terminal kinase/stress activated protein kinase pathway: a role for the mixed lineage kinase 3/protein tyrosine kinase 1, a novel member of the mixed lineage kinase family. *J Biol Chem*. 1996;271:27225-27228.
  48. Ichijo H, Nishida E, Irie K, et al. Induction of apoptosis by ASK1, a mammalian MAPKKK that activates SAPK/JNK and p38 signaling pathways. *Science*. 1997;275:90-94.
  49. Yan M, Dai T, Deak JC, et al. Activation of stress-activated protein kinase by MEKK1 phosphorylation of its activator SEK1. *Nature*. 1994;372:798-800.
  50. Minden A, Lin A, McMachon M, et al. Differential activation of ERK and JNK mitogen-activated protein kinases by Raf-1 and MEKK. *Science*. 1994;266:1719-1723.
  51. Yujiri T, Sather S, Fanger GR, Johnson GL. Role of MEKK1 in cell survival and activation of JNK and ERK pathways. *Science*. 1998;282:1911-1914.
  52. Yujiri T, Ware M, Widmann C, et al. MEK kinase 1 gene disruption alters cell cycle migration and c-Jun NH2-terminal kinase regulation but does not cause a measurable defect in NF-kappa B activation. *Proc Natl Acad Sci U S A*. 2000;97:7272-7277.
  53. Lange-Carter CA, Pleiman CM, Gardner AM, Blumer KJ, Johnson GL. A divergence in the MAP kinase regulatory network defined by MEK kinase and Raf. *Science*. 1993;260:315-319.
  54. Salmeron A, Ahmad TB, Garfile GW, Pappin D, Narsimhan RP, Ley SC. Activation of MEK-1 and SEK-1 by Tpl-2 proto-oncoprotein, a novel MAP kinase kinase kinase. *EMBO J*. 1996;15:817-826.
  55. Yamaguchi K, Shirakabe K, Shibuya H, et al. Identification of a member of the MAPKKK family as a potential mediator of TGF-beta signal transduction. *Science*. 1995;270:2008-2011.
  56. Shibuya H, Yamaguchi K, Shirakabe K, et al. TAB1: an activator of the TAK-1 MAPKKK in TGF-beta signal transduction. *Science*. 1996;272:1179-1182.
  57. Gallo KA, Johnson GL. Mixed-lineage kinase control of JNK and p38 MAPK pathways. *Nat Rev*. 2002;3:663-672.
  58. Tanaka S, Hanafusa H. Guanine-nucleotide exchange protein C3G activates JNK1 by a ras-independent mechanism: JNK1 activation inhibited by kinase negative forms of MLK3 and DLK mixed kinases. *J Biol Chem*. 1998;273:1281-1284.
  59. Fan G, Merritt SE, Kortjenann M, Shaw PE, Holzman LB. Dual leukine zipper bearing kinase (DLK) activates p46 SAPK and p38 MAPK but not Erk2. *J Biol Chem*. 1996;271:24788-24793.
  60. Ikeda A, Hasegawa K, Masaki M, et al. Mixed lineage kinase LZK forms a functional signaling complex with JIP-1, a scaffold protein of the c-Jun NH (2)-terminal kinase pathway. *J Biochem*. 2001;130:773-781.
  61. Dumitru CD, Ceci JD, Tsatsanis C, et al. TNF-alpha induction by LPS is regulated posttranscriptionally via a Tpl2/ERK-dependent pathway. *Cell*. 2000;103:1071-1083.
  62. Dhillon AS, Kolch W. Untying the regulation of the Raf-1 kinase. *Arch Biochem Biophys*. 2002;404:3-9.
  63. Zhang S, Han J, Sells MA, et al. Rho family GTPases regulate p38 mitogen-activated protein kinase through the downstream mediator Pak1. *J Biol Chem*. 1995;270:23934-23936.
  64. Frost JA, Xu S, Hutchison MR, Marcus S, Cobb MH. Actions of Rho family small G proteins and p21-activated protein kinases on mitogen-activated protein kinase family members. *Mol Cell Biol*. 1996;16:3707-3713.
  65. Salojin KV, Zhang J, Delovitch TL. TCR and CD28 are coupled via ZAP-70 to the activation of the Vav/Rac-1/PAK-1/p38 MAPK signaling pathway. *J Immunol*. 1999;163:844-853.
  66. Bar-Sagi D, Hall A. Ras and Rho GTPases: a family reunion. *Cell*. 2000;103:227-238.
  67. Takai Y, Sasaki T, Matozaki T. Small GTP-binding proteins. *Physiol Rev*. 2001;81:153-208.
  68. Ridley AJ. Rho family proteins: coordinating cell responses. *Trends Cell Biol*. 2001;11:471-477.

69. Weston CR, Lambright DG, Davis RJ. Map kinase signaling specificity. *Science*. 2002;296:2345-2347.
70. Enslin H, Davis RJ. Regulation of MAP kinases by docking domains. *Biol Cell*. 2001;93:5-14.
71. Sharrocks AD, Yang SH, Galanis A. Docking domains and substrate-specificity determination for MAP kinases. *Trends Biochem Sci*. 2000;25:448-453.
72. Chang CI, Xu BE, Akella R, Cobb MH, Goldsmith EJ. Crystal structures of MAP kinase p38 complexed to the docking sites on its nuclear substrate MEF2A and activator MKK3b. *Mol Cell*. 2002;9:1241-1249.
73. Nagata Y, Moriguchi T, Nishida E, Tokodoro K. Activation of p38 Map kinase pathway by erythropoietin and interleukin-3. *Blood*. 1997;90:929-934.
74. Nosaka Y, Arai A, Miyasaka N, Miura O. CrkL mediates Ras-dependent activation of the Raf/ERK pathway through the guanine nucleotide exchange factor C3G in hematopoietic cells stimulated with erythropoietin or interleukin-3. *J Biol Chem*. 1999;274:30154-30162.
75. Nagata Y, Nishida E, Tokodoro K. Activation of JNK signaling pathway by erythropoietin, thrombopoietin, and interleukin-3. *Blood*. 1997;89:2664-2669.
76. Arai A, Kanda E, Miura O. Rac is activated by erythropoietin or interleukin-3 and is involved in activation of the Erk signaling pathway. *Oncogene*. 2002;21:2641-2651.
77. Jacobs-Helber SM, Penta K, Sun Z, Lawson A, Sawyer ST. Distinct signaling from stem cell factor and erythropoietin in HCD57 cells. *J Biol Chem*. 1997;272:6850-6853.
78. Nagata Y, Takahashi N, Davis RJ, Tokodoro K. Activation of p38 MAP kinase and JNK but not ERK is required for erythropoietin-induced erythroid differentiation. *Blood*. 1998;92:1859-1869.
79. Sui X, Krantz SB, You M, Zhao Z. Synergistic activation of MAP kinase (ERK1/2) by erythropoietin and stem cell factor is essential for expanded erythropoiesis. *Blood*. 1998;92:1142-1149.
80. Jacobs-Heller SM, Ryan JJ, Sawyer ST. JNK and p38 are activated by erythropoietin (EPO) but are not induced in apoptosis following withdrawal in EPO-dependent HCD57 cells. *Blood*. 2000;96:933-940.
81. Okuda K, Sanghera JS, Pelech SL, et al. Granulocyte-macrophage colony-stimulating factor, interleukin-3 and steel factor induce rapid tyrosine phosphorylation of p42 and p44 MAP kinase. *Blood*. 1992;79:2880-2887.
82. Foltz IN, Lee JC, Young PR, Schrader JW. Hemopoietic growth factors with the exception of interleukin-4 activate the p38 mitogen-activated protein kinase pathway. *J Biol Chem*. 1997;272:3296-3301.
83. Foltz IN, Schrader JW. Activation of the stress-activated protein kinases by multiple hematopoietic growth factors with the exception of interleukin-4. *Blood*. 1997;89:3092-3096.
84. Ronjuckarin P, Miyakawa Y, Fox NE, et al. The roles of phosphatidylinositol 3-kinase and protein kinase C zeta for thrombopoietin-induced mitogen-activated protein kinase activation in primary murine megakaryocytes. *J Biol Chem*. 2001;276:41014-41022.
85. Miyazaki R, Ogata H, Kobayashi Y. Requirement of thrombopoietin-induced activation of ERK for megakaryocyte differentiation and of p38 for erythroid differentiation. *Ann Hematol*. 2001;80:284-291.
86. Ronjuckarin P, Drachman JG, Kaushansky K. Thrombopoietin-induced activation of the mitogen-activated protein kinase (MAPK) pathway in normal megakaryocytes: roles in endomitosis. *Blood*. 1999;94:2676-2685.
87. Filippi M-D, Porteu F, Le Pesteur F, et al. Requirement for mitogen-activated protein kinase activation in the response of embryonic stem cell-derived hematopoietic cells to thrombopoietin *in vitro*. *Blood*. 2002;99:1174-1182.
88. Tamura K, Suto T, Senfleben U, et al. Requirement for p38alpha in erythropoietin expression: a role for stress kinases in erythropoiesis. *Cell*. 2000;102:221-231.
89. Adams RH, Porras A, Alonso G, et al. Essential role of p38alpha MAP kinase in placental but not embryonic cardiovascular development. *Mol Cell*. 2000;6:109-116.
90. Mudgett JS, Ding J, Guh-Siesel L, et al. Essential role for p38alpha mitogen-activated protein kinase in placental angiogenesis. *Proc Natl Acad Sci U S A*. 2000;97:10454-10459.
91. Allen M, Svensson L, Roach M, Hambor J, McNeish J, Gabel CA. Deficiency of the stress kinase p38alpha results in embryonic lethality: characterization of the kinase dependence of stress responses of enzyme-deficient embryonic stem cells. *J Exp Med*. 2000;191:859-870.
92. Platanius LC, Fish EN. Signaling pathways activated by interferons. *Exp Hematol*. 1999;27:1583-1592.
93. Platanius LC. Interferons: laboratory to clinic investigations. *Curr Opin Oncol*. 1995;7:560-565.
94. Broxmeyer HE, Lu L, Platzer E, et al. Comparative analysis of the influences of human gamma, alpha and beta interferons on human multipotential (CFU-GEMM), erythroid (BFU-E) and granulocyte-macrophage (CFU-GM) progenitor cells. *J Immunol*. 1983;131:1300-1305.
95. Raefsky EL, Platanius LC, Zoumbos NC, Young NS. Studies of interferon as a regulator of hematopoietic cell proliferation. *J Immunol*. 1985;135:2507-2512.
96. Broxmeyer HE, Cooper S, Rubin BY, Taylor MW. The synergistic influence of human interferon-gamma and interferon-alpha on suppression of hematopoietic progenitor cells is additive with the enhanced sensitivity of these cells to inhibition by interferons at low oxygen tension *in vitro*. *J Immunol*. 1985;135:2502-2506.
97. Delforge A, Vandenplas B, Lagenaux L, et al. Influence of recombinant alpha and gamma interferons on the *in vitro* proliferation of myeloid and leukemic progenitors. *Eur J Haematol*. 1990;44:307-311.
98. Ganser A, Carlo-Stella C, Greher J, Volkens B, Hoelzer D. Effect of recombinant interferons alpha and gamma on human bone marrow-derived megakaryocytic progenitor cells. *Blood*. 1987;70:1173-1179.
99. Gugliotta L, Bagnara GP, Catani L, et al. *In vivo* and *in vitro* inhibitory effect of alpha-interferon on megakaryocyte colony growth in essential thrombocythaemia. *Br J Haematol*. 1989;71:177-181.
100. Means RT Jr, Krantz SB. Inhibition of human erythroid colony-forming units by tumor necrosis factor requires beta interferon. *J Clin Invest*. 1993;91:416-419.
101. David M, Petricoin E, Benjamin C, et al. Requirement for MAP kinase (ERK2) activity in interferon  $\alpha$ - and interferon  $\beta$ -stimulated gene expression through STAT proteins. *Science*. 1995;269:1721-1723.
102. Uddin S, Fish EN, Sher DA, Gardziola C, White MF, Platanius LC. Activation of the phosphatidylinositol 3'-kinase serine kinase by IFN $\alpha$ . *J Immunol*. 1997;158:2390-2397.
103. Uddin S, Mazchrzak B, Woodson J, et al. Activation of the p38 Map kinase by Type I IFNs. *J Biol Chem*. 1999;274:30127-30131.
104. Uddin S, Sharma N, Majchrzak B, et al. The Rac1/p38 Map kinase pathway is required for IFN $\alpha$ -dependent transcriptional activation but not serine phosphorylation of Stat-proteins. *J Biol Chem*. 2000;275:27634-27640.
105. Verma A, Deb DK, Sassano A, et al. Activation of the p38 Map kinase pathway mediates the suppressive effects of Type I interferons and trans-forming growth factor beta on normal hematopoiesis. *J Biol Chem*. 2002;277:7726-7735.
106. Verma A, Deb DK, Sassano A, et al. Cutting edge: activation of the p38 mitogen-activated protein kinase signaling pathway mediates cytokine-induced hematopoietic suppression in aplastic anemia. *J Immunol*. 2002;168:5984-5988.
107. Young NS, Maciejewski J. The pathophysiology of acquired aplastic anemia. *N Engl J Med*. 1997;336:1365-1372.
108. Young NS. Hematopoietic cell destruction by immune mechanisms in acquired aplastic anemia. *Semin Hematol*. 2000;37:3-14.
109. Tanaka T, Kyrokawa M, Ueki K, et al. The extracellular signal-regulated kinase pathway phosphorylates AML1, an acute myeloid leukemia gene product and potentially regulates its trans-activation ability. *Mol Cell Biol*. 1996;16:3967-3979.
110. Blalock WL, Pearce M, Steelman LS, et al. A conditionally-active form of MEK1 results in autocrine transformation of human and mouse hematopoietic cells. *Oncogene*. 2000;19:526-536.
111. Hoyle PE, Moyer PW, Steelman LS, et al. Differential abilities of the Raf family of protein kinases to abrogate cytokine dependency and prevent apoptosis in murine hematopoietic cells by a MEK1-dependent mechanism. *Leukemia*. 2000;14:642-656.
112. Blalock WL, Moyer PW, Chang F, et al. Combined effects of aberrant MEK1 activity and BCL2 overexpression on relieving the cytokine dependency of human and murine hematopoietic cells. *Leukemia*. 2000;14:1080-1096.
113. Towatari M, Iida H, Tanimoto M, et al. Constitutive activation of mitogen-activated protein kinase pathway in acute leukemia cells. *Leukemia*. 1997;11:479-484.
114. Kim SC, Hahn JS, Min YH, et al. Constitutive activation of extracellular signal-regulated kinase in human acute leukemias: combined role of activation of MEK, hyperexpression of extracellular signal-regulated kinase, and downregulation of phosphatase, PAC1. *Blood*. 1999;93:3893-3899.
115. Milella M, Kornblau SM, Estrov Z, et al. Therapeutic targeting of the MEK/MAPK signal transduction module in acute myeloid leukemia. *J Clin Invest*. 2001;108:851-859.
116. Morgan MA, Dolp O, Reuter CW. Cell-cycle-dependent activation of mitogen-activated protein kinase kinase (MEK-1/2) in myeloid leukemia cell lines and induction of growth inhibition and apoptosis by inhibitors of RAS signaling. *Blood*. 2001;97:1823-1834.
117. Ajenjo N, Aaronson DS, Ceballos E, et al. Myeloid leukemia cell growth and differentiation are independent of mitogen-activated protein kinase ERK1/2 activation. *J Biol Chem*. 2000;275:7189-7197.
118. Laughi P, Tabilio A, Pinelli S, et al. Expression and activation of SHC/MAP kinase pathway in primary acute leukemia blasts. *Hematol J*. 2001;2:70-80.
119. Jarvis WD, Fornari FA Jr, Tombs RM. Evidence for involvement of mitogen-activated protein kinase, rather than stress-activated protein kinase, in potentiation of 1-beta-D-arabinofuranosylcytosine-induced apoptosis by interruption of protein kinase C signaling. *Mol Pharmacol*. 1998;54:844-856.
120. Yu C, Wang S, Dent P, Grant S. Sequence-dependent potentiation of placlitazell-mediated apoptosis in human leukemia cells by inhibitors of the mitogen-activated protein kinase kinase/mitogen-activated protein kinase pathway. *Mol Pharmacol*. 2001;60:143-154.
121. Dent P, Jarvis WD, Birrer MJ, et al. The roles of signaling by the p42/p44 mitogen-activated protein (MAP) kinase pathway; a potential route to radio- and chemo-sensitization of tumor cells resulting in the induction of apoptosis and loss of clonogenicity. *Leukemia*. 1998;12:1843-1850.

122. Reuter CWM, Morgan MA, Bergmann L. Targeting the Ras signaling pathway: a rational, mechanism-based treatment for hematologic malignancies? *Blood*. 2000;96:1655-1669.
123. Bos JL, Verlaan-de-Vries M, van-Der-Emb AJ, et al. Mutations in N-Ras predominate in acute myeloid leukemia. *Blood*. 1987;69:1237-1241.
124. Farr CJ, Saiki RK, Erlich HA, McCormick F, Marshall CJ. Analysis of Ras gene mutations in acute myeloid leukemia by polymerase chain reaction and oligonucleotide probes. *Proc Natl Acad Sci U S A*. 1988;85:1629-1633.
125. Iida M, Towatari M, Nakao A, et al. Lack of constitutive activation of MAP kinase pathway in human acute myeloid leukemia cells with N-Ras mutation. *Leukemia*. 1999;13:585-589.
126. Karp JE, Lancet JE, Kaufmann SH, et al. Clinical and biologic activity of the farnesyltransferase inhibitor R115777 in adults with refractory and relapsed acute leukemias: a phase I clinical-laboratory correlative trial. *Blood*. 2001;97:3361-3369.
127. Milella M, Estrov Z, Kornblau SM, et al. Synergistic induction of apoptosis by simultaneous disruption of the Bcl-2 and MEK/MAPK pathways in acute myelogenous leukemia. *Blood*. 2002;99:3461-3464.
128. Groffen J, Stephenson JR, Heisterkamp N, et al. Philadelphia chromosomal breakpoints are clustered within a limited region, *bcr*, on chromosome 22. *Cell*. 1984;36:93-99.
129. Daley GQ, Van Etten RA, Baltimore D. Blast crisis in a murine model of chronic myelogenous leukemia. *Science*. 1990;247:824-830.
130. Lugo TC, Pendergast AM, Muller AJ, Witte ON. Tyrosine kinase activity and transformation potency of *bcr-abl* oncogene products. *Science*. 1990;247:1079-1082.
131. Cortez D, Reuther G, Pendergast AM. The *Bcr-Abl* tyrosine kinase activates mitogenic signaling pathways and stimulates G1-to-S phase transition in hematopoietic cells. *Oncogene*. 1997;15:2333-2342.
132. Voss J, Posern G, Hannermann JR, et al. The leukaemic oncoproteins *Bcr-Abl* and *Tel-Abl* (ETV6/*Abl*) have altered substrate preferences and activate similar intracellular signalling pathways. *Oncogene*. 2000;19:1684-1690.
133. Zou X, Calame K. Signaling pathways activated by oncogenic forms of *Abl* tyrosine kinase. *J Biol Chem*. 1999;274:18141-18144.
134. Dan S, Naito M, Tsuruo T. Selective inhibition of apoptosis in Philadelphia chromosome-positive chronic myelogenous leukemia cells by an inhibitor of *BCR-ABL* tyrosine kinase. *Cell Death Diff*. 1998;5:710-715.
135. Kang CD, Yoo SD, Hwang BW, et al. The inhibition of ERK/MAPK not the activation of JNK/SAPK is primarily required to induce apoptosis in chronic myelogenous leukemic K562 cells. *Leuk Res*. 2000;24:527-534.
136. Yu C, Krystal G, Varticovski L, et al. Pharmacologic mitogen-activated protein/extracellular signal-regulated kinase kinase/mitogen-activated protein kinase inhibitors interact synergistically with ST1571 to induce apoptosis in *Bcr/Abl*-expressing human leukemia cells. *Cancer Res*. 2002;62:188-199.
137. Barragan M, Bellosillo B, Campas C, et al. Involvement of protein kinase C and phosphatidylinositol 3 kinase pathways in the survival of B-cell chronic lymphocytic leukemia cells. *Blood*. 2002;15:2969-2976.
138. Ringhausen I, Schneller F, Bogner C, et al. Constitutively activated phosphatidylinositol 3 kinase (PI-3K) is involved in the defect of apoptosis in B-CLL: association with protein kinase C delta. *Blood*. 2002;100:3741-3748.
139. Kawachi K, Ogasawara T, Yasuyama M. Activation of extracellular signal-regulated kinase through B-cell antigen receptor in B-cell chronic lymphocytic leukemia. *Int J Hematol*. 2002;75:508-513.
140. Epling-Burnette PK, Bai F, Painter S, et al. Survival signals of leukemic NK cells depend on the activation of MAPK and Ras [abstract]. *Blood*. 2002;100:732a.
141. Cripe LD, Gelfanov VM, Smith EA, et al. Role for c-jun N-terminal kinase in treatment-refractory acute myeloid leukemia (AML): signaling to multi-drug-efflux and hyperproliferation. *Leukemia*. 2002;16:799-812.
142. Laurent G, Blau CA. Signaling pathways activated by daunorubicin. *Blood*. 2001;97:2535-2540.
143. Stadheim TA, Kucera GL. c-Jun N-terminal kinase/stress-activated protein kinase (JNK/SAPK) is required for mitoxantrone- and anisomycin-induced apoptosis in HL-60 cells. *Leuk Res*. 2002;26:55-65.
144. Yu R, Shtil AA, Tan TH, Roninson IB, Kong AN. Adriamycin activates c-jun N-terminal kinase in human leukemia cells: a relevance to apoptosis. *Cancer Lett*. 1996;107:73-81.
145. Alsayed Y, Uddin S, Mahmud N, et al. Activation of Rac1 and the p38 Map kinase pathway in response to all-*trans*-retinoic acid. *J Biol Chem*. 2001;272:4012-4019.
146. Miranda MB, McGuire TF, Johnson DE. Importance of MEK-1/-2 signaling in monocytic and granulocytic differentiation of myeloid cell lines. *Leukemia*. 2002;16:683-692.
147. Yen A, Roberson MS, Varvayanis S, Lee T. Activation of mitogen-activated protein (MAP)/extracellular signal-regulated kinase (ERK)-dependent MAP kinase activation needed to elicit HL-60 cell differentiation and growth arrest. *Cancer Res*. 1998;58:3163-3172.
148. Tallman MS, Nabhan C, Feusner JH, Rowe JM. Acute promyelocytic leukemia: evolving therapeutic strategies. *Blood*. 2002;99:759-767.
149. Verma A, Mohindru A, Deb DK, et al. Activation of Rac1 and the p38 Map kinase pathway in response to arsenic trioxide. *J Biol Chem*. 2002;277:44988-44995.
150. Srinivasa SP, Doshi PD. Extracellular signal-regulated kinase and p38 mitogen-activated protein kinase pathways cooperate in mediating cytokine-induced proliferation of a leukemic cell line. *Leukemia*. 2002;16:244-253.
151. Korus M, Mahon GM, Cheng L, Whitehead IP. p38 MAPK-mediated activation of NFκB by the RhoGEF domain of *Bcr*. *Oncogene*. 2002;21:4601-4612.
152. Mayer IA, Verma A, Grumbach IM, et al. The p38 MAPK pathway mediates the growth inhibitory effects of interferon-α in *BCR-ABL*-expressing cells. *J Biol Chem*. 2001;276:28570-28577.
153. Witt O, Sand K, Pekrun A. Butyrate-induced erythroid differentiation of human K562 leukemia cells involves inhibition of ERK and activation of p38 MAP kinase pathways. *Blood*. 2000;95:2391-2396.
154. Raitano AB, Halpern JR, Hambuch TM, Sawyers CL. The *Bcr-Abl* leukemia oncogene activates Jun kinase and requires Jun for transformation. *Proc Natl Acad Sci U S A*. 1995;92:11746-11750.
155. Burgess GS, Williamson EA, Cripe LD, et al. Regulation of the c-jun gene in p210 *BCR-ABL* transformed cells corresponds with activity of JNK, the c-jun N-terminal kinase. *Blood*. 1998;92:2450-2460.
156. Dickens M, Rogers JS, Cavanagh J, et al. A cytoplasmic inhibitor of the JNK signal transduction pathway. *Science*. 1997;277:693-696.
157. Hess P, Pihan G, Sawyers CL, Flavell RA, Davis RJ. Survival signaling mediated by c-Jun NH(2)-terminal kinase in transformed B lymphoblasts. *Nat Genet*. 2002;32:201-205.
158. Xu X, Heidenreich O, Kitajima I, et al. Constitutively activated JNK is associated with HTLV-1 mediated tumorigenesis. *Oncogene*. 1996;13:135-142.
159. Ng PW, Iha H, Iwanaga Y, et al. Genome-wide expression changes induced by HTLV-1 Tax: evidence for MLK-3 mixed lineage kinase involvement in Tax-mediated NF-κB activation. *Oncogene*. 2001;20:4484-4496.
160. Pedersen IM, Buhl AM, Klausen P, Geisler CH, Jurlander J. The chimeric anti-CD20 antibody rituximab induces apoptosis in B-cell chronic lymphocytic leukemia cells through a p38 mitogen activated protein-kinase-dependent mechanism. *Blood*. 2002;99:1314-1319.
161. Liu RY, Fan C, Liu G, Olshaw NE, Zuckerman KS. Activation of p38 mitogen-activated protein kinase is required for tumor necrosis factor-α-supported proliferation of leukemia and lymphoma cell lines. *J Biol Chem*. 2000;275:21086-21093.
162. Giri DK, Aggarwal BB. Constitutive activation of NF-κB causes resistance to apoptosis in human cutaneous T cell lymphoma Hut-78 cells: autocrine role of tumor necrosis factor and reactive oxygen intermediates. *J Biol Chem*. 1998;273:14008-14014.
163. O'Connell MA, Cleere R, Long A, O'Neill LA, Kelleher D. Cellular proliferation and activation of NF-κB are induced by autocrine production of tumor necrosis factor alpha in the human T lymphoma line HuT 78. *J Biol Chem*. 1995;270:7399-7404.
164. Kotlyarov A, Neininger A, Scubert C, et al. MAPKAP kinase 2 is essential for LPS-induced TNF-α biosynthesis. *Nat Cell Biol*. 1999;1:94-97.
165. Kalas W, Kisielow P, Strzadala L. Inhibition of MEK induces fas expression and apoptosis in lymphomas overexpressing Ras. *Leuk Lymphoma*. 2002;43:1469-1474.
166. Vockerodt M, Haier B, Buttgerit P, Tesch H, Kube D. The Epstein-Barr virus latent membrane protein 1 induces interleukin-10 in Burkitt's lymphoma cells but not in Hodgkin's cells involving the p38/SAPK2 pathway. *Virology*. 2001;283:198.
167. Moore KW, de Waal Malefyt R, Coffmann RL, O'Garra A. Interleukin-10 and the interleukin-10 receptor. *Annu Rev Immunol*. 2001;19:683-765.
168. Chen SY, Lu J, Shih YC, Tsai CH. Epstein-Barr virus latent membrane protein 2A regulates c-Jun protein through extracellular signal-regulated kinase. *J Virol*. 2002;76:9556-9561.
169. Mathas S, Hinz M, Anagnostopoulos I, et al. Aberrantly expressed c-Jun and JunB are a hallmark of Hodgkin lymphoma cells, stimulate proliferation and synergize with NF-κB. *EMBO J*. 2002;21:4104-4113.
170. Mukhopadhyay A, Fiumara P, Li Y. Receptor activator of nuclear factor-κB ligand activates mitogen-activated protein kinases signaling pathways in Hodgkin and Reed-Sternberg cells [letter]. *Blood*. 2002;99:3485-3486.
171. Fiumara P, Snell V, Li Y, et al. Functional expression of receptor activator of nuclear factor kappa B in Hodgkin disease cell lines. *Blood*. 2001;98:2784-2790.
172. Hallek M, Bergsagel PL, Anderson KC. Multiple myeloma: increasing evidence for a multistep transformation process. *Blood*. 1998;91:3-21.
173. Pratt G. Molecular aspects of multiple myeloma. *J Clin Pathol Mol Pathol*. 2002;55:273-283.
174. Klein B, Zhang XG, Lu ZY, et al. Interleukin-6 in human multiple myeloma. *Blood*. 1995;85:863-872.
175. Klein B, Zhang XG, Jourdan M, et al. Paracrine rather than autocrine regulation of myeloma-cell growth and differentiation by interleukin-6. *Blood*. 1989;73:517-526.
176. Klein B. Cytokine, cytokine receptors and transduction signals, and oncogenes in multiple myeloma. *Semin Hematol*. 1995;32:4-19.

177. Kawano M, Hirano T, Matsuda T, et al. Autocrine generation and requirement of BSF-2/IL-6 for human multiple myeloma. *Nature*. 1988;332:83-85.
178. Zhang XG, Bataille R, Widjenes J, Klein B. Interleukin-6 dependence of advanced malignant plasma cell dyscrasia. *Cancer*. 1992;69:1373-1376.
179. Bataille R, Jourdan M, Zhang XG, Klein B. Serum levels of interleukin 6, a potent myeloma cell growth factor, as a reflect of disease severity in plasma cell dyscrasias. *J Clin Invest*. 1989;84:2008-2011.
180. Hilbert D, Kopf M, Mock B, Kohler G, Rudicoff S. Interleukin 6 is essential for in vivo development of B lineage neoplasms. *J Exp Med*. 1995;182:243-248.
181. Bataille R, Barlogie B, Lu ZY, et al. Biologic effects of anti-interleukin 6 murine monoclonal antibody in advanced multiple myeloma. *Blood*. 1995;86:685-691.
182. Georgi-Hemming P, Wiklund HJ, Ljungren O, Nilsson K. Insulin-like growth factor I is a growth and survival factor in human multiple myeloma cell lines. *Blood*. 1996;88:2250-2258.
183. Jelinek DF, Witzig TE, Arendt BK. A role for insulin-like growth factor in the regulation of IL-6 responsive human myeloma cell line growth. *J Immunol*. 1997;159:487-496.
184. Ge NL, Rudikoff S. Insulin-like growth factor I is a dual effector of multiple myeloma cell growth. *Blood*. 2000;96:4091-4095.
185. Ogata A, Chauhan D, Urashima M, et al. IL-6 triggers multiple myeloma cell growth via the Ras dependent mitogen activated protein kinase cascade. *J Immunol*. 1997;159:2212-2220.
186. Corradini P, Ladetto M, Voena C, et al. Mutational activation of N- and K-ras mutations in plasma cell dyscrasias. *Blood*. 1993;81:2708-2713.
187. Liu P, Leong T, Quam L, et al. Activating mutations of N- and K-ras oncogenes in multiple myeloma show different clinical associations: analysis of the Eastern Cooperative Oncology Group phase III trial. *Blood*. 1996;88:2699-2706.
188. Zhang B, Fenton RG. Proliferation of IL-6-independent multiple myeloma does not require the activity of extracellular signal-regulated kinases (ERK1/2). *J Cell Physiol*. 2002;193:42-54.
189. Qiang Y-W, Kopantzev E, Rudicoff S. Insulinlike growth factor-I signaling in multiple myeloma: downstream elements, functional correlates, and pathway cross-talk. *Blood*. 2002;99:4138-4146.
190. Pene F, Claessens YE, Muller O, et al. Role of the phosphatidylinositol 3-kinase/Akt and mTOR/p70S6-kinase pathways in the proliferation and apoptosis in multiple myeloma. *Oncogene*. 2002;21:6587-6597.
191. Tu Y, Gardner A, Lichtenstein A. The phosphatidylinositol 3-kinase/AKT kinase pathway in multiple myeloma plasma cells: role of cytokine-dependent survival and proliferative responses. *Cancer Res*. 2000;60:6763-6770.
192. Neufeld G, Cohen T, Gengrinovitch S, Poltorak Z. Vascular endothelial growth factor (VEGF) and its receptors. *FASEB J*. 1999;13:9-22.
193. Yaccoby S, Barlogie B, Epstein J. Primary myeloma cells growing in SCID-hu mice: a model for studying the biology and treatment of myeloma and its manifestation. *Blood*. 1998;92:2908-2913.
194. Podar K, Tal Y-T, Davies FE, et al. Vascular endothelial growth factor triggers signaling cascades mediating multiple myeloma cell growth and migration. *Blood*. 2001;98:428-435.
195. Hideshima T, Akiyama M, Hayashi T, et al. Targeting p38 MAPK inhibits multiple myeloma cell growth in the bone marrow milieu. *Blood*. 2002;101:703-705.
196. Chauhan D, Pandey P, Ogata A, et al. Dexamethasone induces apoptosis of multiple myeloma cells in a JNK/SAP kinase independent mechanism. *Oncogene*. 1997;15:837-843.
197. Chauhan D, Hideshima T, Treon T, et al. Functional interaction between retinoblastoma protein and stress-activated protein kinases in multiple myeloma cells. *Cancer Res*. 1999;59:1192-1195.
198. Chauhan D, Kharbanda S, Ogata A, et al. Interleukin-6 inhibits Fas-induced apoptosis and stress-activated protein kinase activation in multiple myeloma cells. *Blood*. 1997;89:227-234.
199. Xu F-H, Sharma S, Gardner A, et al. Interleukin-6-induced inhibition of multiple myeloma cell apoptosis: support for the hypothesis that protection is mediated via inhibition of JNK/SAPK pathway. *Blood*. 1998;92:241-251.
200. Anderson KC. Targeted therapy for multiple myeloma. *Semin Hematol*. 2001;38:286-294.
201. Le Couill S, Pellat-Deceunynck C, Harousseau J-L, et al. Farnesyl transferase inhibitor R115777 induces apoptosis of human myeloma cells. *Leukemia*. 2002;16:1664-1667.
202. Hideshima T, Richardson P, Chauhan D, et al. The proteasome inhibitor PS-341 inhibits growth, induces apoptosis, and overcomes drug resistance in human multiple myeloma cells. *Cancer Res*. 2001;61:3071-3076.
203. LeBlanc R, Catley LP, Hideshima T, et al. Proteasome inhibitor PS-341 inhibits human myeloma cell growth in vivo and prolongs survival in a murine model. *Cancer Res*. 2002;62:4996-5000.
204. Mitsiades N, Mitsiades CS, Richardson PG, et al. The proteasome inhibitor PS-341 potentiates sensitivity of multiple myeloma cells to conventional chemotherapeutic agents: therapeutic applications. *Blood*. Prepublished on November 7, 2002, as DOI 10.1182/blood-2002-06-1768.
205. Orłowski RZ, Stinchcombe TE, Mitchell BS, et al. Phase I trial of the proteasome inhibitor PS-341 in patients with refractory hematologic malignancies. *J Clin Oncol*. 2002;20:4420-4427.
206. Richardson P, Barlogie B, Berenson J, et al. A phase II multicenter study of the proteasome inhibitor bortezomib (VELCADE, formerly PS-341) in multiple myeloma patients with relapsed/refractory disease [abstract]. *Blood*. 2002;100:104a.
207. Zangari M, Barlogie B, Prather J, et al. Marked activity also in del 13 multiple myeloma of PS 341 and subsequent thalidomide in a setting of resistance to post-autotransplant salvage therapies [abstract]. *Blood*. 2002;100:105a.
208. Dai Y, Landowski TA, Rosen ST, Dent P, Grant S. Combined treatment with the checkpoint abrogator UCN-01 and MEK1/2 inhibitors potently induces apoptosis in drug-sensitive and -resistant myeloma cells through an IL-6-independent mechanism. *Blood*. 2002;100:3333-3343.
209. Goldman JM, Druker BJ. Chronic myeloid leukemia: current treatment options. *Blood*. 2001;98:2039-2042.
210. Druker BJ. Perspectives in the development of a molecularly targeted agent. *Cancer Cell*. 2002;1:31-36.