

Transcription Factors Runx1 to 3 Are Expressed in the Lacrimal Gland Epithelium and Are Involved in Regulation of Gland Morphogenesis and Regeneration

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PURPOSE. Lacrimal gland (LG) morphogenesis and repair are regulated by a complex interplay of intrinsic factors (e.g., transcription factors) and extrinsic signals (e.g., soluble growth/signaling factors). Many of these interconnections remain poorly characterized. Runx-related (Runx) factors belong to a small family of heterodimeric transcription factors known to regulate lineage-specific proliferation and differentiation of stem cells. The purpose of this study was to define the expression pattern and the role of Runx proteins in LG development and regeneration.

METHODS. Expression of epithelial-restricted transcription factors in murine LG was examined using immunostaining, qRT-PCR, and RT²Profiler PCR microarrays. The role of Runx transcription factors in LG morphogenesis was studied using siRNA and ex vivo LG cultures. Expression of Runx transcription factors during LG regeneration was assessed using in vivo model of LG regeneration.

RESULTS. We found that Runx factors are expressed in the epithelial compartment of the LG; in particular, Runx1 was restricted to the epithelium with highest level of expression in ductal and centroacinar cells. Downregulation of Runx1 to 3 expression using Runx-specific siRNAs abolished LG growth and branching and our data suggest that Runx1, 2, and 3 are partially redundant in LG development. In siRNA-treated LG, reduction of branching correlated with reduction of epithelial proliferation, as well as expression of cyclin D1 and the putative epithelial progenitor cell marker cytokeratin-5. Runx1, Runx3, and cytokeratin-5 expression increased significantly in regenerating LG and there was modest increase in Runx2 expression during LG differentiation.

CONCLUSIONS. Runx1 and 2 are new markers of the LG epithelial lineage and Runx factors are important for normal LG morphogenesis and regeneration.

Keywords: lacrimal gland regeneration, stem and progenitor cells, Runx1, Runx2, qRT PCR microarrays, Runx3

Runx-related (Runx) factors belong to a small family of heterodimeric transcription factors that control stem cell fate and regulate lineage-specific proliferation and differentiation.¹ All Runx factors bear a conserved DNA-binding and heterodimerization domain²; however, they have different expression patterns and roles in tissue morphogenesis and homeostasis. Mouse genetic studies show that *Runx1* (also known as AML1/Cbfa2) is essential for hematopoiesis,^{3,4} *Runx2* (also known as AML3/Cbfa1) is required for osteogenesis,⁵ and *Runx3* (also known as AML2/Cbfa3) is involved in gut development, neurogenesis, and lung alveolar differentiation.^{6–9} Runx proteins can act as repressors or activators depending on the cellular context.^{10,11} Runx proteins also contribute significantly to the transduction of fibroblast growth factor (FGF), Notch, transforming growth factor β , and Wnt

signals,^{12–15} which control stem cell function and tissue regeneration.

Recent reports indicate that Runx proteins play a role in regulation of stem cells in epithelial derivatives.^{16–18} In particular, Runx1 to 3 are expressed in hair follicles where they regulate morphogenesis and stem cell survival.^{19–22} Runx1 is involved in regulation of epithelial cell adhesion, migration, and epithelial-mesenchymal cross talk.^{16,23}

The expression and/or role of Runx proteins in development and regeneration of major ocular glands, lacrimal gland (LG) and meibomian gland (MG), has not been previously studied. LG is an exocrine-type gland that accounts for the bulk of the aqueous portion of the precocular tear film.²⁴ Murine LG development starts at approximately E13.5 as an invagination of conjunctival epithelium into the surrounding mesenchyme.

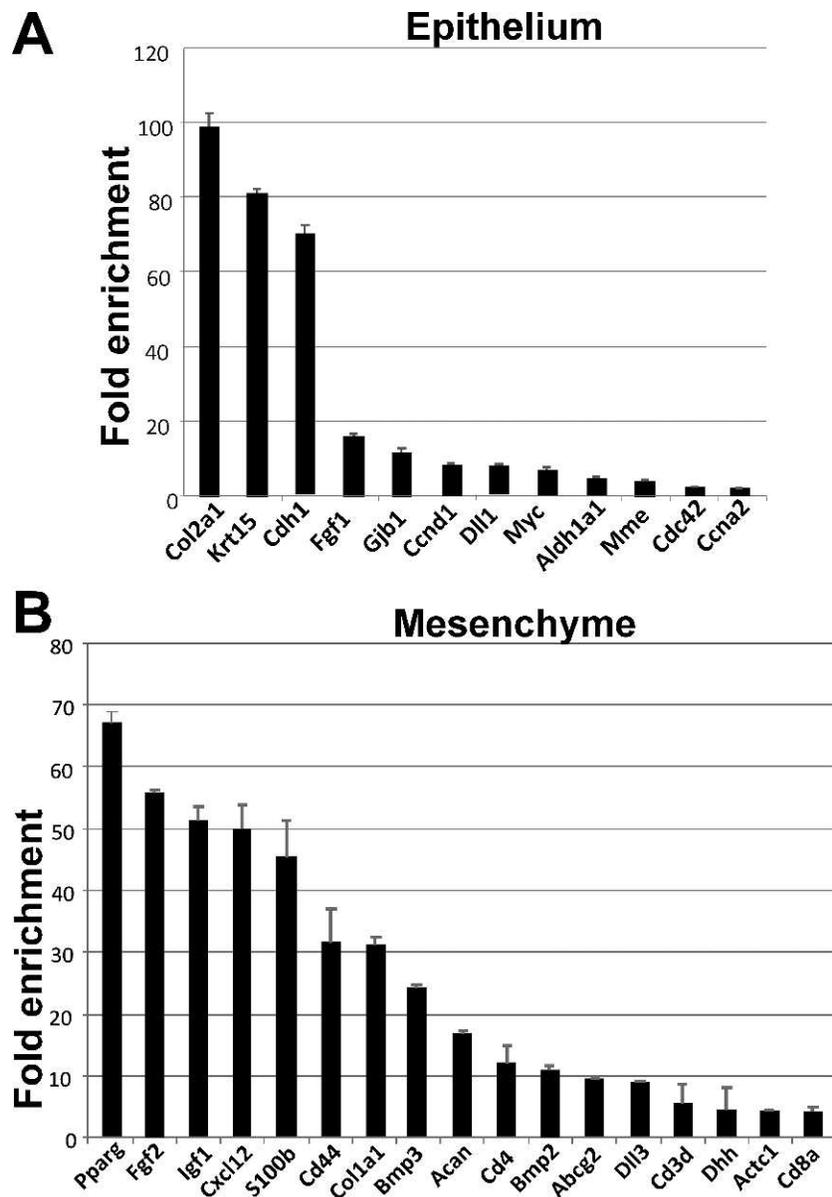


FIGURE 1. Stem cell PCR array revealed genes differentially expressed in LG epithelium and mesenchyme. Total RNA was prepared from E16.6 enzymatically separated LG epithelium and mesenchyme. Relative expression was calculated using the household genes provided in the array. *Bars* represent means \pm SEM of two experiments. LG epithelium was enriched with expression of collagen, type II, alpha 1 (Col2a1), keratin 15 (Krt5), cadherin 1 (Cdh1), fibroblast growth factor 1 (Fgf1), gap junction protein, beta 1 (Gjb1), Cyclin D1 (Ccnd1), delta-like 1 (Dll1), myelocytomatosis oncogene (Myc), aldehyde dehydrogenase family 1, subfamily A1 (Aldh1a1), membrane metalloendopeptidase (Mme), cell division cycle 42 homolog (Cdc42), Cyclin A2 (Ccna2). LG mesenchyme was enriched with expression of peroxisome proliferator activated receptor gamma (Pparg), fibroblast growth factor 2 (Fgf2), insulin-like growth factor 1 (Igf1), chemokine (C-X-C motif) ligand 12 (Cxcl12), S100 protein, beta polypeptide (S100b), CD44 antigen (CD44), aldehyde dehydrogenase family 1, subfamily A1 (Aldh1a1), bone morphogenetic protein 3 (Bmp3), aggrecan (Acan), CD4 antigen (CD4), bone morphogenetic protein 2 (Bmp2), ATP-binding cassette, subfamily G (WHITE), member 2 (Abcg2), delta-like 3, (Dll3), CD3 antigen, delta polypeptide (CD3d), desert hedgehog (Dhh), actin, alpha, cardiac muscle 1 (Actc1), CD8 antigen, alpha chain (CD8a).

Subsequently, epithelial ducts form an elaborate network through a process known as branching morphogenesis.^{25,26} The branching pattern and function of the LG is regulated by epidermal growth factor, FGFs, bone morphogenetic proteins (BMPs), Wnts, and numerous transcription factors.²⁷⁻³² Recent studies indicate that, similar to other exocrine glands (pancreas, salivary, mammary),³³⁻³⁶ the LG has a high regenerative potential and is able to repair itself even after substantial damage.³⁷ During LG regeneration, the epithelial component of the gland undergoes epithelial-mesenchymal

transition.^{24,38} In this process, epithelial cells lose cell junctions, polarity, and epithelial-specific markers, and acquire migratory phenotype and expression of mesenchymal markers.³⁷ When gland remodeling is completed, cells return to an epithelial phenotype and form new LG ductal and acinar structures.³⁷ There is also evidence for a population of proliferating nestin-positive stem cells that expand during LG regeneration; a subset of these cells bear markers of myoepithelial cells, suggesting a common progenitor for myoepithelial and epithelial lineages.³⁷ It is possible that LG

regeneration involves both dedifferentiation of mature epithelial cells, and activation, proliferation, and migration of epithelial stem cells.

Inflammation of the lacrimal gland, such as in Sjögren's syndrome, graft versus host disease, or other pathological conditions, can induce LG destruction. Unfortunately, these pathological conditions are also associated with a decline in LG regenerative ability. Inflammation may impact the stem cell niche, or change expression of important regulators of LG repair.^{24,38} Defining the mechanism(s) and factors that control LG morphogenesis and regeneration is important for developing new strategies to treat LG pathologies. Currently, we have limited understanding of the factors that induce progenitor cell proliferation and differentiation, and maintain the differentiated state once regeneration is completed.

In this study, we identified several transcription factors with a specific pattern of expression in the epithelial or mesenchymal cell lineage of the LG. We found that Runx1 and Runx2 were restricted to the LG epithelial compartment and Runx3 was weakly expressed in both epithelial and mesenchymal compartments. Downregulation of Runx1 to 3 expression in LG organ cultures significantly reduced LG growth, branching, and epithelial proliferation; thus Runx factors are necessary for normal LG morphogenesis. Runx1 and 3 expression increased during LG regeneration/remodeling and correlated with upregulation of cytokeratin 5 (Krt5), while Runx2 expression remained largely unchanged. We conclude that Runx proteins are LG epithelial cell lineage markers and regulate LG morphogenesis. Moreover, Runx1 and 3 may be important for LG regeneration.

MATERIALS AND METHODS

Mice

Imprinting control region (ICR) (CD-1) out-bred timed-pregnant mice (Harlan Laboratories, Hayward, CA) were used for isolation of embryonic LGs for organ cultures and quantitative (qRT-PCR) arrays. Reactivable Runx1 knockout mice (*Runx1^{Re/Re}*) have been previously described^{39,40} and were used for studies of Runx1 expression and function. Female BALB/c mice (Taconic, Germantown, NY) were used for regeneration studies and qRT-PCR arrays. All experiments were in accordance with the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research and were approved by The Scripps Research Institute and Tufts Medical Center Institutional Animal Care and Use Committees. ICR mice were sacrificed and embryos were kept in dissection medium (high-glucose minimum essential media [MEM] supplemented with 10% fetal bovine serum [FBS] and antibiotic/antimycotic) on ice prior to LG isolation. BALB/c mice were anesthetized and the exorbital LGs were left untreated (control) or were injected with 2 μ L volume of recombinant human IL-1 α (1 μ g, a generous gift from the BRB Preclinical Repository of the National Cancer Institute), as previously described.⁴¹ Mice were sacrificed at various times postinjection and LGs removed and processed for RNA extraction.

Culture Media

LG dissection was performed in high-glucose MEM supplemented with 10% FBS and antibiotic-antimycotic (cat.# 15240062; Invitrogen, Carlsbad, CA).

LG organ cultures were maintained in CMRL Medium-1066 (cat.# 11530037; Invitrogen) supplemented with Insulin-Transferrin-Selenium-Supplement (cat.# 51300044; Invitro-

TABLE 1. Genes Expressed in the LG Epithelium at E16.5

Lacrimal Gland Epithelium		E16.5
Gene	Fold-Up Regulation	P Value
Dnmt3b	6.306	0.00309
Foxp1	7.336	0.000959
Myc	6.786	0.000038
Pax6	17.530	0.000005
Runx1	11.926	0.001072
62	17.441	0.003939
Sox6	7.988	0.000005
Sox9	4.957	0.004121
Tert	2.266	0.004974
Vdr	7.369	0.000321

LGs were isolated at E16.5 and epithelium and mesenchyme were separated and gene expression in the epithelium and mesenchyme was compared in qRT-PCR stem cell transcription factors array.

gen), human transferrin-40 μ g/mL (cat.# 0030124SA; Invitrogen), 0.1% Albumax I (cat.# 11020021; Invitrogen), and 150 μ g/mL ascorbic acid (prepared fresh) (cat.# 07157; STEMCELL Technologies Inc., Vancouver, Canada).

Antibodies

The following primary antibodies were used for immunostaining: mouse monoclonal E-cadherin antibodies (Clone 36/E-cadherin; BD Pharmingen, San Jose, CA; or clone DECMA-1; Millipore, Billerica, MA), mouse monoclonal antibody to Pecam (CD31, clone MEC 13.3; BD Pharmingen), rat monoclonal antibody against Ser²⁸ phosphohistone-H3 (P-H3) (Clone HTA28; Sigma-Aldrich, St. Louis, MO), laminin antibody produced in rabbit (cat.# L 9393; Sigma-Aldrich), and cytokeratin 5 (Krt5) rabbit polyclonal antibody (cat.# AF138; Covance, San Diego, CA). Complimentary secondary antibodies were obtained from Invitrogen.

X-Gal Staining

Tissue was fixed with 1.25% paraformaldehyde (PFA) and 1.25% glutaraldehyde solution and processed for X-Gal staining as described in the Supplementary Methods.

TABLE 2. Genes Expressed in the LG Mesenchyme at E16.5

Lacrimal Gland Mesenchyme		E16.5
Gene	Fold-Up Regulation	P Value
Foxp2	7.672	0.007084
Gata6	40.564	0.000308
Klf2	8.576	0.021912
Klf4	6.208	0.000546
Nfatc1	6.294	0.002887
Nr2f2	9.021	0.000207
Pax1	14.392	0.001801
Pitx2	48.605	0.00389
Pparg	38.927	0.000666
Zfp2	13.236	0.02511
Dlx1	14.540	0.028748
Esr1	23.755	0.011947
Gata1	5.480	0.029089

LGs were isolated at E16.5 and epithelium and mesenchyme were separated and gene expression in the epithelium and mesenchyme was compared in qRT-PCR stem cell transcription factors array.

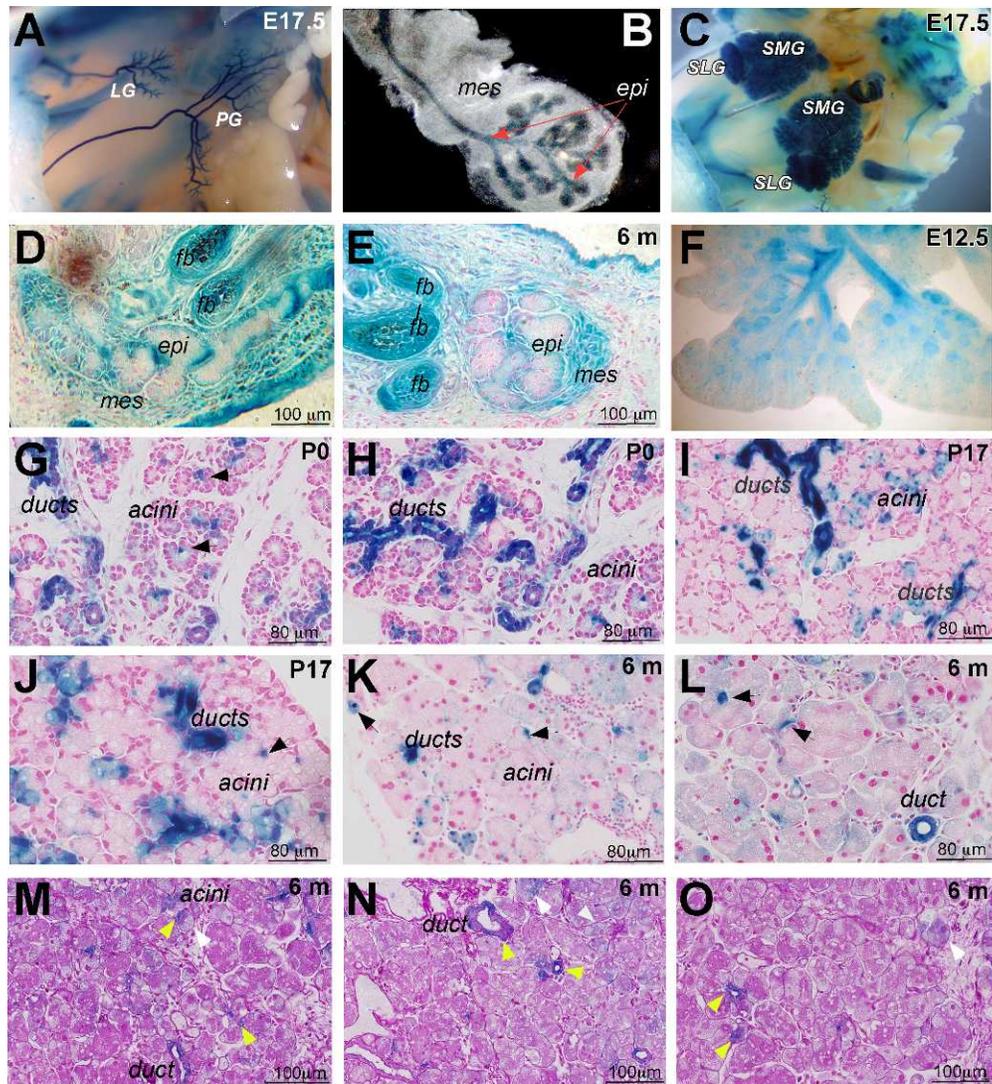


FIGURE 2. Analysis of Runx1 expression in the lacrimal gland of heterozygous Runx1 knockin-lacZ mice. (A, B) Whole-mount X-Gal staining of mouse embryo at E17.0. Runx1 expression is restricted to the epithelium of the LG and PG; (B) shows an excised LG. *Red arrows* mark epithelial component of the lacrimal gland. (C) Runx1-lacZ is expressed in the SLG and SMG. (D, E) Runx1 is also expressed in the epithelial and mesenchymal components of the meibomian gland (mei) and lung epithelium (F). (G–L) Expression pattern of Runx1 in the LGs at postnatal development day 0 (P0; [G, H]), P17 (I, J), 6-month-old (K, L) old mice. Runx1 expression is restricted to the ducts and centroacinar cells (*black arrows*) within the secretory acini. LGs obtained from Runx1 knockin-lacZ mice were stained with X-Gal and paraffin sections were prepared. Nuclei were stained with fast red to visualize LG morphology. (M–O) PAS reaction was also used for staining of sections of Runx1-LacZ LGs. Strong LacZ staining was seen within all PAS⁺ ducts, including a main duct (m. duct) and centroacinar cells (*yellow arrows*) and within some PAS⁻ acini/cells (*white arrows*).

Immunoblotting

Total protein was prepared from small interfering RNAs (siRNAs)-treated LG organ cultures (approximately 10–15 LGs per each condition) using radioimmunoprecipitation assay (RIPA) lysis buffer and sonicated. Equal aliquots of protein were resolved by SDS-PAGE, transferred to polyvinylidene difluoride membrane (PVDF), and probed with antibody to Cyclin D1 (cat.# 2922; Cell Signaling, Boston, MA). β -actin antibody (cat.# 4970; Cell Signaling) was used as a loading control. Bands were quantified by densitometry, and the cyclin-D1/ β -actin ratio calculated. Two experiments were performed, and the results from Runx1, 2, and 3 siRNA-treated cultures were normalized to the values from the control siRNA-treated cultures.

Isolation of LG Epithelium and Mesenchyme

Epithelial and mesenchymal components of E16.5 LGs were separated as previously described.^{31,42,43} Briefly, LGs were dissected and treated with trypsin/pancreatin solution on ice for 40 minutes, and the epithelial structure was mechanically separated from the mesenchyme. To prepare trypsin/pancreatin solution, trypsin (cat.# 85450C-25G; Invitrogen) and pancreatin (P3292; Sigma) were dissolved in Tyrode solution (see Supplementary Methods) to a final concentration of 2.25% wt/vol and 0.75% wt/vol respectively. Solution was filter sterilized.

qRT-PCR and qRT-PCR Microarray

Three independent experiments were performed to obtain triplicates for each microarray. For each experiment, LGs were

isolated from at least two litters of embryos. RNA was isolated using RNeasy Mini kit (cat.# 74104; SABiosciences, Qiagen, Valencia, CA); cDNA was made using RT² First Strand Kit (cat.# 330401; SABiosciences, Qiagen). Pathway-specific PCR arrays or individual primer PCR assays were performed using RT² SYBR Green qPCR Mastermix (SABiosciences, Invitrogen) (see Supplementary Methods for primers and arrays). Quantitative RT-PCR was performed on the ABI 7300 system (Life Technologies, Grand Island, NY) and data analyzed using online normalization and analysis tools (provided in the public domain, <http://sabiosciences.com/pcrarraydataanalysis.php>).

RNA Interference

The siRNAs targeting Runx1, Runx2, and Runx3, as well as negative control (scrambled) and penetration control (CY3-conjugated) siRNAs were from Ambion, Invitrogen. RNA interference (RNAi) was tested using at least two different siRNAs for each of Runx1, Runx2, or Runx3. Control and Runx siRNAs were delivered using Lipofectamine RNAiMAX Transfection Reagent (Invitrogen). Transfection efficiency was determined to be 95% to 98% using the Cy3-labelled siRNA. Details of RNAi and qRT-PCR are described in the Supplementary Methods. Gene expression profiles were generated by normalizing the Ct (threshold cycle) of genes of interest with a housekeeping gene 29S and then comparing gene expression in LGs treated with control siRNA and combinations of Runx1, 2, and 3 siRNAs. Approximately 10 to 15 LGs per each condition were used. Each experiment was repeated at least three times.

Statistical Analysis

For each experiment, approximately 10 to 15 LGs per condition/treatment were used. For gene or protein expression studies, LG tissue samples were pooled together. All experiments were repeated two to three times. Data are expressed as means \pm SD from two to three experiments. Paired Student's *t*-test was used to determine the statistical significance of differences between experimental (Runx siRNA-treated) and control (control siRNA-treated) LGs. *P* values less than 0.05 were considered statistically significant.

RESULTS

Expression of Stem Cell Markers in Separated Epithelial and Mesenchymal Components of Developing LG

Remodeling of tissue after injury recapitulates many of the cellular events that govern embryonic development, including stem/progenitor cell activation.⁴⁴ Thus, to begin to identify novel markers of LG progenitors that may be involved in regeneration, we first examined stem cell-associated signaling pathways and regulators in embryonic LG using stem cells and stem cell transcription factors qRT-PCR arrays. To identify markers specific for the epithelial and mesenchymal lineages, we prepared RNA from enzymatically separated E16.5 LG epithelial and mesenchymal components. Complete separation of epithelial and mesenchymal cells was confirmed by measuring expression of Barx2 (which is expressed only in LG epithelium) and Barx1 (which is expressed only in LG mesenchyme).²⁸ Comparison of epithelial and mesenchymal cells using the Stem Cell Array (PAMM-405A; SABiosciences, Qiagen), showed that epithelial cells were enriched for cell cycle regulators, such as Cyclin D1 (CcnD1), cell division cycle 42 homolog (Cdc42), Cyclin A2 (CcnA2), and regulator of cell proliferation myelocytomatosis oncogene (Myc) (Fig. 1). These markers are consistent with our observation that epithelium is more proliferative than mesenchyme (see Supplementary Fig. S1). LG mesenchyme preferentially expressed growth factors such as FGF2 and FGF3; BMP1, 2, and 3; growth differentiation factor 3 (GDF3); and insulin-like growth factor-1. Mesenchyme was also enriched in hematopoietic cell lineage markers (glycoproteins, cluster differentiation 4 and 8) CD4 and CD8a, mesenchymal progenitor cell markers, such as Acan (Agc1), and various adhesion and matrix proteins (Fig. 1). These findings suggest that mesenchyme of developing LG undergoes substantial remodeling and is a source of growth factors important for epithelial morphogenesis.^{30,31}

To define a transcription factor signature for LG epithelium, we compared epithelial and mesenchymal cells using the Stem Cell Transcription Factor PCR Array (PAMM-501Z; SABiosciences, Qiagen). Sox6 and Sox9 (SRY-box containing transcription factors 6 and 9, stem cell markers), Foxp1 (forkhead transcription factor-1, regulator of epithelial injury response), Six2 (Sixe oculis-related homeobox), Runx1 (runt family

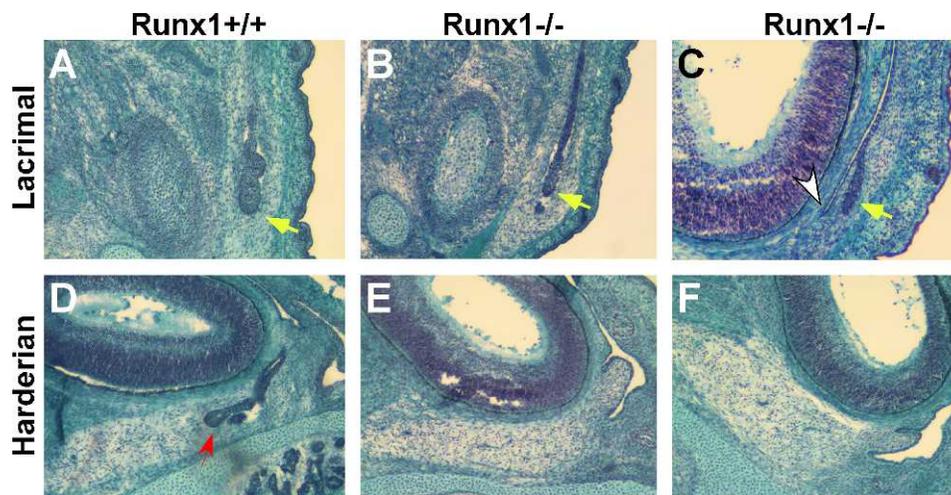


FIGURE 3. Mutation in the Runx1 gene reduces LG budding and perturbs HG development in embryos at E16.5. (A–C) LG; (D–F) HG development in WT (Runx1+/+) (A, D) and mutant (Runx1-/-) (B, C, E, F) embryos. Yellow arrows indicate LG budding, white arrow conjunctiva, and red arrow HG epithelium component in WT embryos.

TABLE 3. Analysis of Gene Expression in Ocular Glands and Other Branched Organs

Expression	Lacrimal Gland	Harderian Gland	Meibomian Gland	Lung	Submandibular Gland
Runx1	+++	+++	+++	+++	+++
Runx2	+++	+++	+++	++	+++
Runx3	++	+	+	+	+
Barx1	++	+++	+	N/A	+++
Barx2	+++	+++	++	+	+++
Pax6	+++	+/-	+++	+	-

+++ , highly expressed; ++ , intermediate level of expression; + , low level of expression; +/- , very low expression; N/A , not determined; - , no expression.

transcription factor), and Pax6 (paired homeobox factor) were exclusively expressed in LG epithelium (Table 1). Specificity of Pax6 for the LG epithelial compartment has been previously reported.^{27,28} Relative to mesenchyme, LG epithelium was also enriched in Myc, and the epigenetic regulators DnmtB (DNA methyltransferase 3B), and Tert (Telomerase reverse transcriptase) (Tables 1, 2).

Several transcriptional regulators were restricted to LG mesenchyme (Table 2) or enriched in mesenchyme relative to epithelium, including Foxp2, which is also found early in developing conjunctiva,⁴⁵ Pax1, and Kruppel-like factors, Klf2 and Klf4. The latter factors are involved in trafficking and differentiation of CD8⁺ and CD4⁺ T cells, which are important players in animal models of autoimmunity.⁴⁶

Runx1 Is a New Marker of the LG Epithelial Cell Lineage

Our study provides the first evidence that Runx1 expression is restricted to the LG epithelium. In other tissues, Runx1 is involved in stem cell specification, proliferation, and differentiation,^{21,23,46} and influences tissue homeostasis.⁴⁶ Runx1 was recently shown to be involved in differentiation of the epithelial secretory cell lineage in colon.⁴⁶ Moreover, absence of *Runx1* was associated with global changes in expression of inflammation-related genes.^{3,46,47} Thus, we decided to further investigate Runx expression and function during LG development.

A reversible Runx1 knockin-LacZ mouse was used to examine Runx1 expression and function in LG morphogenesis.^{39,40} During embryonic development, Runx1 was expressed throughout the epithelium of LG, parotid glands (PG), submandibular (SMG), and sublingual glands (SLG) (Figs. 2A–C). Runx1-LacZ was also expressed in ductal and mesenchymal components of the MG, in the follicle bulge region (fb) of eyelashes (Figs. 2D, 2E), and in other branched organs, such as lung (Fig. 2F) and mammary gland (not shown). In early postnatal (P0–P17) and adult (6-month-old) mice, Runx1 expression remained restricted to the epithelial component of the LG with high level of expression in ductal epithelium in a subset of cells within the acini called centroacinar cells (Figs. 2G–J, black arrows), which represent the extension of the intercalated duct cells. Centroacinar cells in other branched glands are reported to be progenitor-like and involved in tissue regeneration.^{48,49} Runx1-LacZ also marked cells in ducts that morphologically and positionally resemble myoepithelial cells (MECs) (not shown). The expression of Runx1 within LG ducts, centroacinar cells, and MECs suggests a specific function in epithelial progenitor populations. Adult Runx1-LacZ LG sections were stained using the Periodic Acid-Schiff (PAS) reaction, which identifies a subset of acinar and ductal cells.⁵⁰ PAS-positive (PAS⁺) epithelial cells (pink, see Figs. 2M–O) were seen in main and intercalated ducts and within acini (Figs. 2M, 2N). Runx-LacZ marked all PAS⁺ ducts and centroacinar cells

(Figs. 2M–O, yellow arrows) and some PAS⁺ acinar cells (Figs. 2M, 2N, white arrows).

Role of Runx1 Gene Function in LG Development

Runx1-null mice die at E11.0–12.5 due to impaired endothelial and hematopoietic stem cell function. In *Tie2-Cre;Runx1^{Fl^{ox}:lacZ/Fl^{ox}:lacZ}* embryos, Cre recombinase is expressed under the control of the endothelial/hematopoietic-specific *Tie2* promoter,^{51–53} resulting in selective reactivation of *Runx1* in hematopoietic cells. The gross appearance of rescued *Tie2-Cre;Runx1^{Fl^{ox}:lacZ/Fl^{ox}:lacZ}* embryos was indistinguishable from that of wild-type (WT) embryos. Serial paraffin sections of E16.5–17.0 embryos showed only slight difference between *Cre;Runx1^{Fl^{ox}:lacZ/Fl^{ox}:lacZ}* and WT glands: WT LG started to branch and had several small buds (Fig. 3A), while many *Cre;Runx1^{Fl^{ox}:lacZ/Fl^{ox}:lacZ}* LGs contained only one bud (Figs. 3B, 3C) that was short and close to conjunctiva (Fig. 3C). At E16.5, harderian glands (HG) emerge from the nasal part of conjunctival epithelium; HG were evident in WT embryos (Fig. 3D, red arrow), but absent in *Cre;Runx1^{Fl^{ox}:lacZ/Fl^{ox}:lacZ}* embryos (Figs. 3E, 3F). Despite the embryonic delays, adult *Cre;Runx1^{Fl^{ox}:lacZ/Fl^{ox}:lacZ}* LG and HG morphology was essentially normal with only slight variation in gland sizes (not shown). Thus Runx1 controls timing of primary bud outgrowth, but may be redundant in later development.

Role of Runx Genes in LG Development

To examine possible redundancy of Runx genes, Runx 1, 2, and 3 expression was measured by qRT-PCR in adult ocular glands and other branched organs (lung and submandibular gland) (Table 3). All three Runx genes were expressed in LG and HG; Runx1 and Runx2 were restricted to the epithelial component of the glands, whereas Runx3 mRNA was detected in both epithelium and mesenchyme (Table 4).

To further define the function of Runx genes in LG development, we inhibited their expression using either single siRNAs targeted against each Runx isoform or a combination of siRNAs to target all isoforms. The Runx-targeted siRNAs were applied to LG organ cultures prepared at E15 (single-bud stage,

TABLE 4. Analysis of Runx1 to 3 Expression in Separated LG Epithelium and Mesenchyme

Expression	LG Epithelium	LG Mesenchyme
Runx1	+++	-
Runx2	++	-
Runx3	+	+
Barx1	-	+++
Barx2	+++	-

+++ , highly expressed; ++ , intermediate level of expression; + , low level of expression; - , no expression.

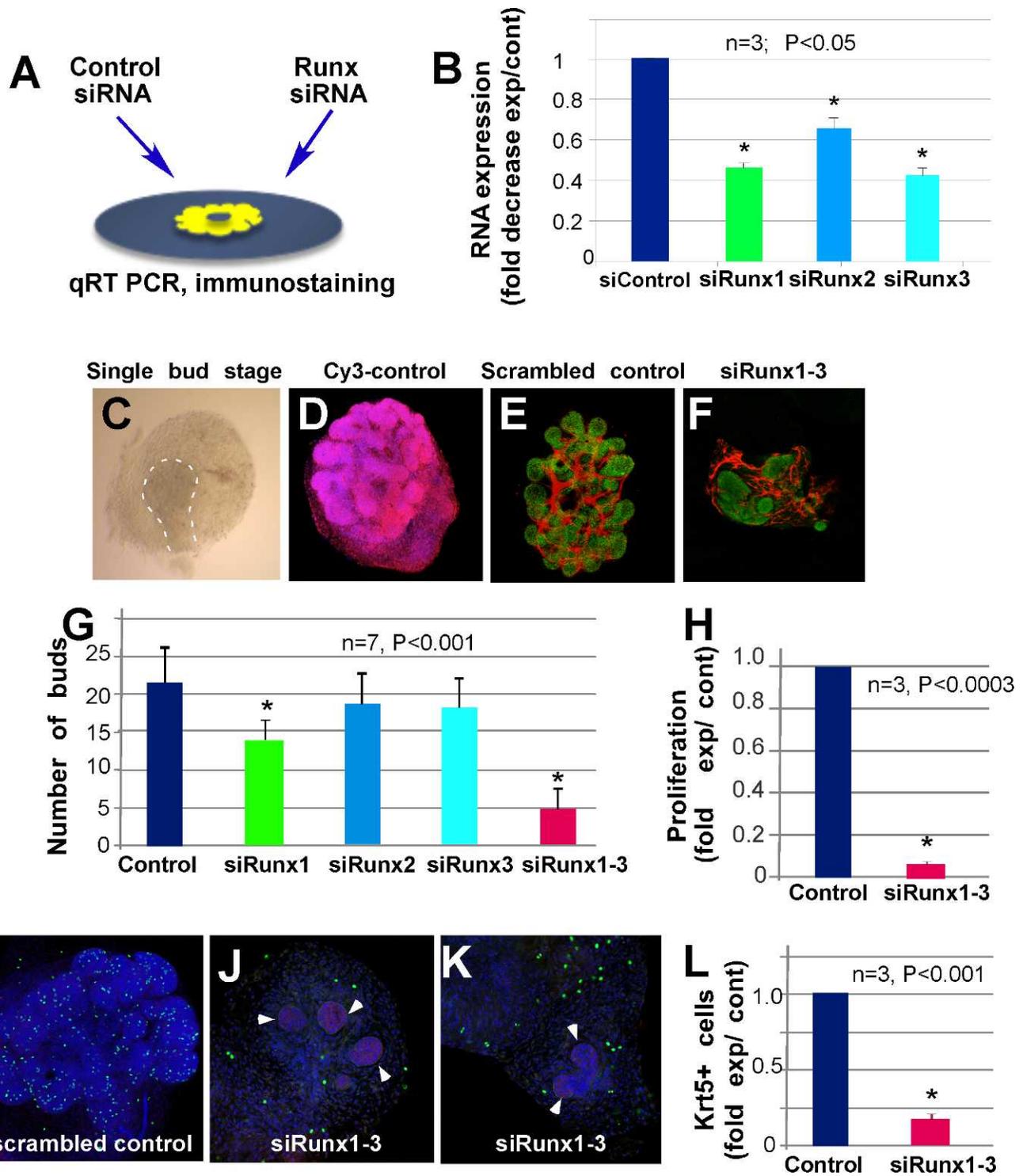


FIGURE 4. Knockdown of Runx1, 2, and 3 reduces LG epithelial growth, cell proliferation, and number of Krt5-expressing progenitor cells. (A) Scheme of ex vivo siRNA treatment. LG were isolated at one bud stage (also see [C]) at E15.5 and cultured on a filter floating on the surface of culture medium. Control and Runx siRNAs were delivered using RNAiMAX (Invitrogen). (B) Runx1, 2, and 3 RNAs fold decrease after specific siRNA application is shown relative to control-treated LGs. (C) Example of LG at one bud stage used for siRNA application. (D) CY3-labeled control RNA is found in all cells of the LG. (E) Scrambled control siRNA application does not affect LG branching. (F) Combined specific Runx1, 2, and 3 siRNA applications strongly reduce LG growth/branching. (G) Quantification of buds in siRNA-treated glands shows significant inhibition of branching after application of Runx1, or combinations of Runx1 and 2 or Runx1, 2, and 3 siRNAs (E-cadherin is *green*, PECAM is *red*). (H-K) The number of proliferating cells is decreased in Runx-siRNA treated LGs (H) quantification of H-3P antibody-stained proliferating cells in control and Runx1, 2, and 3 siRNA-treated LG buds. Control (I) and Runx1, 2, and 3 siRNA-treated (J-K) LG were stained with H-3P antibody (*green*), E-cadherin (*red*), and 4',6-diamidino-2-phenylindole (*blue*) (*white arrowheads* show epithelium). LGs treated with Runx1, 2, and 3 siRNA show almost no cell proliferation in the epithelial component of the gland, while proliferating cells are still present in mesenchyme. (L) Number of Krt5-expressing cells is strongly reduced in LGs treated with Runx1, 2, and 3 siRNA. Asterisks denote statistical significance in the results compared to the control.

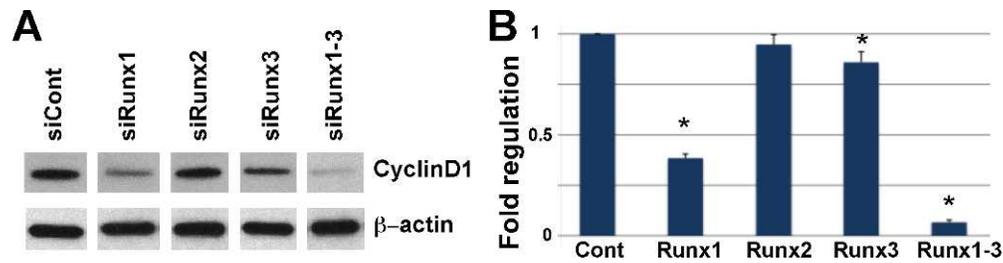


FIGURE 5. Knockdown of Runx1, 2, and 3 expression results in decreased CyclinD1 protein. (A) Protein was prepared from siRNA-treated LG organ cultures, resolved by SDS-PAGE, and probed with the cyclin-D1 or β -actin antibodies. (B) Graphic representation of densitometric analyses of Western blots from two independent experiments (mean \pm SD of Optical Density [OD] of bands). For analysis of the Western blot images ImageJ 1.46 (National Institutes of Health, Bethesda, MD) has been used. The relative densities of the loading-control bands and the sample bands were calculated. Adjusted density for each sample lane was calculated by dividing the sample relative density by the loading-control relative density for each lane separately. Asterisks mark significant statistical difference in the level of CyclinD1 of experimental siRNA treated lacrimal glands compared to control treated glands.

Figs. 4A, 4C). Scrambled siRNA acted as a control and CY3-conjugated control siRNA was used to evaluate tissue penetration (Figs. 4D, 4E). Labeled siRNA (penetration control) was found in all cells of treated LGs (Fig. 4D). Reduction of Runx RNA was confirmed by qRT-PCR (Fig. 4B). Application of Runx1 siRNA alone induced a small but statistically significant decrease in LG branching (two-tailed $P = 0.0007$) (Fig. 4G), while application of Runx2 (two-tailed $P = 0.4251$) or Runx3 siRNA (two-tailed $P = 0.5423$) had no effect (Fig. 4G). However, LG treated with combined Runx1 and Runx2 siRNAs (two-tailed $P < 0.0001$) or Runx1, Runx2, and Runx3 siRNAs (two-tailed $P = 0.0001$) showed greatly reduced branching and irregular bud shape relative to control siRNA treated LGs (Figs. 4E–G).

Runx proteins are reported to regulate stem cell proliferation.²³ To assess cell proliferation in LGs treated with Runx siRNAs, we used an phospho-histone-3 antibody (Ser28-H3-P, referred to here as H3-P) that detects phosphorylated histones at the onset of mitosis.⁵⁴ Combined application of siRNA to all three Runx genes almost completely abolished proliferation in the epithelial component of the gland (Figs. 4H–K), although proliferating cells were still detected in the mesenchyme (Figs. 4J, 4K). Control siRNAs had no effect on cell proliferation (Figs. 4I, 4H). We hypothesize that perturbation of Runx1, 2, and 3 during LG morphogenesis decreases the number of proliferating progenitor cells (either epithelial stem cells or MECs). Consistent with this, expression of cytokeratin-5 (Krt5), a reported marker of epithelial progenitors in several tissues,^{55–57} was strongly reduced in Runx-siRNA-treated LG (Fig. 4L). To explore the mechanism of Runx regulation of cell proliferation, we measured CyclinD1 protein (a highly conserved cyclin protein family member expressed in the

epithelium [see Fig. 1]) in siRNA-treated LG by immunoblotting. CyclinD1 was modestly downregulated by Runx1 siRNA treatment and strongly downregulated by application of all three Runx siRNAs; application of single Runx2 or Runx3 siRNAs had no significant effect on CyclinD1 levels (Fig. 5).

Runx1 and 3 Are Upregulated During Early Stages of LG Regeneration

To elicit LG regeneration, we used the established IL-1 α injection method. IL-1 α induces a strong immune response, extensive inflammation, and LG damage, followed by a regenerative phase that restores morphology by 7 to 10 days after injury.³⁷ If Runx genes are expressed in epithelial progenitor cells or cells undergoing active remodeling during LG regeneration, we predicted that Runx expression would increase during the regeneration phase. LGs were injected with IL-1 α or saline (vehicle control), and collected at day 1, 2, 3, 7, or 21 after injection for RNA extraction. Levels of Runx1 to 3, Krt5 (epithelial progenitor marker), RBM3 (member of the glycine-rich RNA-binding protein family, ubiquitously expressed in the LG), and PrL27 (noncoding RNA, poly[A]-bearing) and PrS13 (16S ribosomal RNA gene) housekeeping mRNAs were determined by qRT-PCR (Table 5, Fig. 6, graphic representation of Table 5). Runx1 and Runx3 were upregulated in regenerating LG. Runx2 mRNA remained largely unchanged during the period of LG remodeling, but was slightly increased during epithelial differentiation/maturation (Fig. 6). As expected, Rbm3 expression was largely unchanged. Significantly, changes in Runx1 and Runx3 expression during LG regeneration correlated with changes in expression of Krt5, peaking during the first 3 days and declining from 7 to 21 days back to

TABLE 5. Changes in Gene Expression During Lacrimal Gland Regeneration

	Day 1		Day 2		Day 3		Day 7		Day 21	
	Fold Regulation	P0 Value								
Runx1	2.2342	0.000174	3.0275	0.000005	2.7089	0.000054	1.6359	0.00017	1.358	0.019599
Runx2	0.931	0.130836	1.2128	0.212677	0.9869	0.555818	1.1843	0.282192	2.1496	0.003509
Runx3	2.8746	0.000221	4.6128	0.00001	4.1988	0.000004	2.5115	0.00028	1.006	0.430019
Krt5	1.6831	0.006834	2.9337	0.000223	3.1363	0.000064	2.3936	0.005639	1.7249	0.029249
RBM3	1.1965	0.353178	1.2148	0.199725	0.9289	0.542553	1.0698	0.726245	0.9809	0.466395

Expression of Runx1, Runx2, Runx3, Krt5, and Rbm3 in IL-1 injected and control LGs was studied by qRT-PCR. Fold regulation was calculated by comparing of the level of expression of each gene in IL-1-treated glands with its expression in saline-injected glands. Gene expression was normalized to PrL27 and PrS13 housekeeping genes. The experiment was performed in triplicate. The $\Delta\Delta C_t$ -based fold regulation and P0 value calculations were performed using the RT² Profiler PCR Array Data Analysis software (SABiosciences, Qiagen). Statistically significant changes and corresponding P0 values are shown in bold.

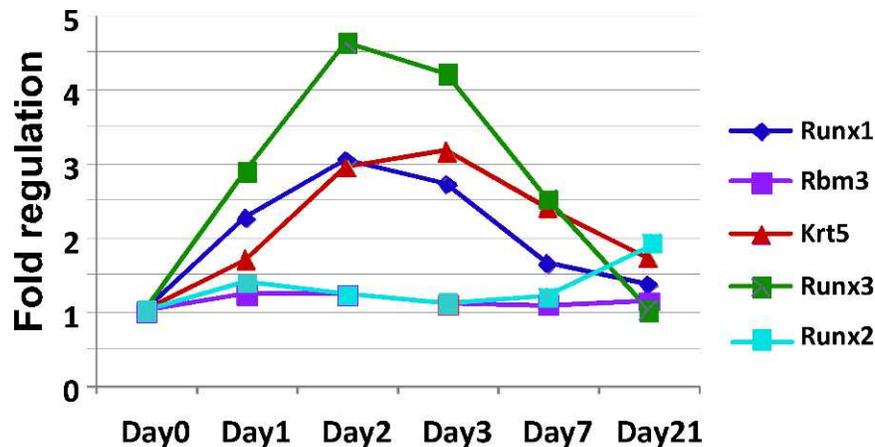


FIGURE 6. Graphic representation of Table 5. LGs were injected with IL-1 or saline (vehicle control) and collected at day 1, 2, 3, 7, or 21 after injection for RNA extraction. Levels of Runx1, Runx2, Runx3, Krt5, Rbm3, and PrL27 and PrS13 mRNAs were determined by qRT-PCR. Increase in Runx1 and Runx3 expression correlates with increase in expression of epithelial progenitor cell marker Krt5, while expression of Runx2 and ubiquitously expressed Rbm3 remains unchanged during regeneration.

the baseline level in uninjured glands. Thus, Runx1 and Runx3 are induced during LG regeneration and may be involved in expansion of epithelial progenitor cells.

DISCUSSION

Here we studied the expression and function of Runx factors during LG development and regeneration. We identified Runx1 and Runx2 as novel specific epithelial markers of the LG, and showed that Runx3 is expressed in both LG epithelium and mesenchyme. Consistent with our data, recent publications show that Runx proteins are expressed in epithelial cells in salivary and mammary glands, prostate, intestine, and lung.^{16,18,19,23,46,58} Recent lineage tracing studies of skin epithelia using *Runx1-CreER* knockin mice reported that embryonic Runx1+ placode cells show long-term contribution to multiple hair follicle epithelial lineages,²¹ suggesting a role for Runx1 in multipotential epithelial stem cells. The elevated level of Runx1 expression in LG centroacinar and ductal cells, upregulation of Runx1 expression during LG regeneration, and the reported contribution of Runx1 in controlling multiple stem cell lineages, suggests that high level of Runx1 expression might mark LG epithelial progenitor cell population(s). However, currently, the identity of LG stem/progenitor cells remains unclear and it is unknown whether the ductal, acinar, and MEC lineages originate from a common multipotent stem cell or multiple lineage-specific stem/progenitor cells.

The dramatic impairment of LG epithelial proliferation and branching by blockage of Runx1 to 3 correlates with other reports that Runx proteins regulate lineage-specific cell proliferation, differentiation, and cell migration.⁵⁹⁻⁶³ RUNX functions are also associated with several cancers and other human pathologies; Runx1 and Runx2 proteins may act as oncogenes by directly promoting cell proliferation^{62,64} and Runx1 has been implicated in promoting cell cycle progression through induction of cyclin D proteins.⁶⁵ Indeed, we show that Runx proteins regulate cyclin D1 expression at least during LG development. Interestingly cyclin D1 and CDK4 were recently reported to induce Runx2 and Runx3 phosphorylation, ubiquitylation, and proteasomal degradation, suggesting complex interaction between Runx and cell cycle proteins.⁶⁶

Runx proteins have been implicated in regulation of multiple developmental and regenerative processes, including cell adhesion and matrix production, cell movement, and

cytoskeletal organization. For example, Runx1 directly regulates E-cadherin and matrix metalloproteinase 9 (MMP9) expression.⁶⁷ Downregulation of E-cadherin and upregulation of MMP9 is closely linked with epithelial mesenchymal transition (EMT)³⁸ during LG regeneration. Although knock-down of Runx expression during LG morphogenesis only modestly affected E-cadherin expression (not shown), changes in adhesion and matrix remodeling may be part of the mechanism by which Runx proteins control morphogenesis and regeneration.

Defining new biomarkers of the LG epithelial lineage and of potential stem/progenitor cells is critical to understanding the LG regenerative process, and may aid the development of cell-based interventions for LG degeneration.

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