Uveal melanoma (UM) is the most common primary intraocular malignancy in adults.1 Tumors arise from melanocytes within the uveal tract, which is composed of the iris, the ciliary body, and the choroid. The standard treatment for patients with primary UM is organ-preserving radiation therapy (RT), delivered primarily as plaque brachytherapy, proton-beam therapy, or stereotactic external-beam RT.2 However, UM tumors require large doses of radiation per fraction for external-beam RT and high total RT dose (~80 Gy) to achieve tumor ablation. Large fraction-sizes and high total doses of RT can damage adjacent normal tissue structures of the eye leading to vision-threatening complications, including radiation retinopathy, papillopathy, ischemia, and neovascular glaucoma.3–7

The discovery of BRAF somatic mutations in approximately 60% of cutaneous melanoma patients has led to successful development of targeted therapies that have shown significant clinical benefit resulting in approval by the Food and Drug Administration of two agents: vemurafenib and dabrafenib.8–10 However, BRAF mutations are rare in UM.11 Instead, activating somatic mutations in the GNAQ gene have recently been shown to be present in approximately 50% of UM patients.12 The GNAQ gene encodes for the GTP-binding G-protein \( \alpha_q \) subunit, which mediates signaling between G-protein-coupled receptors and phospholipase C\( \beta \) (PLC\( \beta \)).13 GNAQ mutations in UM most commonly occur in codon 209 within the GTPase catalytic domain,11 resulting in a loss of intrinsic GTPase activity and constitutive activation of the \( \alpha_q \) protein. This in turn leads to increased activation of PLC\( \beta \), which cleaves phosphatidylinositol biphosphate to generate inositol triphosphate and diacylglycerol (DAG). DAG production activates the conventional and novel protein kinase C (PKC) families of proteins, resulting in enhanced growth and apoptotic escape.14

Importantly, recent studies using RNA interference-mediated downregulation of various PKC isoforms have shown that PKC\( \alpha \), PKC\( \beta \), PKC\( \epsilon \), PKC\( \theta \), and PKC\( \delta \) are functionally important for viability of GNAQ\( ^{mut} \) UM cells (Poulaki V, et al. IOVS 2012;53:ARVO E-Abstract 6871).15,16 Consistent with the important role of PKC signaling in mediating the oncogenic effects of mutant \( \alpha_q \) in UM, the PKC inhibitors enzastaurin,
PKC Inhibitors Sensitize GNAQ Mutant UM Cells to IR

Clonogenic Survival Assay

Radiosensitization was established with the standard clonogenic assay. Twenty-four hours after IR, the medium was changed and cells were incubated at 37°C for another 14 days to allow colony formation. At the end of the assay, colonies were fixed and stained with 6% formaldehyde and 0.5% crystal violet. Colonies containing more than 50 cells were counted. Surviving fractions were calculated as (mean colony counts)/[(cells inoculated) × (plating efficiency)], in which plating efficiency was defined as (mean colony counts)/[cells inoculated for nonirradiated controls). Statistically significant differences in survival curves were analyzed using the SPSS 19.0 software package (SPSS, Inc., Chicago, IL, USA) by means of a weighted, stratified, linear regression, according to the linear-quadratic formula \[ S(D)/S(0) = \exp(D + JD^2). \]

The sensitivity enhancement ratio (SER) for each PKC inhibitor was calculated as the ratio of surviving fraction of vehicle-treated cells to corresponding PKC inhibitor–treated cells at 6 Gy.

Cell-Cycle Analysis

Cell proliferation (percentage of S-phase cells) and cell-cycle distribution (DNA content) was determined 18 hours after IR using the Click-iT EdU Assay kit (Invitrogen) and TOPRO-3 as a total DNA counterstain (Invitrogen). Samples were analyzed by flow cytometry (FACScan Canto II; BD Biosciences, San Jose, CA, USA), with a minimum of 20,000 events. Data were analyzed with FlowJo software (Tree Star, Inc., Ashland, OR, USA).

Quantitative Real-Time PCR

RNA was extracted from cells 18 hours after IR using the RNasy Mini kit (Qiagen, Germantown, MD, USA). RNA was reverse transcribed into cDNA (SuperScript III First-Strand Synthesis SuperMix; Invitrogen), which was quantified by real-time PCR (StepOne PLUS Real-Time PCR System; Applied Biosystems, Foster City, CA, USA). TaqMan Gene Expression Assays (Applied Biosystems) were used to detect the expression levels of the following genes: CDC25A, CCND1, CDKN1A, CDKN1B, TOP2A, and TP53BP1. Gene expression was normalized to ACTB using the ∆∆Ct method. Statistical differences between treatment groups were evaluated by Student’s t-test.

Immunofluorescence and Image Analysis

Three and 18 hours after IR, cells were fixed in 4% paraformaldehyde (Electron Microscopy Sciences, Hatfield, PA, USA) and permeabilized with 0.5% Triton X-100 (Amresco, Solon, OH, USA). Nonspecific antibody binding was blocked by 1% bovine serum albumin. Primary antibodies against γH2AX (1:2000; Bethyl Laboratories, Montgomery, TX, USA) were detected by using anti-rabbit secondary antibodies labeled with Alexa Fluor 488 (Invitrogen); 4′,6-diamidino-2-phenylindole (DAPI, 1:1000; Sigma-Aldrich) was used for nuclei counterstaining.

Cells were imaged using the Cell Lab IC-100 Image Cytometer (IC-100; Beckman Coulter, Brea, CA, USA), equipped with a ×40/0.90NA objective. The imaging camera (Hamamatsu, Bridgewater, NJ, USA) was set to capture 8-bit images at 1 × 1 binning (1344 × 1024 pixels, 6.5 um²/pixel) with two images captured per field (DAPI, AlexaFluor 488). In general, 36 images were captured per coverslip.

Images were analyzed for γH2AX intensity using custom algorithms developed with the Pipeline Pilot (v8.0) software platform (Accelrys, San Diego, CA, USA) in a similar workflow as previously described. The background signal was...
PKC Inhibitors Sensitize GNAQ Mutant UM Cells to IR

RESULTS

PKC Inhibitors Enhance IR-Induced Reduction in Cell Viability, Cell Proliferation, and Clonogenic Survival of GNAQmt UM Cells

We hypothesized that small-molecule PKC inhibitors used at significantly lower concentrations than their half maximal inhibitory concentration would enhance IR-induced antitumor activity in UM cells. To test this hypothesis, we compared the impact of treatment with IR alone, PKC inhibitors alone, or PKC inhibitors combined with IR on GNAQmt cells (Mel202, 92.1) UM cells. GNAQmt/BRAFmt cells, an atypical UM cell line more likely derived from a cutaneous melanoma, served as controls. Cells were treated with DMSO, BIM (0.5 μM) or AEB071 (0.5 μM) for 3 hours followed by 0, 2, 4, or 6 Gy of IR. Cell viability and proliferation were determined 120 hours after IR with trypan blue dye, and radiosensitization was established with the standard clonogenic assay. Compared with IR alone, both PKC inhibitors combined with IR significantly decreased cell viability (Fig. 1A), cell proliferation (Fig. 1B), and clonogenic survival (Fig. 1C) of GNAQmt, but not GNAQmt/BRAFmt melanoma cells.

Cell viability (Fig. 1A) was not largely affected by IR alone. Compared with IR or PKC inhibitor monotherapy, combination therapy demonstrated a further significant and IR dose-dependent reduction in viability of GNAQmt cells. The viability of GNAQmt/BRAFmt melanoma cells was not affected by PKC inhibitors or by combination therapy.

Cell proliferation (Fig. 1B) was significantly decreased by IR alone in both GNAQmt and GNAQmt/BRAFmt cells. Compared with IR or PKC inhibitor monotherapy, combination therapy demonstrated a further significant reduction in proliferation of GNAQmt cells. The proliferation of GNAQmt/BRAFmt melanoma cells was not affected by PKC inhibitors.

Combination therapy significantly reduced the clonogenic survival of GNAQmt, but not GNAQmt/BRAFmt melanoma cells (Fig. 1C). Radiosensitization was statistically determined by measuring the SER, defined as the ratio of surviving fraction of vehicle-treated cells to corresponding PKC inhibitor–treated cells at 6 Gy. GNAQmt Mel202 cells exhibited a SER of 4.07 and 3.75 for BIM and AEB071, respectively. GNAQmt 92.1 cells showed a similar effect with SER of 2.64 for BIM and 3.16 for AEB071. SER ranged from 0.98 for AEB071 to 1.01 for BIM in GNAQmt/BRAFmt OCM3 cells, indicating no radiosensitizing effect by PKC inhibitors in GNAQmt/BRAFmt cells. Thus, our results suggest PKC inhibitors increase radiosensitivity in GNAQmt UM cell lines. As AEB071 is currently being evaluated in phase I clinical trials for metastatic UM, we focused further studies on the effects of AEB071.

PKC Inhibitor AEB071 Increases IR-Induced Cell Cycle Arrest in GNAQmt UM Cells

We next examined the effect of combining PKC inhibitors with IR on cell-cycle distribution in GNAQmt (Mel202, 92.1) and GNAQmt/BRAFmt (OCM3) cells. Cells were treated with DMSO or AEB071 (0.5 μM) for 3 hours followed by 0 or 6 Gy of IR. Cell-cycle distribution was determined 18 hours after IR by flow cytometry. Compared with IR alone, AEB071 combined

Western Blotting

The following antibodies were purchased from commercial sources and used for Western blotting: mouse monoclonal p53 (DO-1; Santa Cruz Biotechnology, Santa Cruz, CA, USA), rabbit polyclonal p21 (C-19; Santa Cruz Biotechnology), rabbit monoclonal cyclin D1 (9262; Cell Signaling, Danvers, MA, USA), and rabbit polyclonal phospho-Chk2 (Thr68; Cell Signaling).

Cells were lysed in RIPA buffer (Pierce, Rockford, IL, USA) containing Protease Inhibitor Cocktail (Roche, Indianapolis, IN, USA). Protein concentration was determined by bicinchonic acid assay (Pierce). Cell lysates were resolved in NuPAGE 4% to 12% Bis-Tris SDS-polyacrylamide gels (Invitrogen). After separation, proteins were transferred to Immobilon-P polyvinylidine fluoride membranes (Millipore, Billerica, MA, USA). Membranes were blocked for 1 hour with 5% milk (in tris-buffered saline with 0.1% Tween-20). Primary antibodies were incubated overnight at 4°C, followed by secondary antibodies for 1 hour at room temperature. Immunoreactive bands were visualized by SuperSignal West Femto chemiluminescence reagents (Pierce). β-actin (1:4500; Santa Cruz Biotechnology) was used as a loading control.
with IR decreased the proportion of proliferating cells (S-phase) in GNAQ<sup>mt</sup>, but not GNAQ<sup>mt</sup>/BRAF<sup>mt</sup> melanoma cells (Fig. 2).

IR alone decreased the proportion of GNAQ<sup>mt</sup> and GNAQ<sup>mt</sup>/BRAF<sup>mt</sup> cells in S phase, and increased the proportion of GNAQ<sup>wt</sup> and GNAQ<sup>wt</sup>/BRAF<sup>mt</sup> cells in G<sub>y</sub>/M phases (Fig. 2). AEB071 alone decreased the proportion of GNAQ<sup>mt</sup> cells in S phase, and increased the proportion of GNAQ<sup>mt</sup> cells in G<sub>1</sub> phase (Figs. 2A, 2B). Combination therapy demonstrated the greatest decrease in the proportion of proliferating cells (S-phase) in GNAQ<sup>mt</sup> cell lines. The cell-cycle distribution in GNAQ<sup>mt</sup>/BRAF<sup>mt</sup> cells was not affected by AEB071 (Fig. 2C). These experiments suggest that the impacts observed on proliferation and clonogenicity (Fig. 1) were mediated in part by the combined effects of PKC inhibition and IR on cell-cycle progression.

**PKC Inhibitor AEB071 Augments IR-Induced DNA Damage–Associated γH2AX Staining Intensity in GNAQ<sup>mt</sup> UM Cells**

To investigate the impact of AEB071 and IR on the DNA damage response, we used high-throughput microscopy and automated image analysis to measure induction and resolution of the phosphorylated histone protein H2AX (γH2AX) in GNAQ<sup>mt</sup> Mel202 and GNAQ<sup>mt</sup>/BRAF<sup>mt</sup> OCM3 cells. γH2AX is directed to sites flanking DNA double-strand breaks (DSBs) during the DNA damage response. As such, γH2AX protein expression is a sensitive indicator of IR-induced DNA DSBs. In these experiments, γH2AX intensity (sum of pixel intensity in the nucleus) was analyzed by customized image analysis tools, which automatically identify cells and nuclei to extract fluorescence-based measurements. In Mel202 cells (Fig. 3A), IR increased γH2AX intensity by 3.1-fold 3 hours after treatment. Combined IR and AEB071 treatment further increased γH2AX intensity by 40% relative to IR alone (4.3-fold relative to DMSO controls). Eighteen hours after IR, levels of γH2AX declined by 67% compared with 3-hour levels in both GNAQ<sup>mt</sup> and GNAQ<sup>wt</sup>/BRAF<sup>mt</sup> OCM3 cells, levels of γH2AX did not differ significantly between cells subjected to IR alone (3.6-fold) or combined treatment (3.6-fold; Fig. 3B). Eighteen hours after IR, levels of γH2AX declined by 67% compared with 3-hour levels in both
PKC Inhibitor AEB071 Modulates IR-Induced Changes in Protein Expression and Posttranslational Modification in GNAQmut UM Cells

To characterize the impact of combination therapy on proteins involved in DNA damage response, cell-cycle progression, and cell survival, we performed proteomic profiling by RPPA analysis in GNAQmut Mel202 cells (Supplementary Fig. S1). Cells were treated with DMSO or AEB071 (0.5 μM) for 3 hours followed by 0 or 6 Gy of IR. When analyzing proteins involved in the DNA damage response at 18 hours after 0 or 6 Gy of IR exposure (Fig. 5A), we found significant increases in Rad50 expression and increased nuclear factor (NF)-κB phosphorylation after IR alone. Treatment with PKC inhibitor alone resulted in significantly increased expression of Rad50, ERCC1, and XRCC1, and significantly decreased phosphorylation of NF-κB. Compared with individual treatments, combined treatment with AEB071 and IR resulted in significantly increased Chk2 phosphorylation at threonine 68 and increased expression levels of 53BP1, BRCA2, Rad50, and total Chk2. Additionally, combined treatment resulted in significantly decreased phosphorylation of NF-κB. These data suggest persistent DNA damage signaling after combination therapy. Among the proteins involved in cell-cycle progression (Fig. 5C), AEB071 treatment alone resulted in significantly increased total levels of the cyclin-dependent kinase inhibitor p27; decreased phosphorylation of Rb; and decreased levels of total Rb, Myc, and CDK1. These effects persisted in the presence of IR, indicating cell-cycle arrest on combined treatment. Among proteins involved in cell growth (Fig. 5B), the phosphorylation of 4EBP1 and S6 was significantly decreased after IR alone and

PKC Inhibitors Enhance IR-Induced Gene Expression Changes Affecting Cell Cycle and DNA Damage Repair in GNAQmut UM Cells

We next sought to characterize changes in the expression of genes associated with cell-cycle progression and DNA damage in response to combination therapy in GNAQmut (Mel202, 92.1) and GNAQmut/BRAFmut (OCM3) cells. Cells were treated with DMSO, BIM (1 μM), or AEB071 (0.5 μM) for 3 hours followed by 0 or 6 Gy of IR. RNA was isolated 18 hours after IR, and mRNA expression was measured by quantitative real-time PCR. Compared with IR alone, combination therapy in GNAQmut cells significantly reduced the expression of positive regulators of cell-cycle progression, including CDC25A (Fig. 4A) and CCND1 (Fig. 4B), and significantly increased the expression of negative regulators of cell-cycle progression, including CDKN1A (Fig. 4C) and CDKN1B (Fig. 4D). The expression of genes implicated in DNA damage response, including TOP2A (Fig. 4E) and TP53BP1 (Fig. 4F), was also significantly increased after combined treatment compared with IR alone, except in Mel202 cells where a similar increase in the expression level of TP53BP1 after IR alone and after combined treatment was observed. Conversely, the expression of these genes in GNAQmut/BRAFmut melanoma cells was not largely affected by PKC inhibitors or by combination therapy compared with IR alone.

PKC Inhibitors Sensitize GNAQ Mutant UM Cells to IR

Figure 2. PKC inhibitor AEB071 increases IR-induced cell-cycle arrest in GNAQmut UM cells. (A) GNAQmut Mel202, (B) GNAQmut 92.1, and (C) GNAQmut OCM3 cells were treated with DMSO or AEB071 (0.5 μM) for 3 hours followed by 0 or 6 Gy of IR. Eighteen hours after IR, cell-cycle distribution, including percentage of S-phase cells (left) and 2N/4N DNA content (right), were detected by flow cytometry.
PKC Inhibitors Sensitize GNAQ Mutant UM Cells to IR

IR-alone treatment, PKC inhibitor alone, and after combined treatment. In the IR-alone group, among the highest-scoring interactants in the upregulated group was NF-κB p53/6. Conversely, in the PKC-treated groups (alone and combined with IR), NF-κB p53/6 was among the most downregulated interactants, potentially providing a mechanistic clue into the radiosensitizing properties of the PKC inhibitor AEB071. Additionally, compared with IR alone, the upregulated network after combined treatment preserved proteins involved in the DNA damage response, and gained proapoptotic factors and inhibitors of the cell-cycle progression. Conversely, the downregulated network after combined treatment consisted of the NF-κB pathway and mediators of cell growth. These data establish a reciprocal dynamic between the PKC inhibitor and IR, wherein separate as well as common mechanisms of cell survival are inhibited by the combined therapy. Additional validation of protein expression levels of select proteins by immunoblot analysis supported our gene expression and RPPA results (Supplementary Fig. S2).

DISCUSSION

RT is the standard treatment for patients with primary UM. However, the large doses of RT required to achieve tumor control can affect normal tissues adjacent to the tumor and often lead to vision-threatening complications, including radiation retinopathy, papillopathy, ischemia, and neovascular glaucoma.3–7 The most frequently observed radiation complication is radiation retinopathy, which has been described in up to 50% of patients treated with RT.35,57 Unfortunately, the options for management of radiation retinopathy and other common radiation-related complications remain limited.57

Figure 3. The PKC inhibitor AEB071 augments IR-induced DNA damage-associated γH2AX staining intensity in GNAQmut UM cells. (A) GNAQmut Mel202 and (B) GNAQmut OCM3 cells were treated with DMSO or AEB071 (0.5 μM) for 3 hours followed by 0 or 6 Gy of IR. Three and 18 hours after IR, cells were fixed, stained with anti-γH2AX antibody, and examined by high-throughput microscopy (upper). DAPI (nucleus) and γH2AX were pseudo-colored in red and green, respectively. Binary nuclear and cellular masks were generated by a combination of watershed and threshold image transformations. The γH2AX sum of pixel intensities within nuclear masks was used to quantify γH2AX staining intensity (lower). Values are normalized to DMSO controls at each time point. Error bars indicate SEM (n > 50 cells/condition collected over two independent experiments). Scale bar: 20 μm; a indicates statistically different from DMSO control; b indicates statistically different from IR alone; and c indicates statistically different from AEB071 alone.

Figure 4. PKC inhibitors enhance IR-induced gene expression changes in GNAQmut UM cells. The GNAQmut (Mel202, 92.1) and GNAQwt (OCM3) cells were treated with DMSO, BIM (1 μM), or AEB071 (0.5 μM) for 3 hours followed by 6 Gy of IR. RNA was isolated 18 hours after IR, and mRNA expression was measured for the following genes by quantitative real-time PCR: (A) CDC25A, (B) CCND1, (C) Cdkn1A, (D) Cdkn1B, (E) Top2a, (F) Tp53bp1. Bars represent mean ± SD from three biological replicates; *P < 0.05 for comparing IR only to PKC inhibitor combined with IR at 6 Gy.
FIGURE 5. PKC inhibitor AEB071 modulates IR-induced changes in protein expression and posttranslational modification in GNAQ-mutant UM cells. GNAQ-mutant Mel202 cells were treated with DMSO or AEB071 (0.5 μM) for 3 hours followed by 0 or 6 Gy of IR. Cell lysates were collected 18 hours after IR, and the expressions of proteins were determined by RPPA. Heat maps represent normalized expression values for each protein (red, high; green, low). Each treatment was done in triplicate, which corresponds to three distinct heat map boxes per treatment condition. Effects on the expression of select proteins controlling (A) DNA damage response, (B) cell growth, (C) cell-cycle regulation, and (D) apoptosis are presented. (E) Network analysis was performed to identify the most represented signaling networks on each treatment condition using NetWalker software.
PKC Inhibitors Sensitize GNAQ Mutant UM Cells to IR

IOVS | April 2014 | Vol. 55 | No. 4 | 2137

Therefore, there is an unmet clinical need for therapeutic agents capable of selectively enhancing the sensitivity of UM cells to radiation. Such agents would potentially allow for reductions in the total dose and/or fraction size of RT delivered to the eye, and would represent an important clinical advance by minimizing the frequency of radiation-induced vision impairment.

In this study, we investigated the radiosensitizing effects of the small-molecule PKC inhibitors BIM and AEB071 in UM cell lines expressing mutant GNAQ. Melanoma cells harboring wild-type GNAQ served as controls. We found that, compared with IR or PKC inhibitors alone, combined treatment resulted in significantly enhanced antitumor activity against GNAQmt, but not GNAQwt/BRAFmt cells, evidenced by decreased cell viability, decreased cell proliferation, and decreased clonogenic survival. Consistent with these observations, combination therapy resulted in the highest reduction in the S-phase fraction of GNAQmt cells. Together, these results demonstrate that small-molecule PKC inhibitors specifically enhance the effect of IR in GNAQmt cells, highlighting a unique vulnerability in this class of tumors.

To determine the molecular mechanisms underlying the reduced clonogenic potential of GNAQmt UM cells on the combined treatment, we measured the induction and resolution of γH2AX foci, a surrogate marker of DNA DSBs. Based on nuclear γH2AX intensity, DNA DSBs were induced to a greater extent and persisted longer in GNAQmt cells treated with combination therapy compared with IR alone. Conversely, PKC inhibition did not affect γH2AX intensity or dynamics in GNAQwt/BRAFmt melanoma cells. This finding suggests that combination therapy may render GNAQmt tumors more susceptible to the effects of DNA DSBs by delaying DNA damage resolution. Consistent with these observations, GNAQmt cells treated with combination therapy showed persistently increased Chk2 phosphorylation and increased levels of other DNA damage-associated proteins, such as BRCA2, Rad50, and 53BP1, 18 hours after IR, a time at which these proteins had returned to baseline in cells treated with IR alone, suggesting completed repair processes. In sum, these data indicate that PKC inhibitors are able to potentiate IR-induced DNA damage and interfere with the rate of DNA repair in GNAQmt cells. This mechanism is common among potent radiosensitizers.

Our results also demonstrate that small-molecule PKC inhibitors combined with IR collaborate to promote apoptotic and proapoptotic effects of IR in GNAQmt cells through induction of proteins involved in cell-cycle arrest, cell-growth arrest, and apoptosis. Activating phosphorylation of NF-κB on serine 536 was central to the upregulated network of mutation present in patients with primary UM (~45%), whereas mutations affecting the GNAQ R183 position are much less frequent (~3%). Apart from GNAQ Q209 and R183 mutations, UM patients also harbor mutations in GNAI1 gene, which codes for the Gα11 subunit of the GTP-binding G-protein. The close functional relationship between Gαq and Gα13, together with constitutive activation of PKCs in both GNAQmt and GNAI1mt UM cells, provides rationale that UM cells harboring GNAQ or GNAI1 mutations are selectively sensitive to PKC inhibitors. This has indeed been demonstrated in several previous studies (Poulaki V, et al. IOVS 2012;53:ARVO E-Abstract 6871). Recent work by Chen et al., using two different PKC inhibitors across a panel of six different UM cell lines harboring GNAQ or GNAI1 mutations, as well as melanocyte cell lines stably overexpressing GNAQ or GNA11, demonstrated selective growth inhibition of these cells, whereas melanoma cell lines harboring mutations in other genes were not sensitive to PKC inhibition, regardless of whether they were derived from uveal or cutaneous melanoma. Although PKC inhibition causes selective growth inhibition in both cells carrying an activating GNAQ or GNAI1 mutation, whether the combination of PKC inhibitors with IR will result in the same radiosensitization in cells harboring GNAI1 (and GNAQ R183) mutations requires experimental validation. As our control GNAQwt/BRAFmt OCM5 cell line was recently identified as an atypical UM cell line more likely derived from a cutaneous melanoma, we cannot exclude the possibility that the observed cooperative effects of PKC inhibitors and IR are relevant to all UM cells and not restricted to UM cells carrying GNAQ mutations. However, this possibility would not obviate the potential therapeutic benefit of combination therapy in patients with UM.

The results of previous attempts to specifically radiosensitize UM tumors to radiation have been limited in success. Intravitreal injections of VEGF pathway inhibitors (e.g., bevacizumab, ranibizumab, pegaptanib sodium) have recently been tested for radiosensitizing effects in the treatment of primary UM; however, they have yielded inconsistent results. These studies did, however, demonstrate the feasibility of combining drugs delivered intravitreally with RT. The preclinical data presented in this study support the hypothesis that PKC small-molecule inhibitors should be considered as part of a combined modality approach with RT in the treatment of primary GNAQmt UM. As phase I clinical trials of the small-molecule PKC inhibitor AEB071 are ongoing in metastatic UM, a clinical trial using PKC inhibitors in combination with RT in primary GNAQmt UM is warranted. This approach would represent an important advance in genotype-driven personalized targeted therapy and promises to potentially improve patient outcome while minimizing vision-threatening toxicities associated with RT.
PKC Inhibitors Sensitize GNAQ Mutant UM Cells to IR

Acknowledgments

The authors acknowledge the joint participation by the Adrienne Helis Malvin Medical Research Foundation and the Diana Helis Henry Medical Research Foundation through their direct engagement in the continuous active conduct of medical research in conjunction with Baylor College of Medicine.

SEM is a Dan L. Duncan Scholar, a Caroline Wiess Law Scholar, and a member of the Dan L. Duncan Cancer Center (supported by the National Cancer Institute [NCI] Cancer Center Support Grant P30CA125125) at Baylor College of Medicine. Additional support was provided by K01 DK096093 (SMH), and the Pilot/Feasibility Program of the Diabetes Research Center (P30-DK079638) at Baylor College of Medicine (SMH). NM is a Dan L. Duncan Scholar, a Caroline Wiess Law Scholar, and a member of the Dan L. Duncan Cancer Center (supported by the NCI Cancer Center Support Grant P30CA125125) at Baylor College of Medicine. His work is also supported by the Conquer Cancer Foundation of the American Society of Clinical Oncology (ASCO) Career Development Award, a Pilot/Feasibility Program of the Diabetes and Endocrinology Research Center (P30-DK079638) at Baylor College of Medicine, and by the Prostate Cancer Foundation. Imaging resources were supported by Specialized Cooperative Centers Program for Reproductive Research U54 HD007495 (FJ DeMayo), P30 DK-56338 (MK Estes), P30 CA-125123 (CK Osborne); and the Dan L. Duncan Cancer Center of Baylor College of Medicine. Supported by the Cytometry and Cell Sorting Core at Baylor College of Medicine with funding from the National Institutes of Health (AI036211, CA125123, and RR024574) and the expert assistance of Joel M. Sederstrom.

Disclosure: J.Z. Cerne, None; S.M. Hartig, None; M.P. Hamilton, None; S.A. Chew, None; N. Missiades, None; V. Poulaki, None; S.E. McGuire, None.

References

PKC Inhibitors Sensitize GNAQ Mutant UM Cells to IR