

# The Genetic Causes of Nonsyndromic Congenital Retinal Detachment: A Genetic and Phenotypic Study of Pakistani Families

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**PURPOSE.** To evaluate consanguineous pedigrees from Pakistan with a clinical diagnosis of nonsyndromic congenital retinal nonattachment (NCRNA) and identify genes responsible for the disease as currently only one NCRNA gene is known (atonal basic helix-loop-helix transcription factor 7: *ATOH7*).

**METHODS.** We implemented a three-step genotyping platform: single nucleotide polymorphism genotyping to identify loss of heterozygosity regions in patients, Retinal Information Network panel screening for mutations in currently known retinal genes. Negative patients were then subjected to whole exome sequencing.

**RESULTS.** We evaluated 21 consanguineous NCRNA pedigrees and identified the causal mutations in known retinal genes in 13 out of our 21 families. We found mutations in *ATOH7* in three families. Surprisingly, we then found mutations in familial exudative vitreoretinopathy (FEVR) genes; low-density lipoprotein receptor-related protein 5 mutations (six families), tetraspanin 12 mutations (two families), and *NDP* mutations (two families). Thus, 62% of the patients were successfully genotyped in our study with seven novel and six previously reported mutations in known retinal genes.

**CONCLUSIONS.** Although the clinical diagnosis of all children was NCRNA with severe congenital fibrotic retinal detachments, the molecular diagnosis determined that the disease process was in fact a very severe form of FEVR in 10 families. Because severe congenital retinal detachment has not been previously associated with all the FEVR genes, we have thus expanded the phenotypic spectrum of FEVR, a highly variable retinal detachment phenotype that has clinical overlap with NCRNA. We identified seven novel mutations. We also established for the first time genetic overlap between the Iranian and Pakistani populations. We identified eight NCRNA families that do not harbor mutations in any known retinal genes, suggesting novel causal genes in these families.

**Keywords:** *ATOH7*, *LRP5*, *TSPAN12*, *NDP*, FEVR, NCRNA, blindness, retinal detachments

Nonsyndromic congenital retinal nonattachment (NCRNA, OMIM #221900) is an autosomal recessive, devastating congenital form of retinal detachment leading to permanent blindness from birth. Children with NCRNA have congenital blindness caused by partial to complete retinal detachments. The main clinical findings are congenital insensitivity to light, nystagmus, microphthalmia, shallow anterior chamber (AC), and a massive fibrovascular mass behind the lens.<sup>1,2</sup> Nonsyndromic congenital retinal nonattachment is reported to be clinically and genetically heterogeneous. The retina and blood vessels in one or both eyes display variable clinical and histologic anomalies. Systemic malformations associated with the disease have been shown in some individuals; however, no consistent association has been noticed.<sup>1</sup>

Autosomal recessive NCRNA is common in North Khorasan, Iran, among descendants of a Kurdish population. In this founder population, the locus was mapped to chromosome 10q21.<sup>1,3</sup> Subsequently, a homozygous (hmz) 6523-bp deletion was found in atonal basic helix-loop-helix (BHLH) transcription factor 7 (*[ATOH7]* also known as *Math5*), in a cluster of three conserved noncoding elements that spans a remote *cis* regulatory element 20-kb upstream from *ATOH7*.<sup>1</sup> Atonal BHLH transcription factor 7 encodes a member of the BHLH family of transcription factors that plays an important role in the development of retinal ganglion cell (RGC) and optic nerve formation.<sup>4,5</sup> It was determined that the absence of RGCs creates neovascular growth of fetal blood vessels in the vitreous that results in early retinal detachment.<sup>1</sup> Thus, the intrinsic mechanism of retinal cell fate determination is disrupted in



NCRNA, which in turn leads to early onset retinal detachment and subsequent blindness.<sup>1</sup>

In addition to NCRNA, mild and partial acquired retinal detachments can occur in patients with familial exudative vitreoretinopathy (FEVR), a rare progressive genetic disorder affecting retinal blood vessel development.<sup>6</sup> This disorder is characterized by partial vascularization of the peripheral retina and defective vascular differentiation.<sup>6</sup> All modes of inheritance are seen in FEVR patients.<sup>6</sup> The clinical presentation of FEVR patients is highly variable; most have mild disease, including neovascularization, exudation, dragged vessels and macula, retinal tears, and rhegmatogenous retinal detachments, which is not seen until the first or second decade of life.<sup>6,7</sup> Usually, FEVR is an acquired condition (not congenital such as NCRNA). Some patients present with falciform retinal folds extending from the optic nerve to the temporal periphery, which sometimes comes in contact with the lens, causing leukocoria.<sup>8,9</sup>

Mutations in five genes have been reported to cause FEVR; *NDP*, a gene encoding Norrie disease protein (X-linked), *FZD4*, a gene encoding frizzled-4 (autosomal dominant and recessive), *LRP5*, a gene encoding low-density lipoprotein receptor-related protein 5 (autosomal dominant and recessive), *TSPAN12*, a gene encoding tetraspanin-12 (autosomal dominant and recessive), and *ZNF408*, a gene encoding zinc finger protein 408 (autosomal dominant).<sup>6,8,10-12</sup> Individuals affected by mutations in the *LRP5* gene often have reduced bone mineral density, osteopenia, and osteoporosis in addition to vision loss.<sup>6,8,13</sup> Mutations of FEVR are associated with a wide range of phenotypes, ranging from mild retinal folds and avascular zones to later onset retinal detachments. Currently, FEVR mutations are not associated with congenital total retinal detachments and complete congenital blindness.

Because *ATOH* is currently the only gene associated with NCRNA, we aimed to identify the genetic causes of NCRNA in a Northern Pakistani population cohort, and at the same time validate a new gene discovery protocol established by our laboratories. In this study, we identified that mutations in genes associated previously with FEVR can result in the NCRNA phenotype. Our study suggests that although NCRNA and FEVR are classified as distinct diseases, a margin of overlap exists between the two. Thus, we expanded the phenotypic spectrum of FEVR. We also determined that a three-step gene discovery protocol established by our laboratories is highly effective for the identification of new genes and the elimination of previously known disease associated genes in Mendelian disorders.

## METHODS

This study was approved by the institutional review boards of the participating centers, according to the tenets of the Declaration of Helsinki. Informed consent was obtained from all patients at the initial visit in Pakistan at the El Shifa Trust Eye Hospital, where two of our clinical collaborators saw the babies (AK, SS) and another oversaw the DNA extractions (RQ); the consent form contained a paragraph about genetic analyses of retinal genes, including whole exome sequencing. Twenty-one NCRNA families (28 affected members) were recruited, phenotyped, and genotyped from Pakistan and included in this study. Entry criteria were as follows: the diagnosis of NCRNA was given to patients with the history of congenital blindness because of partial to complete retinal detachment from infancy, shallow anterior chamber, leukocoria, and abnormal B-scan. All children included in this study were born full term, free from uncomplicated pregnancies, and did not have any known medical issues.

All families were questioned for detailed, ocular, and visual histories, and pedigrees were drawn. Ophthalmic examinations were performed on all affected patients. Blood samples from all available NCRNA family members were collected at the Al Shifa Trust Eye Hospital Department of Pediatric Ophthalmology and Strabismus, Rawalpindi, Pakistan. Extraction of DNA from the NCRNA families and additional blood samples from 100 random unrelated healthy Pakistani control individuals were provided by the COMSATS Institute of Information Technology Department of Biosciences, Rawalpindi, Pakistan, for determination of allele frequency in the general Pakistani population.

## DNA Extraction and Primer Design

Genomic DNA was extracted from peripheral blood leukocytes with an extraction kit (FlexiGene; QIAGEN, Hilden, Germany) and a blood kit (QIAamp DNA; QIAGEN), according to the instructions by the manufacturer. DNA quantity and quality was verified by spectrophotometer (NanoDrop 1000; Thermo Fisher Scientific, Wilmington, DE, USA). We used a PCR and Sanger sequencing approach to identify mutations in candidate genes. Primers were designed by Exon Primer (provided in the public domain by the Institute for Human Genetics, Technical University of Munich, Germany (<http://ihg.gsf.de/ihg/Exon-Primer.html>)) and by Primer3 online program.<sup>14,15</sup> To ensure the completeness and quality of the sequences and for detection of potential mutations located in splice sites, a minimal distance between primer and exon/intron boundary was selected of at least 60 bp when primers were designed. Sequences for the used primers are available upon request.

## Step 1: Single Nucleotide Polymorphism (SNP) Microarrays

The genomes of 11 out of our 17 consanguineous NCRNA families were analyzed for homozygous chromosomal regions by using SNP homozygosity mapping (Infinium HD 660K; Illumina, Inc., San Diego, CA, USA), in accordance with the protocol provided by the manufacturer. Homozygous regions were visualized and identified with a commercial software package (Bead Studio pLink; Illumina, Inc.) and a freeware spreadsheet, EXCLUDEAR. Chromosomal segments were accepted as having significant homozygous regions if they contained  $\geq 300$  consecutive homozygous SNPs, as the likelihood that this would occur by chance is less than 1:100 with a particular focus to identified homozygous regions. All the known retinal genes (the retinal genes listed in Retinal Network information website) that resided in homozygous chromosomal segments were analyzed for mutations using Sanger sequencing.

## Step 2: RetNet Known Retinal Disease Gene Panel Screening by Targeted Next Generation Sequencing (NGS)

Fourteen patients diagnosed with NCRNA were screened by our recently developed RetNet known retinal disease gene panel,<sup>16</sup> which contains all the currently known retinal disease genes (163) by targeted NGS. The chip helped us to screen all retinal genes listed in the Retinal Information Network website and create a candidate retinal disease gene list for future study.

## Step 3: Whole Exome Sequencing

We performed whole exome capture following the manufacturer's protocol (Illumina, Inc.). We used 2  $\mu$ g of link-ligated DNA for the capture reaction. Exome capture (SureSelect V4; Agilent Technologies, Santa Clara, CA, USA) was performed according to the company's standard protocol. The amplified

TABLE 1. Candidate Genes for NCRNA Families in Their IBD Regions

Family No.	Chromosome	Candidate Gene	IBD Rank	Successfully Genotyped With Candidate Genes	Excluded Genes
NCRNA4-5	10	<i>ATOH7</i>	Middle	Yes	
NCRNA5-4	10	<i>ATOH7</i>	Biggest	Yes	
NCRNA6-3	10	<i>ATOH7</i>	2nd biggest	Yes	
NCRNA7-4	X	<i>NDP</i>	2nd biggest		<i>NDP</i>
	11	<i>LRP5</i>	13th biggest	Yes	
	11	<i>ZNF408</i>	15th biggest		<i>ZNF408</i>
NCRNA8-3	7	<i>TSPAN12</i>	Last		<i>TSPAN12</i>
NCRNA9-3	7	<i>TSPAN12</i>	Biggest	Yes	
NCRNA11-3	7	<i>TSPAN12</i>	Smallest		<i>TSPAN12</i>
NCRNA13-3	7	<i>TSPAN12</i>	Biggest		<i>TSPAN12</i>
NCRNA17-2	7	<i>TSPAN12</i>	7th biggest	Yes	
NCRNA17-3			2nd biggest		
NCRNA19-3	7	<i>TSPAN12</i>	2nd biggest		<i>TSPAN12</i>
NCRNA19-6			9th biggest		
NCRNA19-3	11	<i>FZD4</i>			<i>FZD4</i>
NCRNA19-4			Biggest		
NCRNA19-6					
NCRNA19-3	11	<i>LRP5</i>	3rd biggest	Yes	
NCRNA19-4			2nd biggest		
NCRNA19-6			2nd biggest		

Known retinal genes have been checked in the families according their location in IBD rank. Table shows IBD rank and successfully genotyped families with the genes identified in the IBD regions. Seven families harbored mutated genes in their top IBD regions, the rest of known genes were excluded from the study for the given families.

capture product was sequenced with commercial software (HiSeq 2000; Illumina, Inc.) to pinpoint the genetic variants responsible for the disease in those patients who did not carry mutations in known genes. Sequence capture was performed in accordance with the protocol of the manufacturer (Illumina, Inc.) in the sequencing platform of the McGill University and Genome Quebec Innovation Centre. Details of the protocol are described by Coussa et al.<sup>17</sup>

### Mutation Analysis

Known retinal genes within the homozygous regions, retinal detachment genes shown in RetNet (Retinal Information Network, tables of genes and loci causing inherited retinal diseases), and all putative mutations found by RetNet panel screening were Sanger-sequenced. Complete sequencing of the candidate genes within the homozygous regions was done to exclude all possible mutations. If a patient had several homozygous regions carrying different known genes for the given disease, the screening of the genes was started from a gene located in the biggest homozygous segment.

### Sanger Sequencing

Sanger sequencing was used to identify mutations in candidate genes. Primers and PCR conditions are available upon request. The assays were performed by the sequencing platform of the McGill University and Genome Quebec Innovation Centre using forward or reverse sequencing primers designed for PCR reactions that covered a designated exons and splice sites of the exons. The results were analyzed using an NGS software program (Sequencher; Gene Codes Corporation, Ann Arbor, MI, USA).

### In Silico Mutational Analyses

For novel mutations found, we performed in silico analyses predicting the variants to be damaging by SIFT,<sup>18</sup> Blosum62,<sup>19</sup> and Polyphen-2.<sup>20</sup> We then studied conservation of the residue

in homologous proteins of lower animals and we verified that the variant has not been reported in the Exome Variant Server (NHLBI GO Exome Sequencing Project), dbSNP135, or 1000 Genome datasets. Finally, segregation of the mutations between family members was then performed and we excluded the newly found variant from normal controls that are culturally matched to the original family.

### RESULTS

In collaboration with a research group in Islamabad, Pakistan, we assembled 21 consanguineous pedigrees from Pakistan with an NCRNA diagnosis. Consanguineous families have homozygous regions, one of which contains the causal gene with homozygous mutations. This fact allows for a genetic strategy that is well established, called homozygosity mapping and inheritance by descent (IBD) studies. Our approach to finding the causal genes in NCRNA patients was to first identify all the significant homozygous regions in the probands (patients) and then look for overlap of all these regions with all affected members of that family, followed by searching for overlap with other families. This three-step process allows us to narrow and decrease the number of homozygous regions. The remaining overlapping regions can then be probed for candidate genes that are expressed in retina, using available databases.

In a preliminary study of our cohort, we screened families for known NCRNA and retinal detachment genes (including *FEVR* genes) which resided in one of the top five homozygous segments documented by the SNP array. We hypothesized that the causal gene for NCRNA resides in one of the top five largest intervals of our identified homozygous regions in consanguineous families and a new causal gene underlying the disease may locate in the overlapping homozygous regions of different family members with the same disease phenotype.

Analyzes of homozygous regions of the patients and direct sequencing of the candidate genes located in top five regions

TABLE 2. Summary of Analysis Done by Bioinformatics Tools for Novel Mutations

Family	Gene and Mutation	Polyphen2	SIFT	Blosum62 Score
NCRNA4	<i>ATOH7</i> c.G125C p. Arg42 Pro	Probably damaging (score 0.999)	Affect protein function (score of 0.00)	-2
NCRNA7	<i>LRP5</i> c.T235C p.Trp79Arg	Probably damaging (score 1.000)	Affect protein function (score of 0.00)	-3
NCRNA11	<i>NDP</i> c.T205G p.Cys69Gly	Probably damaging (score 0.997)	Affect protein function* (score of 0.00)	-3

Summary of analysis done by bioinformatics tools to predict the pathogenic nature of the novel mutations; In SIFT the amino acid change is predicted to be damaging if the score is  $\leq 0.05$ , and tolerated if the score is  $> 0.0515$ . On the other hand, in the PolyPhen-2 score, variants with scores of 0.0 are predicted to be benign (tolerated) and values closer to 1.0 are considered as deleterious.<sup>17</sup> Blosum62 scores range from +3 to -3; negative scores are presumed to be damaging.<sup>16</sup>

helped us to successfully genotype seven NCRNA families with known genes located in the regions of interest (Table 1).

### Clinical Findings

The affected individuals presented with the history of congenital blindness and presented with searching roving eye movements. Anterior segment examination showed small eyes. Some patients had cloudy corneas due to iridocorneal adhesion, shallow anterior chamber due to forward displacement of lens iris diaphragm, and irregular pupils secondary to posterior synechiae and leukocoria (white pupils); but there was no consistent elevation of intraocular pressure. Fundoscopy, when possible, revealed a dense fibrovascular mass behind the lens in each eye, originating from the optic nerve and attached behind the lens and peripheral fundus. In some cases, B-scan ultrasonography showed hyperechoic shadows involving the entire vitreous cavity. Other cases showed total retinal detachments and fibrous bands originating from optic nerve posteriorly and attached anteriorly behind the lens.

No other sensory or neurologic defects were detected in the affected individuals. Their auditory, olfactory, endocrine, and intellectual functions were intact. Together, these clinical findings suggest bilateral persistence of the fetal blood vessels, with complete congenital detachment of the neural retina and intravitreal vascular proliferation during infancy. The obligate heterozygotes are clinically normal. They have good vision, with no structural or functional eye pathology.

**Known Genes for NCRNA; ATOH7.** In our 21 Pakistani NCRNA families, only three families had *ATOH7* in their largest IBD regions (our mutation analysis confirmed this). Surprisingly, two of these families harbored the reported deletion in

*ATOH7*, which is the well-known, common mutation in the founder population of North Khorasan, in Iran.<sup>1</sup> Eight out of our 21 NCRNA families did not harbor mutations in *ATOH7*, nor in any other retinal detachment gene in their homozygous regions, suggesting that we have families with new gene/s for NCRNA. To be sure, we screened all our patients for *ATOH7* exonic and promoter mutations by Sanger sequencing. Two families (NCRNA5 and NCRNA6) were found to have the well-known 6523-bp deletion located 20 kb upstream from *ATOH7* (phenotypes of the patients are shown in Fig. 1). Segregation analyses for these families were conducted as described by Ghasvand et al.<sup>1</sup> (see Fig. 2). We also found a novel homozygous missense mutation p. Arg42Pro, altering a highly conserved arginine for a proline residue in *ATOH7* in the third family (NCRNA4). This is the first missense mutation reported

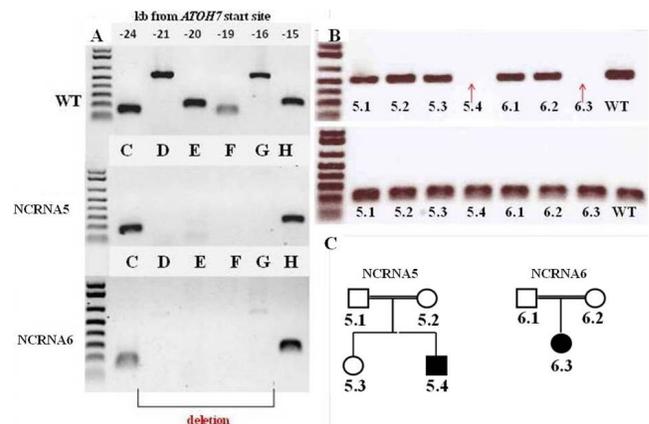


FIGURE 2. Polymerase chain reaction-based genotyping of homozygous 6523-kb deletion in 5' *ATOH7* in NCRNA5 and NCRNA6 families. (A) A polymerase chain reaction-based deletion confirmation in the NCRNA5 and NCRNA6 families conducted as described by Ghasvand et al.<sup>1</sup> Letters (C, D, E, F, G, H) represent six adjacent amplicons in an NCRNA subject that amplify conserved noncoding elements ~20 kb upstream of *ATOH7* transcription start site. As it can be seen from the figure, C and H amplicons are present in both wild-type (WT) and mutant subjects, while the rest of the amplicons are missing in NCRNA5 and NCRNA6 family members. (B) Cosegregation results between family members using primers designed by Ghasvand et al.<sup>1</sup> In the first panel, primers for G and H amplicons are pooled, whereas in the second panel, only primers for H amplicons were used to verify if deletion of the area is present in the members of the family. Red arrows show deletion in affected family members compared with a WT control band. (C) Pedigrees of NCRNA5 and NCRNA6 families.

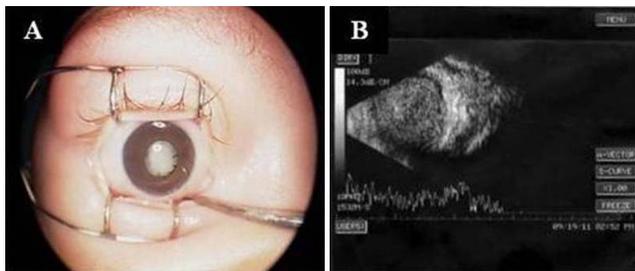


FIGURE 1. Anterior segment and B scan of NCRNA5-4. Family NCRNA5 was found to have the well-known 6523-bp deletion located 20 kb upstream from *ATOH7*. Anterior segment with leukocoria (A) and disorganized B scan showing retrolental mass with total vitreous condensation (B) caused by the variation in *ATOH7* gene in NCRNA5-4.

TABLE 3. Summary of Successfully Genotyped NCRNA Families

Family	Chromosome	Causal Gene	Change	Reported Mutations	Cosegregation
NCRNA2	11	<i>LRP5</i>	c.G1828C p. Gly610Arg hmz	Yes	Yes
NCRNA3	11	<i>LRP5</i>	c.G1300A p.Asp434Asn hmz	Yes	Yes
NCRNA4	10	<i>ATOH7</i>	c.G125C p. Arg42Pro hmz	New	Yes
NCRNA5	10	<i>ATOH7</i>	6523-bp deletion hmz	Yes	Yes
NCRNA6	10	<i>ATOH7</i>	6523-bp deletion hmz	Yes	Yes
NCRNA7	11	<i>LRP5</i>	c.T235C p.Trp79Arg hmz	New	Yes
NCRNA9	7	<i>TSPAN12</i>	IVS4-2A>G hmz	New	Yes
NCRNA11	X	<i>NDP</i>	c.T205G p.Cys69Gly hmz	New	Yes
NCRNA14	X	<i>NDP</i>	c.336_402IVS4del hmz	New	Yes
NCRNA16	11	<i>LRP5</i>	c.G425A p.Arg142Gln htz	Yes	Not able
NCRNA17	7	<i>TSPAN12</i>	c.423del T p. Pro141fsX21 hmz	New	Yes
NCRNA19	11	<i>LRP5</i>	c.G4202A p.Gly1401Asp hmz	Yes	Yes
NCRNA20	11	<i>LRP5</i>	c.T1622C p.Leu541Pro hmz	New	Not able

From 21 consanguineous NCRNA pedigrees 13 family have been successfully genotyped with mutations in known retinal genes; three families have mutations in *ATOH7* gene, and the rest of the families have mutations in *FEVR* genes; *LRP5* mutations (in six families); *TSPAN12* mutations (in two families); and *NDP* mutations (in two families). We did not find any damaging mutations in known retinal genes in eight families.

in *ATOH7*. In silico analyses predicts that this mutation is probably damaging (Table 2) and conservation analysis shows the significance of the residue (Fig. 3). All identified mutations are shown in Table 3.

p. R42P

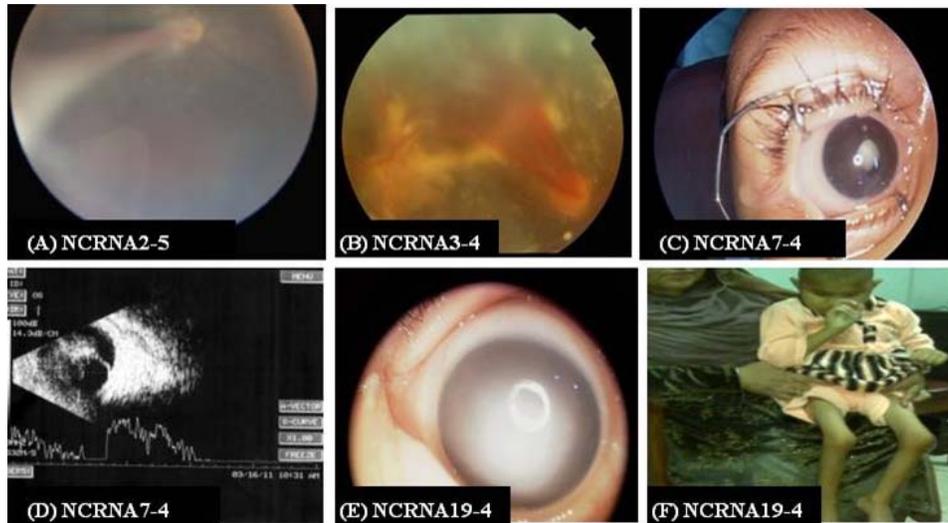
H. SAPIENS	GRLESAARRRLAANARERR
P. TROGLODYTES	GRLESAARRRLAANARERR
B. TAURUS	GRLESAARRRLAANARERR
C. LUPUS	GRLESAARRRLAANARERR
M. MUSCULUS	GRLESAARRRLAANARERR
R. NORVEGICUS	GRLESAARRRLAANARERR
G. GALLUS	ERMESAARRRLAANARERR
X. TROPICALIS	VRLEGSTKRRLAANARERR
D. RERIO	EKFESAMRRRLAANARERR

FIGURE 3. Mutation conservation analysis of *ATOH7*. We found a novel homozygous missense mutation p. Arg42Pro, altering a highly conserved arginine for a proline residue in *ATOH7* in the NCRNA4 family; this is the first missense mutation reported in *ATOH7*. In silico analyses predicts that this mutation is probably damaging (Table 2) and conservation analysis shows the significance of the residue.

**Mutations in *LRP5*.** The gene *LRP5* encodes a transmembrane low-density lipoprotein receptor and plays a key role in eye development, in skeletal homeostasis, and many bone density-related diseases (*LRP5*, OMIM #603506).

Although the *LRP5* gene is known for mild retinal detachments and folds and was located in IBD regions of the patients, *FEVR* was not considered as the primary diagnosis since these patients presented with congenital blindness. However, *LRP5* was the only attractive retinal candidate gene in the region, and Sanger sequencing identified the mutations in the gene.

We found five homozygous and one heterozygous (htz) mutations in the *FEVR* gene *LRP5*; namely, two members of each NCRNA2 (NCRNA2-5 and NCRNA2-6) and NCRNA3 (NCRNA3-4 and NCRNA3-7) families were carrying the reported p.Gly610Arg and p.Asp434Asn homozygous mutations, respectively. A member from a third family (NCRNA7-4) had a novel homozygous p.Trp79Arg mutation. We then identified the reported homozygous p.Gly1401Asp mutation in three members of a single family (NCRNA19-3, 19-4, and 19-6). Panel screening with RetNet then identified htz and hmz *LRP5* mutations in an additional two families (NCRNA16 and NCRNA20, respectively); NCRNA16 has the previously reported p.Arg142Gln change and the NCRNA20 family carries a



**FIGURE 4.** Eye phenotype for NCRNA2, NCRNA3, NCRNA7 and NCRNA19 families and skeletal abnormalities found in a NCRNA19 family member. (A) The fundus photo shows the fibrovascular fold arising from the optic nerve and extending to the retrolental area in the temporal region in NCRNA2-5; family NCRNA2 has been identified to carry a homozygous p. Gly610Arg alteration in *LRP5* gene. (B) Fundus photograph of NCRNA3-4 show hemorrhage retinal fold with a fibrovascular mass behind the lens which is a typical presentation of NCRNA1<sup>1</sup> caused by p.Asp434Asn mutation in *LRP5* gene in NCRNA3-4 proband. (C, D) Anterior segment and B-scan for NCRNA7-4. (C) Total leukocoria. (D) B-scan shows atrophy of the detached retina. (E) A member of NCRNA19 family with p. Gly1401Asp mutation showing dense corneal opacity. (F) Skeletal abnormalities (NCRNA19-4).

novel p.Leu541Pro alteration. Cosegregation of these mutations in the family members could not be checked.

A summary of the *LRP5* mutations is given in Table 3. Since the p.Gly1401Asp mutation in *LRP5* found in the NCRNA19 family members has previously been reported to cause osteoporosis-pseudoglioma,<sup>10</sup> we checked bone mineral density (BMD) in patient NCRNA19-4 and the result confirmed that the patient has reduced BMD and osteoporosis. The rest of the families were not able to come back for the given test.

The eye phenotype in our study consisted of congenital blindness with congenital retinal detachments and corneal opacity, posterior synechia, shallow AC, due to anteriorly displaced lens and iris, leukocoria, retrolenticular fibrovascular plaque, retinal folds (in some patients), and complete retinal detachment, unlike the current phenotypes associated with *LRP5* type FEVR (Fig. 4). The altered residues are all evolutionarily conserved (Fig. 5). We then documented delayed growth and skeletal abnormalities in some patients in these families (Fig. 4). Pedigrees for successfully genotyped NCRNA2, NCRNA3, NCRNA7 and NCRNA19 family positive for homozygous *LRP5* mutations are shown in Figure 6.

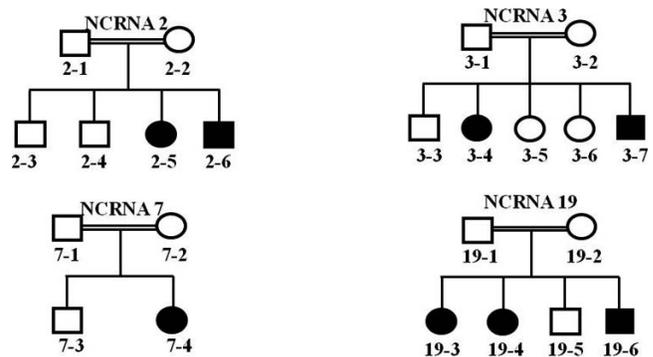
To confirm the molecular diagnosis, we performed NGS in these patients and excluded other possible mutations. It is important to note that this is the first report relating *LRP5* mutations with the NCRNA phenotype.

**Mutations in *TSPAN12*.** The gene *TSPAN12* encodes a cell-surface protein that is characterized by the presence of four hydrophobic domains and is predicted to mediate signal transduction events that play a role in the regulation of cell development, activation, growth and motility (*TSPAN12*, OMIM#613138). Tetraspanin-12 is currently known to cause autosomal dominant and recessive FEVR,<sup>21-23</sup> but not autosomal recessive NCRNA, and it is not known to cause severe retinal detachments.

We identified two new *TSPAN12* mutations for the first time in NCRNA. We confirmed a novel homozygous splice site mutation, IVS4-2A>G and a novel homozygous frameshift mutation, p. Pro141fsX21 in *TSPAN12* in two of our families (NCRNA9 and NCRNA17; Table 3). The Berkeley Drosophila Genome Project-splice site predictor analysis tool showed that the IVS4-2 A>G mutation found in NCRNA9 deletes the acceptor site. Cosegregation (the tendency for closely linked

	p.W79R
H. SAPIENS	FQFSKGA <b>W</b> VY <b>W</b> TDVSEEA <b>W</b> IKQ
P. TROGLODYTES	FQFSKGA <b>W</b> VY <b>W</b> TDVSEEA <b>W</b> IKQ
B. TAURUS	FQFSKGA <b>W</b> VY <b>W</b> TDVSEEA <b>W</b> IKQ
C. LUPUS	FQFSKGA <b>W</b> VY <b>W</b> TDVSEEA <b>W</b> IKQ
M. MUSCULUS	FQFSKGA <b>W</b> VY <b>W</b> TDVSEEA <b>W</b> IKQ
R. NORVEGICUS	FQFSKGA <b>W</b> VY <b>W</b> TDVSEEA <b>W</b> IKQ
G. GALLUS	FQYSQGIV <b>W</b> VY <b>W</b> TDVSEEA <b>W</b> IKQ
X. TROPICALIS	FLYSQGV <b>W</b> IY <b>W</b> TDVSEES <b>W</b> IKQ
D. RERIO	FLFSEGL <b>W</b> IF <b>W</b> TDVSEEA <b>W</b> IKQ

**FIGURE 5.** Mutation conservation analysis for *LRP5*. We identified a novel homozygous p.Trp79Arg mutation in family NCRNA7. Conservation analysis showed that the altered residue is evolutionarily conserved in many species.



**FIGURE 6.** Successfully genotyped NCRNA2, NCRNA3, NCRNA7 and NCRNA19 family pedigrees positive for homozygous *LRP5* mutations. Four families carrying mutation in *LRP5* gene were consanguineous.



FIGURE 7. Phenotypes of patients with *TSPAN12* mutations. (A, C) Anterior segment photographs show shallow AC, posterior synechiae, white pupils. (B) Total vitreous condensation. (D) Retrolental mass and atrophic retina with thin atrophic band attachment to optic nerve, confirming severe disease (red arrow shows site of the optic nerve).

genes and genetic markers to segregate together) analysis showed perfect cosegregation of both mutations in the families. We revisited the phenotypes in those patients and found leukocoria, corneal opacities, a shallow AC, posterior synechiae, and total retinal detachment (Fig. 7). Six families had *TSPAN12* as a candidate gene in their significant homozygous regions (Table 1), but only NCRNA9 and NCRNA17 had mutations in this gene. Pedigrees of the families positive for *TSPAN12* mutations are shown in Figure 8.

Thus, for the first time, we found recessive mutations in *TSPAN12* in clinically diagnosed NCRNA patients.

**Mutations in *NDP*.** The gene *NDP* encodes a protein that plays a key role in retinal vascularization and activates the Wnt signaling pathway with the help of its coreceptors, *FZD4* and *LRP5*. The gene is also thought to be implicated in neural cell differentiation and proliferation, as well as neuroectodermal cell-cell interaction (*NDP*, OMIM#300658). Mutations in *NDP* are known to cause X-linked exudative vitreoretinopathy.<sup>24</sup>

With the help of the RetNet panel and WES, we found *NDP* mutations in two families (NCRNA11 and NCRNA14). A novel homozygous 244-bp deletion in *NDP*, which starts from the end of the third exon, deleting 67-bp from exon three and 177-bp for 3' UTR, was identified in one of our families. Another novel mutation, p. Cys69Gly, was identified in a second family (NCRNA11, Table 3). This novel mutation is predicted to be

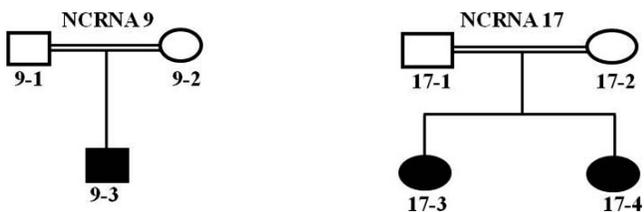


FIGURE 8. Pedigrees of patients with *TSPAN12* mutations. Pedigrees of families carrying mutations in *TSPAN12* gene.

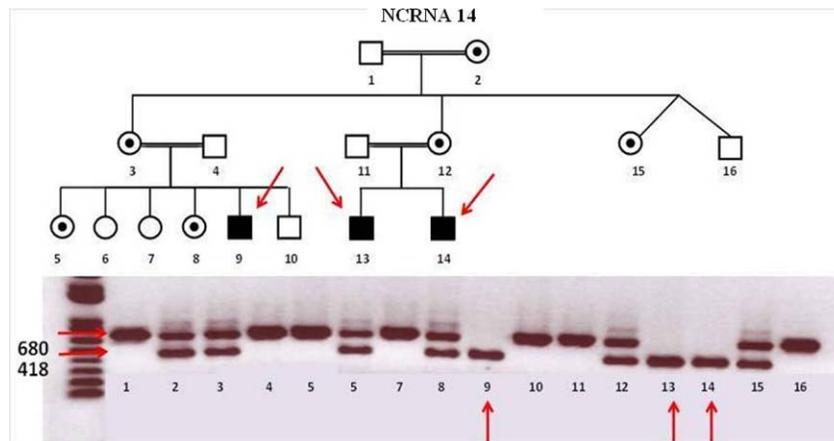
probably damaging by an online polymorphism phenotyping tool, PolyPhen,<sup>17</sup> with a score of 0.997 (Table 2) and the Cys residue is evolutionally highly conserved (Fig. 9). Both the deletion and the missense mutations cosegregate within the families. Segregation of the mutation between the NCRNA11 family members is shown in Figure 10.

Phenotypes of the patients with *NDP* mutations is consistent with the phenotype of previously described NCRNA families in Iran<sup>1</sup> and also similar to the phenotypes of families carrying mutations in *ATOH7*, *LRP5*, and *TSPAN12* genes described in this study; bilateral corneal opacities, shallow AC, posterior synechiae, retrolenticular fibrovascular plaque, and complete congenital retinal detachment.

Genetic causes of NCRNA in the remaining eight families were not identified after extensive SNP, WES, and RetNet panel testing and are still under investigations for novel candidate NCRNA genes.

	p. C69G
H. SAPIENS	VLLARCEGHCSQASRSEPL
P. TROGLODYTES	VLLARCEGHCSQASRSEPL
B. TAURUS	VLLARCEGHCSQASRSEPL
C. LUPUS	VLLARCEGHCSQASRSEPL
M. MUSCULUS	VLLARCEGHCSQASRSEPL
R. NORVEGICUS	VLLARCEGHCSQASRSEPL
G. GALLUS	VLLARCEGRCSQTSRSEPM
X. TROPICALIS	VLLARCEGRCSQTSRSDPL
D. RERIO	VLLARCEGHCSHTSRSDPL

FIGURE 9. Mutation conservation analysis for *NDP* variation. A novel mutation, p. Cys69Gly was identified in NCRNA11 family. This novel mutation is predicted to be probably damaging with a score of 0.997 (PolyPhen, Table 2) and the Cys residue is evolutionally highly conserved.



**FIGURE 10.** Family NCRNA14 pedigree and cosegregation result for *NDP* gene deletion. A novel homozygous 244-bp deletion in *NDP* gene (gene deletion: 43808839–43809101) was identified in NCRNA14 family. Mutation cosegregates within the family members. Red arrows show the affected members of the family. Family members with double bands (correspond to the sizes 418 and 680, WT and mutant bands, respectively) are heterozygous female carriers in the pedigree.

## DISCUSSION

Nonsyndromic congenital retinal nonattachment (OMIM #221900) is a rare genetic disorder defined by congenital retinal detachments and children are completely blind at birth. Often, the retinas are found to be fibrotic and appear as a white mass behind the crystalline lens.<sup>1–3</sup> The goal of this study was to dissect an inbred cohort of clinically diagnosed NCRNA patients, to determine the genetic causes of NCRNA, in the hope of getting a molecular or genetic entry point into adult retinal detachments, a much more common condition, without any known molecular determinants.

In this study, 28 Pakistani blind children from 17 consanguineous and 4 nonconsanguineous families with the diagnosis of NCRNA were analyzed with variety of gene-identifying techniques, including SNP microarrays searching for homozygous regions. All the patients were from Northern Pakistan where consanguineous marriages are very prevalent. As expected, the majority of the patients in our cohort carried significant homozygous regions in their genomes as a result of consanguineous marriages commonly practiced in the region.

In 7 (41%) of 17 consanguineous families, homozygosity mapping successfully identified the region and subsequently a disease-causing mutation in the only currently known NCRNA gene or other retinal disease genes. It is important to note that the mutated gene was in the largest or second largest homozygous region in 6 of the 11 families (Table 1) successfully genotyped using homozygosity mapping. In accordance with our previous study,<sup>25</sup> the data obtained in this current study suggest that the ranking of the IBD regions (continuous regions over which two haplotypes are identical by descent) can be very helpful to identify which homozygous segment contains the possible underlying causal and mutant gene. In the rest of the families, the known NCRNA and retinal disease genes located in significant homozygous regions were excluded from the study. The additional six successfully genotyped families were identified by RetNet panel screening (Table 3).

Although the clinical diagnosis and strict entry criteria of all children was NCRNA, in eight families, the molecular diagnosis determined that the disease process was in fact a new and severe form of FEVR. Familial exudative vitreoretinopathy is a rare disease and usually characterized by a much milder, acquired phenotype characterized by retinal folds, inferotemporal dragging of the optic disc and macula, increased vessels

in the equatorial region, and a peripheral avascular zone.<sup>26</sup> Familial exudative vitreoretinopathy was first described by Criswick and Schepens in 1969<sup>27</sup>; it is currently not known to be associated with the severe developmental and congenital retinal detachment in babies and children. Therefore, we have expanded the phenotypic spectrum of FEVR to a severe retinal detachment phenotype that clinically overlaps with NCRNA. In addition, we identified the identical large deletion (6523 bp) in the *ATOH7* gene found in the Kurdish founder population of Northern Iranian with a high incidence of NCRNA,<sup>1</sup> suggesting genetic overlap between the Iranian and Pakistani populations. Therefore, we suggest that this founder mutation in *ATOH7* gene links the two populations.

Our molecular genetics study of clinically evaluated and diagnosed patients with NCRNA revealed five important patterns. First, our work illustrates that NCRNA and FEVR overlap clinically; FEVR can be so severe that it can mimic NCRNA. Therefore, we show for the first time that the NCRNA phenotype can be caused by FEVR mutations. Second, we found the *ATOH7* deletion links the Pakistani population to the Iranian population. Possible explanations for this could be a common ancestor for both populations or substantial interbreeding by the patients' ancestors who all lived in the same general large geographical area. Third, we have found seven novel mutations: (1) a hmz c.125G>C, p.Arg42Pro mutation in *ATOH7*, (2) a hmz c.235T>C, p.Trp79Arg and (3) a hmz c.1622T>C, p.Leu541Pro mutation in *LRP5*, (4) a hmz splice site mutation IVS4-2 A>G and (5) a hmz frameshift mutation c.423del T, p.Pro141fsX21 in *TSPAN12*, and (6) a hmz c.205T>G, p.Cys69Gly and (7) a hmz 244-bp c.336\_402IVS4-del deletion in *NDP*. These families were informed about their results and were genetically counseled about family planning and future treatment options. Fourth, we confirmed that recessive mutations in *TSPAN12* can cause much more severe FEVR phenotype than the previously known dominant mutations related to the autosomal dominant FEVR in accordance with a previous study.<sup>18</sup> A summary of the successfully genotyped NCRNA patients are given in Table 3. In addition, we used our newly developed RetNet known retinal disease gene panel, which for the first time contains all the currently known retinal disease genes by targeted NGS which allowed us to screen all the genes listed in Retinal Information Network website and create candidate retinal disease gene list for future study, in case if they are involved in new unknown mechanisms.

## CONCLUSIONS

Although the clinical diagnosis of all children was NCRNA, the molecular diagnosis determined that the disease process was in fact a severe new form of FEVR in 13 families. However, the phenotype was severe congenital retinal detachment, previously not known to be associated with FEVR mutations. Therefore, we have expanded the phenotypic spectrum of FEVR, and we also show that the disease spectrum is much wider than previously established. Out of 21 Pakistani families with clinically diagnosed NCRNA, 10 families were discovered to have mutations in three FEVR genes: *LRP5* (six families), *TSPAN12* (two families), and *NDP* (two families), and three families were found to have mutations in *ATOH7*. Thus, eight NCRNA families in our cohort do not carry mutations in any known gene, which suggests that *ATOH7* is not the only gene to cause NCRNA in children. These NCRNA families likely have new causal genes. We show that SNP studies—followed by RetNet, WES, and Sanger confirmation—is a powerful protocol to identify the genetic cause of NCRNA. Finally, we established a genetic link between the Iranian and Pakistani populations, through a common founder deletion in *ATOH7*.

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## References

- Ghiasvand NM, Rudolph DD, Mashayekhi M, Brzezinski JA IV, Goldman D, Glaser T. Deletion of a remote enhancer near *ATOH7* disrupts retinal neurogenesis, causing NCRNA disease. *Nat Neurosci*. 2011;14:578-586.
- Phillips CI, Stokoe NL. Congenital hereditary bilateral nonattachment of retina: a sibship of two males. *J Pediatr Ophthalmol Strabismus*. 1979;16:358-363.
- Ghiasvand NM, Shirzad E, Naghavi M, Vaez Mahdavi MR. High incidence of autosomal recessive nonsyndromal congenital retinal nonattachment (NCRNA) in an Iranian founding population. *Am J Med Genet*. 1998;78:226-232.
- Brown NL, Patel S, Brzezinski J, Glaser T. Math5 is required for retinal ganglion cell and optic nerve formation. *Development*. 2001;28:2497-2508.
- Wang SW, Kim BS, Ding K, et al. Requirement for Math5 in the development of retinal ganglion cells. *Genes Dev*. 2001;15:24-29.
- Gilmour DE. Familial exudative vitreoretinopathy and related retinopathies. *Eye (Lond)*. 2015;29:1-14.
- Pendergast SD, Trese MT. Familial exudative vitreoretinopathy. Results of surgical management. *Ophthalmology*. 1998;105:1015-1023.
- Warden SM, Andreoli CM, Mukai S. The Wnt signaling pathway in familial exudative vitreoretinopathy and Norrie disease. *Semin Ophthalmol*. 2007;22:211-217.
- Warburg, M. Heterogeneity of congenital retinal non-attachment, falciform folds and retinal dysplasia: a guide to genetic counseling. *Hum Hered*. 1976;26:137-148.
- Robitaille J, MacDonald ML, Kaykas A, et al. Mutant frizzled-4 disrupts retinal angiogenesis in familial exudative vitreoretinopathy. *Nat Genet*. 2002;32:326-330.
- Toomes C, Bottomley HM, Jackson RM, et al. Mutations in *LRP5* or *FZD4* underlie the common familial exudative vitreoretinopathy locus on chromosome 11q. *Am J Hum Genet*. 2004;74:721-730.
- Jiao X, Ventruto V, Trese MT, Shastry BS, Hejtmancik JF. Autosomal recessive familial exudative vitreoretinopathy is associated with mutations in *LRP5*. *Am J Hum Genet*. 2004;75:878-884.
- Ai M, Heeger S, Bartels CF, Schelling DK; Osteoporosis-Pseudoglioma Collaborative Group. Clinical and molecular findings in osteoporosis-pseudoglioma syndrome. *Am J Hum Genet*. 2005;77:741-753.
- Untergasser A, Cutcutache I, Koressaar T, et al. Primer3- new capabilities and interfaces. *Nucleic Acids Res*. 2012;40:e115.
- Koressaar T, Remm M. Enhancements and modifications of primer design program Primer3. *Bioinformatics*. 2007;23:1289-1291.
- Wang F, Wang H, Tuan HF, et al. Next generation sequencing-based molecular diagnosis of retinitis pigmentosa: identification of a novel genotype-phenotype correlation and clinical refinements. *Hum Genet*. 2014;133:331-345.
- Coussa RG, Otto EA, Gee HY, et al. WDR19: an ancient, retrograde, intraflagellar ciliary protein is mutated in autosomal recessive retinitis pigmentosa and in Senior-Loken syndrome. *Clin Genet*. 2013;84:150-159.
- Kumar P, Henikoff S, Ng PC. Predicting the effects of coding non-synonymous variants on protein function using the SIFT algorithm. *Nat Protoc*. 2009;4:1073-1081.
- Henikoff S, Henikoff JG. Amino acid substitution matrices from protein blocks. *Proc Natl Acad Sci U S A*. 1992;89:10915-10919.
- Adzhubei IA, Schmidt S, Peshkin L, et al. A method and server for predicting damaging missense mutations. *Nat Methods*. 2010;248-249.
- Poulter JA, Davidson AE, Ali M, et al. Recessive mutations in *TSPAN12* cause retinal dysplasia and severe familial exudative vitreoretinopathy (FEVR). *Invest Ophthalmol Vis Sci*. 2012;53:2873-2879.
- Poulter JA, Ali M, Gilmour DE, et al. Mutations in *TSPAN12* cause autosomal-dominant familial exudative vitreoretinopathy. *Am J Hum Genet*. 2010;86:248-253.
- Nikopoulos K, Gilissen C, Hoischen A, et al. Next-generation sequencing of a 40 Mb linkage interval reveals *TSPAN12* mutations in patients with familial exudative vitreoretinopathy. *Am J Hum Genet*. 2010;86:240-247.
- Chen ZY, Battinelli EM, Fielder A, et al. A mutation in the Norrie disease gene (*NDP*) associated with X-linked familial exudative vitreoretinopathy. *Nat Genet*. 1993;5:180-183.
- den Hollander AI, Lopez I, Yzer S, et al. Identification of novel mutations in patients with Leber congenital amaurosis and juvenile RP by genome-wide homozygosity mapping with SNP microarrays. *Invest Ophthalmol Vis Sci*. 2007;48:5690-5698.
- Boonstra FN, van Nouhuys CE, Schuil J, et al. Clinical and molecular evaluation of probands and family members with familial exudative vitreoretinopathy. *Invest Ophthalmol Vis Sci*. 2009;50:4379-4385.
- Criswick VG, Schepens CL. Familial exudative vitreoretinopathy. *Am J Ophthalmol*. 1969;68:578-594.