Deletion of Seventeen Amino Acids at the C-Terminal End of Aquaporin 0Causes Distortion Aberration and Cataract in the Lenses of AQP0\(^{AC/AC}\) Mice

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Purpose. Investigate the effects of the absence of 17 amino acids at the C-terminal end of Aquaporin 0 (AQP0) on lens transparency, focusing property, and homeostasis.

Methods. A knockin (KI) mouse model (AQP0\(^{AC/AC}\)) was developed to express AQP0 only as the end-cleaved form in the lens. For this, AQP0 was genetically engineered as C-terminally end-cleaved with amino acids 1 to 246, instead of the full length 1 to 263 of the wild type (WT). After verifying the KI integration into the genome and its expression, the mouse model was bred for several generations. AQP0 KI homozygous (AQP0\(^{AC/AC}\)) and heterozygous (AQP0\(^{AC/AC}\)) lenses were imaged and analyzed at different developmental stages for transparency. Correspondingly, aberrations in the lens were characterized using the standard metal grid focusing method. Data were compared with age-matched WT, AQP0 knockout (AQP0 \(^{-/-}\)), and AQP0 heterozygous (AQP0 \(^{+/-}\)) lenses.

Results. AQP0\(^{AC/AC}\) lenses were transparent throughout the embryonic development and until postnatal day 15 (P15) in contrast to age-matched AQP0 \(^{-/-}\) lenses, which developed cataract at embryonic stage itself. However, there was distortion aberration in AQP0\(^{AC/AC}\) lens at P5; after P15, cataract began to develop and progressed faster surpassing that of age-matched AQP0 \(^{-/-}\) lenses. AQP0\(^{+/-}\) lenses were transparent even at the age of 1 year in contrast to AQP0 \(^{-/-}\) lenses; however, there was distortion aberration starting at P15.

Conclusions. A specific distribution profile of intact and end-cleaved AQP0 from the outer cortex to the inner nucleus is required in the lens for establishing refractive index gradient to enable proper focusing without aberrations and for maintaining transparency.

Keywords: AQP0, lens transparency, spherical aberration, cataractogenesis, C-terminal cleaved AQP0

The ocular lens needs to be transparent for proper vision. Structurally, the lens has a monolayer of epithelial cells at the anterior surface, beneath which lie multiple layers of fiber cells. The epithelial cells at the equator transform and differentiate into the secondary fiber cells. It is a continuous process. Because there is no cellular turnover, all the fiber cells produced in a life span are contained within the lens. As new fiber cells are added over the older differentiating fiber cells in the outer cortex, inner cortical, and outer nuclear fiber cells undergo maturation and compaction for which loss of cellular nutrients and removing the metabolic wastes is essential. These processes help to organize the fiber cells in a precise manner to minimize light scattering. The oldest fiber cells occupy the center of the ellipsoidal crystalline lens, which grows peripherally. Constant addition of the fiber cells increases the bulk and could lead to spherical aberration (SA), which creates multiple focal points and blurred vision. To circumvent this, the lens adjusts the refractive index gradient (RING),\(^{1,2}\) which varies from 1.386 to 1.406 from the cortex to the nucleus in humans\(^^{3,4}\) and 1.35 to 1.55\(^^{5}\) in mouse. RING adjustment is a cooperative process mediated by several proteins and post-translational modifications.

Aquaporin (AQP) water channels are integral membrane proteins belonging to the superfamily of Aquaporins. They are expressed in microbes, plants, and animals and are critical for cellular development, growth, and homeostasis. AQP5 allow transcellular passage of water (aquaporins) or water and small solutes like glycerol (aquaglyceroporins). Water channels, gap junction channels, ion transporters, and cotransporters are involved in creating a microcirculation within the lens to compensate for its avascular nature by providing the necessary nutrients and removing the metabolic wastes.\(^{5-8}\)

In mammals, 13 different AQP genes have been identified. The mammalian lens expresses AQP0, AQP1, and AQP5. Mammalian AQP0 is a 28-kDa protein that is profusely expressed in the lens contributing approximately 45% of the total membrane proteins of the fiber cells.\(^^{9}\) It is a multifunctional protein. Water permeability,\(^{10-14}\) and cell-to-cell adhesion (CTCA)\(^{15-20}\) functions of AQP0 are important for maintaining lens transparency.\(^{12,15,16,21}\) AQP0 displays a characteristically low water permeability,\(^{11,12,14}\) which is several-fold less compared with its lens counterparts AQP1 and AQP5. Mutations and knockout of AQP0 alter ocular growth and result in dominant cataracts due to the loss of lens homeostasis.
Intact AQP0 is Necessary for Lens Transparency

**Figure 1.** Strategy to generate a AQP0*ΔC/ΔC* by introducing a stop codon after amino acid 246. (A) WT: Schematic structure of WT mouse AQP0 gene showing exons 1-4 (as rectangular vertical or horizontal boxes) and the connecting introns. Vector: Exons 3 and 4 with introns (highlighted in blue and red) as well as a Neo selection gene were amplified by PCR and cloned into the vector (details in [B]). Black dotted lines on either side denote vector sequences. Asterisk indicates an in-frame translation stop codon predicted to truncate AQP0 after the amino acid Asparagine-246. KI-Neo: The recombinant vector (with Exons 3 and 4, introns highlighted in blue and red and the Neo selection gene) was transfected into mouse embryonic stem cells and positive clones were selected using the Neo selection marker. KI: KI selection gene) was transfected into mouse embryonic stem cells and positive embryonic stem cells selected were injected into mouse blastocysts to develop AQP0 KI mouse model (AQP0*ΔC/ΔC*). (B) Schematic of the introduction of the stop codon through a point mutation incorporated into a primer pair (to delete the 17 amino acids after the 246th), and the KI targeting vector design. The C-terminal end deletion was engineered by overlap extension PCR using the primers as indicated. Red rectangle represents the deletion region. SA, short homology arm; LA, long homology arm. N1, N2, PT2, PT3, PT4, P6, and T75 are forward or reverse primers used, as indicated.

whereas, knockout of AQP125 or AQP526 does not cause cataract under normal conditions. The cataract phenotype due to alteration in AQP0, highlights the importance of the unique function of this protein for lenticular homeostasis.2,16,23,27–34

particularly important PTM is N- and/or C-terminal end-truncation, which begins at the inner cortex of the lens. Presence of truncated forms increases as the fiber cells age. Higher concentrations of such forms were noticed in the nuclear region of the lens than in the cortex. Presence of cleaved forms of AQP022,41–45 crystallin,56,47 connexins (Cx5048,51 and Cx4652,53), and beaded filament proteins54,55 in the fiber cells does not affect lens transparency. N-terminal end-cleavage sites, which remove 2 to 6 and 2 to 12 amino acids of AQP0 in the normal lens have been reported.42,45,56,57 C-terminal end-cleavage sites at residues 228, 234, 239, 243, 245, 246, 247, 249, and 259 in human AQP0 have been identified using mass spectrometry.43,45

Among the three AQP0s expressed in the lens, only AQP0 undergoes gradient loss of N- and C-terminal ends.33,45,57 The cleavage of N- and C-terminal ends of AQP0 has long been considered as a consequence of the onset of senescence. However, this type of PTM occurs as early as 2 years after birth in the human lens.1 There is a dearth of investigations aimed at understanding the significance of the presence of intact and cleaved forms of AQP0, connexins, and crystallins in gradients in the lens. We developed an AQP0 knockout (KI) mouse model (AQP0*ΔC/ΔC*) by engineering the AQP0 gene to express a predominant posttranslationally C-terminal end-cleaved form having amino acids 1 to 246. This mouse model was further investigated to understand the necessity for the presence of intact AQP0 (1–263 amino acids) with reference to lens optical quality and focusing. Our results clearly demonstrate that absence of intact AQP0 in the lens causes distortion aberration and cataract.

**Materials and Methods**

**Animals**

The wild type (WT) and mouse models used in this investigation are in C57BL/6J (The Jackson Laboratory, Bar Harbor, ME, USA) inbred strain which does not carry the CP49 gene mutation. WT (AQP0+/+), AQP0 heterozygous (AQP0+/−), AQP0 knockout (AQP0−/−), and a newly developed C-terminally end-deleted AQP0 mutant KI in homozygous (AQP0*ΔC/ΔC*) and heterozygous (AQP0*ΔC/+ΔC+) genotypes were used. WT mouse in FVB strain was used as a positive control for CP49 mutation. For animal procedures, the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research, the National Institutes of Health’s (NIH; Bethesda, MD, USA) Guide for the Care and Use of Laboratory Animals and protocols approved by Stony Brook University Animal Care and Use Committee were followed.

**Generation of AQP0-1-246 Mutant Knockin (AQP0*ΔC/ΔC*) Mouse Model**

A truncated mutant AQP0 KI mouse model AQP0*ΔC/ΔC* was developed through inGenious Targeting Laboratory, Inc. (Ronkonkoma, NY, USA). This model expresses a major form of C-terminally end-cleaved AQP0 (that lacks amino acids 247–265), which is observed in human, bovine, and mouse lens nuclear regions.22,43,45 The schematic diagram (Figs. 1A, 1B) depicts the strategy used for developing AQP0*ΔC/ΔC* mutant knock-in mouse model (details on the KI mouse model development are given in the Supplementary Section).

**Verification of the Absence of CP49 Mutation in the Knockin Mouse Model**

KI mice (AQP0+/ΔC and AQP0*ΔC/ΔC+) were genotyped by PCR to verify the absence of the mutation in the beaded filament...
proteins of WT, AQP0 validate CP49 protein expression, total lens membrane
AQP0 was translated into a histogram using SigmaPlot 10 software
The PCR product amplified by primers revneo3b and MOPE4 was used to screen mice for the introduced stop codon. (A) A 676-bp fragment was amplified using revneo3b/MOPE4 primers. MOPE4 is located on the long homology arm (LA, see Fig. 1B), downstream of the deletion. Revneo3b is located inside the Neo cassette. Among the eight agouti-colored chimeras tested, three (#889, 890, and 893) were positive. The PCR product amplified by primers revneo3b and MOPE4 was sequenced to confirm the presence of the introduced stop codon and the fidelity of the amplified fragment. (B) Screening for Neo deletion. A primer set Neo deletion primer 1 (NDEL1) and NDEL2 was used to screen mice for the deletion of the Neo cassette. The PCR product for the wild-type is 233 bp. A second band with 307 bp indicates Neo deletion in the chimeras (after Neo deletion, one set of LoxP-FRT sites remain [74 bp] in the KI chimeras). (C) Screening for presence of FLP transgene. A primer set, FLP1 and FLP2, used to screen mice for presence of the FLP transgene amplified a positive product of 725 bp; (D) FLP-positive and Neo-negative mice were crossed with C57 WT to eliminate FLP transgene. KI mice #889, 890, and 893 were FLP positive mutant chimeras were crossed with C57 WT mice to eliminate FLP transgene; deletion of the FLP transgene was confirmed by genomic PCR screening of offspring using primer set NDEL1 and NDEL2. Mutant chimeras showed a PCR product of 233 bp for the WT allele and a 307-bp amplicon for the mutant allele. These procedures were performed basically as described by Varadaraj et al. (details are provided in the Supplementary Section). Statistical Analysis SigmaPlot 10 software was used for Student’s t-tests. P values ≤ 0.05 were considered significant.

RESULTS
Introduction of a stop codon after amino acid 246 in mouse AQP0 was achieved by homologous recombination. Genomic sequencing showed that there was no disruption in the native AQP0 locus (data not shown). To verify the presence of the introduced mutation, genomic DNA from eight mouse chimeras with agouti coat color (#888, 889, 890, 891, 892, 893, 894, and 895) was PCR amplified using primers revneo3b located inside the Neo cassette and MOPE4 located on the long homology arm. Among the eight chimeras tested, #889, 890, and 893 amplified a predicted 676-bp DNA segment (Fig. 2A); the amplicons were sequenced for verification of the introduced stop codon and deletion. To delete the Neo gene in the mutant chimeras, #889, 890, and 893 were crossed with FLP transgenic mice. Deletion of the Neo cassette was confirmed by genomic PCR screening of offspring using primer set NDEL1 and NDEL2. Mutant chimeras showed a PCR product of 233 bp for the WT allele and a 307-bp amplicon for the Neo deletion of the mutant allele (Fig. 2B). Presence of the FLP gene was confirmed in all three chimeras by PCR, which amplified a 725-bp fragment (Fig. 2C). The three positive mutant chimeras were crossed with C57 WT mice to eliminate FLP transgene; deletion of the FLP transgene was confirmed by genomic PCR (Fig. 2D). The FLP-negative mice were crossed with WT C57 mouse for more than 15 generations before characterizing the phenotype. Interbreeding between heterozygous (AQP0+/ΔC) parents produced offspring in a Mendelian ratio, as determined by PCR analysis (data not shown).

KI mouse pups were genotyped to authenticate the absence of CP49 deletion mutation. WT and KI (AQP0+/ΔC and AQP0ΔC/ΔC) mice genomic DNAs amplified a 320-bp PCR
monoclonal AQP0 antibody (Santa Cruz Biotechnology, Inc.) (more information is given in the Supplementary Section).
AQP0 exhibited much severe cataract compared with age-matched AQP0+/+ mice (Fig. 5D, left column). At P20 the lenses of AQP0−/− were opaque, whereas AQP0+/− showed normal lens transparency similar to that of WT at P5 (Fig. 5B, left column). However, at P10 AQP0−/− showed normal lens transparency similar to that of WT at P5 (Fig. 5D, left column). At P15 the transparency significantly decreased to an opaque condition. Immunohistochemical analysis of lens sections at different ages showed that WT expressed the 28.2-kDa protein corresponding to intact AQP0, whereas AQP0−/− lens expressed only the cleaved form of 26.5 kDa (Figs. 4A, 4B). Quantification of the immunoreactive bands representing the intact and cleaved forms in AQP0−/− lenses showed no statistical significance (P > 0.05) in antibody binding. Immunohistochemical analysis of lenses of P5 sagittal and cross sections of WT and AQP0−/− showed fiber cell membrane localization of C-terminal end-cleaved AQP0, very similar to the intact AQP0 expressed in the fiber cell plasma membrane of the WT lens (Fig. 4B); quantification bar graph showed comparable intensity in antibody binding between WT and KI lens cross sections.

After verifying normal trafficking and membrane localization of AQP0 KI protein, we imaged the lenses at different stages, such as embryonic day 18 (E18), P5, P10, P15, and P20 to examine whether there is any phenotypic difference compared with age-matched WT and AQP0+/− lenses. At E18, lenses of AQP0−/− resembled those of the WT, in contrast to AQP0−/− lenses, which showed lens opacity. Representative images are given as Figure 5A (left column). AQP0+/− showed normal lens transparency similar to that of WT at P5 also (Fig. 5B, left column). However, at P10 AQP0−/− lens began to show opacity at the nuclear region (Fig. 5C, left column) and by P15 the transparency significantly decreased (Fig. 5D, left column). At P20 the lenses of AQP0−/− exhibited severe cataract compared with age-matched AQP0+/− lenses (Fig. 5E). Lens transparency was quantified using MALDI-TOF MS and Western blotting analyses. Both modes of protein expression analyses showed that WT expressed the 28.2-kDa protein corresponding to intact AQP0, whereas AQP0−/− expressed 28.2 and 26.5 kDa polypeptides corresponding to intact and cleaved form; AQP0+/− lens expressed only the cleaved form of 26.5 kDa (Figs. 4A, 4B). Quantification of the immunoreactive bands representing the intact and cleaved forms in AQP0−/− lenses showed no statistical significant difference (P > 0.05) in antibody binding. Immunoblotting of total lens membrane proteins of WT, AQP0−/−, and AQP0+/− mice using CP49 antibody verified the expression of CP49 (Fig. 3B).

To examine whether there is any phenotypic difference compared with age-matched WT and AQP0+/− lenses, we conducted MALDI/TOF/MS analysis of 5-day-old mouse lens proteins. AQP0+/− lenses expressed the 28.2-kDa protein corresponding to intact AQP0, whereas AQP0−/− expressed 28.2 and 26.5 kDa polypeptides corresponding to intact and cleaved form. Immunoblotting of total lens membrane proteins of WT, AQP0−/−, and AQP0+/− mice using CP49 antibody verified the expression of CP49 (Fig. 3B). Protein expression profile of WT, AQP0−/−, or AQP0+/− mouse lenses at P5 was characterized by MALDI-TOF MS and Western blotting analyses. Both modes of protein expression analyses showed that WT expressed the 28.2-kDa protein corresponding to intact AQP0, whereas AQP0−/− expressed 28.2 and 26.5 kDa polypeptides corresponding to intact and cleaved form; AQP0+/− lens expressed only the cleaved form of 26.5 kDa (Figs. 4A, 4B). Quantification of the immunoreactive bands representing the intact and cleaved forms in AQP0−/− lenses showed no statistical significant difference (P > 0.05) in antibody binding. Immunohistochemical analysis of lenses of P5 sagittal and cross sections of WT and AQP0−/− showed fiber cell membrane localization of C-terminal end-cleaved AQP0, very similar to the intact AQP0 expressed in the fiber cell plasma membrane of the WT lens (Fig. 4B); quantification bar graph showed comparable intensity in antibody binding between WT and KI lens cross sections.
using pixel brightness intensity, which is inversely proportional to transparency (Fig. 5F). Development of lens opacity, which began after P5 in AQP0\(^{+/+}\) almost doubled with each 5-day postnatal progression in age. By P20, it surpassed the severity of cataract in AQP0\(^{+/+}\) lens. The difference in the severity of cataract between the AQP0\(^{+/+}\) and AQP0\(^{D^+/D^+}\) lenses was statistically significant. Star in Figure 5F denotes the statistically significant difference compared with the WT lenses (P < 0.001) and the triangle indicates significance compared with AQP0\(^{+/+}\) (P < 0.05).

We tested the lenses for their ability to focus the lines of a copper grid. At E18, the transparent AQP0\(^{D^+/D^+}\) lens focused...
the gridlines well like the WT lens and in contrast to the AQP0⁺/⁻ lens (Fig. 5A, right column). At P5, WT lenses focused copper grid lines sharply (Fig. 5B, right column) and AQP0⁺/⁻ lenses showed SA due to the cataractous nature of the lens. The AQP0⁺/⁻/⁻ lens, which appeared transparent produced distorted grid lines especially at the nuclear region. At P10 and P15, AQP0⁺/⁻/⁻ displayed spreading of the central nuclear aberration to the periphery (Figs. 5C, 5D). AQP0⁺/⁻ lenses showed increase in the severity of SA throughout the lens. By P20 (Fig. 5E), lens cataract was severe in AQP0⁺/⁻/⁻ and AQP0⁺/⁻/⁻, notably more in the latter.

The changes we observed in the AQP0⁺/⁻/⁻/⁻ lenses, prompted us to examine the lenses of AQP0⁺/⁻ mice. At early developmental stages, such as P5 and P10, AQP0⁺/⁻ lenses with one copy of intact AQP0 did not show lens opacity and aberration like AQP0⁻/⁻/⁻ lenses (data not shown). At P15, dark-field images showed comparable lens transparency (Fig. 6A, left column). However, by P15, both AQP0⁻/⁻/⁻ and AQP0⁻/⁻/⁻ lenses showed aberration, which was apparent at a much higher degree in the former. AQP0⁻/⁻/⁻ lenses showed a distinct pattern of central nuclear aberration as revealed by grid focusing (Fig. 6A, right column). Adult 2- (Fig. 6B) and 12-month-old (Fig. 6C) AQP0⁺/⁻/⁻ lenses showed increased light scattering and central nuclear aberration (red circle), in contrast to global SA in AQP0⁺/⁻/⁻ lens. Figure 6D shows that lens transparency of AQP0⁺/⁻/⁻ continued to deteriorate with progression in age and the deterioration was statistically significant (*P* < 0.05) compared with the WT at each age shown; however, in AQP0⁺/⁻/⁻ lens transparency was significantly higher than in AQP0⁺/⁻/⁻. Optical quality of WT, AQP0⁺/⁻, AQP0⁻/⁻, AQP0⁺/⁻/⁻ and AQP0⁻/⁻/⁻ lenses was analyzed from the images of the lenses focusing the electron microscopy grid. The grid line pattern magnified by WT lenses at the tested ages showed a positive barrel distortion aberration (Figs. 5, 6) in which the straight lines bend outward from the image center. The grid line pattern was significantly distorted throughout the lenses in AQP0⁻/⁻/⁻ and AQP0⁻/⁻. In homozygous KI (AQP0±/±) lenses,
the much-distorted grid line pattern displayed two distinct zones at P5 (Fig. 5B). Zone I in the cortex showed a barrel distortion aberration (positive radial distortion) as in the WT lenses (Fig. 5B) and Zone II at the lens nucleus showed an abnormally increased barrel distortion aberration compared with WT-lens nucleus. These distortion aberration zones progressively increased in size with the age of the lenses (compare Figs. 5B and 5C). P10 (Fig. 5C) lens exhibited a tendency toward transitioning to a pincushion distortion aberration (negative radial distortion) in the nuclear region. At P15, AQP0<sup>MC/MC</sup> showed three distinct aberration zones, I, II, and III (Fig. 5D). At this stage, the nuclear Zone II clearly exhibited a pincushion distortion aberration and a new Zone III was established between the Zones I and II (i.e., at the corticonuclear junction) with a greater barrel distortion aberration than that of cortical Zone I. In the heterozygous (AQP0<sup>−/−</sup>) lenses, only two distortion aberration zones start to appear at age P15 (Fig. 6A), as in the case of AQP0<sup>MC/MC</sup> lenses at P5 (Fig. 5B). These distortion aberration zones slowly progressed with the age of the lens (compare Figs. 6A–C). While P15 (Fig. 6A) lens showed more of a barrel distortion, the 2-month-old lens clearly showed a pincushion distortion (Fig. 6B). There was no distinct Zone III in AQP0<sup>−/−</sup> mouse lenses (Fig. 6C). The distortion aberration zones are shown as a schematic model (Fig. 7).

To find out whether the end-cleavage process is part of lens development or a consequence of senescence, we performed Western blotting of membrane proteins extracted from lens fiber cells of P10 pups and adult WT mice. Membrane proteins extracted from AQP0-transfected MDCK cells served as negative control for posttranslationally end-cleaved AQP0. As anticipated, protein samples from MDCK cells transfected with mouse intact AQP0 served as specific band of 28.2 kDa for expression of the transfected AQP0 indicating the absence of posttranslational truncation. Protein samples from MDCK cells exhibited a negative control for posttranslationally end-cleaved AQP0. As anticipated, protein samples from MDCK cells transfected with mouse intact AQP0 served as specific band of 28.2 kDa for expression of the transfected AQP0 indicating the absence of posttranslational truncation. As expected, Western blotting of membrane proteins extracted from lens fiber cells of P10 pups and adult WT mice showed two immunoreactive bands of 28.2 and 25.6 kDa, as observed for P10 lenses (Fig. 8). These results demonstrate that end-truncation begins at a very early age in life and is not a senescence-dependent phenomenon.

**DISCUSSION**

Starting from the outer cortex and leading into the inner nucleus, lens has an array of gradients. Along this gradient, the intracellular pH reads 7.2 to 6.8<sup>nM</sup> Ca<sup>2+</sup> from 300 to 700 nM<sup>g</sup>, protein gradient (less to more), water gradient (more to less) membrane proteins like AQP0, and cytosolic proteins like crystallins as intact soluble (more to less), N- and/or C-terminal end-cleaved insoluble forms (less to more), and refractive index gradient (RING: 1.386–1.406 in human).<sup>3,5</sup> In accordance with this gradient trend, N- and/or C-terminal ends of AQP0, connexin, and crystallin proteins are progressively cleaved toward the center of the lens and remain there throughout life. The purpose behind these events remained puzzling. In a previous study using an AQP0<sup>−/−</sup> and a transgenic model that expresses AQP1 in the fiber cells of AQP0<sup>−/−</sup> mouse, we hypothesized that intact and end-cleaved AQP0 must be playing a crucial role in adjusting the lens RING.<sup>22</sup> The current study corroborates that by demonstrating distortion aberration in the lenses of the AQP0<sup>MC/MC</sup> mouse model that expresses only cleaved AQP0. Arrangement of intact and cleaved forms in a gradient from the outer cortex to the inner nucleus must be the norm to ensure sharp focusing.

AQP0 is essential for embryonic and postnatal development. This is highlighted in AQP0<sup>−/−</sup> lens in which loss of lens transparency is fast and severe starting from the early developmental stages (Fig. 5). However, in AQP0<sup>MC/MC</sup> lens cataract development is either absent or slow in the initial developmental stages but is accelerated in the postnatal stages (Fig. 5E); absence of intact AQP0 during these stages, results in cataract that is more severe than in AQP0<sup>−/−</sup>/C0 mouse lens there was no distinct Zone III in AQP0<sup>−/−</sup>/C0, lens cortex in which loss of lens transparency is fast and severe starting from the early developmental stages (Fig. 5). However, in AQP0<sup>MC/MC</sup> lens cataract development is either absent or slow in the initial developmental stages but is accelerated in the postnatal stages (Fig. 5E); absence of intact AQP0 during these stages, results in cataract that is more severe than in AQP0<sup>−/−</sup>/C0, suggesting that presence of C-terminal end-cleaved AQP0 in the absence of the intact form could alter lens homeostasis significantly. It can be speculated that alterations in the expression level of intact AQP0 in the differentiating cortical fiber cells and abnormal accelerated N- and C-terminal end-cleavage during senescence could cause lens opacity and cataract due to the loss of homeostasis.

E18 lens showed severe cataract in AQP0<sup>−/−</sup>/C0 whereas the age-matched AQP0<sup>MC/MC</sup> lens was transparent without any aberrations (Fig. 5A). In the AQP0<sup>MC/MC</sup> mouse lens there was gradual development of distortion aberration until P15 followed by cataractogenesis (Figs. 5, 6). Absence of intact AQP0 does not appear to affect lens homeostasis significantly during early developmental stages in the AQP0<sup>MC/MC</sup> lens but is essential during the later stages of postnatal lens development and for transparency. A notable result is the presence of distortion aberration in early postnatal lenses, such as in P5, suggesting that presence of C-terminal end-cleaved AQP0 in the absence of the intact form could alter lens homeostasis significantly.
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...even though there was no cataract. Because distortion aberration is a manifestation of alterations in the RING, it can be inferred that albeit the AQP0\textsuperscript{WT/MC} lens manages to be transparent and prolong cataractogenesis, starting at an early age it is unable to establish and maintain the proper RING to avoid distortion aberration. Our data show that in the cortex, presence of the 17 amino acids at the C-terminal end of AQP0 is essential for RING development to ensure transparency and sharp focusing of the lens.

In AQP0\textsuperscript{WT/C} mouse lens, presence of 50% of intact and cleaved forms caused loss of transparency, increased SA, and cataract. Presence of 100% of the cleaved forms (as in a WT lens) and 50% intact AQP0 in the AQP0\textsuperscript{WT/MC} prolonged the onset of cataract. However, absence of intact AQP0 in the ratio as is present in the WT, did create distortion aberration at P15. AQP0\textsuperscript{MC/MC} developed three distinct distortion (aberration) zones. This could be due to the loss of the arrangement of intact AQP0 and postranslationally end-cleaved forms in a specific manner from the cortex (more intact AQP0 and less cleaved-AQP0) to the nucleus (less intact AQP0 and more cleaved-AQP0), which is required for lens RING development. Recent studies on a connexin 46 (Cx46) mutant mouse model (Cx46fs380) showed a barrel distortion and a pincushion distortion in the lenses. The phenotypic difference in distortion aberration between AQP0\textsuperscript{MC/MC} and Cx46fs380 mouse models could be due to the functional difference of these proteins in the fiber cell membrane. Also, the mutant Cx46 did not traffic to the membrane whereas the C-terminal end-cleaved AQP0 trafficked to the plasma membrane (Fig. 4B). Our results show that intact and cleaved forms of AQP0 in the appropriate ratio in the lens is essential for the establishment and maintenance of RING, transparency, and homeostasis.

The lens grows continuously. Normal end-cleavage of AQP0 is not aging-related (senescence-related) but it is associated with fiber cell maturation process to adjust the lens RING. For example, in early-postnatal young lenses, such as in P10, differentiating fiber cells are more and contain only intact AQP0, the number of which is more than that in the mature fiber cells that carry both intact and cleaved AQP0. The opposite is true in the adult lenses. P10 lenses have less end-cleaved AQP0 due to the small volume of mature fiber cells than adult lenses that contain a large volume of mature fiber cells and more end-cleaved AQP0. The difference in the ratios of intact versus cleaved AQP0 between young and adult is not due to aging but due to the amount of differentiating and mature fiber cells in the lens. Mature adult contains more mature fiber cells in the inner cortex and nucleus and therefore the volume of cleaved forms is more. The 17 amino acid C-terminal end-cleaved AQP0 also showed a similar trend in human lens; however, there was senescence-related abnormally accelerated rate of C-terminal end-cleavage in AQP0. The study by Kort limbinis et al. on human lens AQP0 shows that normal C-terminal end-cleavage is a part of the fiber cell maturation process and an increase in intact versus cleaved forms is mainly due to the size of the lens; second, the aging process could induce abnormally accelerated end-cleavage and lead to aging-related blurred vision.

With its virtues of low water permeability, high CTCAs property, and capability of undergoing end-cleavage, AQP0 could play a major role in the formation and maintenance of RING. Intact AQP0 forms small patches of square array thin junctions in the cortex and cleaved forms facilitate formation of large patches of thin junctions in the nuclear region, potential to regulate fiber cell adhesion efficiency. The C-terminal ends of intact AQP0 at the outer cortex interact with other proteins, such as beaded filament proteins filensin and CP49, to provide anchorage; fiber cell architecture, and lens biomechanics. Interaction of adhesion protein N-Cadherin and cytoskeletal elements during fiber cell elongation has been reported. CTCA is necessary to maintain the proper RING, which is low at the outer cortex and high in the central nucleus where it is more or less constant. Absence of C-terminal end of AQP0 in the KI lens could be facilitating the formation of large patches of thin junctions globally in the lens and such increased CTCA of unfavorable mesures at the outer cortex could be correspondingly altering the refractive index at this region. Our data suggest that presence of intact AQP0 is a necessity to establish the RING conducive for proper focusing. To adjust the refractive index, end-cleavage should occur gradually, starting from the inner cortex, concomitant with dehydration of the fiber cells. Slow and steady processes like these within the fiber cells confer optical and biomechanical properties suitable for focusing light from objects near or far.

In conclusion, a conducive ratio of intact and C-terminal end-cleaved forms of AQP0 must be maintained in a spatial and temporal distribution manner to maintain lens transparency, RING and homeostasis in the continually growing lens for precise focusing. Until the questions of how the end-cleavages occur in the lens and what are the mechanisms involved are deciphered, developing a reciprocal mouse model with only intact AQP0 in the lens would remain a far-fetched goal.

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