Maternal Pertussis Immunization and Immunoglobulin G Levels in Early- to Late-Term and Preterm Infants

Maarten M. Immink, MSc; Mireille N. Bekker, MD, PhD; Hester E. de Melker, PhD; Gerco den Hartog, PhD; Nynke Y. Rots, PhD; Pieter G. M. van Gageldonk, BSc; Floris Groenendaal, MD, PhD; Elisabeth A. M. Sanders, MD, PhD; Nicoline A. T. van der Maas, MD, MSc; for the Dutch Maternal Pertussis Vaccine Investigation Group

Abstract

IMPORTANCE Maternal tetanus, diphtheria, andacellular pertussis (Tdap) vaccination protects newborns against severe pertussis. Data on transplacental antibody transfer on Tdap vaccination before 24 weeks' gestation remain scarce and are particularly relevant for preterm infants to increase the time interval for maternal antibody transfer.

OBJECTIVE To assess noninferiority of anti–pertussis toxin (anti-PT) immunoglobulin G (IgG) antibody levels at age 2 months in early- to late-term infants following Tdap vaccination between 20 0/7 and 24 0/7 weeks' gestation compared with 30 0/7 and 33 0/7 weeks' gestation and compared with preterm infants.

DESIGN, SETTING, AND PARTICIPANTS This prospective, multicenter cohort study included pregnant women aged 18 years or older in birthing centers and hospitals in the Netherlands between August 2019 and November 2021 who received Tdap vaccination between 20 0/7 and 24 0/7 weeks' gestation. Women with imminent premature birth were recruited if they had received maternal Tdap vaccination between 20 and 24 weeks' gestation. Blood samples were collected from mothers at delivery, from the umbilical cord, and from infants at age 2 months. Data from infants' blood samples at age 2 months were compared with a reference cohort (recruited between January 2014 and February 2016) of early- to late-term infants of the same age whose mothers had received Tdap vaccination between 30 0/7 and 33 0/7 weeks' gestation.

EXPOSURE Maternal Tdap vaccination between 20 0/7 and 24 0/7 weeks' gestation or 30 0/7 and 33 0/7 weeks' gestation.

MAIN OUTCOMES AND MEASURES The primary outcome was the geometric mean concentration (GMC) of anti-PT IgG antibodies in early- to late-term infants (≥ 37 0/7 weeks' gestation) at age 2 months, comparing maternal Tdap vaccination between 20 0/7 and 24 0/7 weeks' vs 30 0/7 and 33 0/7 weeks' gestation (reference cohort). Anti-PT GMC in 2-month-old infants born preterm (<35 0/7 weeks' gestation) compared with early- to late-term infants after maternal Tdap vaccination between 20 and 24 weeks' gestation was a secondary outcome.

RESULTS In total, 221 women who delivered 239 offspring were enrolled in the study: 66 early- to late-term infants (median gestational age [GA], 40.6 weeks [IQR, 39.8-41.0 weeks]; 38 [57.6%] male) and 73 preterm infants (median GA, 32.1 weeks [IQR, 29.5-33.0 weeks]; 42 [54.5%] female) had blood samples collected at 2 months of age. Anti-PT GMC was 14.7 IU/mL (95% CI, 10.6-20.4 IU/mL) in early- to late-term infants following maternal Tdap vaccination between 20 0/7 and 24 0/7 weeks' gestation compared with 27.3 IU/mL (95% CI, 20.1-37.1 IU/mL) in 55 infants in the reference group (median GA, 40.3 [IQR, 39.1-41.0]; 33 [60.0%] female). The mean anti-PT GMC in preterm infants in
Abstract (continued)

the study group was 11.2 IU/mL (95% CI, 8.1-15.3 IU/mL) (P = .23 compared with early-to-
late-term infants).

CONCLUSIONS AND RELEVANCE In this cohort study, 2-month-old preterm and early-to-late-term infants showed significantly lower anti-PT antibody levels following maternal Tdap vaccination between 20 O/7 and 24 O/7 weeks’ gestation compared with 30 O/7 and 33 O/7 weeks’ gestation; preterm and early-to-late-term infants had similar anti-PT antibody levels, but both groups showed significantly lower antibody levels compared with the reference group. Epidemiological research should investigate whether maternal Tdap vaccination before 24 weeks’ gestation provides sufficient protection against clinical pertussis, particularly in preterm infants, as long as no correlate of protection is available.


Introduction

According to the World Health Organization (WHO), 81% of infants worldwide (105 million) received 3 doses of a diphtheria, tetanus, and pertussis vaccine in 2021, protecting them against vaccine-preventable diseases that may cause serious, even fatal, illness and disability. Despite high vaccine coverage, pertussis remains endemic in many countries. Newborns and infants too young to be fully vaccinated are at the highest risk of severe complications. To protect newborns and infants in the first months of life, maternal vaccination with a tetanus, diphtheria, and acellular pertussis (Tdap) vaccine from 20 weeks’ gestation onward has been offered to all pregnant women in the Netherlands since December 2019. Infant diphtheria, tetanus, and pertussis (DTaP), inactivated poliovirus (IPV), Haemophilus influenzae type b, and hepatitis B vaccinations are given at 3, 5, and 11 months of age (2 + 1 dose schedule) for protection against pertussis provided that the mother received Tdap vaccination during pregnancy. An extra vaccination at 2 months of age (3 + 1 dose schedule) after maternal Tdap vaccination is advised if an infant is born before 37 weeks’ gestation or if the time interval between maternal vaccination and delivery is shorter than 2 weeks, since transfer of immunity against pertussis on maternal Tdap vaccination may be insufficient.

During pregnancy, maternal immunoglobulin G (IgG) antibodies are actively transferred across the placenta, mediated by the neonatal Fc receptor expressed on syncytiotrophoblast cells. This saturable process initiates at approximately 13 to 17 weeks’ gestation and increases throughout gestation. Around 33 to 36 weeks’ gestation, fetal IgG antibody levels exceed maternal IgG serum levels and increase to 150% of maternal levels near the due delivery date. Tdap vaccination in the third trimester enhances maternal antipertussis IgG antibody levels in newborns. Maternal Tdap vaccination was reported to prevent 70% to 90% of clinically confirmed pertussis cases and about 90.5% of pertussis hospitalizations in newborns and infants younger than 3 months of age in the UK from 2013 to 2018.8,9

There is no consensus on the optimal timing of maternal Tdap vaccination to achieve the highest antibody transfer. Most studies suggest that Tdap vaccination early in the third trimester results in the highest anti-pertussis toxin (anti-PT) IgG antibody levels at birth, while a Swiss study favored second-trimester vaccination, potentially due to a longer time interval between Tdap vaccination and delivery.10,11 Recently, it was estimated that a period of 7.5 weeks or more before delivery optimizes antibody transfer.6 Tdap vaccination before 24 weeks’ gestation may therefore be particularly relevant for preterm offspring, the group most vulnerable for severe pertussis. Preterm infants have a hospitalization rate for pertussis that is 1.5-times higher than predicted based on the total proportion of infants in the national UK birth cohort.12 Offering maternal Tdap vaccination from 20 weeks’ gestation also widens the opportunity for pregnant women to receive the vaccine, but few
studies have reported antibody levels after maternal Tdap vaccination at or before 24 weeks’ gestation or in preterm infants. These studies had insufficient power to draw firm conclusions.

In this study, pertussis-specific IgG antibody levels after maternal Tdap vaccination between 20 0/7 and 24 0/7 weeks’ gestation were evaluated in early- to late-term (hereafter, term) and preterm offspring with follow-up until 2 months of age. We primarily assessed whether maternal Tdap vaccination between 20 0/7 and 24 0/7 weeks’ gestation would be associated with similar anti-PT antibody levels in term infants at 2 months of age compared with maternal Tdap vaccination between 30 0/7 and 33 0/7 weeks’ gestation. Therefore, data were compared with those from a reference study (recruitment between January 2014 and February 2016) including 55 term infants following maternal Tdap vaccination between 30 0/7 and 33 0/7 weeks’ gestation. Additionally, we compared antibody levels in term and preterm infants following maternal Tdap vaccination between 20 0/7 and 24 0/7 weeks’ gestation.

Methods

Study Participants
In this prospective, multicenter cohort study, antenatal care practitioners working in birthing centers or hospitals recruited pregnant women aged 18 years or older between August 2019 and November 2021. The study design and procedures were previously described. In brief, women were included through 2 recruitment routes; from August 2019, healthy pregnant women were invited to participate and received Tdap vaccination between 20 0/7 and 24 0/7 weeks’ gestation as part of the study. In addition, after 2019, once the Dutch National Immunisation Programme (NIP) offered Tdap vaccination to all pregnant women from 20 weeks’ gestation onward, women with imminent preterm labor were recruited on presentation at the hospital provided that they received Tdap vaccination between 20 0/7 and 24 0/7 weeks’ gestation. These women were vaccinated through the NIP, unrelated to this study but with the same Tdap vaccine as used in the study. Women were excluded if they had received Tdap vaccination within the past 2 years or if there was a known or suspected underlying condition that could interfere with study results. Other exclusion criteria were previously described. Mother-infant pairs were followed up until 2 months after delivery. Data on Bordetella pertussis–specific IgG antibodies from mother-infant pairs in the study were compared with data from the reference study performed between January 2014 and February 2016 that comprised term infants at age 2 months after maternal Tdap vaccination between 30 0/7 and 33 0/7 weeks’ gestation. Both studies used identical vaccines and study procedures for collection and timing of collection of blood samples. Laboratory procedures were performed in the same laboratory using identical procedures. This study was conducted in accordance with the Declaration of Helsinki and approved by the Central Committee on Research Involving Human Subjects in the Netherlands. Oral and written informed consent was obtained from parents or legal guardians. The study followed the Strengthening the Reporting of Observational Studies in Epidemiology (STROBE) reporting guideline.

Maternal Vaccine
Pregnant women received a Tdap vaccine (Boostrix) containing adsorbed B pertussis antigens (ie, inactivated PT, filamentous hemagglutinin [FHA], pertactin [Prn], diphtheria toxoid [DT], and tetanus toxoid [TT]). The Tdap vaccine was administered as a single 0.5-mL intramuscular injection in the deltoid muscle.

Blood Sampling
Finger-stick blood samples (≤300 μL) were collected from mothers within 24 hours after delivery. Umbilical cord blood samples (≤2 mL) were collected at delivery, and heel-stick blood samples from infants (≤300 μL) were collected during home visits before primary vaccination at age 2 months (±5 days). For preterm infants, who often start receiving vaccinations between 6 and 9 weeks in the
Netherlands, blood samples were collected before the first vaccination. Serum samples were stored at −20 °C awaiting analyses.

**Laboratory Analyses**

Immunoglobulin G antibody concentrations against PT, FHA, Prn, DT, and TT were measured by bead-based fluorescent multiplex immunoassay using Luminex xMAP technology (ThermoFisher Scientific), as previously described. For the *B. pertussis* antigens, the assay was calibrated against the WHO international standard for pertussis antiserum (serum reference 06/140), interpolated using a 5-parameter fit, and expressed in international units (IU/mL).

**Statistical Analysis**

Anti-PT IgG antibody levels following maternal Tdap vaccination are associated with prevention of clinical pertussis. Our primary outcome was to assess noninferiority of anti-PT antibody levels in term infants at 2 months of age following maternal Tdap vaccination between 20 0/7 and 24 0/7 weeks' gestation compared with Tdap vaccination between 30 0/7 and 33 0/7 weeks' gestation (reference cohort). The lower limit of the 95% CI of the geometric mean concentration ratio (GMR) between the main and the reference cohorts was set at 0.5 or greater for noninferiority. Secondary outcomes were the geometric mean concentration (GMC) of PT IgG levels in preterm infants at 2 months' postnatal age after maternal Tdap vaccination between 20 0/7 and 24 0/7 weeks' gestation compared with the term cohort after maternal Tdap vaccination between 20 and 24 weeks' gestation and the IgG antibody levels against all Tdap vaccine antigens (ie, PT, FHA, Prn, DT, and TT) in blood samples from infants at 2 months of age, the umbilical cord, and mothers at delivery among term and preterm mother-infant pairs (eFigure in Supplement 1).

To assess our primary research question and allow 80% power and an α of 5%, 58 term and 54 preterm mother-infant pairs were required. We aimed for inclusion of 60 pairs in each group to allow loss to follow-up regarding the available blood samples.

For the scope of this study, we defined preterm as birth between 24 0/7 and 34 6/7 weeks' gestation since offspring antibody levels are expected to exceed maternal antibody levels at the end of this time window, and these late-preterm offspring may therefore resemble offspring born at full term regarding transplacental antibody transfer. Term birth was defined as 37 0/7 or more weeks' gestation.

Comparison of baseline characteristics was done using either *t*, Mann-Whitney *U*, or Fisher exact test. The IgG-antibody concentrations against all antigens were log-transformed and computed into GMCs with corresponding 95% CIs. In all groups, including the reference cohort, GMCs at different time points were assessed using generalized estimating equation models with a gaussian distribution with identity link function. An exchangeable correlation structure enabled adjustment for similarities in antibody levels among siblings who were twins or triplets. No additional adjustment was applied. The GMRs were calculated from the GMCs within different groups and expressed with 95% CIs. R, version 2023.03.1 (R Project for Statistical Computing) was used with the geepack package for analyses. Findings were based on available data, and missing data were handled by complete participant analyses. Two-sided *P* < .05 was considered significant.

**Results**

In total, 221 pregnant women who received second-trimester Tdap vaccination were included. They delivered 239 offspring, of whom 148 (61.9%) were term and 91 (38.1%) were preterm. The preterm offspring included 14 pairs of twins and 2 sets of triplets. All 148 term offspring (range, 37 0/7-42 0/7 weeks' gestation) were singletons; 66 of these (28 [42.4%] female; 38 [57.6%] male) had a blood sample collected at 2 months of age. The 91 preterm offspring (range, 25 2/7-34 6/7 weeks' gestation) were born to 73 mothers, and 73 of these offspring (42 [54.5%] female; 35 [45.5%] male) had a blood sample obtained at age 2 months (Figure 1). Detailed demographics of study and
reference mother-infant pairs are shown in Table 1. The median gestational age (GA) at birth was not significantly different between term infants in the study group (40.6 weeks [IQR, 39.8-41.0 weeks]) and the reference group of 55 infants (33 [60.0%] female; 22 [40.0%] male; median GA, 40.3 weeks [IQR, 39.1-41.0 weeks]). Median GA was significantly different for preterm offspring (32.1 weeks [IQR, 29.5-33.0 weeks]) compared with term offspring in both the study cohort and the reference cohort. The median GA at maternal Tdap vaccination in the study groups of term and preterm mother-infant pairs (22.0 weeks [IQR, 20.9-23.1 weeks] and 22.9 weeks [IQR, 22.0-23.4 weeks], respectively) was significantly different from the median GA in the term mother-infant pairs in the reference cohort (31.1 weeks [IQR, 30.5-31.7 weeks]). The median time interval between maternal Tdap vaccination and delivery was 18.3 weeks (IQR, 17.1-19.7 weeks) for term births and 9.4 weeks (IQR, 6.9-10.7 weeks) for preterm births in the study cohort and 9.0 weeks (IQR, 8.1-9.9 weeks) for term births in the reference cohort (Table 1).

At 2 months of age in term infants, the anti-PT GMC after maternal Tdap vaccination between 20 0/7 and 24 0/7 weeks’ gestation (14.7 IU/mL; 95% CI, 10.6-20.4 IU/mL) was significantly lower than the GMC in the reference cohort after Tdap vaccination between 30 0/7 and 33 0/7 weeks’ gestation (27.3 IU/mL; 95% CI, 20.1-37.1 IU/mL) (Table 2 and Figure 2). The GMR was 0.54 (95% CI, 0.34-0.85), with the 2.5% bound of the 95% CI at 0.34 (97.5% bound at 0.85) refuting noninferiority requirements.

At 2 months of age after maternal Tdap vaccination between 20 0/7 and 24 0/7 weeks’ gestation, no significant differences in anti-PT GMCs were observed in preterm infants compared with term infants (11.2 IU/mL [95% CI, 8.1-15.3 IU/mL] vs 14.7 IU/mL [95% CI, 10.6-20.4 IU/mL]) (P = .23) (Table 2 and Figure 3). In term infants at 2 months of age, besides anti-PT levels, the GMC of IgG against Prn was significantly lower after Tdap vaccination between 20 0/7 and 24 0/7 weeks’ gestation compared with 30 0/7 to 33 0/7 weeks’ gestation (59.8 IU/mL [95% CI, 38.4-93.0 IU/mL] vs 110.3 IU/mL [95% CI, 71.6-170.0 IU/mL]). No differences in GMCs were observed for FHA, DT, or TT for term infants in the study compared with the reference cohort (Table 2 and Figure 2). In preterm
infants compared with term infants at age 2 months after maternal Tdap vaccination between 20 0/7 and 24 0/7 weeks’ gestation, GMCs were significantly lower for FHA (48.8 IU/mL [95% CI, 37.3-63.8 IU/mL] vs 83.1 IU/mL [95% CI, 63.6-109.1 IU/mL]) and TT (1.2 IU/mL [95% CI, 1.0-1.5 IU/mL] vs 1.5 IU/mL [95% CI, 1.2-1.9 IU/mL]), whereas no significant differences were observed for Prn and DT (Table 2 and Figure 3).

In umbilical cord serum samples from term offspring, significantly lower GMCs were observed when mothers received Tdap vaccination between 20 0/7 and 24 0/7 weeks’ gestation compared with 30 0/7 and 33 0/7 weeks’ gestation for PT (58.6 IU/mL [95% CI, 46.4-74.2 IU/mL] vs 125.1 IU/mL [95% CI, 94.0-166.3 IU/mL]) and Prn (295.5 IU/mL [95% CI, 216.7-402.8 IU/mL] vs 500.5 IU/mL [95% CI, 322.5-776.7 IU/mL]). No differences were observed for FHA, DT, and TT IgG levels (Table 2 and Figure 3). In umbilical cord serum samples, GMCs from preterm offspring compared with term offspring following maternal Tdap vaccination between 20 0/7 and 24 0/7 weeks’ gestation were significantly lower for FHA (193.5 IU/mL [95% CI, 155.2-241.3 IU/mL] vs 295.2 IU/mL [95% CI, 249.1-349.9 IU/mL]) and Prn (143.7 IU/mL [95% CI, 97.3-212.4 IU/mL] vs 295.5 IU/mL [95% CI, 216.7-402.8 IU/mL]). No differences between term and preterm offspring were observed for PT, DT and TT (Table 2 and Figure 3).

Comparing antibody levels at delivery among mothers of term offspring in the study and reference groups, the anti-PT GMC at delivery was significantly higher in the study group (61.8 IU/mL [95% CI, 46.8-81.7 IU/mL] vs 32.9 IU/mL [95% CI, 26.0-41.6 IU/mL]). No differences for the other Tdap antigens were found (Table 2 and Figure 2). Comparing preterm and term mother-infant pairs following Tdap vaccination between 20 0/7 and 24 0/7 weeks’ gestation, mothers had significantly higher GMCs after preterm than term delivery for all antigens (eg, PT 60.4 IU/mL [95% CI, 44.1-82.7 IU/mL] vs 32.9 IU/mL [95% CI, 26.0-41.6 IU/mL]) except Prn (Table 2 and Figure 3).

Table 1. Baseline Characteristics at the Mother and Infant Level for Preterm and Early- and Full-Term Mother-Infant Pairs

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Participants, by GA at maternal Tdap vaccination</th>
<th>20 0/7-24 0/7 wk</th>
<th>30 0/7-33 0/7 wk</th>
</tr>
</thead>
<tbody>
<tr>
<td>Maternal age at delivery, mean (SD), y</td>
<td>31.4 (3.8)</td>
<td>31.7 (4.0)</td>
<td>32.6 (3.3)</td>
</tr>
<tr>
<td>GA at maternal immunization, median (IQR), wk</td>
<td>22.9 (22.0-23.4)</td>
<td>22.0 (20.9-23.1)</td>
<td>31.1 (30.5-31.7)</td>
</tr>
<tr>
<td>Pregnancy duration, wk</td>
<td>Median (IQR)</td>
<td>32.1 (29.5-33.0)</td>
<td>40.6 (39.8-41.0)</td>
</tr>
<tr>
<td>25 2/7-27 6/7</td>
<td>13 (17.8)</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>28 0/7-31 6/7</td>
<td>25 (34.2)</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>32 0/7-34 6/7</td>
<td>35 (47.9)</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>37 0/7-39 6/7</td>
<td>NA</td>
<td>21 (31.8)</td>
<td>23 (41.8)</td>
</tr>
<tr>
<td>40 0/7-42 0/7</td>
<td>NA</td>
<td>45 (68.2)</td>
<td>32 (58.2)</td>
</tr>
<tr>
<td>Interval between maternal immunization and delivery, median (IQR), wk</td>
<td>9.4 (6.9-10.7)</td>
<td>18.3 (17.1-19.7)</td>
<td>9.0 (8.1-9.9)</td>
</tr>
<tr>
<td>Multiple pregnancy</td>
<td>No</td>
<td>45 (78.1)</td>
<td>66 (100)</td>
</tr>
<tr>
<td>Twins</td>
<td>13 (19.2)</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Triplets</td>
<td>2 (2.7)</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Infant sex</td>
<td>Female</td>
<td>42 (54.5)</td>
<td>28 (42.4)</td>
</tr>
<tr>
<td>Male</td>
<td>35 (45.5)</td>
<td>38 (57.6)</td>
<td>22 (40.0)</td>
</tr>
<tr>
<td>Birth weight, mean (SD), g</td>
<td>1631 (499)</td>
<td>3622 (430)</td>
<td>3446 (481)</td>
</tr>
<tr>
<td>Birth weight percentile corrected for GA, mean (SD)</td>
<td>38.8 (31.5)</td>
<td>53.1 (27.8)</td>
<td>42.4 (28.0)</td>
</tr>
<tr>
<td>Age at blood sample obtainment, mean (SD), d</td>
<td>55.2 (6.2)</td>
<td>61.0 (3.0)</td>
<td>61.4 (2.1)</td>
</tr>
</tbody>
</table>

Abbreviations: GA, gestational age; NA, not applicable; Tdap, tetanus, diphtheria, and acellular pertussis.

* Term birth was defined as GA of 37 0/7 weeks or more and preterm birth as less than 35 0/7 weeks’ gestation.

* Data are presented as number (percentage) of participants unless otherwise indicated.

* In the study cohort, 73 preterm infants born to 60 mothers (due to multiple pregnancies) had a blood sample obtained at 2 months of age, as did 66 term infants.

* In the reference cohort, infants had a blood sample obtained at 2 months of age.

* Seven pairs of dichorionic-diamniotic twins, 4 pairs of monochorionic-diamniotic twins, 2 pairs of monochorionic-monoamniotic twins, and 2 sets of trichorionic-triamniotic triplets. Numbers sum to 60 infants (77 including siblings) for the total number of mother-infant pairs, but only 73 of 77 infants (94.8%) had a blood sample obtained at 2 months of age.

* Birth weight and birth weight percentiles were presented for the firstborn infant only if there were multiple pregnancies.

* Blood samples at age 2 months were obtained as close as possible to infant immunization but may have been obtained earlier than at 2 months of age because, in the Netherlands, routine preterm primary vaccinations are administered between 6 and 9 weeks after birth.
analyses using a Tdap vaccination cutoff of February 27, 2020, the first day of COVID-19 social distancing measures in the Netherlands, were conducted and found no differences in antibody levels after birth among mothers or infants before vs during COVID-19 measures.

**Discussion**

In this prospective cohort study, anti-PT IgG levels in term infants at 2 months of age following maternal Tdap vaccination between 20/7 and 24/7 weeks' gestation were inferior to those in the group with Tdap vaccination between 30/7 and 33/7 weeks' gestation, with an approximate 2-fold reduction in GMCS of anti-PT IgG levels. As long as the mechanisms of protection following maternal Tdap vaccination are not fully understood and no correlate of protection is available, anti-PT IgG levels are often used in studies like ours as surrogate markers for protection. Anti-PT levels in umbilical cord blood are correlated with protection against pertussis, and lower anti-PT IgG levels may point to less protection against pertussis in newborns. We also observed a reduction in anti-Prn antibody levels after maternal Tdap vaccination between 20 and 24 weeks' gestation compared with the reference group.

The GA at which to administer maternal Tdap vaccination for the highest antibody transfer may vary per vaccine antigen. Many studies have suggested that Tdap vaccination between 27/7 and 30/7 weeks' gestation results in maximal pertussis-specific antibody levels and avidity in term.

<table>
<thead>
<tr>
<th>Antibody</th>
<th>GMC (95% CI), IU/mL, by GA at maternal Tdap vaccination</th>
<th>GMR (95% CI) at term, study vs reference</th>
<th>P value</th>
<th>GMR (95% CI), preterm vs term</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>20/7-24/7 wk (study cohort)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mothers at delivery</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Samples, No.</td>
<td>73</td>
<td>138</td>
<td>55</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Anti–pertussis toxin</td>
<td>60.4 (44.1-82.7)</td>
<td>32.9 (26.0-41.6)</td>
<td>61.8 (46.8-81.7)</td>
<td>0.53 (0.35-0.80)</td>
<td>&lt;.001</td>
</tr>
<tr>
<td>Anti–filamentous hemagglutinin</td>
<td>220.5 (171.5-283.5)</td>
<td>161.1 (135.5-191.5)</td>
<td>163.4 (132.5-204.6)</td>
<td>0.99 (0.73-1.34)</td>
<td>.92</td>
</tr>
<tr>
<td>Antipertactin</td>
<td>203.1 (134.6-306.4)</td>
<td>176.5 (129.9-239.8)</td>
<td>286.0 (182.4-448.3)</td>
<td>0.62 (0.35-0.98)</td>
<td>.08</td>
</tr>
<tr>
<td>Anti–diphtheria toxoid</td>
<td>0.6 (0.2-0.8)</td>
<td>0.3 (0.2-0.4)</td>
<td>0.4 (0.3-0.5)</td>
<td>0.85 (0.58-1.24)</td>
<td>.35</td>
</tr>
<tr>
<td>Anti–tetanus toxoid</td>
<td>5.6 (4.6-6.8)</td>
<td>3.3 (2.9-3.8)</td>
<td>3.5 (3.0-4.2)</td>
<td>0.94 (0.74-1.21)</td>
<td>.59</td>
</tr>
<tr>
<td>Neonatal umbilical cord blood</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Samples, No.</td>
<td>86</td>
<td>146</td>
<td>54</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Anti–pertussis toxin</td>
<td>52.8 (40.7-68.6)</td>
<td>58.6 (46.4-74.2)</td>
<td>125.1 (94.0-166.3)</td>
<td>0.47 (0.31-0.72)</td>
<td>&lt;.001</td>
</tr>
<tr>
<td>Anti–filamentous hemagglutinin</td>
<td>193.5 (153.2-241.3)</td>
<td>294.2 (249.1-349.9)</td>
<td>300.9 (261.2-419.3)</td>
<td>0.80 (0.66-1.22)</td>
<td>.43</td>
</tr>
<tr>
<td>Antipertactin</td>
<td>141.7 (97.3-212.4)</td>
<td>295.5 (216.7-402.8)</td>
<td>500.5 (322.5-776.7)</td>
<td>0.59 (0.33-1.05)</td>
<td>.049</td>
</tr>
<tr>
<td>Anti–diphtheria toxoid</td>
<td>0.2 (0.4-0.9)</td>
<td>0.5 (0.4-0.6)</td>
<td>0.6 (0.5-0.9)</td>
<td>0.74 (0.50-1.09)</td>
<td>.09</td>
</tr>
<tr>
<td>Anti–tetanus toxoid</td>
<td>5.2 (4.2-6.4)</td>
<td>6.0 (5.2-6.9)</td>
<td>7.4 (6.2-8.8)</td>
<td>0.81 (0.62-1.04)</td>
<td>.06</td>
</tr>
</tbody>
</table>

**Infants at 2 mo of age**

<table>
<thead>
<tr>
<th>Antibody</th>
<th>GMC (95% CI), IU/mL, by GA at maternal Tdap vaccination</th>
<th>GMR (95% CI) at term, study vs reference</th>
<th>P value</th>
<th>GMR (95% CI), preterm vs term</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>20/7-24/7 wk (study cohort)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mothers at delivery</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Samples, No.</td>
<td>73</td>
<td>66</td>
<td>55</td>
<td>NA</td>
<td>NA</td>
</tr>
<tr>
<td>Anti–pertussis toxin</td>
<td>11.2 (8.1-15.3)</td>
<td>14.7 (10.6-20.4)</td>
<td>27.3 (20.1-37.1)</td>
<td>0.54 (0.34-0.85)</td>
<td>.005</td>
</tr>
<tr>
<td>Anti–filamentous hemagglutinin</td>
<td>48.8 (37.3-63.8)</td>
<td>83.1 (63.6-109.1)</td>
<td>83.7 (67.4-103.9)</td>
<td>0.99 (0.70-1.42)</td>
<td>.97</td>
</tr>
<tr>
<td>Antipertactin</td>
<td>38.9 (24.7-61.5)</td>
<td>59.8 (38.4-93.0)</td>
<td>110.3 (71.6-170.0)</td>
<td>0.54 (0.29-1.01)</td>
<td>.045</td>
</tr>
<tr>
<td>Anti–diphtheria toxoid</td>
<td>0.1 (0.1-0.1)</td>
<td>0.1 (0.1-0.2)</td>
<td>0.1 (0.1-0.2)</td>
<td>0.89 (0.59-1.35)</td>
<td>.57</td>
</tr>
<tr>
<td>Anti–tetanus toxoid</td>
<td>1.2 (1.0-1.5)</td>
<td>1.5 (1.2-1.9)</td>
<td>1.7 (1.4-2.0)</td>
<td>0.92 (0.70-1.21)</td>
<td>.52</td>
</tr>
</tbody>
</table>

Abbreviations: GMC, geometric mean concentration; GMR, geometric mean concentration ratio; NA, not applicable.

* Term birth was defined as a gestational age of 37.0/7 weeks or more and preterm birth as less than 35.0/7 weeks' gestation.
offspring, though these studies provided no data on Tdap vaccination before 24 weeks’ gestation.\textsuperscript{4,7,21-26} In contrast, an observational Swiss study suggested that Tdap vaccination earlier in pregnancy led to higher maternal antibody transfer, potentially because of the longer transfer time before delivery.\textsuperscript{10} The recent Optimising the Timing of Whooping Cough Immunisation in Mums (OptIMUM) randomized clinical trial observed the highest pertussis-specific IgG antibody levels in umbilical cord serum when mothers received the Tdap vaccine early in the third trimester (28-32

Figure 2. Individual Immunoglobulin G (IgG) Antibody Concentrations and Geometric Mean Concentrations (GMCs) After Second- vs Third-Trimester Tetanus, Diphtheria, and Pertussis Vaccination in Early- and Full-Term Mother-Infant Pairs at Different Time Points

A Anti-PT

B Anti-FHA

C Anti-Ptn

D Anti-DT

E Anti-TT

Horizontal lines represent GMCs and vertical bars, 95% CIs. DT indicates diphtheria toxoid; FHA, filamentous hemagglutinin; GA, gestation; Ptn, pertactin; PT, pertussis toxin; TT, tetanus toxoid.

a P < .05.

b P < .01.

c P < .001.
weeks' gestation) compared with earlier than 24 weeks' and 24 to 27 weeks' gestation. Notably, the number of preterm offspring included in that study was too small to draw conclusions for this most vulnerable group (15 [4%]; 5 per study group).6

The reduction in anti-B pertussis antibodies in offspring following maternal Tdap vaccination before 24 weeks' gestation may possibly be explained by the fact that peak levels of anti-B pertussis antibodies following vaccination are achieved when materno-fetal antibody transfer is still

Figure 3. Individual Immunoglobulin G (IgG) Antibody Concentrations and Geometric Mean Concentrations (GMCs) After Second-Trimester Tetanus, Diphtheria, and Pertussis Vaccination in Early- and Full-Term vs Preterm Mother-Infant Pairs at Different Time Points

All women were vaccinated between 20 0/7 and 24 0/7 weeks' gestation (GA). Horizontal lines represent GMCs and vertical bars, 95% CIs. DT indicates diphtheria toxoid; FHA, filamentous hemagglutinin; Prn, pertactin; PT, pertussis toxin; TT, tetanus toxoid.

\[\text{\textit{A} Anti-PT} \]

\[\text{\textit{B} Anti-FHA} \]

\[\text{\textit{C} Anti-Prn} \]

\[\text{\textit{D} Anti-DT} \]

\[\text{\textit{E} Anti-TT} \]

\[\text{\textit{a} P < .05.} \]

\[\text{\textit{b} P < .01.} \]

\[\text{\textit{c} P < .001.} \]
suboptimal. This appears not to be compensated by the increased time of around 9 weeks for transport until delivery.

Based on epidemiological studies, a minimum of 2 to 4 weeks between maternal Tdap vaccination and delivery seems required for protection of term offspring against clinical pertussis. Up to 7 to 8 weeks was estimated to result in optimal antibody transfer. Nevertheless, even in the case of a similar 9-week time interval between maternal Tdap vaccination and delivery in this study, GMCs in preterm offspring following maternal Tdap vaccination between 20 0/7 and 24 0/7 weeks' gestation were at least 2-fold lower than those in term offspring from the reference group following maternal Tdap vaccination between 30 0/7 and 33 0/7 weeks' gestation. Also, while a longer interval in case of maternal Tdap vaccination between 20 and 24 weeks' gestation was associated with almost similar anti-PT antibody levels in term and preterm offspring after a time interval of 9 weeks and 18 weeks, respectively, a significant reduction in anti-PT levels but also anti-FHA and anti-Prn levels compared with Tdap vaccination between 30 0/7 and 33 0/7 weeks' gestation was observed. The OpTIMUM trial also showed significantly lower anti-PT antibodies after maternal Tdap vaccination at 24 0/7 weeks' gestation or earlier.

In a study from Belgium, maternal Tdap vaccination at around 27 weeks' gestation resulted in improved maternal-derived, pertussis-specific antibody levels in preterm offspring, potentially due to receiving the vaccine at a time in pregnancy with a long enough interval before delivery together with a postvaccination antibody peak during the third trimester that has improved antibody transfer compared with the second trimester. This is relevant for most preterm offspring because currently, 84% of all preterm newborns in the Netherlands are delivered at or after 32 weeks' gestation.

In the UK, timing of Tdap vaccination changed in 2016 from 28 weeks' to 16 weeks' gestation onward. A subsequent analysis of data on pertussis cases in the hospital showed that the effectiveness of maternal Tdap vaccination against pertussis-related hospitalization in infants had remained high. It must be noted that the overall maternal Tdap vaccination coverage over the period of the study also increased, which might have contributed to robustly high maternal Tdap vaccine effectiveness rates. With an undefined correlate of protection, studies like ours cannot draw conclusions on clinical effectiveness of maternal Tdap vaccination at or before 24 weeks' gestation. Epidemiological studies on effectiveness with data stratified for GA at birth are required.

While protection against severe pertussis following maternal Tdap vaccination has been confirmed by many observational studies, important knowledge gaps in the protective mechanisms remain. In addition to quantitative antibody levels, quality and functionality of anti-B pertussis antibodies may contribute to protection against clinical pertussis, as may maternal immune cells, such as T cells, are transferred to the offspring during pregnancy and may vary with timing of maternal vaccination. To our knowledge, the present study is the first to investigate transplacental antibody transfer following maternal Tdap vaccination before 24 0/7 weeks' gestation in a large group of preterm- and early- to late-term infants up to the age of their first vaccinations and to compare antibody transfer with a well-defined cohort of 55 mother-infant pairs after Tdap vaccination between 30 0/7 and 33 0/7 weeks' gestation.

Limitations
This study has limitations. Most importantly, the term offspring from the reference cohort were recruited within different periods. The present study was performed partially during nonpharmaceutical COVID-19 interventions, with reduced B pertussis circulation compared with the 2014 to 2016 inclusion period of the reference cohort. Lower endemic B pertussis transmission might result in reduced preexisting antibody levels due to a lack of boosting in women of childbearing age and may have impacted the antibody response to maternal Tdap vaccination. Sensitivity analyses with a Tdap vaccination cutoff at February 27, 2020, the first day of COVID-19 social distancing measures in the Netherlands, yielded no differences in antibody levels in mothers or infants after birth before and during COVID-19-measures in the present study. The vaccination history of mothers in the study and reference cohorts were similar. Starting in 1957 (ie, the start of the
Dutch NIP), a whole-cell pertussis vaccine was used for infant vaccinations. In 1996, an acellular pertussis component was added to the DT-IPV booster dose administered at age 4 years. All participating mothers were born before 2005, the year when the infant acellular pertussis–boosted vaccine replaced infant whole-cell pertussis vaccines. In the near future, more pregnant women will receive the acellular pertussis primary or booster vaccine. This may impact antipertussis immune status and response to maternal Tdap vaccination.

We did not study the antibody response to Tdap vaccination in mothers vaccinated earlier vs later during pregnancy. Potential differences may exist. We found that maternal antibodies at delivery following Tdap vaccination before 24 weeks' gestation were significantly higher with a shorter interval between Tdap vaccination and delivery, suggesting a rapid decline after peak levels following vaccination. Another limitation is the rate of loss to follow-up for samples. We included more term and preterm infants compared with other studies, but many appointments for blood sample obtainment at 2 months of age were cancelled due to COVID-19–related safety measures, resulting in large dropout rates among 2-month-old infants. However, samples from 66 term and 73 preterm infants yielded enough power to assess noninferiority. The dropout group at age 2 months was not selective and not expected to affect results. Finally, we performed a sensitivity analysis to compare mothers with imminent preterm labor recruited on presentation at the hospital with other mothers of preterm offspring vaccinated as part of the study (n = 46 vs 14). We found no statistically significant differences, but differences in clinical baseline factors that may correlate with preterm labor may have occurred. However, we had no clinical data to compare these 2 groups in detail.

Conclusions

In this cohort study, maternal Tdap vaccination between 20 O/7 and 24 O/7 weeks’ gestation compared with 30 O/7 and 33 O/7 weeks’ gestation was associated with significantly lower anti-PT antibody levels in 2-month-old term and preterm infants despite a similar interval between maternal vaccination and delivery in preterm infants after Tdap vaccination between 20 O/7 and 24 O/7 weeks’ gestation and term infants after 30 O/7 and 33 O/7 weeks’ gestation. Further epidemiological research should determine whether maternal Tdap vaccination before 24 weeks’ gestation provides sufficient protection against clinical pertussis both in term and preterm infants as long as no correlate of protection is available.
Concept and design: Immink, Bekker, de Melker, Rots, Sanders, van der Maas.

Acquisition, analysis, or interpretation of data: All authors.

Drafting of the manuscript: Immink, Sanders, van der Maas.

Critical review of the manuscript for important intellectual content: Bekker, de Melker, den Hartog, Rots, van Gageldonk, Groenendaal, Sanders, van der Maas.

Statistical analysis: Immink, Groenendaal, van der Maas.

Obtained funding: van der Maas.

Administrative, technical, or material support: Immink, den Hartog, van Gageldonk, van der Maas.

Supervision: Bekker, Sanders, van der Maas.

Conflict of Interest Disclosures: None reported.

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Role of the Funder/Sponsor: ZonMw had no role in the design and conduct of the study; collection, management, analysis, and interpretation of the data; preparation, review, or approval of the manuscript; and decision to submit the manuscript for publication.

Group Information: Members of the Dutch Maternal Pertussis Vaccine Investigation Group are listed in Supplement 2.

Data Sharing Statement: See Supplement 3.

Additional Contributions: We thank the women who participated in this study and all instances that assisted with participant recruitment and performed study procedures. Jan van de Kastelee, PhD, and Mirjam Knol, PhD, from the National Institute for Public Health and the Environment in the Netherlands provided statistical advice on the data analyses without compensation.

REFERENCES


SUPPLEMENT 1.
eFigure. Comparisons for the Primary and Secondary Aim

SUPPLEMENT 2.
Dutch Maternal Pertussis Vaccine Investigation Group

SUPPLEMENT 3.
Data Sharing Statement