Impact of Minority Nonnucleoside Reverse Transcriptase Inhibitor Resistance Mutations on Resistance Genotype After Virologic Failure

Jonathan Z. Li,1 Roger Ribaudo,3 Michael J. Kozal,6 Evgenia S. Svarovskaia,5 Jeffrey A. Johnson,4 Anna Maria Geretti,7 Michael D. Miller,5 and Daniel R. Kuritzkes1

1Division of Infectious Diseases, Brigham and Women’s Hospital, Harvard Medical School, Boston, Massachusetts; 2Irsi Caixa AIDS Research Institute and Lluita Contra la SIDA Foundation, Badalona, Spain; 3Harvard School of Public Health, Boston, Massachusetts; 4Yale University School of Medicine and VA Connecticut Healthcare System, New Haven, Connecticut; 5Gilead Sciences, Inc, Foster City, California; 6Centers for Disease Control and Prevention, Atlanta, Georgia; 7Institute of Infection and Global Health, University of Liverpool, United Kingdom; 8Division of Infectious Diseases and Hospital Epidemiology, University Hospital Zurich, University of Zurich, Switzerland; 9Department of Biomedicine, Aarhus University, Denmark; 10University of Minnesota, Minneapolis; and 11Department of Infectious Diseases, Aarhus University, Skejby, Denmark

Drug-resistant human immunodeficiency virus type 1 (HIV-1) minority variants increase the risk of virologic failure for first-line nonnucleoside reverse transcriptase inhibitor (NNRTI)-based regimens. We performed a pooled analysis to evaluate the relationship between NNRTI-resistant minority variants and the likelihood and types of resistance mutations detected at virologic failure. In multivariable logistic regression analysis, higher NNRTI minority variant copy numbers, non-white race, and nevirapine use were associated with a higher risk of NNRTI resistance at virologic failure. Among participants on efavirenz, K103N was the most frequently observed resistance mutation at virologic failure regardless of the baseline minority variant. However, the presence of baseline Y181C minority variant was associated with a higher probability of Y181C detection after virologic failure. NNRTI regimen choice and preexisting NNRTI-resistant minority variants were both associated with the probability and type of resistance mutations detected after virologic failure.

Keywords. HIV-1 drug resistance; minority variants; virologic failure; resistance genotyping.

Human immunodeficiency virus (HIV) drug resistance mutations present below 10%–20% of the viral population are not reliably detected by genotyping techniques that use population (Sanger) sequencing [1, 2]. In a recent pooled analysis, we demonstrated that the presence of baseline drug-resistant minority variants more than doubles the risk of virologic failure in patients initiating a first-line nonnucleoside reverse transcriptase inhibitor (NNRTI)-based regimen [3]. However, it is unknown how these baseline minority variants relate to the risk and type of resistance mutations detected at virologic failure.

The mechanism by which these detectable drug-resistant minority variants mediate treatment failure would at first glance be straightforward and involve the expansion of virus harboring that resistance mutation. Such a pattern has been detected in studies of resistance developing against a CCR5-antagonist [4] and raltegravir [5]. In addition, there can be a relatively high concordance of preexisting minority variants with mutations detected after virologic failure of raltegravir-based regimens [6] and NNRTI-based regimens in treatment-experienced patients [7]. However, these studies were of limited sample size and data from treatment-naive individuals do not always support such a straightforward interpretation [8]. We used data collected during a previously described pooled analysis of drug-resistant minority variants to evaluate the relationship between baseline NNRTI-resistant minority variants, antiretroviral therapy (ART) regimen, and other factors on the likelihood and types of
resistance mutations detected by population sequencing after virologic failure.

METHODS

This analysis derives from a sub-study of a previously reported pooled analysis of HIV type 1 (HIV-1) drug-resistant minority variants on the risk of virologic failure for treatment-naive individuals initiating an NNRTI-based regimen [3]. Patient-level data were obtained from all studies to exclude participants with any evidence of pre-ART drug resistance and to standardize the definition of virologic failure. A total of 240 participants are included from seven studies with genotypic resistance results after virologic failure [8–14].

All of the studies evaluated baseline K103N minority variants, and 6 of the 7 studies evaluated baseline Y181C minority variants as well [8–13]. A detailed description of the assays, limits of detection, and minority variants detected can be found in the original report [3]. Mutations conferring NRTI and NNRTI resistance were defined as those with a Stanford HIV resistance DB score ≥10 for the participant’s ART regimen. Medication adherence was available from 3 studies and was determined either by self-reported medication adherence over the previous 4 or 7 days [9], or by a clinic-and was determined either by self-reported medication adherence over the previous 4 or 7 days [9], or by a clinic-based pill count [14].

Fisher exact tests, Wilcoxon rank sum tests, and Cochran-Mantel-Haenszel statistics (stratified by study) were used to compare factors associated with either the presence of genotyping results or resistance at virologic failure. Minority variant copy number was estimated as the product of percentage of minority variant and level of HIV-1 RNA. Individuals without detectable minority variants were assigned a minority variant copy number of 0%; in a sensitivity analysis, a minority variant copy number equivalent to 10% of the assay limit of detection was imputed. A multivariable logistic regression, stratified by study, with backward elimination was performed to evaluate factors that predicted the risk of NNRTI resistance at virologic failure. Variables included in the regression model were determined a priori with the exception of ethnicity, which was found to be a significantly associated with the risk of NNRTI resistance at virologic failure on univariate analysis. Statistical analysis and the creation of figures were performed using SAS 9.2 and GraphPad Prism 5.

RESULTS

Genotypic resistance data after virologic failure were available from 240 of the 319 (75%) participants in the original pooled analysis with virologic failure [3]. The median time from virologic failure to resistance genotyping was 22 days (interquartile range [IQR], 0–49 days). More than half of the participants had NNRTI resistance detected at the time of virologic failure (Supplementary Table 1). Those with detectable NNRTI resistance at the time of virologic failure had lower baseline CD4+ cell counts (205/mm³ vs 261/mm³, \( P = .008 \)), and were more commonly receiving a nevirapine-based regimen \( (P = .03) \). The NNRTI component of the ART regimen was variable with 9 different combinations represented. The most common NNRTI backbone used for individuals on either efavirenz or nevirapine was zidovudine/lamivudine (AZT/3TC). Of those on AZT/3TC and efavirenz, 50% (49/99) had detectable NNRTI resistance at virologic failure compared with 73% (8/11) of those on AZT/3TC and nevirapine \( (P = .21) \). There were significant differences in the distribution of races/ethnicities \( (P = .003) \) with whites comprising a smaller proportion of those with NNRTI resistance at virologic failure than blacks. Overall mean ART adherence rates were similar between those with and without NNRTI resistance at virologic failure. No significant differences in baseline characteristics between individuals with or without resistance testing at virologic failure were detected with one exception (Supplementary Table 2). Those without genotyping data had higher rates of ART adherence \( (P < .01) \).

A significantly higher proportion of those with \( \geq 1\% \) NNRTI minority variants had detectable NNRTI resistance at virologic failure compared to either individuals harboring \(<1\% \) NNRTI minority variants or no detectable minority variants (92% with \( \geq 1\% \) minority variants vs 49% with \(<1\%, \ P = .002 \) and 92% \( \geq 1\% \) minority variants vs 58% without, \( P = .01 \)). A similar outcome was seen when participants were stratified based on harboring \( \geq 0.5\% \) vs \(<0.5\% \) minority variants. Among those with detectable minority variants at baseline, increasing copy numbers of NNRTI resistance mutations was associated with a higher probability of resistance at virologic failure (Figure 1A). Interestingly, individuals with no detectable minority variants had an intermediate outcome. This result is likely due to the varying limits of detection for the assays included in this pooled analysis [3]. Thus, individuals without detectable minority variants based on a less sensitive assay may, in fact, harbor low-frequency mutations that might have been detectable by a more sensitive test. We therefore performed a sensitivity analysis using both measured and imputed minority variant copy number. Individuals without detectable minority variants were assigned an imputed minority variant copy number equivalent to 10% of the assay limit of detection. Results of this analysis closely mirrored those of the measured values alone (Figure 1B).

In multivariable logistic regression analysis, factors that were independently associated with higher odds of NNRTI resistance at virologic failure included having a higher baseline NNRTI minority variant copy number, nevirapine use, and nonwhite ethnicity (Supplementary Table 3). Baseline viral load, CD4+ count, and ART adherence were not found to
be significant predictors of NNRTI resistance at virologic failure.

We evaluated the relationship between the NNRTI-resistant minority variants detected at baseline and the resistance mutations that emerged at virologic failure. Participants were categorized into those receiving efavirenz and those receiving a nevirapine-based regimen. Individuals receiving an efavirenz-based regimen were found to have K103N as the most common NNRTI resistance mutation detected at virologic failure regardless of the baseline resistance pattern (Figure 2A). However, the presence of baseline Y181C was associated with a higher rate of Y181C detection at virologic failure (18% vs 3%, P = .01). Y181C was the most commonly detected NNRTI resistance at virologic failure for those receiving a nevirapine-based ART regimen, although there were relatively few participants receiving nevirapine (Figure 2B). In those individuals with no baseline NNRTI resistance mutation but resistance on virologic failure, Y181C was detected in 75% (9 of 12) of participants receiving nevirapine as compared to 4% (3 of 79) of those receiving efavirenz (P < .001).

In the original pooled analysis, 228 participants had pre-ART assessment of minority M184V mutations, and 10 were found to have an M184V minority variant. Of these 10, virologic failure occurred in 4 participants, and M184V was found in the virologic failure genotype of 2. By contrast, M184V was detected by viral genotyping at virologic failure in 21 of 80 participants without preexisting M18V minority variant (P = .30). The 2 participants with preexisting K65R minority variants did not have virologic failure.
DISCUSSION

In this study, we found that among treatment-naïve patients initiating an NNRTI-based regimen, the presence of NNRTI-resistant minority variants, nonwhite ethnicity, and nevirapine use were all associated with an increased risk of NNRTI resistance detected at virologic failure. Interestingly, the type of NNRTI resistance that emerged at virologic failure frequently differed from those detected as minority variants at baseline. This finding was unexpected as the dose-dependent relationship of baseline minority variants with both risk of virologic failure and detectable resistance at virologic failure initially suggested a straightforward explanation for their effects. There are several potential explanations for this discrepancy. The presence of one minority variant could predispose to the development of additional resistance mutations. Despite the relatively short time from the date of virologic failure to resistance genotyping, there may have been sufficient time since the end of virologic suppression for selection of more fit resistance variants [13]. It is possible that earlier virologic sampling in patients with baseline Y181C minority variant may have detected variants containing both Y181C and K103N prior to K103N becoming the dominant species. Alternatively, the detection of baseline drug-resistant minority variant could be a marker of greater underlying viral diversity and may be associated with the presence of other undetected resistance mutations that eventually become the dominant species.

We found that the type of minority variant mutation at baseline clearly influenced which resistance mutations detected at virologic failure as participants on efavirenz were more likely to have Y181C detected at virologic failure when that mutation was present as a baseline minority variant. Which resistance mutation emerged at virologic failure also was strongly correlated with the NNRTI regimen. Our results support the observation both in vitro and in vivo that the Y181C mutation offers relatively high levels of resistance and fitness preservation in the setting of nevirapine exposure [15–17]. The same association has been found of the K103N resistance mutation in the setting of efavirenz use [17, 18].

A number of studies have now shown that nonwhite ethnicity is associated with increased risk of virologic failure [19, 20]. We found that nonwhite ethnicity was also associated with a higher risk of NNRTI resistance at the time of virologic failure. One potential explanation may lie in ethnic-specific distributions of genetic polymorphisms (eg, in CYP2B6) that affect antiretroviral medication (ARV) metabolism and drug concentrations. These genotypes have been shown to affect the risk of virologic failure and likely affect the risk of resistance emergence given that slow-metabolizer genotypes are more frequent in nonwhite participants and allow for longer periods of functional monotherapy after treatment discontinuation [21].

Due to its low cost, nevirapine continues to be one of the most commonly used ARVs, especially in developing countries. In this study, nevirapine use was also independently associated with a higher risk of NNRTI resistance at the time of virologic failure. However, this finding should be interpreted with caution given the variation in the NRTI backbones, which could modify the risk of treatment failure and resistance emergence.

This study has several limitations. First, we combined data from 7 studies using assays with varying limits of minority variant detection [3]. We performed a sensitivity analysis using an imputed proportion of minority variants for those without detectable minority variants using 10% of the assay limit of detection. The results were consistent with the analysis performed using measured minority variant proportion alone. In addition, one of the studies evaluating an efavirenz-based regimen did not test for the presence of the Y181C minority variant [14]. This omission may have led to an underestimation of the association between baseline Y181C minority variant presence and Y181C detection by standard genotyping at virologic failure. Finally, only a small proportion of the total study population was tested for the presence of NRTI resistance, which limits our ability to evaluate the impact of baseline NRTI-resistant minority variants on their emergence during treatment failure.

We have now demonstrated that for individuals initiating a first-line NNRTI-based regimen, NNRTI-resistant minority variants increase both the risk of virologic failure and NNRTI resistance detection at the time of treatment failure. Our results also show that the minority variants detected at baseline frequently differ from the resistance mutations observed at virologic failure. Additional studies of viral diversity, linkage analysis of low-frequency resistance mutations, and longitudinal observations during early virologic failure would provide further insights on how drug resistance mutations emerge and evolve during antiretroviral treatment failure.

Supplementary Data

Supplementary materials are available at The Journal of Infectious Diseases online (http://jid.oxfordjournals.org/). Supplementary materials consist of data provided by the author that are published to benefit the reader. The posted materials are not copyedited. The contents of all supplementary data are the sole responsibility of the authors. Questions or messages regarding errors should be addressed to the author.

Notes

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