High-Affinity H7 Head and Stalk Domain–Specific Antibody Responses to an Inactivated Influenza H7N7 Vaccine After Priming With Live Attenuated Influenza Vaccine

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Recent studies have shown that live attenuated influenza vaccines (LAIVs) expressing avian influenza virus hemagglutinins (HAs) prime for strong protective antibody responses to an inactivated influenza vaccine (IIV) containing the HA. To better understand this priming effect, we compared H7 HA head and stalk domain–specific B-cell responses in H7N7 LAIV–primed subjects and non–H7-primed controls after a single dose of H7N7 IIV. As previously reported, H7N7 LAIV–primed subjects but not control subjects generated strong hemagglutination-inhibiting and neutralizing antibody responses to the H7N7 IIV. Here, we found that the quantity, epitope diversity, and affinity of H7 head–specific antibodies increased rapidly in only H7N7 LAIV-primed subjects after receipt of the IIV. However, all cohorts generated a vigorous, high-affinity, stalk-specific antibody response. Consistent increases in circulating memory B-cell frequencies after receipt of the IIV reflected the specificity of high-affinity antibody production. Our findings emphasize the value of LAIVs as a vehicle for prepandemic vaccination.

Keywords. Avian influenza; H7 hemagglutinin; live attenuated; vaccine; antibody affinity; anti-stalk antibodies; memory B cells.

A key feature of pandemic influenza A viruses is a hemagglutinin (HA) that is novel to most of the human population. Novel HAs carried by pandemic viruses are typically derived from avian influenza viruses, which represent a source of at least 16 different HA subtypes (H1–H16). Avian influenza viruses expressing the H5 and H7 HA subtypes have raised pandemic concerns because of their ability to spread directly from birds to humans and cause severe disease [1, 2].

Vaccination strategies that generate protective levels of antibodies (Abs) against novel HA subtypes are key for pandemic preparedness. This reflects the major protective role of anti-HA Abs [3]. However, a challenge is the poor immunogenicity of novel avian HAs in adults. The hemagglutination inhibition (HI) and microneutralization (MN) Ab titers attained in prime-boost studies of unadjuvanted, inactivated H5 and H7 subvirion vaccines, even at high HA doses, have been at best modest [4–8]. Responses to live attenuated influenza vaccines (LAIVs) expressing H5 or H7 have also been poor [9–12].

Despite weak Ab induction by H5 or H7 LAIV alone, recent studies have shown that these vaccines effectively prime for a protective Ab response to a single dose of...
doses of an H7N7 LAIV derived from the H7N7 A/Netherlands/2/2000 (NL/00), which is antigenically similar to the H7 of NL/03 [17]. The current study analyzed sera and peripheral blood mononuclear cells (PBMCs) collected at the time of IIV administration (day 0) and on days 7, 14, and 28. PBMCs were frozen until required. Serum HI and MN titers were determined against the IIV virus [12].

**Generation of H7 HA1 and HA2 Domains**

The HA0 gene segments of H7N7 NL/03 and H7N9 A/Shanghai/1/2013 (SH/13) were chemically synthesized. DNAs encoding HA1 (1–320) and HA2 (331–540) of NL/03 and SH/13 were cloned, expressed, and purified as previously described [18]. HA1 proteins were properly folded and contained a high percentage of functional oligomers [18].

**Affinity Measurement by Surface Plasmon Resonance (SPR)**

Steady-state equilibrium binding of serum Ab after H7N7 IIV administration was monitored at 25°C, using a Bio-Rad ProteOn SPR biosensor [19, 20]. The recombinant HA globular head domain (rHA1-His6) or stalk domain (rHA2-His6) of NL/03 or SH/13 was coupled to a GLC sensor chip. The spatial density of antigen on the chip surface was adjusted to measure only monovalent Ab binding. Ab off-rate constants, which describe the fraction of antigen-Ab complexes that decay per second, were determined directly from sample interactions with rHA1 or rHA2 proteins, using SPR in the dissociation phase [19, 20].

**H7 Gene-Fragment Phage Display Library (GFPDL) Construction and Panning With Sera**

The HA0 gene segment of H7N7 NL/03 was used for construction of a GFPDL [21, 22]. Equal volumes of pooled polyclonal human sera from each cohort were used for each round of GFPDL panning. GFPDL selection was carried out in solution (with protein A/G) [20, 21, 23].

**Enzyme-Linked Immunospot (ELISpot) Assay for Ab-Secreting Cells**

Ab-secreting cells (ASCs) were enumerated by an ELISpot assay performed largely as previously described [24]. PBMCs were analyzed directly (day 7 after IIV receipt) or after in vitro stimulation (days 0, 14, and 28 after IIV receipt). The following reagents were used to coat plates for antigen-specific ASC enumeration: baculovirus-expressed trimeric HA ectodomains [25] from the H7N7 influenza strain NL/00, the H7N3 strain A/chicken/British Columbia/CN-6/2004 (BC/04), and the H7N9 strain A/Anhui/1/2013 (AN/13); a trimeric HA globular head protein from the H7N9 strain SH/13 [26]; chimeric HAs [27] consisting of an H4 head domain and H7 stalk domain (cH4/7) [26] or an H9 head domain and H1 stalk domain (cH9/1) [28]; and baculovirus-expressed full-length HAs from the
H7N7 strain NL/03 (BEI Resources Repository, National Institute of Allergy and Infectious Diseases, National Institutes of Health), the H1N1 strain A/California/04/2009 (CA/09; BEI Resources Repository), and the H3N2 strain A/Wisconsin/67/2005 (WI/05; Protein Sciences, Meriden, Connecticut). The chimeric Has primarily identified Ab binding to the HA stalk domain, since they carried exotic head domains that are novel to most humans [26, 28]. Wells were coated with goat antihuman immunoglobulin G (IgG; Life Technologies, Carlsbad, California) to measure total IgG ASCs. Negative control wells were coated with bovine serum albumin.

MBC Analysis
Measurement of antigen-specific MBC frequencies was based on previously described methods [29, 30]. PBMCs obtained on days 0, 14, and 28 after IIV administration were rested overnight and then stimulated for 6 days in complete medium containing 1 µg/mL R848 (Sigma Aldrich, St. Louis, Missouri), 10 ng/mL interleukin 2 (Life Technologies), and 55 µM 2-mercaptoethanol (Life Technologies). Antigen-specific and total IgG ASCs generated from stimulated MBC precursors were enumerated by an ELISpot assay. Antigen-specific IgG MBCs are shown as a percentage of total IgG+ MBCs, calculated as the percentage of antigen-specific ASCs among the total number of IgG ASCs. The limit of sensitivity for MBC frequency determination was set at 2 antigen-specific IgG ASCs/5 x 10^5 stimulated PBMCs.

Statistical Analysis
Differences were examined by paired t tests (for within-cohort comparisons) and by 2-sample t tests (for between-cohort comparisons), using data on a log scale. The corresponding nonparametric sign test and Wilcoxon rank sum test were used for confirmation of findings. Correlations were tested by Pearson correlation analysis and confirmed by Spearman correlation analysis. P values of <.05 were considered statistically significant.

RESULTS
HI and MN Responses
Babu et al [12] reported the shedding and immunogenicity of H7N7 LAIV and the HI and MN Ab responses to H7N7 IIV in the H7N7 LAIV–primed and unprimed cohorts. One or 2 doses of H7N7 LAIV did not generate a detectable H7-specific HI or MN response [12]. H7N7 LAIV–primed subjects in the current analysis represent a subset of subjects from the previous study [12] who were selected on the basis of sample availability. Of these, 6 of 7 responded to the H7N7 IIV, with HI and MN titers of ≥40, including 2 subjects who received only 1 of 2 scheduled H7N7 LAIV doses (Figure 1A and 1B). In contrast, no subject in the H2N3 LAIV–primed and unprimed cohorts had detectable HI or MN responses following IIV receipt ([12] and data not shown). There was no relationship between HI and MN responses to H7N7 IIV and H7N7 LAIV growth during priming. H7N7 LAIV shedding was detected in only 3 of 7 subjects after the first dose and in no subjects after the second dose.

Analysis of Serum Ab Binding to H7 HA1 and HA2
Polyclonal serum Ab binding to the HA1 (head) and HA2 (stalk) subunits of H7 was evaluated by SPR. Individual subject samples from days 0 and 14 were tested for binding to homologous HA1 and HA2 from H7N7 NL/03 and to a variant HA1 from H7N9 SH/13 (Supplementary Figure 1). Binding to the recombinant proteins was present on day 0, likely reflecting antigenic cross-reactivity between H7 and the HAs of seasonal influenza viruses (Supplementary Figure 1). Ab binding to NL/03 HA1 was equally low in all cohorts on day 0 and provided no evidence of prior H7-specific B-cell responses in H7N7 LAIV–primed subjects (Figure 2A). Binding to the homologous

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**Figure 1.** Hemagglutination inhibition (HI) and microneutralization (MN) antibody (Ab) titers after H7N7 inactivated influenza vaccination in H7N7 live attenuated influenza vaccine (LAIV)–primed subjects. Serum HI and MN titers against H7N7 NL/03 were determined at intervals after the inactivated vaccine. Each line connects data points for an individual subject (n = 7). Data points for subjects who received 2 LAIV doses are shown as filled circles; open circles identify data points for the 2 subjects who received only 1 H7N7 LAIV dose. Two subjects did not provide serum on day 28. Abbreviation: IIV, inactivated influenza vaccine.
HA1 increased substantially in the H7N7 LAIV–primed cohort from days 0 to 14. This increase was also significant in the unprimed cohort but was much smaller than in the H7N7 LAIV–primed cohort. On day 14, total anti-HA1 Ab binding was significantly higher in the H7N7 LAIV–primed cohort than in the other cohorts. Binding to the variant HA1 closely resembled the result for the homologous HA1, demonstrating extensive cross-reactivity by Abs induced by the IIV in the H7N7

Figure 2. Serum antibody (Ab) binding to H7 HA1 and HA2 proteins in H7N7 live attenuated influenza vaccine (LAIV)–primed, H2N3 LAIV–primed, and unprimed cohorts before and after H7N7 inactivated influenza vaccination. Total Ab binding to properly folded functional H7N7 NL/03 HA1-His6 (A), H7N9 SH/13 HA1-His6 (B), and H7N7 NL/03 HA2 (C) was measured by surface plasmon resonance. Maximum resonance unit (RU) values for Ab binding are shown for individual H7N7 LAIV–primed (circles), H2N3 LAIV–primed (triangles), and unprimed (diamonds) subjects on days 0 (open symbols) and 14 (filled symbols). Differences were examined by paired t tests (for within-cohort comparisons) and 2-sample t tests (for between-cohort comparisons), using data on a log scale. P values are shown for all within-cohort comparisons and for between-cohort comparisons that were significantly different (P < .05). Abbreviation: HA, hemagglutinin.

Figure 3. HA1-specific antibody (Ab) binding affinity following H7N7 inactivated influenza vaccination is enhanced by H7N7 live attenuated influenza vaccine (LAIV) priming. Off-rate constants are shown for polyclonal serum Ab binding to the H7 HA1 (A) and HA2 (B) proteins of influenza H7N7 NL/03. Sera were collected from H7N7 LAIV–primed (circles), H2N3 LAIV–primed (triangles), and unprimed (diamonds) cohorts at the time of inactivated vaccine administration (day 0; open symbols) and after 14 days (filled symbols). Ab off-rate constants, shown for individual subjects, were determined by surface plasmon resonance and provide a measure of net binding affinity. Differences were examined by paired t tests (for within-cohort comparisons) and 2-sample t tests (for between-cohort comparisons), using data on a log scale. P values are shown for all within-cohort comparisons and for between-cohort comparisons that were significantly different (P < .05). Abbreviation: HA, hemagglutinin.
LAIV–primed cohort (Figure 2B). HA2 binding significantly increased in all cohorts after the IIV (Figure 2C).

SPR determinations of dissociation rates for serum Abs bound to antigen are independent of Ab concentration and provide a measure of the net affinity of polyclonal Ab binding. The off rates for Abs bound to H7 HA1 decreased significantly from days 0 to 14 in the H7N7 LAIV–primed cohort and the unprimed cohort, indicating increased Ab affinity maturation (Figure 3A). However, Ab affinity for HA1 on day 14 was significantly stronger in the H7N7 LAIV–primed cohort than in the other cohorts. Ab affinity for H7 HA2 increased significantly in all cohorts from days 0 to 14, indicating that Ab affinity had not been maximized by exposure to the HA2 of seasonal influenza viruses (Figure 3B). Production of high-affinity Abs specific for HA1 in the H7N7 LAIV–primed cohort (in particular) and for HA2 in all cohorts after the IIV suggests activation of preexisting MBCs. Both the quantity and net affinity of HA1-binding Abs (but not HA2-binding Abs) on day 14 correlated with HI and MN titers (Supplementary Figure 2 and data not shown).

Analysis of Epitope Profiles of H7-Specific Serum Abs

GFPDL analysis of serum Abs was used to relate H7N7 LAIV priming to the repertoire of H7-specific Abs generated by the IIV. Pooled sera collected on days 0 and 14 were used to pan a GFPDL library displaying H7 HA peptides. The number of bound phages increased substantially from days 0 to 14 in the H7N7 LAIV–primed cohort but changed minimally in the other cohorts (Figure 4A). In the H7N7 LAIV–primed cohort, day 0 serum Abs bound predominantly to epitopes in the C-terminus portion of HA1 and to epitopes in HA2 (Figure 4B). Importantly, HA1 epitopes bound by day 14 Abs in the H7N7 LAIV–primed cohort were mostly in multiple large unique sequences that encompassed the receptor-binding domain (RBD). These sequences are likely to contain conformational epitopes targeted by neutralizing Abs [21, 22]. The number of bound RBD-spanning sequences increased from 2 on day 0 to 9 on day 14 (Figure 4). Day 14 Abs in the H7N7 LAIV–primed cohort also bound a substantially greater number of phages that expressed HA2 epitopes. In the other cohorts, the number of bound HA2 epitope–expressing phages

![Figure 4](https://academic.oup.com/jid/article-abstract/212/8/1270/2193302)
increased from day 0 to day 14, but binding to HA1 epitopes remained relatively weak. Overall, these findings indicate that H7N7 LAIV primed for a B-cell response to the IIV that was largely directed against RBD-associated epitopes and HA2 epitopes, whereas the response in the other cohorts was primarily against epitopes in HA2.

**Analysis of Ab-Secreting Plasmablasts (PBs)**

Strong Ab production following intramuscular influenza vaccination is associated with a brief wave of circulating Ab-secreting PBs that is thought to reflect MBC activation [15, 31]. PBMCs collected from the H7N7 LAIV-primed and unprimed cohorts on days 0 and 7 after IIV administration were analyzed for antigen-specific IgG PBs. Antigen-specific PBs were not detected on day 0. Both cohorts generated a strong H7 stalk-specific PB response to the IIV (Figure 5). H7 head-specific PBs were present in all subjects in the H7N7 LAIV-primed cohort but at lower frequencies than stalk-specific PBs. Some unprimed subjects had low numbers of H7 head-specific PBs, but most full-length H7-specific PBs in the unprimed cohort were specific for the stalk region. Low frequencies of PBs specific for the nonvaccine component H1 were present in both cohorts, suggesting some cross-reactive PB induction. Day 7 PB data were in agreement with the SPR analysis of HA1 and HA2 binding and the GFPDL epitope mapping performed using day 14 polyclonal sera (Figures 2 and 4, respectively).

**Analysis of Memory B Cells**

Collectively, our analysis of serum Ab and PB responses to the IIV suggested activation of H7 head–specific MBCs in the H7N7 LAIV–primed cohort and H7 stalk–specific MBCs in all cohorts. To test this directly, we measured antigen-specific MBC frequencies in PBMCs on day 0. MBCs were also analyzed on days 14 and 28, when postvaccination frequencies were likely to be maximal [31].

On day 0, most subjects in all cohorts had circulating MBCs specific for the full-length H1 and H3 HAs of recent seasonal influenza viruses and for the H1 stalk region (Figure 6 and Supplementary Figure 3). Generally, a smaller proportion of subjects in all cohorts had MBCs specific for the H7 stalk region. H7 head–specific MBCs were rarely detected in any cohort on day 0. In all cohorts, the frequencies of MBCs measured against full-length H7 and the H7 stalk were similar, indicating that few were specific for the head domain.

In all cohorts, there was a marked increase in the frequencies of H7 stalk–specific MBCs from days 0 to 14 (Figure 6). H7 head–specific MBC frequencies were also increased on day 14 in most subjects in the H7N7 LAIV–primed cohort and in a smaller proportion of subjects in the other cohorts. Full-length H7–specific MBC frequencies increased in all cohorts, reflecting the increased frequencies of head- and/or stalk-specific MBCs. A consistent increase in full-length H3–specific MBC frequencies in all cohorts from days 0 to 14 suggests conservation of at least some stalk epitopes in H7 and H3 (Supplementary Figure 3). H7 stalk–specific MBC frequencies in all cohorts remained substantially higher on day 28 than on day 0, but measurable H7 head–specific MBC frequencies in some subjects on day 14 had fallen to undetectable levels by day 28 (Figure 6). The frequencies of MBCs specific for the nonvaccine component H1 generally remained similar on days 0, 14, and 28 in all cohorts. Overall, the expansion of MBC populations largely reflected the specificity of high-affinity Ab production.

**DISCUSSION**

In the current study, we sought to better understand the impact of H7N7 LAIV priming on the H7-specific B-cell response to H7N7 IIV. Our analysis discriminated between responses to the H7 head and stalk domains. We show that H7N7 LAIV priming promotes a strong, diverse, and high-affinity Ab response to the H7 head domain, including epitopes in the RBD, after H7N7 IIV administration. Both the magnitude and net affinity of this response correlated with protective Ab levels. We also demonstrate strong H7 stalk–specific Ab responses to the IIV in H7N7 LAIV–primed and non–H7-primed subjects.
A key question is the nature of immune memory generated by H7N7 LAIV priming that influences the H7-specific B-cell response to the IIV. Our analysis indicates that the response to the IIV in H7N7 LAIV–primed subjects reflects more-extensive affinity maturation of H7 head–specific Abs than in non–H7-primed subjects. One possibility is that the IIV activated affinity-matured, H7 head–specific MBCs that were induced during H7N7 LAIV priming. This is suggested by the presence of circulating H7 head–specific PBs in all H7N7 LAIV–primed subjects on day 7 after the IIV (Figure 5). Circulating HA–specific PBs on day 7 after influenza vaccination are thought to be the products of activated MBCs, since they carry highly mutated immunoglobulin variable region genes [15, 31]. The activated MBCs may have differentiated directly into ASCs or entered germinal centers for additional rounds of affinity maturation before ASC formation [32]. Germinal center remodeling of the specificities of preexisting MBCs might account for the induction of day 7 PBs specific for the nonvaccine component H1 (Figure 5), without expansion of H1–specific MBC populations (Figure 6) [33]. We did not detect H7 head–specific MBCs in the circulation of most of the H7N7 LAIV–primed subjects at the time of H7 IIV administration (Figure 6). However, this does not necessarily indicate the absence of these cells, since the major MBC repositories are in secondary lymphoid tissues [34, 35]. Our analysis of H7 head–specific MBCs (Figure 6) suggests that small MBC populations generated by vaccination might be only transiently detected in the circulation.

The effect of H7N7 LAIV priming might also reflect the induction of H7–specific CD4+ T cells. Recently, Nayak et al [36] demonstrated that subjects primed with nonreplicating forms of H5 generated a long-maintained increase in H5–specific CD4+ T cells and responded to an H5 boost with a more robust expansion of H5–specific CD4+ T cells than did non–H5-primed subjects. Furthermore, the H5–specific CD4+ T-cell expansion correlated with MN Ab production. However, the extent to which MN Ab production reflected the availability of T-cell

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**Figure 6.** Circulating memory B cells (MBCs) in H7N7 live attenuated influenza vaccine (LAIV)–primed (A), H2N3 LAIV–primed (B), and unprimed (C) cohorts before and after H7N7 inactivated influenza vaccination. Antigen-specific immunoglobulin G (IgG) MBC frequencies were determined in peripheral blood mononuclear cells (PBMCs) collected on days 0, 14, and 28. MBC analysis was based on in vitro stimulation to generate IgG antibody-secreting cells (ASCs), followed by enumeration of antigen-specific and total IgG ASCs. Antigen-specific MBC frequencies are shown as the percentage of total IgG MBCs. MBCs specific for H7 from NL/00, H7 head only from SH/13, chimeric ch4/7, and H1 from CA/09 are shown for each cohort. MBCs specific for H7 from NL/03, H3 from WI/05, and chimeric ch9/1 are shown in Supplementary Figure 3. PBMCs for all sampling times were not available for all subjects. To illustrate kinetic patterns, data points from consecutive samplings are connected for individual subjects. The numbers of data points per sampling time are 5–6 for the H7N7 LAIV–primed cohort, 5 for the H2N3 LAIV–primed cohort, and 7–9 for the unprimed cohort. Abbreviation: IIV, inactivated influenza vaccine.
help or preexisting H5-specific MBCs is unclear. Healthy adults who have only been exposed to the HA5 of seasonal influenza viruses have CD4+ T cells that cross-react with H5 and H7 HAs [36–38]. In our analysis, the vigorous stalk-specific Ab response to the H7N7 IIV by non–H7-primed subjects likely reflects a T-cell–dependent process involving CD4+ T cells and stalk-specific MBCs generated through encounters with seasonal influenza viruses. The implication is that H7 head–specific Ab production in non–H7-primed subjects is primarily limited by a deficiency in head-specific MBCs rather than by a shortage of T-cell help for H7-specific B-cell responses.

The H7N7 IIV used in the current study elicited very few significant HI and MN Ab responses when administered in a prime-boost regimen to H7-naive subjects, even at high doses [6]. Our analysis demonstrates that a single dose of this vaccine generates a rapid and vigorous H7 stalk–specific Ab response in H7N7 LAIV–primed and non–H7-primed subjects. Recent reports of human Ab responses to H5 and H7 IIVs described similar findings [26, 39]. It has been proposed that strong anti-stalk Ab responses are generated when stalk-specific MBCs respond in the absence of competition from MBCs specific for the immunodominant head region [15]. Recently, Ellebedy et al [16] demonstrated that the first dose of an H5 IIV elicited a stronger serum Ab response to the stalk than to the H5 head. However, a second dose of the same vaccine generated a vigorous antihemagglutinin response but only a feeble antistalk response. This is consistent with the first dose expanding the head-specific MBC population so that it outcompeted the stalk-specific MBC population after the second dose. In our analysis, however, subjects primed to generate a strong anti–H7 head Ab response to the H7N7 IIV also generated a strong antistalk Ab response. Additional studies are required to determine whether this relates to the timing of the boost [5, 36, 40, 41] or to the character of HA-specific B-cell and T-cell memory generated when LAIV is used as the priming vehicle.

Our analysis extends previous studies demonstrating that LAIVs expressing avian HAs prime for protective HI and MN Ab responses to a single dose of the HA in the form of an unadjuvanted IIV [12, 13]. We show that H7N7 LAIV primes for a high-affinity H7 head–specific Ab response to H7N7 IIV without down-modulation of the antistalk Ab response. Collectively, our analysis indicates high-affinity H7 head–specific MBC induction by H7N7 LAIV, although we were unable to directly demonstrate these MBCs at the time of IIV administration. Notably, the 2 subjects who received only a single H7N7 LAIV dose could not be distinguished from the 2-dose recipients in their response to the H7N7 IIV. Thus, a single dose of LAIV expressing an avian HA might be sufficient as a highly effective, pre-pandemic priming strategy.

Supplementary Data

Supplementary materials are available at The Journal of Infectious Diseases online (http://jid.oxfordjournals.org). Supplementary materials consist of data provided by the author that are published to benefit the reader. The posted materials are not copyrighted. The contents of all supplementary data are the sole responsibility of the authors. Questions or messages regarding errors should be addressed to the author.

Notes

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