Characterization and comparison of iron oxyhydroxide precipitates from biotic and abiotic groundwater treatments

Katarzyna R. Arturi, Christian B. Koch and Erik G. Søgaard

ABSTRACT

Removal of iron is an important step in groundwater treatment for drinking water production. It is performed to prevent organoleptic issues and clogging in water supply systems. Iron can be eliminated with a purely physico-chemical (abiotic) method or biotically with the help of iron-oxidizing bacteria (FeOB). Each of the purification methods requires different operating conditions and results in formation of iron oxyhydroxide (FeOOH) precipitates. Knowledge about the differences in composition and properties of the biotic and abiotic precipitates is desirable from a technical, but also scientific point of view. In this study, a broad spectrum of analytical methods was employed to describe in detail the differences between FeOOH produced in a number of Danish waterworks using either the purely physico-chemical or the biological variation of the iron removal method. According to the results from X-ray diffraction (XRD), small angle X-ray spectroscopy, and visible light microscopy, the precipitate formed with the help of FeOB was more microcrystalline and consisted of agglomerated particles. The purely physico-chemical precipitate was more amorphous and, typically, was composed of smaller particles.

Key words | abiotic iron oxyhydroxides, biotic iron oxyhydroxides, groundwater treatment, iron oxidizing bacteria, iron removal

INTRODUCTION

The ordinary technique for removal of dissolved iron from groundwater, typically referred to as a purely physico-chemical (or abiotic) treatment, involves aeration of groundwater followed by catalytic oxidation of ferrous iron Fe(II) to ferric iron Fe(III). The simultaneous oxidation and precipitation of ferric oxyhydroxides (FeOOH) take place on the surface of sand grains in in-depth sand filters (Sharma et al. 1999, 2001). For groundwaters with high iron concentrations, the purely physico-chemical iron removal is often not sufficient, and Fe can be eliminated biologically (biotically) by the means of iron-oxidizing bacteria (FeOB), a group of morphologically heterogeneous chemolithotrophic procaryotes living in iron-rich environments (Sharma et al. 2005; Li et al. 2006). Such conditions can be found in many waterworks in Denmark, especially in the southwestern part of Jutland (Søgaard et al. 2000, 2001). The biotic and abiotic iron removal techniques are similar in their application in waterworks (aeration followed by filtration), but require different levels of oxygen in the processed raw water to become optimized (10 mg/L and 1 mg/L for the purely physico-chemical and biological iron removal, respectively) (Pacini et al. 2014). While high iron and low oxygen contents promote the biologic iron removal and retard its physico-chemical counterpart (van Beek et al. 2016), the removal of iron in waterworks employing FeOB is known to be a combination of both processes, resulting
in precipitates of mixed origin. The extent of each process and the final composition of the precipitate depend on the applied process conditions (pH, flow, oxygen content, temperature).

Most of the studies regarding biotic FeOOH precipitates focus on the mechanisms for their formation and the bacterial metabolism (Hallberg & Ferris 2004; Ferris 2005; Bird et al. 2011). The mechanisms for the formation of FeOOH in the presence of bacteria are still widely debated. These precipitates may often contain FeOB-originating exopolymers operating as nucleation sites upon which FeOOH precipitate readily. However, it has been proposed that iron precipitation is a secondary effect that could be attributed to the metabolic activity of nitrogen mediating bacteria (Kluglein et al. 2014). The composition of the exopolymers has also been disputed widely, and the consensus is that the exopolymers are of organic origin (Chan et al. 2009). The general structure of the biotic precipitate has been identified as poorly ordered ferrithydrate (Kennedy et al. 2005), with few variations (crystalline and amorphous) allowed depending on the conditions (equilibrium and flow, respectively) (Chan et al. 2004). Several authors claim that the amorphous ferrithydrate of biological origin can crystallize into goethite (Kennedy et al. 2005) or lepidocrocite (Rhodon et al. 2002) by aging in water. While the abiotic precipitate has the amorphous structure of ferrithydrate (Cornell & Schwertmann 2003), as is expected from a process with high kinetics rates of precipitation, the properties of both precipitates are typically quite different. The biotic precipitate is denser and closely packed, compared to the loosely bound physico-chemical one. The difference is attributed to the cross-bonding of the FeOOH particles by the carbohydrate containing exopolymers (Søgaard et al. 2000). Biotic and abiotic iron removal treatments used in drinking water production have been compared many times, mostly concerning the iron removal efficiency (Mouchet 1992; Pacini et al. 2005; De Vet et al. 2011). According to the results, the presence of FeOB significantly increases the kinetic rates of iron removal (Søgaard et al. 2000). From both the scientific and the technical points of view, it would be desirable to discriminate between the biotic and abiotic precipitates, and yet, very few studies have focused on the structure and properties of FeOOH solid by-products from drinking water production. In this study, a broad spectrum of analytical tools was employed to expand the knowledge about the structure and properties of biotic and abiotic FeOOH precipitates. The aim was to describe differences between them in dry conditions and to study the influence of the microbiological activity on the final solid iron containing product. The samples were obtained from several Danish waterworks remediating iron from the groundwater by a purely physico-chemical method or by the means of FeOB. The precipitates were compared with regard to their crystallinity, composition, thermal properties, particle size, morphology, magnetic properties, and fine structure.

**MATERIALS AND METHODS**

**Samples**

In this study, precipitates collected at selected Danish waterworks using either the purely physico-chemical or FeOB-based iron removal methods were compared. A high number of waterworks was used in order to cover the natural variation of the precipitates’ compositions and to make the results representative (Table 1). While all the samples were studied to obtain the conclusions, only a limited number of distinct and representative results for each group are presented in the figures and tables. The omitted results were almost identical to some of those shown. In order to confirm the type of precipitate, all samples were examined microscopically for the presence of exopolymers shaped as twisted stalks, which are characteristic for Gallionella ferruginea. In all samples produced by the purely physico-chemical treatment method, the exopolymers were

<table>
<thead>
<tr>
<th>Type</th>
<th>Method</th>
<th>Location</th>
<th>Type</th>
<th>Method</th>
<th>Location</th>
</tr>
</thead>
<tbody>
<tr>
<td>WW</td>
<td>AB</td>
<td>Forum</td>
<td>WW</td>
<td>AB</td>
<td>Jernvedlund</td>
</tr>
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<td>Grindsted (I)</td>
<td>WW</td>
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<tr>
<td>WW</td>
<td>B</td>
<td>Astrup</td>
<td>WW</td>
<td>B</td>
<td>Grindsted (II)</td>
</tr>
<tr>
<td>PP</td>
<td>B</td>
<td>Vognsøl</td>
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</table>

WW, waterworks.  
PP, pilot plant.  
AB, abiotic.  
B, biotic.  
The Vognsøl PP has been described in detail by Søgaard et al. (2001).
absent. With minor deviations, the waterworks samples were produced according to the previously described groundwater treatment procedure: aeration followed by in-depth sand filtration. The samples were collected from the sand filters during backwashing with air to loosen the precipitate from the sand grains, followed by backwashing with water to remove the precipitate from the filter. The filters at the waterworks typically consist of two layers: (1) fine corned sand (100 cm, grain size 1.0–1.5 mm); and (2) sand grains with increasing diameter for the increasing depth of the filter (1.0–1.4 m, grain size 2–8 mm). The velocity of filtration was ≈5–8 m/h. The velocity of backwash was ≈15–18 m/h. The filter medium at the pilot plant (PP) consisted of sand, 0.5 m deep with a grain size of 1.5 ± 3.0 mm. Sand filters on the PP were cultivated with *G. ferruginea* by mixing new sand grains with the sand grains collected at the Astrup waterworks. The iron content of the groundwater was regulated by addition of FeSO₄ (technical grade, JSF, Kemira MiljøA/S). The precipitates were collected as filters through 0.45 μm sterile filters, diluted into range 0–10 mg/L, and analyzed for the major constituents of the precipitates, i.e., for total Fe, Ca, and Mn concentrations. The functional groups present in the samples were identified with a Perkin Elmer PE2000 Fourier transform infrared (FTIR) spectrometer. Infrared spectra were obtained using the KBr pressed pellet technique (≈0.3 mg sample in 300 mg KBr).

**Thermal properties**

Differential scanning calorimetry (DSC 822 Mettler Toledo) and thermogravimetric analysis (TGA/SDT 851 Mettler Toledo) were used to determine the thermal properties and content of the samples. A dynamic method of heating with a heating ramp of 10 K/min was applied for both DSC and TGA. Measurements were performed in inert and oxidative atmospheres. N₂ was used as a protective gas, while air and O₂ were applied as the reactive gas in the inert and oxidative runs, respectively. The freeze-dried powders were weighted (≈5 mg for DSC and ≈10 mg for TGA), placed in aluminum (DSC) or alumina (TGA) bins, sealed, and analyzed. The TGA analysis was preceded by two blank runs. Each measurement was performed in triplicate.

**Particle size and morphology**

Small-angle X-ray scattering (SAXS) data were obtained using an adaptation of a Bruker AXS, Nanostar SAXS system, with a rotating anode X-ray generator, cross-coupled Goebel mirrors, three pinholes, and a Bruker AXS Hi-star Area Detector (Cu – K, = 1.54 Å, 0.3 × 0.3 mm² source point, 6 kW power). The scattering intensity (*I*) was measured, and the data were corrected (background and azimuthal average) to obtain a spectrum of average *I* vs. *q*. The results were used to determine the particle size and fractal dimension of the precipitates. The particle size was determined with Guinier approximation for spherical particles 

\[
\ln(I) = \ln(I_0) - \frac{1}{3} R_g^2 \cdot q
\]

The radius of gyration (*Rg*) was calculated by extrapolation of intensity at zero scattering angle (*I₀*) from the ln(*I*) vs. *q*² plot. The fractal dimension values were estimated in the Porod’s region (*I* ≈ a + b · *q*⁴; a,b-constants). Porod’s law was extended by changing the
power from 4 to 6-$D_F$, as the studied FeOOH particles were considered to have complex, and not entirely smooth, surfaces. The values of fractal dimensions ($D_F$) were extracted from the slope of $\ln(I)$ vs. $\ln(q)$ plot for high $q$-values ($\ln(I) = A - (6D_F) \cdot \ln(q)$). The values were determined with SASFit package (version 0.94.0, Joachim Kohlbrecher and Ingo Bressler, Laboratory for Neutron Scattering, Paul Scherrer Institut, Switzerland). The number of points used in calculations was optimized in order to reduce the value of $q \cdot R_g$ (Guinier’s region) and to minimize the value of $\chi^2 = \sum_{i=1}^{N} \frac{Y_i - C_i}{\sigma_i}^2$ (Porod’s region). A Leica visible light microscope with phase contrast and differential interference contrast was used to obtain images of the precipitates recorded with a camera using photo films. A Philips CM20 transmission electron microscope (TEM) was used to view samples air-dried onto copper grids.

**Magnetic properties**

Magnetic properties of the samples were analyzed with electron paramagnetic resonance (EPR). A Bruker ESP-300E spectrometer operated in the absorption mode at 9.2 GHz, in conjunction with a Varian E-257 variable temperature device and a Hewlett-Packard 5350B frequency counter with a Bruker ER035M NMR gaussmeter were used. The results were recorded as an intensity ($I$, a.u.) vs. magnetic field ($B$, Gauss), and converted to $I$ vs. $g$-values by the formula: $g = (714.484 \cdot \nu) / B$, where $\nu$ is frequency (Hz).

**Fine structure**

Fine structure of the samples was analyzed with Mössbauer spectroscopy. The spectra were obtained using a conventional constant acceleration spectrometer and a $^{57}$Co in a Rh source. The spectrometer was calibrated using a 12 $\mu$m thick foil at room temperature, and isomer shifts were given relative to the centroid on this absorber. Spectra were measured between room temperature and 5 K using closed cycle and flow-type He cryostats. Hyperfine parameters were obtained by fitting a combination of Lorentzian line-shaped doublet and sextet components constrained to be pairwise identical.

**RESULTS AND DISCUSSION**

**Crystallinity**

According to the XRD results, the main constituent of the precipitates, regardless of the applied precipitation method, was amorphous ferricydrite. For the abiotic samples, the amorphous structure was identified as 2-line ferricydrite, while the biotic samples resembled the more crystalline 6-line ferricydrite (Figure 1). The identified peaks characteristic for 6-line ferricydrite included 41.45°, 46.92°, 53.80°, 62.33°, 72.60°, and 74.82°. In both the biotic as well as the abiotic samples, some crystalline minerals were identified, e.g., quartz, chalk, and gypsum. The presence of the first two minerals (or the lack thereof) can be attributed to the composition of the sand filter and the addition of chalk at some of the waterworks. Gypsum was formed by the reaction of sulfate from the groundwater and the chalk. No crystalline forms of iron oxyhydroxides were found in the biotic precipitate. The nature of the registered XRD signal (broad peaks, baseline, high noise level), made it difficult to interpret the data unambiguously and determine the composition with precision. This is not an uncommon phenomenon for XRD analysis of amorphous iron oxides (Carlson & Schwertmann 1981).

**Figure 1** XRD diffractograms. The characteristic peaks of 6-line ferricydrite are marked with X. X and Y represent heterogeneous elements incorporated into the structure of ferricydrite.
Composition

The composition of the precipitates formed during groundwater treatment depends on the composition of the groundwater, as discussed previously (Czekalla et al. 1985). The AAS analysis has shown that the precipitates consisted mainly of FeOOH, minor amounts of calcium, and traces of manganese (Table 2). The abiotic samples were more heterogeneous than the biotic ones, i.e., they contained more calcium. According to the FTIR results (Figure 2), the biotic samples were dominated by many bands indicating the presence of FeOOH: ≈1,000 cm$^{-1}$ (Si-O-Fe and Fe-O-Fe), 538 cm$^{-1}$ and 471 cm$^{-1}$ (Fe-O). Furthermore, the presence of water (-OH stretch) and -COOH groups, which would support the theory about the organic origin of the FeOB stalks, was determined as well (1,633 cm$^{-1}$ and 1,419 cm$^{-1}$, respectively). In addition to the typical FeOOH absorption bands, the abiotic samples exhibited bands at 1,420, 875, and 712 cm$^{-1}$, expressing the presence of calcite (CaCO$_3$).

<table>
<thead>
<tr>
<th>Content (wt. %)</th>
<th>Fe</th>
<th>Ca</th>
<th>Mn</th>
</tr>
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<tr>
<td>Biotic (Waterworks)</td>
<td>43.1</td>
<td>0.4</td>
<td>0.2</td>
</tr>
<tr>
<td>Biotic (Vognsbøl PP)</td>
<td>46.9</td>
<td>0.2</td>
<td>0.0</td>
</tr>
<tr>
<td>Abiotic (Waterworks)</td>
<td>42.2</td>
<td>2.3</td>
<td>0.2</td>
</tr>
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</table>

Thermal properties

The obtained DSC and TGA results (Figures 3 and 4, respectively) have shown that the FeOOH precipitates, when heated, undergo both exothermic and endothermic transformations leading to a significant loss of mass (30–35 wt.%), which is in agreement with previous findings (Oliveira et al. 2005). For both types of precipitates, the DSC curves were dominated by a broad exothermic peak between
200 and 400 °C representing the solid–solid transformation of ferrihydrite into a more crystalline form of iron oxyhydroxides, e.g., hematite. This conversion has begun as an endothermic oxygen uptake at ≈200 °C in the oxygen-rich atmosphere. The process took place through dehydration, dehydroxylation, and rearrangements, and resulted in significant mass loss between 200 and 300 °C for both precipitate types. The biotic ferrihydrite was converted more rapidly, as evidenced by higher mass loss gradients and heat values. Above the temperature of 300 °C, the biotic samples continued to lose mass, and the difference between the results procured with O₂ and N₂ could be attributed to the oxidation of the organic core of exopolymers. Above 400 °C, the mass loss for the biotic samples stagnated, while a substantial decrease in the mass, attributed to the decomposition of CaCO₃, could be noted for the abiotic samples above 700 °C. The pyrolytic decomposition of the samples in the N₂ atmosphere and oxidation with O₂ resulted in similar final mass losses for the respective types of samples.

**Particle size and morphology**

According to the SAXS results (Figure 5), the abiotic precipitate consisted of smaller particles, while the biotic precipitation results in the formation of iron hydroxide agglomerates. Although not all requirements for the model were satisfied ($q < 1/R_g$), probably due to the polydispersity and agglomeration of the particles, it was still possible to compute and compare the values for biotic and abiotic precipitates. The average values of the particle size, which were expressed as $R_g$ (Table 3), were larger than previously calculated, most probably due to the unique method of production (Gilbert et al. 2009). The $D_F$ values were determined to lie in the range 1–2, lower for the biotic samples than for the abiotic ones, which indicates a lower degree of surface complexity. In general, the computed $D_F$ were quite small. Similar results from laboratory samples were obtained by other authors (Gilbert et al. 2007), who explained that the low fractals might be due to the open structure of the clusters. Agglomeration of the particles in the biotic precipitate was seen clearly in the visible light microscopy images presented in Figure 6. The exact structure of the exopolymers observed is shown using TEM (Figure 7).

**Magnetic properties**

The obtained EPR spectra were shaped as broad tops instead of sharp peaks (Figure 8), which could be attributed

### Table 3

<table>
<thead>
<tr>
<th>Sample</th>
<th>$R_g$ (Å)</th>
<th>$D_F$</th>
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<tbody>
<tr>
<td>Biotic</td>
<td>152 ± 2</td>
<td>1.47 ± 0.02</td>
</tr>
<tr>
<td>Abiotic</td>
<td>115 ± 2</td>
<td>2.02 ± 0.02</td>
</tr>
</tbody>
</table>

![Table 3](https://iwaponline.com/aqua/article-pdf/66/2/96/398284/jws0660096.pdf)

Figure 5 | SAXS results showing the linearity of the curves at the low angles in the Guinier’s region.

![Figure 5](https://iwaponline.com/aqua/article-pdf/66/2/96/398284/jws0660096.pdf)

Figure 6 | Light microscopy images (magnification 100×) of the (a) biotic and (b) abiotic iron precipitates.
to the large amounts of paramagnetic iron in the precipitates. At $g \approx 4.2 \ (B \approx 1,500 \text{ G})$, a single peak represents ‘high-spin’ Fe$^{3+}$ in low symmetry (rhombic) and distributed crystal field, as noted previously by Olivié-Lauquet et al. (1999). At $g \approx 2.0$, the presence of manganese is expressed as a sextet pattern characteristic of the hyperfine split $M = -\frac{1}{2} \leftrightarrow +\frac{1}{2}$ central field transition of Mn(II). The minor difference between the EPR spectra of the biotic and purely abiotic samples was related to the exact shapes of the curves. For the biologically precipitated FeOOH samples, the curves were flat, while the precipitates from the purely physico-chemical process resulted in more hill-shaped peaks. This phenomenon could relate to some of the samples’ features, most probably the particle size.

Mössbauer analysis has shown that at $T = 80 \text{ K}$, all the samples exhibit spectra with one doublet only. The parameters of the doublets ($\Delta E_Q = 0.85 \cdot 0.02 \text{ mm} \cdot \text{s}^{-1}$ and $\delta = 0.46 \cdot 0.02 \text{ mm} \cdot \text{s}^{-1}$ with a linewidth of approximately $0.50 \text{ mm} \cdot \text{s}^{-1}$) were indicative of high-spin Fe(III) with oxygen as a ligand in a relatively distorted octahedra. At $T = 15 \text{ K}$, the spectra of the biologically precipitated samples exhibited one sextet with very broad lines (Figure 9). A simple fit of these spectra yielded a magnetic hyperfine splitting of the sextet with a negligible quadruple shift and an outer linewidth $\approx 1.66 \text{ mm} \cdot \text{s}^{-1}$. From the temperature scan of the biologically precipitated samples, a blocking temperature of about $35 \text{ K}$ was estimated. These parameters are typical for poorly ordered ferrihydrite. In contrast, the spectra of the abiotic precipitates contained a doublet component. Here, the samples exhibited behavior either completely dominated by the doublet ($\approx 95\%$, with similar quadruple splitting at $80 \text{ K}$, and blocking temperature below $15 \text{ K}$), or with partially ordered magnetic spectrum with a magnetic hyperfine field at $\approx 25 \text{ K}$. Mössbauer spectroscopy has shown that the biotic samples were more easily ordered by lowering the temperature, probably due to the presence of agglomerates. The abiotic...
samples were not easily ordered due to the smaller particles. Samples from Grindsted I did not show sign of a sextet even at liquid He temperature (5 K). The iron precipitate collected in Forum contained considerable amounts of manganese, which gave rise to a mixed Mössbauer spectrum.

CONCLUSIONS

A broad spectrum of analytical tools can be used to gain knowledge about the differences between the biotic and abiotic FeOOH precipitates. While the majority of the precipitate consisted of Fe, other elements were present as well, e.g., Ca, Mn, and Si, as evidenced by the applied analytical tools. The structure of the studied iron hydroxides depended primarily on the manner of their formation, i.e., the groundwater treatment technique used. The results have shown that the biotic precipitate is slightly more crystalline than its purely physico-chemical counterpart, yet no purely crystalline iron oxyhydroxides were detected. The difference in the mineral structure was confirmed partly by the thermal analysis, which showed faster -OH desorption and recrystallization of the biotically formed FeOOH. The FTIR indicated the presence of organic -COOH functional groups, which would confirm the organic origin of the exopolymers, and their role as a template for a combined biotic and abiotic iron oxidation and precipitation. The presence of stalks resulted in larger particles forming agglomerates, and thus also a denser precipitate. The differences in particle sizes were also indicated by the EPR and Mössbauer spectral analysis. The biologically precipitated flocs contain carbohydrates and smaller amounts of proteins and fats (Ghiorse 1984). The biotic sludge is much more dense than the purely abiotic physico-chemical precipitate, which appears as distinct colloidal FeOOH particles dispersed in the water. Therefore, the frequency of backwashing the sand filters is lower on waterworks using FeOB than on waterworks using aeration and chemical catalytic precipitation. In the first filtration step, a joint removal of iron, manganese, and arsenic can take place with the help of adsorption and co-precipitation, and that process is improved when a biological iron oxidation/precipitation principle is used at the waterworks. Normally, a second filtration step takes care of manganese removal by increasing pH with the help of strong oxidation of the water from the first filtration step.

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REFERENCES


