Pulmonary Responses to *Stachybotrys chartarum* and Its Toxins: Mouse Strain Affects Clearance and Macrophage Cytotoxicity

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We investigated differences in the pulmonary and systemic clearance of *Stachybotrys chartarum* spores in two strains of mice, BALB/c and C57BL/6J. To evaluate clearance, mice were intratracheally instilled with a suspension of radiolabeled *S. chartarum* spores or with unlabeled spores. The lungs of C57BL/6J mice showed more rapid spore clearance than the lungs of BALB/c mice, which correlated with increased levels of spore-associated radioactivity in the GI tracts of C57BL/6J as compared with BALB/c mice. To identify mechanisms responsible for mouse strain differences in spore clearance and previously described lung inflammatory responses, we exposed alveolar macrophages (AMs) lavaged from BALB/c and C57BL/6J mice to *S. chartarum* spores, *S. chartarum* spore toxin (SST), and satratoxin G (SG) in vitro. The *S. chartarum* spores were found to be highly toxic with most cells from either mouse strain being killed within 24 h when exposed to a sporecell ratio of 1:75. The spores were more lethal to AMs from C57BL/6J than those from BALB/c mice. In mice, the SST elicited many of the same inflammatory responses as the spores in vivo, including AM recruitment, pulmonary hemorrhage, and cytokine production. Our data suggest that differences in pulmonary spore clearance may contribute to the differences in pulmonary responses to *S. chartarum* between BALB/c and C57BL/6J mice. Enhanced AM survival and subsequent macrophage-mediated inflammation may also contribute to the higher susceptibility of BALB/c mice to *S. chartarum* pulmonary effects. Analogous genetic differences among humans may contribute to reported variable sensitivity to *S. chartarum*

Key Words: *Stachybotrys chartarum*; rodent; toxicity; acute; safety evaluation; macrophage; immunotoxicology; lung; pulmonary or respiratory system; respiratory toxicology; dose-response; risk assessment.

Inhalation of *Stachybotrys chartarum* has been associated with multiple symptoms, including muscle aches, headaches, cough, pulmonary hemorrhage, dermatitis, and interstitial lung disease (Cooley et al., 1998). Infant deaths from acute idiopathic pulmonary hemorrhage have been hypothesized to be caused in part by this fungus (Dearborn et al., 1999; Etzel et al., 1998). Responsiveness to *S. chartarum* is idiosyncratic; some humans respond, others do not. Because separating environmental from genetic effects is difficult in humans, we used a mouse model (Rosenblum Lichtenstein et al., 2006) and an *in vitro* cell assay to elucidate mechanisms of responses to *S. chartarum* and to clarify the importance of strain differences that reflect genetic components of host responses.

*Stachybotrys chartarum* grows well at room temperature on wet surfaces composed of cellulose-containing materials such as paper, ceiling tiles, and cardboard (Murtoniemi et al., 2003). With appropriate temperature, light, and relative humidity, it produces mycotoxins, including macrocyclic trichothecenes such as satratoxin G (SG), which inhibit protein and DNA synthesis, induce protein degradation, disrupt cellular function, and cause cellular injury and inflammation (Bae et al., 2009; Hastings et al., 2005; Hudson et al., 2005; Kankkunen et al., 2009; McCrae et al., 2007; Shi et al., 2009; Yike et al., 2007). The fungus also releases stachylysin that can lead to pulmonary hemorrhage (Vesper SJ and Vesper MJ, 2002).

Previous experiments used exposures of rats and mice to *S. chartarum* spores intranasally and intratracheally; lung tissue was examined for histological changes and bronchoalveolar lavage for evidence of injury and inflammation (Nikulin et al., 1996, 1997; Rao et al., 2000; Rosenblum Lichtenstein et al., 2006). *Stachybotrys chartarum* has been consistently reported to cause pulmonary hemorrhage and extensive inflammation as well as apoptosis, cytokine release, DNA damage, and changes in gene expression (Bae et al., 2009; Chung et al., 2003b; Dearborn et al., 1999; Islam et al., 2006, 2008; Penttinen et al.,...
In addition to mycotoxins, Stachybotrys chartarum releases SchS34, a 34-kD antigen (Rand and Miller, 2008).

We have previously shown that BALB/c mice respond more to pulmonary exposure to St. chartarum spores than do C57BL/6J or C3H/HeJ mice (Rosenblum Lichtenstein et al., 2006). Lungs from exposed BALB/c mice showed the most hemorrhage, the highest permeability of the air-blood barrier, the most white blood cell recruitment, and the highest inflammatory cytokine and chemokine levels in response to intratracheally instilled spores of St. chartarum. Our data also show that mice have dose-dependent pulmonary responses to St. chartarum and that their genetic background affects the slope of these dose-response curves.

We wanted to know whether strain differences influence clearance kinetics of spores. BALB/c mice clear both Bacillus anthracis and Cryptococcus neoformans from the lungs more efficiently than do C57BL/6J mice (Decken et al., 1998; Lyons et al., 2004). The mucociliary escalator and macrophage activity play important roles in the pulmonary clearance of pathogens in the lungs; multiple environmental factors modulate this role.

In this study, we sought to clarify the roles of pulmonary clearance and macrophage cytotoxicity in modulating responses to St. chartarum. We hypothesized that the observed strain differences in St. chartarum responsiveness are mediated in part by different rates of pulmonary spore clearance. The two mouse strains we employed here, C57BL/6J and BALB/c, are inbred strains developed independently and differ from each other at over 50% of characterized genetic loci (Taconic, 1998a,b). Both strains are TLR2+ and TLR4+ (Matsuguchi et al., 2003; Poltorak et al., 1998). BALB/c mice have been shown to be Th2 dominant, whereas C57BL/6 mice are Th1 dominant as summarized by Rosenblum Lichtenstein et al. (2006). Because alveolar macrophages (AMs) from C57BL/6 mice have been shown to have a higher phagocytic capacity than AMs from BALB/c mice (Su et al., 2001), we hypothesized that C57BL/6J mice might clear spores more efficiently than BALB/c mice, leading to differences in integrated spore dose over time. This may contribute to the increased pulmonary inflammation seen in BALB/c as compared with C57BL/6J mice. We also hypothesized that differences in AM sensitivity to St. chartarum-induced cytotoxicity influence both inflammation and spore clearance.

**MATERIALS AND METHODS**

**Fungal strains and spore suspensions.** A trichothecene-producing strain of St. chartarum was obtained from Harriet Burge (Environmental Microbiology Lab, San Bruno, CA) (Rao et al., 2000) and grown on potato dextrose agar (PDA) plates at 15°C. Spores were vacuumed from the surface of 21-day cultures using a modified filter cassette with a 37-mm, 0.4-μm polycarbonate membrane filter (Poretics Corp., Livermore, CA). After the spores were removed from the filter, they were suspended in D-PBS (Mediatech Inc., Herndon, VA) at the desired concentration, as measured under light microscopy at ×200 in a hemocytometer chamber. Minor hyphal fragment content and negligible spore clumping were observed; most spores were ovoid in shape with a mean size of 6 × 9 μm. Two Stachybotrys chemotypes have been described—an atranone-producing chemotype and a trichothecenes-producing chemotype (Andersen et al., 2002, 2003; Rand et al., 2006). The St. chartarum strain described in this study is a trichothecene-producing chemotype.

**SG ELISA.** SG levels in the spore suspension were measured by a modified competitive direct ELISA (Chung et al., 2003a). Briefly, SG antibody was diluted in PBS (1:10,000), coated on microtiter plates (100 μl/well), and incubated overnight at 4°C. Plates were washed and blocked with 1% (w/v) bovine serum albumin (BSA) in PBS at 25°C for 1 h. SG standards (0–500 ng/ml) were made, and SG horseradish peroxidase conjugate was diluted (1:4000 [v/v]) in 1% (w/v) BSA in PBS. Equal volumes of diluted SG horseradish peroxidase conjugate (50 μl) and standards diluted similarly were then mixed and incubated in microtiter plates at 25°C for 1 h. Plates were washed, and bound peroxidase was determined by 100 μl/well incubation with 3,3',5,5'-tetramethylbenzidine (Neogen, Lansing, MI) at 25°C for 0.5 h. Color reaction was stopped with 2N sulfuric acid (100 μl/well), and plates were read at 450 nm wavelength on ELISA plate reader (Molecular Devices, Menlo Park, CA). SG concentrations in samples were determined from the standard curve using Softmax software (Molecular Devices).

**Dual radiolabeled spores.** Stachybotrys chartarum spores were streaked onto PDA plates supplemented with 1 ml of D-[U-14C] glucose (287 mCi/mmol, 200 μCi/ml; GE Healthcare Life Sciences, Piscataway, NJ) and 1 ml of [65-H] thymidine (23 Ci/mmol, 1 mCi/ml; GE Healthcare Life Sciences). Growth conditions and harvesting were as described above. The dual label enabled tracing of both spore DNA and carbohydrate components such as the cell wall and glycosylated proteins. Spores were washed with saline four times prior to use to remove any unincorporated radiolabel. After the fourth wash, no radiolabel was detectable in the wash fluid.

**Animal care.** Animal use protocols for these experiments were approved by the committee on animal experiments, and all institutional and federal guidelines on animal care and handling were followed. Male BALB/c (Taconic, Germantown, NY) and C57BL/6 (The Jackson Laboratory, Bar Harbor, ME) were housed at the Harvard Animal Facilities for a minimum 1-week acclimation period, fed Purina Mouse Chow and water ad libitum, and kept on a 12-h light-dark cycle. All mice in these experiments were age matched (7–10 weeks). All experiments described below were performed on both mouse strains concurrently.

**In Vivo spore clearance.** Spores suspensions were delivered into the lungs of both mouse strains by intratracheal instillation (IT) at 7.5 ± 1 weeks. Each mouse received 6.25 × 105 (a concentration of 107 spores/ml saline in 62.5 μl saline) spores per 25-g body weight for the radiolabeled clearance experiments. Each mouse received 6.25 × 105 (a concentration of 107 spores/ml saline in 62.5 μl saline) spores per 25-g body weight for the quantitative clearance experiments (visual counting) and 6.25 × 104 spores per mouse (not adjusted for body weight) for the radiolabeled clearance experiments. We used the same volume for all the animals in the experiments to standardize the administered radioactive dose from day to day.

Between six and twelve mice per day were administered spores. On each day, the order of the strains was randomly alternated. At least three animals for each strain and time were sacrificed, and organs were harvested. The amount of radioactivity in the delivered dose was quantified by delivering an equivalent quantity of spores to a vial (n = 4) for digestion (see below) and subsequent counting of radioactivity.

Immediately after the spore instillation and 24 h later, the mice were humanely killed by an intraperitoneal lethal dose of sodium pentobarbital (Anphoxy Products Co., Arcadia, CA) (10 mg/mouse). Mice were then exsanguinated by cutting the abdominal aorta and by puncturing the hemidiaphragm to create a bilateral pneumothorax. The lungs and trachea were removed en bloc, suspended in 4-ml distilled water, and homogenized...
through a stainless steel sieve with 100-µm holes. We counted intact spores on a hemocytometer under a light microscope at ×200 magnification. To calculate the number of spores in the lungs (S_L), we multiplied the total volume (V_L) of homogenized lungs (4000 µl) by the number of visible spores counted (S_H) in the hemocytometer grids then divided by the volume of one hemocytometer grid (V_H) (0.9 µl) multiplied by the total number (N_H) of grids counted (between 4 and 10 per mouse). We used the equation $S_L = \frac{S_H \times V_L}{N_H \times V_H}$.

For radioactivity measurements, mice were humanely killed as described above at 5 min, 8 h, and 1, 2, and 7 days after spore instillation. The lungs and trachea, stomach, cecum, and large intestine were removed and placed into preweighed scintillation vials. The small intestines were removed and cut into pieces and distributed among scintillation vials. Vials were reweighed and tissue weights calculated.

Tissues were dissolved in 1.0 ml Solvable (Perkin Elmer, Boston, MA) and 0.1 ml of 100 mM EDTA, followed by 0.3 ml 30% hydrogen peroxide. The caps were kept loose for 15 min to allow gas to escape. Caps were tightened, and the vials were incubated in a 60°C water bath for 1 h. The tubes were cooled to room temperature, and 15 ml of Ultima Gold (Perkin Elmer) was added. Samples were read in a Wallac Oy WinSpectral Scintillation Counter (Turku, Finland) using the 3H/14C dual label program. The radioactivity detected in the stomach, small intestine, large intestine, and cecum was totaled for each mouse to determine the total radioactivity in the GI tract.

**In Vitro spore cytotoxicity.** Four BALB/c and C57BL/6J mice were lavaged for each of the three data points, as described previously (Rosenblum Lichtenstein et al., 2006). We did a 1-ml wash, followed by eleven 0.75-ml washes with D-PBS. The bronchoalveolar lavage fluid was pooled, resulting in one tube for each mouse strain. Seven different experiments were performed to achieve a sample size of ≥3 per time point. BSA (30%) (Calbiochem, La Jolla, CA) was preadded to the ice-chilled Falcon tube at 30°C to achieve a sample size of one tube for each mouse strain. Seven different experiments were performed to achieve a sample size of seven experiments. Every data point shown is based on two to three replicates from each of seven experiments, including three dose-response and four time course preparations.

Spore suspensions were centrifuged at 200 g for 15 min and the supernatants discarded. The cells were resuspended in 1-ml red blood cell lysis solution (Metcalfe et al., 1986) and held at room temperature for 10 min. Supplemented RPMI-1640 was prepared with the following: 1% l-glutamine (Mediatech Inc.), 1% penicillin 100 U/ml and streptomycin 100 µg/ml (100X) (Sigma, St Louis, MO), and 10% fetal bovine serum, standard (HyClone, Logan, UT). Five microliters of supplemented RPMI-1640 was added to the cells which were then centrifuged at 200 g for 10 min. The supernatant was discarded, and the cells were resuspended in 2 ml of supplemented RPMI-1640. The cells were counted on a hemocytometer and diluted to 750,000 cells/ml. One hundred microliters of the cell suspension was pipetted into each well of a 96-well plate (Greiner bio-one µclear plates), and the strains were alternated from well to well. Plates were incubated at 37°C with 5% CO2 overnight to allow the AMs to attach. Washes in subsequent steps are intended to remove nonadherent cell populations (such as neutrophils) that we have previously shown to be less than 3% of the white cell population in lung lavage fluid in healthy mice (Rosenblum Lichtenstein et al., 2006). To three replicates of each pool were used for each spore dose and time point in each of seven experiments, including three dose-response and four time course experiments. Every data point shown is based on two to three replicates from three to seven different experiments for a total of 6–15 wells per data point.

Spore suspensions were prepared in supplemented RPMI-1640 for each experiment. The spores were diluted to 250,000 spores/ml using a hemocytometer. Spores were serially diluted 1:5 in supplemented RPMI-1640, resulting in concentrations of 5000, 1000, 200, and 40 spores per 75,000 macrophages for the dose-response experiments. A dose of 1000 spores per 75,000 macrophages was used for the time course protocols. In all cases, a negative control sample was prepared that contained media without spores.

After overnight recovery from plating, the media was removed and 100 µl of spore preparations or control media were added to each well. Plates were incubated at 37°C in air with 5% CO2 for 24 h for the dose response and for 2, 5, or 24 h for the time course experiments. At each designated time, the cells were washed twice with 300 µl of D-PBS with calcium and magnesium (Cellgro, Herndon, VA). A 20-ml solution with

![FIG. 1. SST per spore characterization. SG equivalents as measured by ELISA in (A) 10^7 Stachybotrys chartarum spores/ml and (B) full strength SST preparation used in Figure 6. Data are mean ± SD.](https://academic.oup.com/toxsci/article-abstract/116/1/113/1657991)
on a logarithmic scale were log transformed for statistical analyses. For the
visual counting of spores in lung homogenates, ANOVA was performed.
For the experiments measuring radiolabeled spore clearance from the lung, we
used a repeated measures analysis that accounts for both the \(^{3}H\) and \(^{14}C\) labels for
each mouse. For the experiments measuring radioactivity in the GI tract,
multivariate analysis of variance (MANOVA) was used to analyze the data
to account for the use of two different isotopes and our examining multiple
tissues. For \textit{in vitro} cytotoxicity experiments, we used a repeated measures
analysis that allows for residual correlation among observations derived from
the same BAL pool to account for differences among pools and spore
preparations. In the dose-response model, we utilized a broken stick analysis
to describe the change in the slopes of both lines at the 200 spores per well dose.
A broken stick analysis allows us to describe two of the expected linear regions
in a typical dose-response S-curve (Ruppert \textit{et al.}, 2003). In this instance, we
have a relatively flat region at doses too low to cause significant cell death and
and a steeper region where increasing dose causes an approximately linear increase
in cell death when the data are graphed on a log-log scale.

\section*{RESULTS}

\textit{Stachybotrys chartarum} Spore Suspension Contained SG

Because SG is water soluble, AMs and pulmonary epithelium would likely be exposed to soluble SG when
exposed to \textit{S. chartarum} suspensions. We therefore measured the SG concentration in this spore preparation. A 1-ml D-PBS
suspension with \(10^6\) spores had an SG concentration of 1.4 ug/ml or 1.4 pg/spore (Fig. 1), which is comparable with
previous estimates of 1 pg/spore for trichothecenes and \textit{S. chartarum} by Yike and Dearborn (2004).

\textbf{Mouse Strain Differences Influence Pulmonary Clearance of \textit{Stachybotrys chartarum} Spores}

C57BL/6J mice cleared \textit{S. chartarum} spores from the lungs significantly faster than BALB/c mice (Figs. 2 and 3).
Comparable spore numbers were delivered to the lungs of BALB/c and C57BL/6J mice. In C57BL/6J mice, 96.3 \pm 11.1% (mean \pm SE) were recovered, and in BALB/c mice, 81.4 \pm 9.9% were recovered (time 0, Fig. 2). After 1 day, C57BL/6J mice had one-third as many spores remaining in the lungs as the BALB/c mice (Fig. 2; \(p = 0.02\), ANOVA). Because \textit{S. chartarum} spores have limited germination potential in the lung (Pestka \textit{et al.}, 2008), the spore numbers in the lung homogenates are unlikely to reflect spore growth.

One limitation using this methodology is that fibrous material in the homogenates makes the spores difficult to
count accurately. In addition, we do not count spore components, only whole spores. We therefore estimated spore
retention in the lungs using radiolabeled spores. Both strains cleared a significant percentage of the instilled spores and spore
components (\(p < 0.001\)) over the 1-week time course. Consistent with the visual counts, the radiolabel data also
showed that C57BL/6J mice cleared spores or spore components from the lung more rapidly than BALB/c mice during
the first 24 h after instillation (\(p = 0.04\); Figs. 3A and B). (For
\(^{3}H\)-thymidine clearance, the best fit lines are \(\log(y_{C57BL/6J}) = 1.54 - 0.29 \log(\text{hours})\), \(\log(y_{BALB/c}) = 1.64 - 0.10 \log(\text{hours})\), \(\log(y_{C57BL/6J}^{14}C) = 1.14 - 0.88 \log(\text{hours})\), and
\(\log(y_{BALB/c}^{14}C) = 1.50 - 0.68 \log(\text{hours})\)).

\section*{Spores Are Cleared via the GI Tract}

The appearance of radioactivity in the GI tract after IT into the lungs demonstrates clearance via the mucociliary escalator.
Significantly, more spore-associated radioactivity was present in the GI tract of C57BL/6J mice 8 h after IT than BALB/c mice (\(p < 0.05\), MANOVA; Figs 4A and B). Both strains showed significantly more radioactivity in the GI tract at 8 h
than at other times, suggesting that this clearance pathway operates early for both strains (\(p = 0.0001\), MANOVA).
The C57BL/6J mice were thus more efficient at clearing the spores from the lungs via the mucociliary escalator than the
BALB/c mice.

\textit{Stachybotrys chartarum} Kills More AMs from C57BL/6J than BALB/c

C57BL/6J AMs died at lower spore doses (\(p = 0.007\)) and sooner (\(p = 0.04\)) than those from BALB/c mice (Figs. 5A
Although both strains exhibited significant cell death at both the 1000 and 5000 spore doses, significantly more C57BL/6J AMs died at spore doses as low as 1000 spores per well with 75,000 cells (p = 0.0001; repeated measures analysis). Many of the dead cells showed a punctate nuclear morphology in their staining with EthD1 consistent with apoptosis. At 24 h after spore addition, for doses less than 200 spores per 75,000 macrophages, the log(y) = 2.29 – 0.20 (log(dose)) and log(y) = 2.04 – 0.06 (log(dose)), where y represents the percent of live cells and dose represents the number of spores per well. For doses greater than 200 spores per 75,000 macrophages, the log(y) = 4.04 – 0.97 (log(dose)) and log(y) = 3.4731 – 0.68 (log(dose)), where y represents the percent of live cells and dose represents the number of spores per 75,000 macrophages. At doses higher than 200 spores/well, the slopes are significantly different (p = 0.007) between the two mouse strains. C57BL/6J AMs died more rapidly in response to S. chartarum than do BALB/c AMs. The equations describing their cell death after addition of 1000 spores are log(y) = 1.87 – 0.098 (time) for BALB/c and log(y) = 1.84 – 0.086 (time) for C57BL/6J mice. The slopes of these two lines are significantly different (p = 0.04). This strain difference is consistent with C57BL/6J mice producing significantly lower levels of inflammatory cytokines in vivo (Rosenblum Lichtenstein et al., 2006).

SST and SG Kill More AMs from C57BL/6J than BALB/c

Increasing doses of SST cause increased cytotoxicity in AMs from both mouse strains (p = 0.0001; Fig. 6A). This is also true for SG only (p = 0.0001), (Fig. 6B). However, we observed a range of low concentrations where little change is seen, but at higher concentrations there is a linear relationship when percent survival is plotted against concentration. Importantly, we show that SG, although having significant cytotoxicity, does not account for all the cytotoxicity of SST.
Thus, SG is a significant component, but there are also others. The concentration that is lethal for 25% of the cells (LC25) for SST for C57BL/6J is 0.017 \( \mu \text{g/ml} \) SG equivalents and for BALB/c is 0.052 \( \mu \text{g/ml} \) SG equivalents. The LC25 for SG for C57BL/6J is 0.075 \( \mu \text{g/ml} \) and for BALB/c is 0.243 \( \mu \text{g/ml} \). We used LC25 rather than LC50 because LC50 is outside the known linear range of our data for SG in BALB/c macrophages. Based on these numbers, SG accounts for 20–25% of the toxicity of the SST.

After 1 day of treatment with SST, more C57BL/6J AMs die than BALB/c AMs at doses between 0.01 and 0.5 \( \mu \text{g} \) SG equivalents/ml (\( p = 0.02 \)). The slopes between those doses are −0.41 and −0.23 for C57BL/6J and BALB/c, respectively (Fig. 6A). Similarly, after 1 day of treatment with SG, more C57BL/6J AMs die than BALB/c AMs at doses between 0.03 and 0.7 \( \mu \text{g} \) SG/ml (\( p = 0.059 \)). The slopes between those doses are −0.27 and −0.18 for C57BL/6J and BALB/c, respectively (Fig. 6B). The two mouse strains showed differences in sensitivity to both SST and SG, with SG accounting for 15–30% of the cytotoxicity of SST at the calculated LD50.

**DISCUSSION**

C57BL/6J mice cleared trichothecene-producing *S. chartarum* spores from their lungs at a faster rate than did BALB/c mice. When spores are delivered to the lungs of C57BL/6J mice, more of the spores were cleared via the mucociliary escalator and then swallowed and excreted through the GI tract than in BALB/c mice. Others may be degraded by macrophages within the lungs. This more efficient clearance in C57BL/6J mice via innate mechanisms may partially explain the diminished pulmonary injury and inflammation that have been reported in this strain (Rosenblum Lichtenstein et al., 2006). In contrast, spores or spore components in the lungs of BALB/c mice persisted longer and thus may have elicited a greater response. For example, 37 ± 1% of the carbohydrate component of the spores persisted in the lungs of BALB/c mice a week after instillation, whereas only 17 ± 5% remained in the lungs of C57BL/6J mice. This persistence might also explain how an extract of *S. chartarum* can also induce allergic asthma.
in BALB/c mice after repeated pulmonary exposures (Viana et al., 2002). The carbohydrate component of the labeled spores most likely includes mycotoxins and glycosylated proteins that may be causing some of the inflammatory responses. Some of the remaining 14C label may also reflect metabolites of spores, mycotoxins, or proteins. C57BL/6J mice may metabolize spores or their mycotoxins more rapidly, and this may in part mediate the increased clearance rate in the C57/BL6J mice. Whichever spore components the 14C label represents, the additional spore-derived material remaining in the BALB/c mouse lungs over time is most likely mediating the increased inflammation observed in those lungs.

We reported that strain differences in response to S. chartarum are distinct from the strain differences seen in response to lipopolysaccharide or to other fungi (Rosenblum Lichtenstein et al., 2006). C57BL/6J is the least responsive strain to S. chartarum and BALB/c is the most responsive, as indicated by both cellular and cytokine changes in lung lavage fluid (Rosenblum Lichtenstein et al., 2006). The nature of these strain differences seems to be unique to S. chartarum. Pulmonary exposure to Coccidioides immitis, C. neoformans, and Aspergillus fumigatus show that C57BL/6 mice are more sensitive to infection and inflammation and that BALB/c are more resistant (Cox et al., 1988; Hoag et al., 1995; Stephens-Romero et al., 2005; Zaragoza et al., 2007). Moreover, murine endotoxin susceptibility also shows a different pattern of strain responsiveness (Wells et al., 2003). Stachybotrys chartarum is the only fungus that we know of to which BALB/c mice are more sensitive than C57BL/6J mice.

It has been reported that C57BL/6J are generally poor at clearing other types of mold, such as Histoplasma capsulatum, B. anthracis, Coccidioides posadasii, and A. fumigatus, which in part explains their increased susceptibility to these moulds (Ehrchen et al., 2008; Lyons et al., 2004; Stephens-Romero et al., 2005; Wu-Hsieh, 1989). In contrast, we showed that C57BL/6J mice may use bulk transport out of the lungs via the mucociliary escalator as a first step toward efficient clearance of the spores and their constituents from the lungs. BALB/c mice, on the other hand, may retain more spores in the lungs, which more effectively induces cytokine release, cascades of cytokine effects, and infiltration of neutrophils.

C57BL/6J AMs also have significantly increased cell death compared with BALB/c. Increased AM death may lead to less cytokine release because dead cells cannot release inflammatory mediators. This results in less pulmonary injury and inflammation as measured in bronchoalveolar lavage parameters.

Similarly, macrophage apoptosis, which appears to be a contributing mechanism for cell death, has been reported to aid in microbial killing, to downregulate the inflammatory response, and even to aid in pulmonary clearance in pneumococcal and streptococcal models (Dockrell et al., 2003; Marriott and Dockrell, 2006; Marriott et al., 2006). If the greater rate of cell death in AMs from C57BL/6J mice is attributable to apoptosis, then the C57BL/6J mice might be expected to have less pulmonary inflammation and better clearance, a scenario compatible with our results. In addition, in a C. neoformans model, depletion of AMs was shown to be protective against infection (Kechichian et al., 2007).

These two mechanisms in which cell death leads to less inflammation may interact. For example, we showed that BALB/c mice have focal inflammation consisting primarily of AMs after administration of S. chartarum into the lungs (Rosenblum Lichtenstein et al., 2006). This focal inflammation may have blocked some of the mouse’s airways and might physically impede clearance of the spores that caused the initial inflammation.

SG accounts for 15–30% of the toxicity of SST. Other trichothecenes, proteins, and mycotoxins may account for the remainder of the cytotoxicity. In addition, various mycotoxins may act synergistically to account for the toxicity of SST. The significant dose response to SG and SST in AMs from both mouse strains suggests that SG accounts for a significant amount of the cytotoxicity of S. chartarum.

Genetic variation in the human population that leads to decreased rates of AM apoptosis in the presence of S. chartarum could lead to increased spore-induced cytokine release, increased inflammation, decreased spore clearance, and ultimately to a higher pulmonary spore exposure over time, exacerbating the effects of spore exposure. Small differences in rates of apoptosis could be magnified by downstream effects leading to greater differences in response to S. chartarum exposure. In addition, it has been hypothesized that exposure to cigarette smoke might make infants more susceptible to acute idiopathic pulmonary hemorrhage (Etzel et al., 1998). Interestingly, it has been reported that cigarette smoke exposure inhibits the natural activity of the mucociliary escalator (Cohen et al., 1979). Such an environmental cause of reduced clearance might further exacerbate pulmonary hemorrhage in an already susceptible infant.

Through this and previous work, we have demonstrated that different mouse strains respond differently to S. chartarum. We interpret the data as showing that both reduced clearance of and higher macrophage cytotoxicity from the bioactive components of S. chartarum in humans might cause similar differences in susceptibility among the human population. The results from this study are the first evidence that differences in spore clearance and macrophage susceptibility to spore-induced death may contribute to strain differences in susceptibility to S. chartarum seen in vivo. That these differences have a genetic basis is not unlikely.

A Centers for Disease Control (CDC) report concluded that S. chartarum was responsible for acute idiopathic pulmonary hemorrhage in a cluster of infants (Dearborn et al., 1999; Etzel et al., 1998), although there may have been other contributing factors such as environmental tobacco smoke or consequences of water damage including bacterial exposures (Dearborn et al., 1999). The original CDC report and a later case study suggested that African-American babies were more likely than Caucasian
babies to die of acute idiopathic pulmonary hemorrhage after exposure to Stachybotrys (Dearborn et al., 1999, 2002; Etzel et al., 1998). It is unclear if the observed ethnic differences were because of choice of controls or to economic or other environmental factors that are correlates of race, genetic differences, or a combination of both.

Genetic variability in human populations may account for some of the wide variation among individuals responding to mold exposure in contaminated occupational and domestic settings. In addition, other concomitant conditions, such as cigarette smoke exposure or bacterial infection, might further exacerbate poor spore clearance in susceptible populations. Future directions include determination of the genetic loci responsible for differential clearance of Stachybotrys and their interactions with environmental factors. It would be of further interest to test the effects of cigarette smoke or concomitant exposure to other pathogens on spore clearance.

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REFERENCES


