Teaching Point
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Adenovirus nephritis and obstructive uropathy in a renal transplant recipient: case report and literature review

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We report an unusual case of adenoviral nephritis in a 45-year-old woman who presented with fever, gross haematuria, acute kidney injury and obstructive uropathy 17 months following renal transplantation. Adenoviral nephritis was confirmed with immunohistochemistry. We identified 10 other published cases of adenoviral nephritis proven by immunohistochemistry. Obstructive uropathy has been reported only once before in a renal transplant recipient with adenoviral nephritis. Contrary to other reports, this case series shows that renal function may not always recover to baseline following the acute adenoviral disease. Adenoviral nephritis should be considered in the renal transplant patient with fever, haematuria, acute kidney injury and hydronephrosis in both the early and late post-transplant periods.

Keywords: adenovirus nephritis; obstructive uropathy; renal transplant

Introduction

Adenovirus infection in a renal transplant recipient was reported 35 years ago in an autopsy study which showed cytopathic changes of allograft tubular epithelial cells and diffuse interstitial pneumonitis [1]. The most common urologic manifestation of adenoviral infection is haemorrhagic cystitis [2,3], but disseminated disease with multi-organ failure and death has rarely been reported in renal transplant patients [4–7]. Over the last 25 years, there have been sporadic case reports of adenovirus-related renal allograft nephritis. In this report, we describe an unusual case of late onset adenoviral allograft nephritis with obstructive uropathy and we review other cases of adenoviral nephritis proven by biopsy immunohistochemistry.

Case report

A 45-year-old woman had a longstanding history of systemic lupus erythematosus, treated for years with glucocorticoids and intravenous cyclophosphamide. She developed end-stage renal disease in 1998. She underwent a deceased-donor renal transplant on April 2006. The adult donor was a six-antigen mismatch. An intraoperative kidney biopsy showed normal histology. She received thymoglobulin induction, followed by prednisone, tacrolimus and mycophenolic acid. Trimethoprim–sulfamethoxazole and valganciclovir were given for viral and bacterial prophylaxis. Her baseline serum creatinine was 1.0 mg/dl. She developed diabetes mellitus requiring an oral agent.

She was admitted to the hospital 17 months following transplant to evaluate 3 days of gross haematuria, abdominal pain and fevers. Her admission medications included prednisone 5 mg daily, tacrolimus 5 mg twice daily, mycophenolic acid 720 mg twice daily, clonidine 0.2 mg twice daily, glipizide 5 mg daily, valganciclovir 450 mg daily and trimethoprim–sulfamethoxazole daily. On admission, her temperature was 37.8 °C, blood pressure was 130/80 mHg and physical exam showed mild tenderness over the right lower quadrant of the allograft and suprapubic area.

Laboratory tests showed increased azotaemia with blood urea nitrogen 45 mg/dl and creatinine 2.3 mg/dl. Urine was grossly bloody and urinalysis showed numerous red blood cells with casts and 10–20 white blood cells per high power field. Urine bacterial culture and urine cytology examination were unremarkable. Transplant renal sonogram showed moderate hydronephrosis with hydroureter (Figure 1). Abdominal pelvic computerized tomography (CT) showed the hydronephrosis and a left ovarian mass consistent with a dermoid cyst; lymphadenopathy was absent. A percutaneous nephrostomy was placed on hospital Day 3, ciprofloxacin was given prophylactically and mycophenolic acid was discontinued. Serum creatinine decreased to 1.4 mg/dl. Cystoscopy performed on hospital Day 5 showed only a small discrete reddish granular area identified near the neo-orifice of the transplant ureter. A bladder biopsy showed normal uroepithelium with no inflammation or pathologic le-
Procurement. Three days later, the percutaneous nephrostomy was removed. The patient had daily fevers to 39 °C. The tacrolimus dose was adjusted to maintain a 12-h trough level of 4–7 ng/ml. Blood and urine cultures, lupus serologies, PCR assays of the blood for adenovirus, Epstein–Barr virus, cytomegalovirus, BK virus (BKV) and parvovirus B19 were negative. Chest X-ray was unremarkable. She was initially reluctant to consent for a renal biopsy. Neutropenia developed, requiring filgrastim therapy. Subsequently, the serum creatinine increased to 2.4 mg/dl. A repeat sonogram showed resolution of the hydronephrosis.

The patient agreed to a renal allograft biopsy, which was performed on hospital Day 6. The biopsy showed necrotizing interstitial nephritis with tubular necrosis, a single granuloma and an inflammatory infiltrate largely comprised of dense plasmacytoid lymphocytes (Figure 2) which had sufficient cytologic atypia to initiate a work up for post-transplant lymphoproliferative disorder, which was ruled out by negative Epstein–Barr encoded small RNA in situ hybridization. No microorganisms were identified on Giemsa, periodic acid-Schiff and acid-fast bacillus stains. Immunoperoxidase stains for SV 40 large T antigen and C4d were negative.

Meanwhile, repeat CT scan studies of the chest, abdomen and pelvis were unremarkable, a positron emission tomography scan was unremarkable and a bone marrow aspirate and biopsy showed hypocellular marrow with no

**Fig. 1.** Renal transplant ultrasound. The renal ultrasound image shows the renal allograft in the right lower quadrant of the abdomen. The allograft measures 12.7 cm in the long axis and demonstrates moderate hydronephrosis.

**Fig. 2.** Localization of adenoviral antigen in the renal allograft. (A) Renal allograft biopsy revealed interstitial nephritis, with numerous mildly atypical lymphocytes, focal necrosis and haemorrhage. Arrow indicates a tubular epithelial cell nucleus with apparent viral cytopathic changes. Haematoxylin and eosin, original magnification ×400. (B and C) Immunostaining for adenovirus antigens labelled several tubular cell nuclei and, more faintly, adjacent cytoplasm (dark brown diaminobenzidine product; haematoxylin counterstain; original magnification ×400).
Table 1. Adenoviral interstitial nephritis of renal allografts, proven by immunohistochemistry

<table>
<thead>
<tr>
<th>Case</th>
<th>Reference</th>
<th>Age</th>
<th>Sex</th>
<th>Immunosuppression</th>
<th>Disease onset after transplant (months)</th>
<th>Presentation</th>
<th>Peak serum creatinine (mg/dl)</th>
<th>Adenoviral detection</th>
<th>Interstitial nephritis</th>
<th>Follow-up</th>
<th>Serum creatinine (mg/dl)</th>
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Presented are 11 published cases of adenoviral interstitial nephritis, all confirmed by immunohistochemistry. Abbreviations include the following: NR, not reported; ND, not determined; ATG, anti-thymocyte globulin; Aza, azathioprine; BCM, basiliximab; CsA, cyclosporine; DCM, daclizumab; My, mycophenolic acid; OKT3, muromonab; Pr, prednisone; Tac, tacrolimus.
tumour or granulomas. Anti-human leukocyte antigen antibody levels were negative. The oral prednisone dose was increased to 15 mg daily. In the following days, the white cell count returned to normal, serum creatinine decreased slightly to 2.0 mg/dl and fevers abated. The patient was sent home 3 weeks after hospital admission, on prednisone 15 mg daily and tacrolimus 2 mg daily. Mycophenolic acid was not restarted. One week after discharge, a urine culture became positive for adenovirus.

Two months after discharge, creatinine had decreased to 1.7 mg/dl and microscopic haematuria had resolved. Prednisone was reduced to 10 mg daily. A repeat blood adenoviral PCR test was negative. Immunosuppressive therapy was targeted to a lower tacrolimus level 4–6 ng/ml and a slow taper of prednisone, over 3 months, to a target of 5 mg daily.

Subsequently, immunostaining for adenovirus antigens was performed, using antibody directed against adenovirus Type 3 protein antigens (Chemicon-Millipore, Billerica, MA) and lung tissue obtained from a patient who died with adenoviral pneumonia as a positive control tissue.

Adenoviral antigens were detected within tubular epithelial cells (Figure 2). Two years later, the patient was doing well, with a stable serum creatinine of 1.2 mg/dl and normal urinalysis.

Discussion

This case illustrates the difficulty of establishing the diagnosis of adenoviral nephritis in the renal allograft. Atypical features of our patient included late onset of the disease (17 months after transplant) and obstructive uropathy.

A search of the literature revealed 10 other cases of renal transplant-related adenoviral nephritis biopsy proven by immunostaining, all reported since 1998 (Table 1) [4,6,8–15]. Two were children and the others were adults. Seven patients were male and four were female. Most of the cases (eight out of 11) presented within 8 months of the transplant, at a mean of 2.9 months with a range of 1–8 months. Three cases presented later, at 17, 18 and 144 months, respectively. Most of the patients initially received induction immunosuppression and maintenance therapy with glucocorticoids, calcineurin inhibitors and mycophenolic acid. Patients commonly presented with gross haematuria and dysuria (10 out of 11), fever (nine out of 11) and acute renal failure (nine out of 11). Three patients required acute haemodialysis, but one patient failed to regain allograft function and returned to maintenance dialysis 6 months after transplant. As for the other patients, only limited follow-up was available, ranging from 4 to 24 months. Although most patients returned to baseline renal function, two reported patients had significant renal impairment at follow-up with serum creatinine values of 2.0 and 2.4 mg/dl, respectively. The present case series shows that a significant minority (three out of 11, 27%) of patients were left with significant renal impairment following adenoviral nephritis, in contrast to previous reports [3,6].

Renal biopsy with specific immunohistochemical staining for adenovirus is required for definite diagnosis of adenoviral interstitial nephritis [2,7,16], but this assay is not readily available at most institutions. Traditionally, a diagnosis of adenoviral infection was made with a positive viral culture from urine, blood or tissue [2,16]. Recent applications of quantitative real-time PCR assays have improved sensitivity over traditional methods. However, the sensitivity, specificity and predictive value of PCR tests have yet to be established [2,16]. As shown in Table 1, only three out of 10 cases were blood PCR positive and seven out of 10 were urine PCR positive at the time of the diagnosis. In our patient, repeated tests of blood adenovirus PCR were negative.

Surveillance studies of asymptomatic adult renal transplant patients have shown an incidence of adenoviral viraemia by PCR testing of 6.5% and viruria by culture of 11% [2,16]. Asymptomatic viral shedding in the urine makes urinary cultures unreliable in the absence of signs and symptoms of disease activity. At present, there is no consensus regarding treatment of asymptomatic patients with detectable adenoviraemia. Ison concluded that screening of asymptomatic adult transplant solid organ recipient for adenovirus is not useful because progression to disease is infrequent [16].

Characteristic histologic changes of adenoviral interstitial nephritis are listed in Table 1. In the present series, nine out of 11 cases had granulomatous changes and eight out of 11 had necrotizing inflammation, while one case had interstitial nephritis only, without necrotizing changes or granulomas. It may be useful to compare histologic changes seen in adenoviral nephritis with the much more common BKV nephritis. Intranuclear ‘smudge’-type viral inclusion bodies are typical of both BKV and adenoviral infection. Interstitial granulomatous changes have been reported in both BKV and adenoviral nephritis but are more common in adenoviral [7]. In a similar fashion, the necrotizing changes seen in adenoviral infection generally distinguish adenoviral nephritis from BKV nephropathy. Another difference between the two may be the lack of chronic changes with adenoviral nephritis, while BKV nephropathy is commonly associated with tubular atrophy and interstitial fibrosis. This may be related to the more indolent course and delay in diagnosis in BKV disease. Electron microscopy in adenoviral infection shows hexagonal virus particles of 70–80 nm in diameter, closely packed in a crystalline array, whereas BKV has an icosahedral shape with 40–44 nm in diameter [2,7,16,17]. Specific immunohistochemical stains for adenovirus and BKV most easily establish the correct diagnosis. There has been only one case report of concurrent adenovirus and BKV interstitial nephritis in a renal transplant patient [7].

Renal manifestations of adenoviral infection are shown in Table 2. Most common is acute haemorrhagic cystitis [2,3,7]. Acute interstitial nephritis with or without acute kidney injury syndrome is the second most common presenta-
Adenoviral nephritis should be considered in the renal transplant recipient with fever, haematuria, acute kidney injury and obstructive uropathy.

(2) Obstructive uropathy tends to resolve with the resolution of interstitial nephritis.

(3) Granulomas and necrotizing interstitial changes are characteristic of adenoviral nephritis and immunohistochemical demonstration of adenovirus in biopsy tissue should be the standard for diagnosis and treatment guidance.

(4) Reduction of immunosuppression is the most important therapeutic intervention, although more severe cases may benefit from additional antiviral therapy.

(5) Contrary to other case reports, renal function does not always recover to baseline following adenoviral nephritis.

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References


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