Clinical Report

A severe phenotype of Gitelman syndrome with increased prostaglandin excretion and favorable response to indomethacin

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Abstract
Our understanding of Gitelman syndrome (GS) and Bartter syndrome has continued to evolve with the use of genetic testing to more precisely define the tubular defects responsible. GS is caused by mutations in the SLC12A3 gene encoding the Na⁺ − Cl⁻ co-transporter of the distal convoluted tubule (DCT) [2]. To date, >240 SLC12A3 mutations have been identified in GS [3, 4]. As DCT-mediated salt reabsorption accounts for only ∼5% of the filtered sodium load, GS has normally been described as having a mild salt-wasting phenotype that is often not detected until adolescence or early adulthood [3, 5, 6].

Background
Gitelman syndrome (GS) is an autosomal recessive salt-losing tubulopathy (OMIM 263800) characterized by hypokalemic metabolic alkalosis, hypomagnesemia and hypocalciuria [1]. GS is caused by mutations in the SLC12A3 gene encoding the Na⁺ − Cl⁻ co-transporter (NCCT) of the distal convoluted tubule (DCT) [2]. Type, >240 SLC12A3 mutations have been identified in GS [3, 4]. As DCT-mediated salt reabsorption accounts for only ∼5% of the filtered sodium load, GS has normally been described as having a mild salt-wasting phenotype that is often not detected until adolescence or early adulthood [3, 5, 6].

Bartter syndrome (BS) is a heterogeneous autosomal recessive salt-wasting condition that tends to present earlier in childhood with a more severe phenotype including significant salt wasting, polyuria and failure to thrive [7]. Mutations affecting the Na⁺ 2Cl⁻ K⁺ co-transporter (NKCC2; OMIM 601678) or the renal outer medullary potassium channel (ROMK; OMIM 241200) in the thick ascending limb (TAL) often present antenatally with polyhydramnios, prematurity and severe salt wasting requiring significant electrolyte supplementation [8, 9]. This subtype of BS is sometimes referred to as antenatal Bartter syndrome or ‘hyperprostaglandin E syndrome’ following the description of elevated prostaglandin E2 (PGE2) levels in such cases [10]. It was this discovery that led to the successful treatment of these patients with the cyclooxygenase inhibitor indomethacin, which remains an important part of therapy along with salt and water supplementation [11].

There are several other subtypes of BS with distinct phenotypes. Type III BS results from mutations that affect the basolateral chloride channel (ClC-Kb) in the DCT and TAL (OMIM 607364) [12]. Type IVa BS with sensorineural deafness is the result of mutations in the Barttin subunit of the ClC-Ka and ClC-Kb channels, which are expressed in the TAL and inner ear (OMIM 602522) [13]. Digenic mutations of both the ClC-Ka and ClC-Kb channels have also been described causing BS with sensorineural deafness (Type IVb BS; OMIM 613090) [14]. Type V BS results from mutations leading to upregulation of the calcium sensing receptor (CaSR) and therefore hypocalcemia and hypercalciuria in addition to the typical salt-losing phenotype (OMIM 601198) [15, 16]. Most recently, the combination of epilepsy, ataxia, sensorineural deafness and salt wasting tubulopathy similar to GS has been associated with mutations of the inward-rectifying potassium channel Kir 4.1 (EAST syndrome; OMIM 612780) [17]. We describe two female siblings presenting in infancy with hypokalemic metabolic alkalosis, severe failure to thrive, polyuria and increased PGE2 excretion. Both siblings revealed a dramatic clinical response to indomethacin including improvements in growth and polyuria. These features were consistent with a diagnosis of BS. However, genetic studies later confirmed a diagnosis of GS with the identification of compound heterozygous mutations in SLC12A3.

Keywords: child; Gitelman syndrome; indomethacin; infant; prostaglandin
Case report

The first case is a Caucasian female born at term weighing 3.2 kg (appropriate for gestational age). No polyhydramnios was noted antenatally. This was the mother’s first child with no history of spontaneous losses, consanguinity or family history of renal disease. She first came to medical attention at the age of 5 months for failure to thrive with a weight of 4.8 kg and length of 59 cm (height SDS −1.2, weight SDS −2.1; Figure 1). She was solely breast fed and thus supplementary formula feeds were introduced over the next month, but her weight fell further to 4.7 kg. At this time, she was reported to feed hourly with frequent vomiting, but no diarrhea. A history of polyuria was noted by the parents with frequent changing of wet diapers. Gross motor delay was also noted as she had only just begun to roll and had significant head lag. The treating pediatrician arranged for admission to the local hospital for further investigations and management.

On admission, her chemistry profile demonstrated severe hyponatremia (Na+ 124 mmol/L) and a hypokalemia (K+ 2.1 mmol/L), hypochloremic (Cl− 72 mmol/L) metabolic alkalosis (pH 7.59, HCO3− >45 mmol/L). Serum magnesium was low (0.57 mmol/L), Blood urea nitrogen (BUN) and creatinine were within normal limits at 2.8 mmol/L and 29 μmol/L, respectively. Urinalysis was benign with no hematuria, proteinuria or pyuria. Urine specific gravity (SG) was <1.005 suggesting a urinary concentrating defect. Her UCa:Cr ratio, which was initially elevated, has been in the hypocalciuric range (<0.05 mmol:mmol) since the age of 2 years. Interestingly, the UCa:Cr ratio was elevated at 4 months of age which demonstrated mild hyponatremia (132 mmol/L) and metabolic alkalosis (pH 7.64, HCO3− 30 mmol/L). Serum potassium (4.1 mmol/L) and magnesium (0.83 mmol/L) were normal. The BUN was mildly elevated at 5.3 mmol/L, consistent with volume depletion, and creatinine normal at 20 μmol/L.

Her first UCa:Cr ratio at 4 months of age was in the normal range (0.74 mmol:mmol) with a normal serum calcium (2.72 mmol/L).

Sodium chloride supplementation (1 mmol/kg/day) was initiated at 1 week. She did not require potassium chloride supplementation (1 mmol/kg/day) until 5 months of age when she became persistently hypokalemic (K+ 2.9 mmol/L). Indomethacin (2 mg/kg/day) was commenced at 1 year of age given for failure to thrive (height SDS −3.7, weight SDS −3.9). Her growth response to indomethacin was appreciable, but less marked than that of her sister. However, her growth has improved with a gradual increase in the dose to 3 mg/kg/day from 2 years of age (current height SDS −1.96, weight SDS −1.11; Figure 1). Magnesium glucoheptonate supplementation was also started at 2 years of age (3 mg of elemental Mg++/kg/day) with the evolution of hypomagnesemia (serum Mg++ 0.66 mmol/L). She has required multiple admissions to hospital for exaggerated electrolyte imbalances including severe hypokalemia (K+ <2.5 mmol/L) in association with routine childhood infections. Her UCa:Cr is now persistently in the hypocalciuric range (<0.05 mmol:mmol).

Her current treatment regime at 5 years of age includes potassium chloride (3.8 mmol/kg/day divided bid), magnesium glucoheptonate (4.7 mg/kg/day of elemental Mg++ divided tid), indomethacin (3.8 mg/kg/day divided bid) and amiloride (0.8 mg/kg once daily). She maintains a high salt diet similar to her sister. Electrolytes and blood gas have been stable on this regimen (current Na+ 137 mmol/L, K+ 2.72 mmol/L). Of note her serum potassium has improved since the recent addition of amiloride. The most recent serum creatinine was normal at 41 μmol/L (eGFR 94 ml/min/1.73 m²).

Genetic testing

Genetic testing identified bi-allelic SLC12A3 mutations in both siblings: c.473G>A encoding p.R158Q and c.631_642del (p.R211_E214del) (Figure 2). The c.473G>A mutation has been previously described in a child with GS[18] while the c.631_642del mutation has not been previously reported. The SLC12A1, KCNJ1, CLCNKA, CLCNKB, BSND genes were also screened with no mutations identified. The c.473G>A mutation was found to be maternal in origin and c.631_642del paternal. Both parents have normal serum electrolytes. The father is of normal height (180 cm, SDS 0.44); however, the mother is short (150 cm, SDS −2.05).

Following the molecular analysis, both sisters were electively admitted to hospital to determine prostaglandin (PGE2) excretion while off indomethacin therapy. We felt that this admission would clarify the need for long-term indomethacin therapy in both patients, given this therapy.
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Fig. 1. Growth profile of cases. (a) Case 1 (birth—2 years); arrow indicates start of indomethacin. (b) Case 1 (2 years—present). (c) Case 2 (birth—2 years); arrow indicates start of indomethacin. (d) Case 2 (2 years—present); arrow indicates increase in dose of indomethacin.

Fig. 2. SLC12A3 gene displaying known exons with mutations identified in the cases marked.
Further investigations

Indomethacin was held for 48 h prior to admission (washout period), but regular supplements were continued in both siblings. Upon admission, initial bloodwork revealed severe hypokalemia (2.5 mmol/L in both) with metabolic alkalosis (Case 1: HCO₃⁻ 32 mmol/L, Case 2: HCO₃⁻ 33 mmol/L). Both siblings were polyuric upon presentation (Case 1: 5.5 mL/kg/h, Case 2: 8.8 mL/kg/h). Twenty-four-hour urine prostaglandin E₂ (PGE₂) excretion was significantly elevated at 2059 ng/day/1.73 m² in Case 1 and 3609 ng/day/1.73 m² in Case 2 (normal range 48-394 ng/24 h/1.73 m²) [10]. After a bolus of normal saline (20 mL/kg) and following 48 h of intravenous fluids (maintenance rate), urine PGE₂ excretion decreased to normal ranges in both siblings (Case 1: 374 ng/24 h/1.73 m², Case 2: 312 ng/24 h/1.73 m² in Case 2). Based on these results, a decision was made to reintroduce indomethacin at their previous doses. It was thought unlikely that the siblings would be able to consume sufficient oral fluids and salt supplementation to replicate the effects of the intravenous volume expansion achieved in hospital. The polyuria and electrolyte/acid–base derangements again improved within 1 week of restarting therapy.

Discussion

The classic description of GS is of a benign salt-wasting condition most often diagnosed in adulthood [1, 3]. However, in more recent literature, GS appears to be responsible for a wider spectrum of disease than initially appreciated [4, 19, 20]. A limited number of reports have emerged describing children affected as early as the neonatal period [21, 22]. Also mentioned within larger cohort descriptions of GS are cases presenting at <2 years of age with symptoms including growth delay, hypotonia and muscular spasms from severe hypomagnesemia [4, 19, 20]. Previous case series have suggested that males may be more severely affected than females [23].

Aside from the severity of our described GS cases, there are some interesting biochemical findings worthy of discussion, in particular the demonstration of raised urinary PGE₂ excretion in both siblings. Increased PGE₂ is typically associated with NKCC2 or ROMK channel mutations resulting in antenatal BS where patients present in the neonatal period with a history of polyhydramnios, prematurity and severe salt wasting. In contrast, patients with GS tend to have normal PGE₂ levels as described by Lüthy et al. [6]. It has been thought that this discrepancy in PGE₂ levels explains why children with BS normally require non-steroidal anti-inflammatory drug therapy in addition to salt and fluid supplementation whereas children with GS do not [6, 20]. Nonetheless, improved growth in response to indomethacin therapy in GS has been described in the literature [21, 24]. Our demonstration of raised urine PGE₂ level confirms that indomethacin is a rational therapy in patients with more severe salt-wasting phenotypes associated with SLC12A3 mutations.

Another feature in both patients was the initial absence of hypocalciuria, which has previously been considered a discriminating feature of GS [6, 20, 25]. More recent case series of patients with SLC12A3 mutations have demonstrated variability in urine calcium excretion [4, 23]. Interestingly, Case 1 demonstrated hypercalciuria upon presentation and Case 2 had normal urinary calcium excretion. To our knowledge, hypercalciuria has never been reported in GS. Both cases developed hypocalciuria with time. Thus, we confirm that urinary calcium excretion cannot be relied upon as a diagnostic feature in GS and that some patients with initially high or normal urinary calcium excretion will eventually develop hypocalciuria.

We would like to acknowledge some limitations in the genetic analysis of the described cases. We were unable to perform quantitative or semi-quantitative techniques such as multiplex ligation probe amplification screening. As a result, it is possible that a large deletion, duplication or rearrangement could have been missed [4]. Also, we did not screen for mutations in the CaSR given the absence of hypocalcemia and development of hypocalciuria.

In conclusion, given the range of phenotypic expression in the salt-losing tubulopathies, genetic testing is the only method capable of confirming the precise nature of the underlying tubular defect. Previously proposed discriminating features such as urine calcium excretion, hypomagnesemia and age of presentation have all been shown to be variable in GS and may evolve with time as illustrated by our cases. Despite this, we would suggest that the identification of SLC12A3 mutations should not preclude pediatric patients with a more severe salt-wasting phenotype from a trial of indomethacin.

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References


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