Role of Translocator Protein Density, a Marker of Neuroinflammation, in the Brain During Major Depressive Episodes

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IMPORTANCE The neuroinflammatory hypothesis of major depressive disorder is supported by several main findings. First, in humans and animals, activation of the immune system causes sickness behaviors that present during a major depressive episode (MDE), such as low mood, anhedonia, anorexia, and weight loss. Second, peripheral markers of inflammation are frequently reported in major depressive disorder. Third, neuroinflammatory illnesses are associated with high rates of MDEs. However, a fundamental limitation of the neuroinflammatory hypothesis is a paucity of evidence of brain inflammation during MDE. Translocator protein density measured by distribution volume (TSPO V₅) is increased in activated microglia, an important aspect of neuroinflammation.

OBJECTIVE To determine whether TSPO V₅ is elevated in the prefrontal cortex, anterior cingulate cortex (ACC), and insula in patients with MDE secondary to major depressive disorder.

DESIGN, SETTING, AND PARTICIPANTS Case-control study in a tertiary care psychiatric hospital from May 1, 2010, through February 1, 2014. Twenty patients with MDE secondary to major depressive disorder and 20 healthy control participants underwent positron emission tomography with fluorine F 18-labeled N-(2-(2-fluoroethoxy)benzyl)-N-(4-phenoxy pyridin-3-yl)acetamide ([18F]FEPPA). Patients with MDE were medication free for at least 6 weeks. All participants were otherwise healthy and nonsmokers.

MAIN OUTCOMES AND MEASURES Values of TSPO V₅ in the prefrontal cortex, ACC, and insula.

RESULTS In MDE, TSPO V₅ was significantly elevated in all brain regions examined (multivariate analysis of variance, $F_{15,23} = 4.5$ [P = .001]). The magnitude of TSPO V₅ elevation was 26% in the prefrontal cortex (mean [SD] TSPO V₅, 12.5 [3.6] in patients with MDE and 10.0 [2.4] in controls), 32% in the ACC (mean [SD] TSPO V₅, 12.3 [3.5] in patients with MDE and 9.3 [2.2] in controls), and 33% in the insula (mean [SD] TSPO V₅, 12.9 [3.7] in patients with MDE and 9.7 [2.3] in controls). In MDE, greater TSPO V₅ in the ACC correlated with greater depression severity ($r = 0.63$ [P = .005]).

CONCLUSIONS AND RELEVANCE This finding provides the most compelling evidence to date of brain inflammation, and more specifically microglial activation, in MDE. This finding is important for improving treatment because it implies that therapeutics that reduce microglial activation should be promising for MDE. The correlation between higher ACC TSPO V₅ and the severity of MDE is consistent with the concept that neuroinflammation in specific regions may contribute to sickness behaviors that overlap with the symptoms of MDE.
Mood in humans, and direct induction of the central immune system in rodents is associated with the sickness syndrome of anhedonia, weight loss, and anorexia, which overlap with the diagnostic criteria for MDE. Also in MDD, several markers of peripheral inflammation, including levels of C-reactive protein, interleukin 6 (IL-6), and tumor necrosis factor (TNF), are frequently increased. Conditions that create neuroinflammation, such as traumatic brain injury, systemic lupus erythematosus, and multiple sclerosis, are associated with prevalence rates of MDE as high as 50%, suggesting a link between brain inflammation and mood symptoms.

Whether brain inflammation occurs during a current MDE remains unclear because most postmortem investigations of neuroinflammation sampled patients with MDD and a history of MDE or individuals who committed suicide (suicide completers) with varied diagnoses. Within such studies, the samples of patients with current MDE were small. Van Otterloo et al reported no difference in the density of activated microglia in the white matter of the orbitofrontal region in 10 patients with MDD. Dean et al sampled 10 patients with MDD and found significantly increased levels of the transmembrane form of TNF in the dorsolateral prefrontal cortex (PFC) but no difference in levels of this form of TNF in the anterior cingulate cortex (ACC) and no difference in the soluble form of TNF in either region. Steiner et al reported increased density of quinolinic acid–positive cells, a marker influenced by microglial activation, in the ACC of 7 patients with MDE. Microarray studies have had mixed results, with a positive finding by Shelton et al of increased proinflammatory and anti-inflammatory cytokine messenger RNA in Brodmann area 10 in 14 patients with MDD. In contrast, several other microarray studies, most of which sampled adjacent regions of the PFC, did not identify this result. Among investigations in suicide completers, a study reported greater HLA-DR staining, a marker of microglial activation, in the dorsolateral PFC and ACC, and a second study reported greater levels of IL-6, TNF, and IL-1β in Brodmann area 10. Neither study of suicide found a relationship to MDD (or MDE), but fewer than 10 patients with MDD were included in each study. The mixed results among postmortem investigations in MDD have been attributed to issues of variation in brain regions sampled, inclusion of patients with early- and late-onset MDD, comorbidity of other psychiatric disorders and addiction, and, with the exception of the microarray studies, small sample size, although lack of focus on sampling the state of MDE may be important for investigations of neuroinflammation.

To determine whether neuroinflammation occurs in MDE secondary to MDD, positron emission tomography (PET) may be applied to measure translocator protein (TSPO) binding in vivo. Translocator protein is an 18-kDa protein located on the outer mitochondrial membranes in microglia, and increased expression of TSPO occurs when microglia are activated during neuroinflammation. Recently, a new generation of PET radiotracers was developed with superior quantification of TSPO binding. Among these, fluorine F 18–labeled N-(2-(2-fluoroethoxy)benzyl)-N-(4-phenoxypyridin-3-yl)acetamide (18F)FEPPA has excellent properties, including a high selective affinity for TSPO, increased binding during induced neuroinflammation, and a high ratio of specific binding relative to free and nonspecific binding.

To date, one neuroimaging study applied carbon 11–labeled N-(2-methoxybenzyl)-N-(4-phenoxypyridin-3-yl)acetamide ([11C]PBR28) PET to investigate TSPO levels in MDD, with negative findings. This earlier study assessed whether TSPO levels were elevated in a sample of 10 patients with MDD who underwent scanning once under a variety of states (treated, untreated, symptomatic, or partially symptomatic); hence, results of that study cannot be considered definitive for determining whether the level of TSPO binding is elevated in MDE. Scores on the Montgomery-Åsberg Depression Rating Scale on the day of PET scanning ranged from 5 to 30, indicating that the severity ranged from almost asymptomatic to moderately symptomatic. Other issues limit interpretation of that study, including potential bias of ongoing antidepressant use, heterogeneity of combined sampling of early- and late-onset MDD, and incomplete information regarding a TSPO polymorphism (rs6971) known to influence binding of the new generation of TSPO PET radioligands, including [18F]FEPPA.

In the present study, 18F FEPPA PET was applied to measure TSPO total distribution volume (V_T), an index of TSPO density, during MDE in patients with MDD compared with healthy, age-matched control participants. The main hypothesis was that TSPO V_T would be elevated in MDE in the PFC, ACC, and insula. The PFC and ACC were chosen because of their role in mood regulation circuitry and affect dysregulation in MDD. The insula is a strong candidate for mediating some of the sickness behaviors in MDD because it is activated in response to an immune challenge and may participate in homeostatic regulation and interoceptive signaling in MDD. The second hypothesis was that greater severity of symptom measures related to the sickness syndrome would be associated with greater elevation of TSPO V_T in these regions.

Methods

All participants provided written informed consent after all procedures were fully explained. The protocol and informed consent forms were approved by the Research Ethics Board of the Center for Addiction and Mental Health, Toronto, Ontario, Canada.

Participants

Twenty patients with a current MDE secondary to MDD (hereinafter termed patients with MDE) and 20 age-matched healthy controls completed the study. Participants were recruited from the Toronto-area community and a tertiary care psychiatric hospital (Centre for Addiction and Mental Health) from May 1, 2010,
Translocator Protein Density in MDE

Table 1. Demographic Characteristics of Study Participants

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Patients With MDE (n = 20)</th>
<th>Healthy Controls (n = 20)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Female sex, No. (%)</td>
<td>12 (60)</td>
<td>11 (55)</td>
</tr>
<tr>
<td>Age, mean (SD), y</td>
<td>34.0 (11.3)</td>
<td>33.6 (12.8)</td>
</tr>
<tr>
<td>TSPO genotype, No. of participants*</td>
<td>15</td>
<td>14</td>
</tr>
<tr>
<td>HAB</td>
<td>5</td>
<td>6</td>
</tr>
<tr>
<td>BMI, mean (SD)</td>
<td>23.4 (5.4)</td>
<td>24.8 (2.9)</td>
</tr>
<tr>
<td>17-Item HDRS score, mean (SD)*</td>
<td>20.0 (3.8)</td>
<td>NA</td>
</tr>
<tr>
<td>Age at first MDE, mean (SD), y</td>
<td>15.7 (5.2)</td>
<td>NA</td>
</tr>
<tr>
<td>Previous MDE, No. (%)</td>
<td>6 (3)</td>
<td>NA</td>
</tr>
<tr>
<td>Previous antidepressant trial, No. (%)</td>
<td>9 (45)</td>
<td>NA</td>
</tr>
<tr>
<td>No previous antidepressant trial, No. (%)</td>
<td>11 (55)</td>
<td>NA</td>
</tr>
</tbody>
</table>

Abbreviations: BMI, body mass index (calculated as weight in kilograms divided by height in meters squared); HAB, high-affinity binding; HDRS, Hamilton Depression Rating Scale; MAB, mixed-affinity binding; MDE, major depressive episode; NA, not applicable; TSPO, translocator protein.

\* Indicates single-nucleotide polymorphism rs6971 of the TSPO gene known to influence fluorine F 18-labeled N-(2-(2-fluoroethoxy)benzyl)-N-(4-phenoxypyridin-3-yl)acetamide binding.

Scores were derived on the day of scanning, with data missing for 1 patient with MDE.

Addition through February 1, 2014. All participants ranged in age from 18 to 72 years and were nonsmokers in good physical health. None of the participants had a history of autoimmune disease, and all were free of illness for at least 2 weeks. Patients with MDE had early-onset MDD (first MDE prior to age 45 years). Health or MDE was confirmed using the Structured Clinical Interview for DSM-IV (Table 1). Healthy controls were age matched within 4 years to the patients with MDE. Exclusion criteria for all participants included pregnancy; the use of any herb, drug, or medication within the past 6 weeks, except for oral contraceptives; and any history of neurologic illness or injury. All participants underwent urine drug screening, and women received a urine pregnancy test on the PET scanning day.

Patients with MDE were administered the 17-item Hamilton Depression Rating Scale (17-item HDRS) on enrollment and on the PET scanning day. For enrollment, a minimum score of 17 on the 17-item HDRS was required. All patients with MDE were medication free for at least 6 weeks before the PET scan (9 patients had completed ≥1 previous antidepressant trial). Other exclusion criteria consisted of concurrent active Axis I disorders, including current alcohol or substance dependence, MDE with psychotic symptoms, bipolar I or II disorder, and borderline or antisocial personality disorder. Depression severity was measured as the total score on the 17-item HDRS, which is also strongly correlated with sickness behaviors of low mood and anhedonia. Additional measures included body mass index (BMI) and levels of several peripheral inflammatory markers in serum (IL-1β, IL-6, TNF, and C-reactive protein) (eAppendix in the Supplement).

Image Acquisition and Analysis

Each participant underwent [18F]FEPPA PET scan conducted at the Research Imaging Centre at the Centre for Addiction and Mental Health. For this scan, intravenous [18F]FEPPA was administered as a bolus (mean [SD], 180.5 [14.5] MBq [to convert to millicuries, multiply by 0.02703]). The [18F]FEPPA was of high radiochemical purity (>96%) and high specific activity (mean [SD], 119 [125] TBq/mmol). Manual and automatic arterial blood sampling (programmable blood sampler PBS-101; Veenstra Instruments) was performed to determine the ratio of radioactivity in whole blood to radioactivity in plasma and the unmetabolized radioligand in plasma needed to create the input function for the kinetic analysis. The scan duration was 125 minutes after the injection of [18F]FEPPA. The PET images were obtained using a 3-dimensional brain scanner (HRRT; CPS/Siemens). All PET images were corrected for attenuation using a single photon point source, cesium 137 (half-life, 30.2 years; energy, 662 keV) and were reconstructed using a filtered back-projection algorithm, with a Hann filter at Nyquist cutoff frequency.

Each participant underwent 2-dimensional axial proton-density magnetic resonance imaging acquired with a 1.5-T scanner (Signa; General Electric) (section thickness, 2 mm; repetition time, >5300 milliseconds; echo time, 13 milliseconds; flip angle, 90°; number of excitations, 2; acquisition matrix, 256 × 256; and field of view, 22 cm). Regions of interest were automatically generated using the in-house software (ROMI) as previously described. Time activity curves were used to estimate TSPO VT using a 2-tissue compartment model that has been shown previously to be an optimal model to quantitate TSPO VT with [18F]FEPPA PET.

DNA Extraction and Polymorphism Genotyping

The binding affinity of the second generation of radiotracers for TSPO, including [18F]FEPPA, is known to be affected by a codominantly expressed single-nucleotide polymorphism (rs6971; C→T) in exon 4 of the TSPO gene (NCBI Entrez Gene 706). Individuals with high-affinity binding (Ala147/Ala147) and mixed-affinity binding (Ala147/Thr147) account for more than 90% of the population in North America. The polymorphism rs6971 was genotyped as described previously. One patient with MDE had low-affinity binding (Ala147/Thr147) and was not included in the analysis.

Statistical Analysis

For the primary hypothesis, we analyzed PET data by multivariate analysis of variance (MANOVA), with TSPO VT in the PFC, ACC, and insula as the dependent variables and diagnosis and genotype as the fixed factors. Main effects were considered significant at the conventional $P < .05$. Effects in each region, analyzed by univariate ANOVA, were considered significant after Bonferroni correction ($P < .017$).

As a secondary analysis, we performed a MANOVA that included every brain region sampled (eg, all cortical and subcortical regions) to assess the effect of diagnosis on TSPO VT. A partial correlation controlling for the rs6971 genotype was used in a secondary analysis to quantify the relationship between TSPO VT in the primary regions of interest and the severity of symptoms of MDE measured by the total 17-item HDRS score. The HDRS score was missing in 1 patient with MDE who...
Secondary to Major Depressive Disorder (MDD) diagnosis, patients with MDE compared with the controls (main effect of global brain effect of diagnosis with elevated TSPO VT in the and several other cortical and subcortical regions indicated a Table 2 and (Fgenotype (Figure 1) (effect of diagnosis in the MANOVA, compared with healthy controls after controlling for the effect of binding had greater TSPO VT compared with individuals with high-affinity binding rs6971 genotype expression was not significantly different between healthy controls and patients with MDE. The total 17-item HDRS score was positively correlated with TSPO VT in the ACC after correcting for the rs6971 genotype (r = 0.63 [P = 0.005]) (Figure 2). Similar correlations were found in the insula and PFC, but these did not survive Bonferroni correction (insula, r = 0.57 [P = 0.01]; PFC, r = 0.46 [P = 0.06]). In the patients with MDE but not in the healthy controls (eAppendix in the Supplement), BMI was significantly and negatively correlated with TSPO VT in the insula after correcting for rs6971 genotype (r = −0.61 [P = 0.006]). The relationship between BMI and TSPO VT was also present in the ACC (r = −0.55 [P = 0.02]) and the PFC (r = −0.49 [P = 0.03]), but neither survived Bonferroni correction (for further details on the relationship to clinical characteristics, see the eTable in the Supplement). In the patients with MDE, none of the serum markers of inflammation had a significant positive correlation with TSPO VT in the primary regions of interest (Table 3). Discussion This study is the first, to our knowledge, to detect microglial activation, as indicated by increased TSPO VT, in a substantial sample of patients with MDE. Although the finding was prominent in the anterior regions of the PFC, ACC, and insula, it was also present throughout all the regions assayed. The highest levels of TSPO VT occurred in patients with MDE with the

Results We observed a global effect of diagnosis on TSPO VT (Figure 1 and Table 2). A MANOVA including all subregions of the PFC and several other cortical and subcortical regions indicated a global brain effect of diagnosis with elevated TSPO VT in the patients with MDE compared with the controls (main effect of diagnosis, F15,23 = 4.5 [P = .001]). We also evaluated the regions selected in our hypothesis. Using the effect of diagnosis in the ANOVA by region, patients with MDE had significantly greater TSPO VT in the PFC (F1,37 = 8.1 [P = .007]), ACC (F1,37 = 12.2 [P = .001]), and insula (F1,37 = 12.3 [P = .001]) compared with healthy controls after controlling for the effect of genotype (Figure 1) (effect of diagnosis in the MANOVA, F3,35 = 4.7 [P = .007]; elevations in magnitude of 26%, 32%, and 33%, respectively). In both groups, the effect of the rs6971 polymorphism was significant (effect of genotype in the MANOVA, F3,35 = 4.5 [P = .009]) in which individuals with high-affinity binding had greater TSPO VT compared with individuals with mixed-affinity binding. Mean scores on the 17-item HDRS indicated moderate to severe MDE (Table 1). Differences in TSPO VT between the patients with MDE and healthy controls remained significant if age was applied as a covariate (eAppendix in the Supplement). The frequency of mixed- and high-affinity binding rs6971 genotype expression was not significantly different between healthy controls and patients with MDE.
Table 2. ANOVA of Regional TSPO VT by Diagnosis and TSPO Genotype

<table>
<thead>
<tr>
<th>Region of Interest</th>
<th>TSPO VT, Mean (SD), mL/cm³</th>
<th>Diagnosis</th>
<th>Genotype</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Patients With MDE</td>
<td>Healthy Controls</td>
<td></td>
</tr>
<tr>
<td></td>
<td>HAB (n = 15)*</td>
<td>Total (N = 20)</td>
<td>HAB (n = 14)*</td>
</tr>
<tr>
<td>MPFC</td>
<td>13.6 (3.1)</td>
<td>9.8 (2.1)</td>
<td>11.4 (.002)</td>
</tr>
<tr>
<td>VLPC</td>
<td>14.9 (2.9)</td>
<td>11.3 (2.4)</td>
<td>9.1 (.005)</td>
</tr>
<tr>
<td>DLPC</td>
<td>13.6 (3.2)</td>
<td>10.9 (2.4)</td>
<td>6.5 (.02)</td>
</tr>
<tr>
<td>OFC</td>
<td>14.4 (2.9)</td>
<td>9.6 (2.8)</td>
<td>7.8 (.008)</td>
</tr>
<tr>
<td>Frontal pole</td>
<td>13.3 (3.0)</td>
<td>8.3 (2.3)</td>
<td>9.1 (.005)</td>
</tr>
<tr>
<td>ACC</td>
<td>13.5 (2.9)</td>
<td>8.0 (2.3)</td>
<td>12.2 (.001)</td>
</tr>
<tr>
<td>Insula</td>
<td>14.2 (3.0)</td>
<td>10.2 (2.2)</td>
<td>12.3 (.001)</td>
</tr>
<tr>
<td>Temporal cortex</td>
<td>14.4 (2.8)</td>
<td>10.9 (2.2)</td>
<td>8.7 (.006)</td>
</tr>
<tr>
<td>Parietal cortex</td>
<td>15.0 (3.1)</td>
<td>11.5 (2.2)</td>
<td>8.9 (.005)</td>
</tr>
<tr>
<td>Occipital cortex</td>
<td>14.5 (3.0)</td>
<td>11.0 (2.1)</td>
<td>7.0 (.01)</td>
</tr>
<tr>
<td>Hippocampus</td>
<td>12.8 (2.5)</td>
<td>9.4 (2.3)</td>
<td>7.5 (.009)</td>
</tr>
<tr>
<td>Thalamus</td>
<td>16.9 (3.6)</td>
<td>11.8 (2.2)</td>
<td>13.6 (.001)</td>
</tr>
<tr>
<td>Dorsal putamen</td>
<td>12.3 (2.6)</td>
<td>9.8 (2.3)</td>
<td>14.1 (.001)</td>
</tr>
<tr>
<td>Dorsal caudate</td>
<td>10.9 (2.6)</td>
<td>8.2 (1.9)</td>
<td>6.7 (.01)</td>
</tr>
<tr>
<td>Ventral striatum</td>
<td>12.2 (3.2)</td>
<td>7.9 (2.1)</td>
<td>6.9 (.01)</td>
</tr>
<tr>
<td>ACC</td>
<td>14.4 (2.9)</td>
<td>8.0 (2.3)</td>
<td>13.2 (.001)</td>
</tr>
<tr>
<td>MPFC</td>
<td>13.5 (2.9)</td>
<td>10.2 (2.2)</td>
<td>12.3 (.001)</td>
</tr>
<tr>
<td>OFC</td>
<td>14.2 (3.0)</td>
<td>8.8 (2.1)</td>
<td>12.9 (3.7)</td>
</tr>
<tr>
<td>DLPFC</td>
<td>13.6 (3.2)</td>
<td>9.5 (2.7)</td>
<td>8.7 (.006)</td>
</tr>
<tr>
<td>VLPFC</td>
<td>14.4 (2.9)</td>
<td>9.6 (2.8)</td>
<td>7.8 (.008)</td>
</tr>
</tbody>
</table>

Abbreviations: ACC, anterior cingulate cortex; ANOVA, analysis of variance; DLPFC, dorsolateral prefrontal cortex; HAB, high-affinity binding; MAB, mixed-affinity binding; MDE, major depressive episode; MPFC, medial PFC; OFC, orbitofrontal cortex; TSPO VT, translocator protein density measured by distribution volume; VLPFC, ventrolateral PFC.

* Indicates binding to the single-nucleotide polymorphism rs6971 of the TSPO gene known to influence fluorine F18–labeled N-(2-(2-fluoroethoxy) benzyl)-N-(4-phenoxypyridin-3-yl)acetamide binding. A more detailed description of the subregions of the PFC is given in the eAppendix in the Supplement.

Because TSPO is upregulated in activated microglia, elevated TSPO VT implies that greater microglial activation, a potentially targetable process of neuroinflammation, is present during MDE. During activation, microglia transform from a monitoring role into a macrophagelike state, responding to infections or insults by phagocytosis of pathogens and dying cells and recruiting immune cells via cytokine secretion. However, active microglia during MDE may represent a maladaptive response. Identifying greater microglial activation in MDE suggests that selective therapeutic strategies, such as stimulating microglial targets like CX3CR1 to promote a more quiescent state, suppressing the effects of cytokines in the central nervous system, or promoting a shift in microglial activity toward repair-oriented functions by activating purinergic receptors, may hold promise. Reducing microglial activation itself might also have therapeutic utility. Consistent with this viewpoint, minocycline hydrochloride, a second-generation tetracycline antibiotic known to reduce microglial activation and TSPO expression in rodents, can attenuate depressive behaviors in rodents. The present study also suggests that the ability of such interventions to reduce microglial activation may be monitored by techniques such as [18F]FEPPA PET.

We found MDE to be associated with elevated TSPO VT across all brain regions examined, and regional TSPO VT was intercorrelated, although the relationships between TSPO VT and the severity of MDE were most pronounced in the ACC. We propose that although global mechanisms may account for elevated TSPO VT in multiple brain regions in MDD, greater TSPO VT in specific regions and/or their associated circuitry has important implications for the pathophysiological features of MDE, identification of mechanisms contributing to symptom severity in MDE, and clinical targeting of treatment.
may be influential for the expression of particular symptoms within this complex disorder. As with any association between symptoms and a central biomarker, the correlation found between greater TSPO VT and greater 17-item HDRS scores in the ACC can be interpreted as an epiphenomenon secondary to a common origin or as one phenomenon predisposing to the other. We favor a causal mechanism of neuroinflammation contributing toward symptoms because induction of inflammation in humans is associated with depressed mood and because direct induction of central inflammation in rodents is associated with anhedonia.

The function of this region in relation to symptoms of MDE is consistent with the interpretation that the ACC participates in regulating and processing negative emotional responses. In MDD, active MDE symptoms are associated with higher metabolic function in the ACC and direct stimulation of the subgenual ACC results in the reduction of MDE symptoms. The negative relationship between TSPO VT and BMI may be consistent with anorexia after induction of central inflammation. The insula is important in this relationship because it integrates interoceptive and affective signaling and is involved in homeostatically driven responses to food cues. Future studies in preclinical models to induce microglial activation in combinations of regions that include the ACC and insula might clarify the role of this abnormality in relation to depressive behavior.

The lack of correlation between the central and peripheral inflammatory markers is consistent with previous reports. Bromander et al found no correlation between serum and cerebrospinal fluid TNF in patients undergoing knee surgery. Similarly, dissociation between central and peripheral cytokines in preclinical data has been reported after peripheral or central inflammatory stimuli. Peripheral cytokines have been proposed to cross the blood-brain barrier in severe medical illness to induce neuroinflammation and symptoms of depression. However, our results suggest that central inflammation may be present during MDE even when peripheral inflammation is absent.

This study has several limitations, most of which are related to the interpretation of TSPO VT and the use of PET imaging. To the best of our knowledge, the most supported explanation for greater TSPO binding with PET is microglial activation, although TSPO has other roles, such as translocating cholesterol from the outer to the inner mitochondrial membranes for steroid hormone synthesis and participating in the mitochondrial permeability transition pore heterooligomer, which influences predisposition toward apoptosis.

Conclusions

To our knowledge, this study is the first to find evidence of a significant elevation of brain TSPO density, a marker of mi-
Translocator Protein Density in MDE

Research Original Investigation

Translocator Protein Density in MDE

Research Foundation (Dr Setiawan), by grant P30 GM103328 from the National Institutes of Health (Dr Rajkowska), and by the Canadian Foundation for Innovation and the Ontario Ministry for Research and Innovation (Infrastructure).

Role of the Funder/Sponsor: The funding sources had no role in the design and conduct of the study; collection, management, analysis, and interpretation of the data; preparation, review, or approval of the manuscript; and decision to submit the manuscript for publication.

Previous Presentations: This paper was presented in part at the 52nd Annual Meeting of the American College of Neuropsychopharmacology; December 9, 2013, Hollywood, Florida; at the 69th Annual Meeting of the Society of Biological Psychiatry; May 8, 2014, New York, New York; and at the 53rd Annual Meeting of the American College of Neuropsychopharmacology, December 8, 2014; Phoenix, Arizona.

Additional Contributions: Cynthia Xu, MD, Research Imaging Centre, Centre for Addiction and Mental Health, coordinated research for this study. Jan Fan, BSc, Research Imaging Centre, Centre for Addiction and Mental Health, provided research assistance. Nathan Kolla, MD, Research Imaging Centre, Centre for Addiction and Mental Health, Institute of Medical Science, University of Toronto assisted with recruitment and medical coverage. Andrea Tyer, BSc, Research Imaging Centre, Centre for Addiction and Mental Health, Department of Pharmacology, University of Toronto assisted with recruitment and medical coverage. Alvinia Ng, BSc, and Laura Nguyen, BSc, worked as study technicians. Jun Parkes, MSc, Armando Garcia, BSc, Winston Stableford, BSc, and Min Wong, BSc, Research Imaging Centre, served as chemistry staff. Terry Bell, BSc, and Ted Harris-Brands, BSc, Research Imaging Centre, provided engineering support. Dr Kolla was funded from the Canadian Institutes of Health Research and Ms Tyer was funded by Brain Canada; all other contributors are paid employees of the Centre for Addiction and Mental Health.

Correction: This article was corrected on March 4, 2015, to fix Table 2.

REFERENCES


