Association between High Tear Epidermal Growth Factor Levels and Corneal Subepithelial Fibrosis in Dry Eye Conditions

Kavita Rao, William J. Farley, and Stephen C. Pflugfelder

PURPOSE. To compare tear epidermal growth factor (EGF) concentration in dry eye (DE) conditions and determine correlations between EGF levels and severity of symptoms and ocular surface signs.

METHODS. In this prospective case–control study, 35 patients with DE, including subgroups with meibomian gland disease (MGD), Sjögren’s syndrome (SS) aqueous tear deficiency, or neurotrophic keratopathy (NK), and 17 asymptomatic control subjects were evaluated. Symptoms, Schirmer test, fluorescein clearance test (FCT), EGF concentration, dye staining, and the presence of corneal subepithelial fibrosis and meibomian gland (MG) orifice metaplasia were recorded. Tear EGF and the severity of irritation and ocular surface signs were correlated.

RESULTS. Tear EGF was higher in MGD than in the control (P = 0.05) and was lower in SS than in the control (P < 0.0001; MGD (P < 0.05) and NK (P < 0.01) groups. The DE subgroup with results in the FCT > 3 and Schirmer 1 ≥ 8 had higher EGF levels than the group with FCT > 3 and Schirmer 1 < 8 and both groups with good tear clearance (P < 0.01). Tear EGF levels correlated inversely with conjunctival (r = −0.49, P = 0.0032) and corneal (r = −0.39, P = 0.0222) dye staining and positively with MG orifice metaplasia (r = 0.36, P = 0.03) and corneal subepithelial fibrosis (r = 0.5, P = 0.006).

CONCLUSIONS. Tear EGF concentration was increased in eyes with MGD, corneal subepithelial fibrosis, and MG orifice metaplasia. Elevated tear EGF may promote development of corneal subepithelial fibrosis and lid margin changes. (Invest Ophthalmol Vis Sci. 2010;51:844–849) DOI:10.1167/iovs.09-3875

Dry eye syndrome is one of the most common ocular diseases, affecting 14% to 33% of the population worldwide and accounting for a high percentage of patient visits to eye care practitioners.1–3 Evidence suggests that alterations in tear composition are responsible in part for the irritation symptoms and ocular surface disease that develops in DE.4,5 Indeed, a study report showed an association between increased concentrations of inflammatory cytokines in tears and the severity of irritation symptoms and ocular surface disease in DE, regardless of the etiology.6

Epidermal growth factor (EGF), secreted by the lacrimal gland, is one of the most abundant growth factors in human tears. EGF receptors are expressed by the corneal epithelium, keratocytes, and endothelium.7 EGF is a pleiotropic cytokine that can stimulate proliferation, migration, and adhesion of corneal epithelial cells during wound healing.6,8 In conjunction with TGF-β, it stimulates myofibroblastic differentiation and production of extracellular matrix by corneal keratocytes.9,10 In a survey of tear proteins in patients with DE conditions, the highest concentration of EGF was noted to be in the subset of patients with meibomian gland disease (MGD).11 We have reported that the presence of MG orifice metaplasia is associated with decreased tear clearance.11 Corneal complications, such as epithelial basement membrane disease, Salzmann’s nodules, and subepithelial fibrosis are commonly observed in patients with MGD.12 These fibrotic changes are frequently located in the peripheral cornea that is in contact with the upper and lower tear menisci. Based on these findings, we hypothesized that the presence of corneal subepithelial fibrosis and metaplasia of the MG orifice epithelium, two common findings in patients with chronic DE and MG disease, is associated with increased tear EGF concentration.

The purpose of this study was to measure tear EGF concentrations in an asymptomatic control group and in with DE. Furthermore, the correlations between tear EGF concentrations, tear production and clearance, and severity of clinical parameters of DE, including corneal subepithelial fibrosis and MG orifice metaplasia, were determined.

MATERIALS AND METHODS

Patients

This study was approved by the Baylor College of Medicine, Institutional Review Board (IRB). After informed consent, 35 consecutive patients with newly diagnosed DE meeting the inclusion and exclusion criteria were enrolled for the study at the Ocular Surface Center at Baylor College of Medicine, Houston, Texas. The enrollment procedure and study protocol were in compliance with the Declaration of Helsinki. Patients completed the Ocular Surface Disease Index (OSDI) symptom severity questionnaire,13 followed by a complete ocular surface examination of both eyes by one of the investigators (SCP) in the following sequence: tear collection, biomicroscopic examination of the lid margins, meibomian glands and cornea, fluorescein tear break-up time (TBUT), corneal fluorescein staining, fluorescein clearance test, conjunctival lissamine green staining, and Schirmer 1 test. The presence of corneal subepithelial fibrosis (gray subepithelial tissue with elevation of the overlying epithelium) was noted. The TBUT was evaluated 1 minute after instilling 5 μL of 1% sodium fluorescein (Greenpark Pharmacy, Houston, TX) onto the inferior tarsal conjunctiva. After a blink, the precorneal tear film was examined using a biomicroscope with a 10× objective under blue-light illumination. The interval between the blink and the appearance of the first dark spot or discontinuity in the precorneal fluorescein-stained tear layer was re-
Baylor grading scheme.14 Briefly, the number of dots of fluorescein
1% sodium fluorescein. The cornea was viewed through an acetate
through a yellow filter (Wratten 12, Boston 7503; Essilor, Omega,
technique.15 Fifteen minutes after instillation of 1% sodium fluorescein,
clearance was performed with a modification of a previously reported
4. One point was added to the score if there was one area of confluent
/staining was graded in each of five zones on a standardized 5-point
scale,15 and the tear clearance score was graded from 0 to 6. If the
was visually compared with one of the colors of a standardized visual
/staining, and 2 points were added if there were two or more.
corded. Three separate readings were taken in each eye, and the
results were averaged. To evaluate corneal fluorescein staining, the
ocular surface was examined with a biomicroscope 10× objective
through a yellow filter (Wratten 12, Boston 7503; Essilor, Omega,
twice the area of confluent staining in each of five zones on a standardized 5-point
/staining was graded in each of five zones on a standardized 5-point
score was graded in each of five zones on a standardized 5-point
/staining was graded according to the Baylor grading scheme.15 Briefly, the number of dots of fluorescein staining was graded in each of five zones on a standardized 5-point scale: 0 dots; 0 to 5 dots; 1; 6 to 15 dots; 2; 16–30 dots; 3; and >30 dots. 4. One point was added to the score if there was one area of confluent staining, and 2 points were added if there were two or more. Tear clearance was performed with a modification of a previously reported technique.15 Fifteen minutes after instillation of 1% sodium fluorescein, the color of the tear meniscus in the outer one third of the lower lid was visually compared with one of the colors of a standardized visual scale,15 and the tear clearance score was graded from 0 to 6. If the color of the tear meniscus was judged to be between two of the six standard scale colors, then the score was graded between these two standard colors. Lissamine green staining of the nasal and temporal bulbar conjunctiva (graded 0–3 in each area based on the density of the dots) was evaluated 1 minute after instillation of lissamine green from
/staining of the nasal and temporal bulbar conjunctiva was graded 0–3 in each area based on the density of the dots.
/nasal, superior, and inferior) with the outer circumference aligned with the limbus and the intensity of corneal fluorescein staining was graded according to the
/gague.11 Briefly, the number of dots of fluorescein staining was graded in each of five zones on a standardized 5-point scale: 0 dots; 0 to 5 dots; 1; 6 to 15 dots; 2; 16–30 dots; 3; and >30 dots. 4. One point was added to the score if there was one area of confluent staining, and 2 points were added if there were two or more.

<table>
<thead>
<tr>
<th>Group</th>
<th>Symptom Severity Score*</th>
<th>Tear Break-up Time (s)</th>
<th>Conjunctival Staining Score†</th>
<th>Corneal Staining Score‡</th>
</tr>
</thead>
<tbody>
<tr>
<td>DE 1</td>
<td>0–20</td>
<td>&gt;7</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>DE 2</td>
<td>20–70</td>
<td>≤7</td>
<td>≥3</td>
<td>≥6</td>
</tr>
<tr>
<td>DE 3</td>
<td>70–30</td>
<td>≤7</td>
<td>≥3</td>
<td>≥6, including central cornea or filaments</td>
</tr>
<tr>
<td>DE 4</td>
<td>&gt;30</td>
<td>≤7</td>
<td>≥3</td>
<td>≥12</td>
</tr>
</tbody>
</table>

Table 1. Symptom Severity Criteria for DE Conditions

DE 1–4, dry eye severity levels.

* Symptom severity score measured by ocular surface disease index (OSDI) questionnaire.15
† Lissamine green staining of the nasal and temporal bulbar conjunctiva was graded 0–3 in each area based on the density of the dots.
‡ Corneal fluorescein staining was graded according to the Baylor grading scheme14 in each of five zones on a standardized 5-point scale:

Tear Collection

Minimally stimulated tear fluid was collected from the inferior tear meniscus of each eye using a 0.5-μL glass capillary micropipette
(Drummond, Broomall, PA). The tear samples from both eyes (1.0 μL total) were eluted into one tube (Eppendorf, Fremont, CA) containing
9 μL of assay buffer for a final dilution of 1:10, centrifuged for 2 minutes, and immediately transported in an insulated cooler to a
~80°C freezer where they remained frozen until they were used for the immunoassay.

EGF Analysis

EGF levels in these samples were analyzed in a masked fashion with an immunoassay (Beadlyte; Millipore Corp., Billerica, MA) according to
the manufacturer’s protocol. In brief, 10 μL of each tear sample, 25 μL of the 1:10 bead solution, and 10 μL of assay buffer (for blanks) were
incubated at 4°C overnight in wells of a microtiter plate precoated with assay buffer. Plates were washed two times with 200 μL of wash
buffer containing 0.05% preservative (Proclin; Sigma-Aldrich, St. Louis, MO). Human cytokine detection antibody (25 μL) was added to each well, and the samples were incubated at 37°C for 1 hour. Streptavidin
phycoerythrin (25 μL) was added to each well, and the samples were incubated at 37°C for 30 minutes. The plates were washed two times with 200 μL of wash buffer, and 150-μL sheathing fluid was added to all wells and the beads suspended on the plate shaker for 5 minutes.
The plate was then read (100 IS; Luminex, Austin, TX) system and the results were analyzed (Bead View Analysis software; Millipore). The concentration of EGF was calculated from a standard curve of known combinations of recombinant human EGF.

Statistical Analysis

Clinical parameters and tear cytokine concentrations in the study groups were compared by analysis of variance (ANOVA) with Tukey
post hoc testing (Prism; GraphPad, La Jolla, CA). Reported probabilities for multiple comparisons made for the same group were lower than
the Bonferroni-corrected threshold. The mean of measured clinical parameters of both eyes was used for statistical comparison.

Table 2. Demographic Features of Control Subjects and Patients with DE

<table>
<thead>
<tr>
<th>Group</th>
<th>n</th>
<th>Mean Age ± SD (y)</th>
<th>% Female</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>17</td>
<td>45.1 ± 14.7</td>
<td>70</td>
</tr>
<tr>
<td>All DE</td>
<td>35</td>
<td>57.5 ± 13.2</td>
<td>86.1</td>
</tr>
<tr>
<td>MGD</td>
<td>20</td>
<td>55.3 ± 16.4</td>
<td>89</td>
</tr>
<tr>
<td>SS</td>
<td>9</td>
<td>58.7 ± 18.3</td>
<td>100</td>
</tr>
<tr>
<td>NK</td>
<td>6</td>
<td>61.6 ± 6.0</td>
<td>67</td>
</tr>
</tbody>
</table>

* Diabetic neuropathy, n = 2; ophthalmic zoster, n = 2; scleral buckling procedure, n = 1; surgical trigeminal ganglion ablation, n = 1.
RESULTS

Demographic and Clinical Data

The demographic data for control and patients with DE are presented in Table 2. There were no significant differences in age and gender between the patients with DE and the asymptomatic control group (P ≥ 0.5). The clinical findings in the control and DE groups are presented in Table 3.

Compared with the control group, the symptom severity scores were significantly greater in the DE group as a whole, as well as in the MGD, SS, and NK subgroups (P < 0.0001). The Schirmer 1 score were significantly lower (P = 0.0001), and conjunctival the lissamine green and corneal fluorescein staining scores were significantly greater (P < 0.0001) in the DE group and the SS and NK subgroups than in the control group. However, corneal subepithelial fibrosis was more prevalent in the MGD and NK subgroups than in the SS subgroup and the control group. MG orifice metaplasia was significantly greater in the MGD subgroup than in the SS and NK subgroups and the control group (P < 0.0001). Tear fluorescein clearance was delayed in the DE group and in the MGD, SS and NK subgroups compared with that in the control group (P < 0.0001).

The severity parameters for each of the four levels of DE severity are presented in Table 4. Corneal fluorescein staining was significantly greater in the level 4 severity group than in the level 1 and 2 groups (P < 0.001). Conjunctival dye staining was significantly greater in level 4 than in levels 1, 2, and 3 (P < 0.005). MG orifice metaplasia was greater in levels 1 and 2 than in levels 5 and 4 (P < 0.0001).

EGF Concentrations

The measured tear EGF concentrations were in the linear portion of the standard curve. The EGF levels in control subjects and patients with DE are presented in Table 1. The EGF level was 1412.9 ± 595.0 pg/mL in the control group and 2016.8 ± 1977 pg/mL in the entire DE group. The lowest EGF levels were seen in the SS subgroup (310.9 ± 232.34 pg/mL), significantly lower than in the control group (P < 0.0001) and the MGD subgroup than in the SS subgroup and the control group.

Normal control group. The mean score of both eyes was used for the Schirmer 1 test, corneal fluorescein staining, conjunctival staining scores, and BCVA. P, vs. normal control; ¶, vs. level 1; §, vs. level 2; ¶, vs. level 3; and ¶, vs. level 4. Symboled footnotes are described in Table 5.

Table 3. Clinical Features of Normal Subjects and Patients with DE

<table>
<thead>
<tr>
<th>Group</th>
<th>n</th>
<th>Symptom Severity Score*</th>
<th>Schirmer 1 Score (mm)</th>
<th>Conjunctival Staining Score†</th>
<th>Corneal Staining Score‡</th>
<th>Corneal Subepithelial Fibrosis§</th>
<th>Melolobian Gland Orifice Metaplasia¶</th>
<th>FCT¶</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal</td>
<td>17</td>
<td>11.9 ± 2.3</td>
<td>27.0 ± 7.8</td>
<td>0 ± 0</td>
<td>0 ± 0</td>
<td>0 ± 0</td>
<td>1.5 ± 0.7</td>
<td></td>
</tr>
<tr>
<td>All DE</td>
<td>35</td>
<td>34.8 ± 9.4</td>
<td>15.0 ± 11.0</td>
<td>1.7 ± 2.0</td>
<td>4.6 ± 4.4</td>
<td>0.4 ± 0.4</td>
<td>0.47 ± 0.5</td>
<td>4.3 ± 1.0</td>
</tr>
<tr>
<td>MGD</td>
<td>20</td>
<td>33.2 ± 8.7</td>
<td>18.5 ± 11.9</td>
<td>0.5 ± 0.15</td>
<td>3.2 ± 2.7</td>
<td>0.47 ± 0.5</td>
<td>0.78 ± 0.4</td>
<td>3.7 ± 2.8</td>
</tr>
<tr>
<td>SS</td>
<td>9</td>
<td>36.8 ± 9.6</td>
<td>4.79 ± 2.9</td>
<td>4.9 ± 2.01</td>
<td>6.33 ± 5.37</td>
<td>0 ± 0</td>
<td>0.2 ± 0.4</td>
<td>5.1 ± 0.8</td>
</tr>
<tr>
<td>NK</td>
<td>6</td>
<td>41.2 ± 15.6</td>
<td>8.4 ± 3.0</td>
<td>1.67 ± 2.66</td>
<td>7.93 ± 4.8</td>
<td>0.6 ± 0.4</td>
<td>0 ± 0</td>
<td>3.77 ± 1.7</td>
</tr>
</tbody>
</table>

The measured tear EGF concentrations were in the linear portion of the standard curve. The EGF levels in control subjects and patients with DE are presented in Table 1. The EGF level was 1412.9 ± 595.0 pg/mL in the control group and 2016.8 ± 1977 pg/mL in the entire DE group. The lowest EGF levels were seen in the SS subgroup (310.9 ± 232.34 pg/mL), significantly lower than in the control group (P < 0.0001) and the MGD subgroup than in the SS subgroup and the control group. MG orifice metaplasia was significantly greater in the MGD subgroup than in the SS and NK subgroups and the control group (P < 0.0001). Tear fluorescein clearance was delayed in the DE group and in the MGD, SS and NK subgroups compared with that in the control group (P < 0.0001).

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Table 4. Clinical Features of Patients with DE Stratified by Severity

<table>
<thead>
<tr>
<th>Severity Level</th>
<th>n</th>
<th>Symptom Severity Score*</th>
<th>Schirmer 1 Score (mm)</th>
<th>Conjunctival Staining Score†</th>
<th>Corneal Staining Score‡</th>
<th>Corneal Subepithelial Fibrosis§</th>
<th>Melolobian Gland Orifice Metaplasia¶</th>
<th>FCT¶</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>8</td>
<td>30.5 ± 11.2</td>
<td>18 ± 12</td>
<td>0.25 ± 0.46</td>
<td>0.75 ± 1</td>
<td>0.57 ± 0.5</td>
<td>0.71 ± 0.44</td>
<td>4.25 ± 1.13</td>
</tr>
<tr>
<td>2</td>
<td>16</td>
<td>32.6 ± 10.3</td>
<td>14 ± 12</td>
<td>1.5 ± 2.3</td>
<td>3.6 ± 2</td>
<td>0.25 ± 0.4</td>
<td>0.62 ± 0.5</td>
<td>4.16 ± 1.5</td>
</tr>
<tr>
<td>3</td>
<td>7</td>
<td>36.5 ± 13.5</td>
<td>9 ± 8</td>
<td>2.3 ± 2.14</td>
<td>9.5 ± 1.5</td>
<td>0.7 ± 0.4</td>
<td>0.14 ± 0.4</td>
<td>3.45 ± 1.66</td>
</tr>
<tr>
<td>4</td>
<td>4</td>
<td>41 ± 10.0</td>
<td>4.8 ± 1.3</td>
<td>5.63 ± 0.75</td>
<td>13.7 ± 2.3</td>
<td>0.25 ± 0.5</td>
<td>0 ± 0</td>
<td>5.5 ± 0.41</td>
</tr>
<tr>
<td>Control</td>
<td>17</td>
<td>11.9 ± 2.3</td>
<td>27.0 ± 7.8</td>
<td>0 ± 0</td>
<td>0 ± 0</td>
<td>0 ± 0</td>
<td>0 ± 0</td>
<td>1.5 ± 0.7</td>
</tr>
</tbody>
</table>

Control, normal control group. The mean score of both eyes was used for the Schirmer 1 test, corneal fluorescein staining, conjunctival staining scores, and BCVA. P, vs. control; ¶, vs. level 1; §, vs. level 2; ¶, vs. level 3; and ¶, vs. level 4. Symboled footnotes are described in Table 5.
the MGD ($P < 0.05$) and NK ($P < 0.01$) subgroups. EGF levels were significantly higher in the DE subgroup with MGD compared to the control group ($P = 0.03$). The highest EGF levels were seen in patients with NK ($3227.5 \pm 2950$ pg/mL), significantly higher than those in the control group ($P = 0.02$).

**Clinical Parameters and Tear EGF Concentrations Stratified by FTC and Schirmer 1 Scores**

The control subjects and patients with DE were stratified into four subgroups, according to their FTC and Schirmer test scores (Table 5). The subgroup with FTC > 3, Schirmer 1 $\geq$ 8 included 14 patients with MGD and 4 with NK. This subgroup had the highest EGF levels, significantly higher than those in the group with FTC > 3, Schirmer 1 < 8 ($P < 0.001$) and both the groups had a good tear clearance (FCT < 3; $P < 0.01$; Fig. 2). Corneal subepithelial fibrosis and MG orifice metaplasia were also significantly higher in this subgroup than the other three subgroups ($P < 0.0001$). The subgroup with FTC > 3 and Schirmer 1 < 8 included all the patients with SS and two patients with NK. This subgroup had significantly higher conjunctival and corneal dye staining scores than did the other three groups. Symptom severity scores were significantly greater in the both groups with delayed FTC than in the group with FTC < 3 and Schirmer $\geq$ 8.

**Correlation Analyses: EGF Levels and Clinical Parameters**

The rank (Spearman) correlation coefficients between tear epidermal growth factor concentration and clinical parameters are presented in Table 6. No correlation was found between the tear EGF concentration and symptom severity scores. A significant inverse correlation was found between the tear EGF levels and the conjunctival dye staining ($r = -0.49$, $P = 0.0032$) and corneal dye staining scores ($r = -0.39$, $P = 0.022$). MG orifice metaplasia ($r = 0.36$, $P = 0.03$), as well as corneal subepithelial fibrosis ($r = 0.5$, $P = 0.0006$) correlated significantly with tear EGF concentrations. Furthermore, there was a significant correlation between tear EGF levels and the Schirmer 1 scores ($r = 0.47$, $P = 0.0042$; Fig. 3).

The tear EGF level and FTC showed an inverse but not significant correlation ($r = -0.1$, $P = 0.54$). The correlation of tear EGF levels and FTC in all DE subgroups and control subjects is shown in Figure 4. All the patients in the SS group had delayed FTC but low EGF levels compared with the control subjects. Most patients in the MGD subgroup had delayed tear clearance and high EGF levels than did the control subjects. Similarly, the NK subgroup shows delayed tear clearance and high EGF levels.

**DISCUSSION**

DE is a common ocular surface disease that has a variety of clinical presentations that include eye irritation, increased sen-

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**Table 5.** Clinical Parameters and Tear EGF Levels in Control Subjects and Patients with DE Grouped According to FTC and Schirmer 1 Scores

<table>
<thead>
<tr>
<th>Group</th>
<th>FTC Cutoffs</th>
<th>n</th>
<th>Symptom Severity Score$^*$</th>
<th>Conjunctival Staining Score$^†$</th>
<th>Corneal Fibrotic Change$‡$</th>
<th>Meibomian Gland Orifice Metaplasia$§$</th>
<th>EGF Levels$¶$</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>FTC $\leq$ 3, Schirmer $\geq$ 8</td>
<td>19</td>
<td>$15.1 \pm 7.97$</td>
<td>$0 \pm 0$</td>
<td>$0.6 \pm 1.9$</td>
<td>$0 \pm 0$</td>
<td>$0.12 \pm 0.3$</td>
</tr>
<tr>
<td>2</td>
<td>FTC $\leq$ 3, Schirmer &lt; 8</td>
<td>4</td>
<td>$26.3 \pm 12.3$</td>
<td>$0 \pm 0$</td>
<td>$2.6 \pm 0$</td>
<td>$0.25 \pm 0.5$</td>
<td>$0.25 \pm 0.5$</td>
</tr>
<tr>
<td>3</td>
<td>FTC $&gt; 3$, Schirmer $\geq$ 8</td>
<td>18</td>
<td>$35.1 \pm 11.13$</td>
<td>$0.8 \pm 1.1$</td>
<td>$4.0 \pm 3.0$</td>
<td>$0.56 \pm 0.51$</td>
<td>$0.87 \pm 0.34$</td>
</tr>
<tr>
<td>4</td>
<td>FTC $&gt; 3$, Schirmer &lt; 8</td>
<td>11</td>
<td>$37.55 \pm 10.72$</td>
<td>$4.7 \pm 2.4$</td>
<td>$7.18 \pm 4.7$</td>
<td>$0.09 \pm 0.3$</td>
<td>$0.18 \pm 0.4$</td>
</tr>
</tbody>
</table>

$^P$, vs. group 1; $^*$vs. group 2; $^†$vs. group 3; and $^‡$vs. group 4. Symboled footnotes are described in Table 3.
sitivity to air drafts, and blurred vision. A previously reported study found a link between compositional changes in tears and the severity of irritation symptoms and ocular surface disease that develops in DE. Our group has previously reported that tear EGF concentration is increased in patients with DE with MGD compared with those without MGD. In the present study, we found the tear EGF was significantly higher in the DE subgroup with MGD than in the control group and the subgroup with SS. EGF levels were significantly lower in the SS group than the control group and DE subgroups with MGD and NK. An interesting and unexpected finding was the elevated EGF concentration in patients with NK compared with the control group.

EGF was measured by a sensitive bead immunoassay in a minute volume (1 μL) of tears collected from the inferior meniscus with a microcapillary pipette. It is likely that the tear volume was greater in subjects with normal tear production and delayed tear clearance compared with those with reduced aqueous tear production; however, we were able to collect the same volume of tears from all subjects and the measured tear EGF concentrations should represent the actual concentration the cells on the surface of the eye are exposed to. Care was taken to minimally stimulate reflex tearing during tear collection by using dim illumination and avoiding contact with the lid margin or ocular surface. Our group has previously reported that tear EGF concentrations decrease with induction of reflex tearing. It is well recognized that the ability to reflex tear is diminished in SS and NK. Therefore, in the event reflex tearing was induced in the subjects who retained the ability to reflex tear, it would have resulted in a decrease in tear EGF concentration, not the increased concentration we observed.

On subgroup of patients with DE based on tear clearance and Schirmer 1 scores, the subgroup with delayed tear clearance and Schirmer 1 scores ≥ 8 had tear EGF concentrations significantly higher than both subgroups with normal tear clearance, and the subgroup with delayed tear clearance and decreased Schirmer 1 score. Corneal fibrotic changes, as well as MG orifice metaplasia were significantly higher in this group with delayed tear clearance and normal tear production than the other three subgroups. This group with delayed tear clearance, and normal Schirmer 1 score also had significantly higher symptom severity and corneal dye staining scores than the group with normal tear clearance and Schirmer 1 score.

Most of the patients with MGD (14/20) were found in the subgroup with FCT > 3 and Schirmer 1 ≥ 8, and it is probable that the high tear EGF concentrations in this group coupled with the delayed tear clearance are responsible for the significantly higher corneal fibrotic changes and MG orifice metaplasia seen in this group. The altered corneal permeability found in DE may facilitate diffusion of tear EGF to basal corneal epithelial cells and subepithelial keratocytes, which may then be stimulated to produce the extracellular matrix proteins that are responsible for the fibrotic changes.

Similar to previously reported studies, tear EGF concentration showed significant correlation with Schirmer 1 score, indicating it is a marker of lacrimal gland function. A statistically significant positive correlation was noted between tear EGF concentration and clinical severity parameters, including corneal subepithelial fibrosis and MG orifice metaplasia. The high tear EGF concentrations observed in the DE subgroups with MGD and NK may play a role in the corneal fibrotic changes and neovascularization found in both these conditions. Corneal and conjunctival dye staining scores showed a significant negative correlation with tear EGF concentration. This finding may explain the increased corneal and conjunctival dye staining in SS aqueous tear deficiency, the group with the lowest tear EGF levels.

It is known that topical application of EGF to the corneal epithelium leads to epithelial hypertrophy and enhances the division of corneal epithelial cells. Furthermore, EGF has been associated with development of corneal vascularization. Tear EGF is essential for maintaining homeostasis of the ocular surface epithelia and to promote epithelial healing after trauma; however, it is likely that when tear EGF concentration increases beyond a certain level, it may promote hypertrophy of the corneal and MG ductal epithelium resulting in the MG orifice metaplasia that is frequently observed in patients with MGD. Additional research is needed to establish this link firmly and to develop strategies to prevent these corneal and lid margin changes in patients with DE with decreased tear clearance.

**Table 6.** Correlation between Tear EGF Level and Dry DE Clinical Severity Parameters

<table>
<thead>
<tr>
<th>Clinical Parameter</th>
<th>Spearman r</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>Symptom severity score</td>
<td>0.09</td>
<td>0.59</td>
</tr>
<tr>
<td>Conjunctival staining score</td>
<td>−0.49</td>
<td>0.0032</td>
</tr>
<tr>
<td>Corneal staining score</td>
<td>−0.39</td>
<td>0.022</td>
</tr>
<tr>
<td>Meibomian gland orifice metaplasia</td>
<td>0.36</td>
<td>0.03</td>
</tr>
<tr>
<td>Corneal subepithelial fibrosis</td>
<td>0.5</td>
<td>0.0006</td>
</tr>
<tr>
<td>Schirmer 1 score</td>
<td>0.47</td>
<td>0.0042</td>
</tr>
<tr>
<td>FCT score</td>
<td>−0.1</td>
<td>0.34</td>
</tr>
</tbody>
</table>

Significant correlations are in bold. Symptom severity score measured by OSD1 questionnaire.

**Figure 3.** Correlation between tear EGF concentration and the Schirmer 1 score.
References


